

Advanced Microscopy Course 17th Nov 2020 Dr Matthew Stower



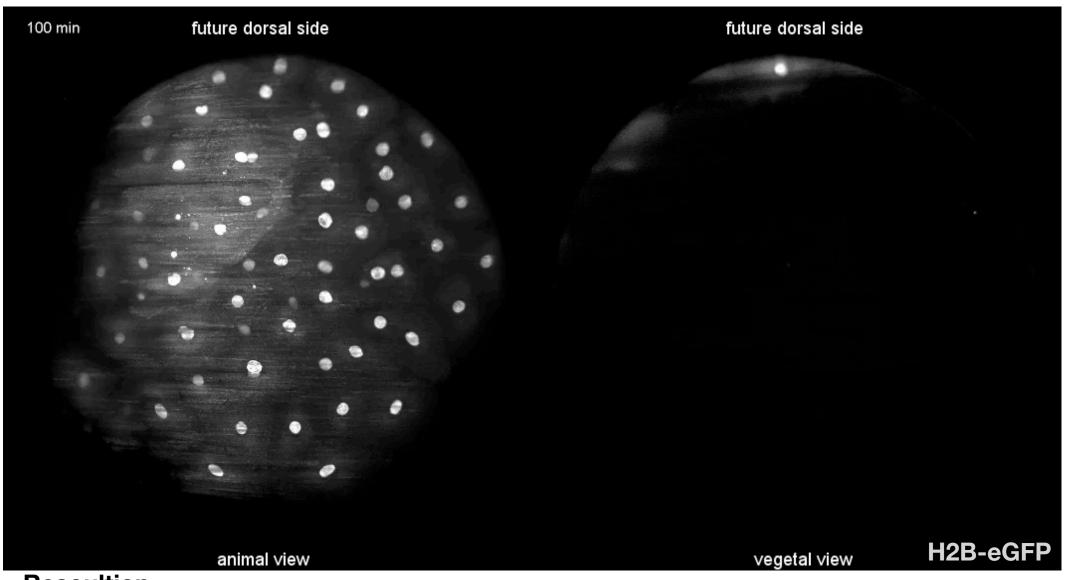




Why use lightsheet imaging?

Live Samples





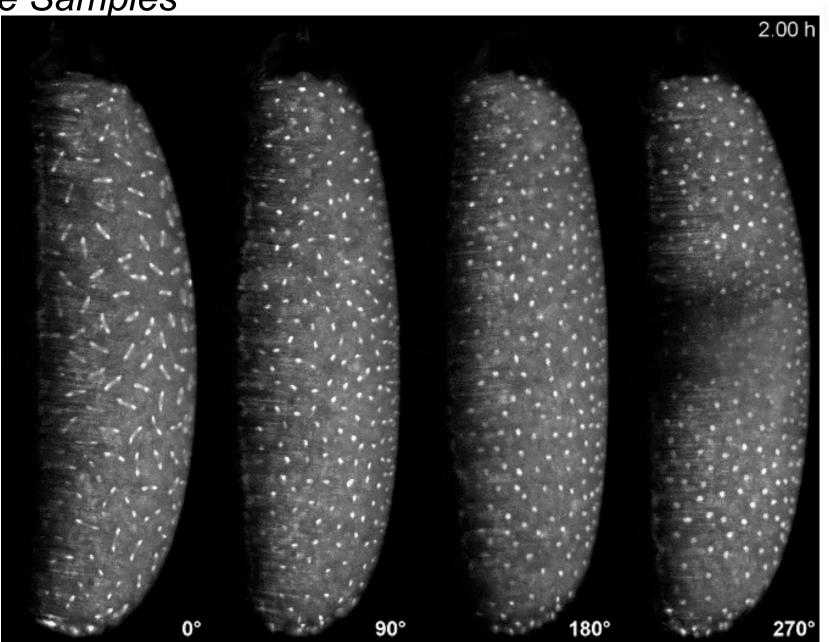
Resoultion lateral 300 nm Axial 1000 nm

16,000 cells

Keller et al., (2008) Science, 322

Why use lightsheet imaging?

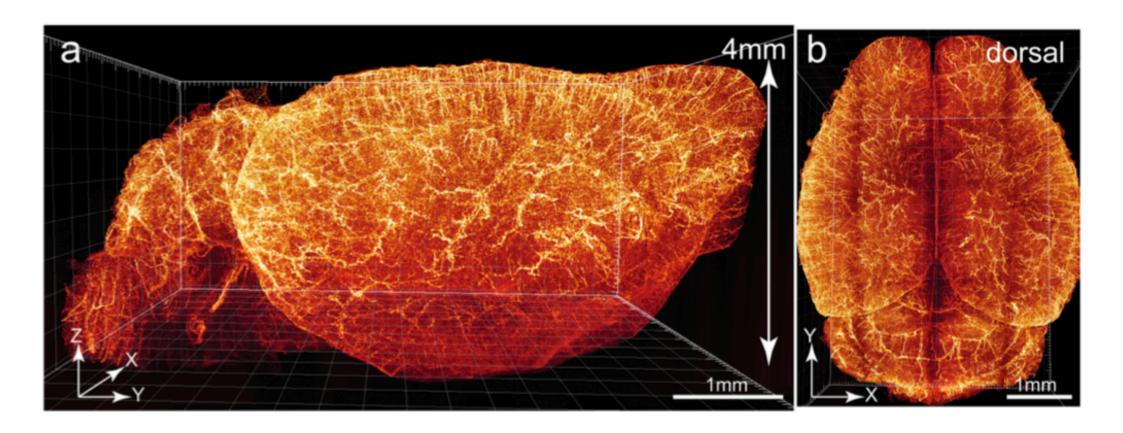
Live Samples



Keller et al., (2010) Nat Meth.

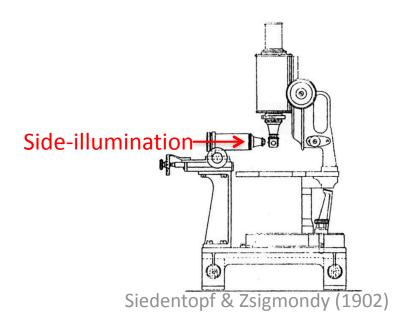
Why use lightsheet imaging?

Large Cleared Samples



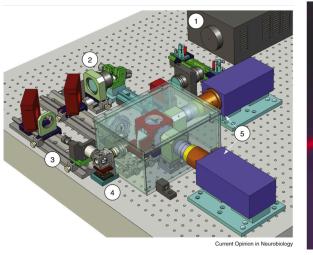
Background

Ultramicroscopy, 1902 bright-field microscopy



Sunlight projected through a **slit-aperture** to observe gold particles

Lightsheet, 1990's flourescence microscopy Ernst Stelzer

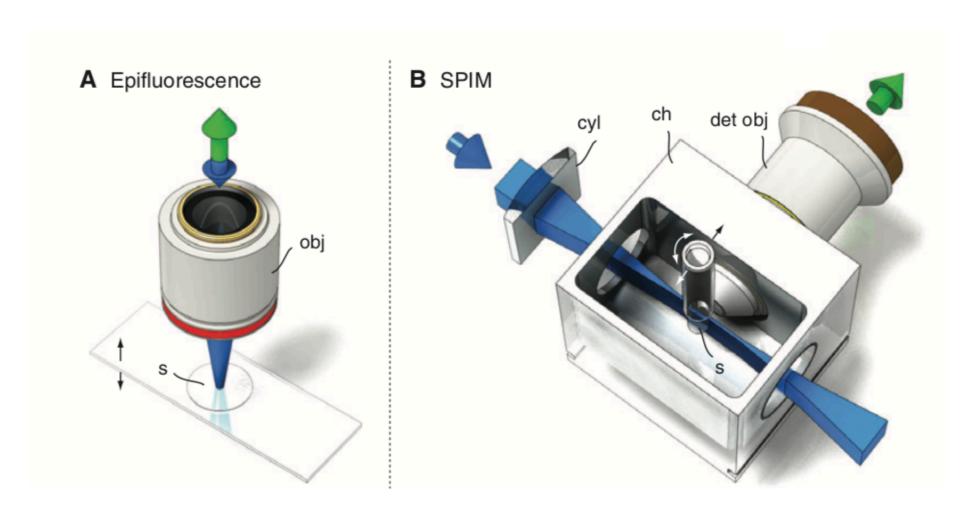




Keller & Stelzer (2008)

Laser lightsheet formed by a cylindrical lens scanned through a **selected plane** of the sample

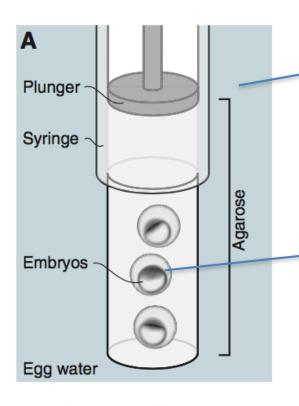
Background



Upright Design

Horizontal Design

Sample Mounting

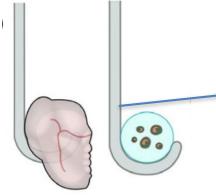


Fluid filled specimen chamber (Water/PBS/Medium)

Normally try to match refractive index sample

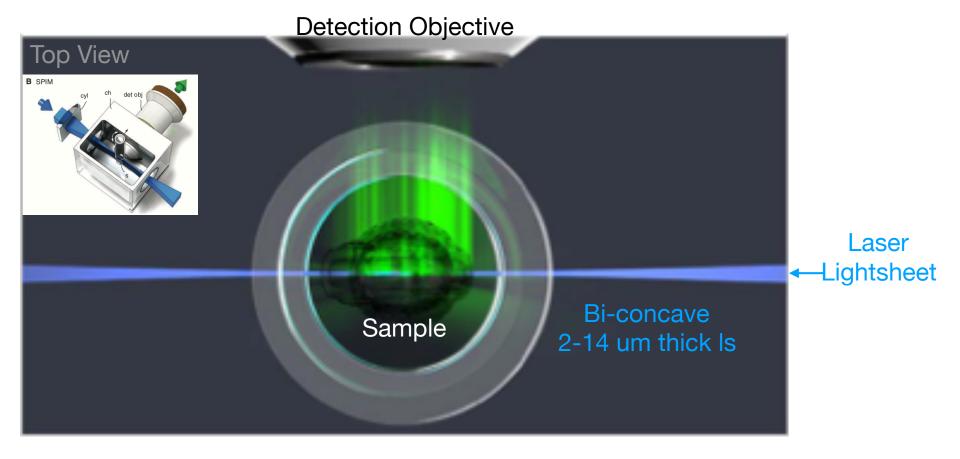
mounting – liquid (water RI 1.33)

Sample immobilised in hydrogel (e.g. agarose) and suspended from motorised stage.



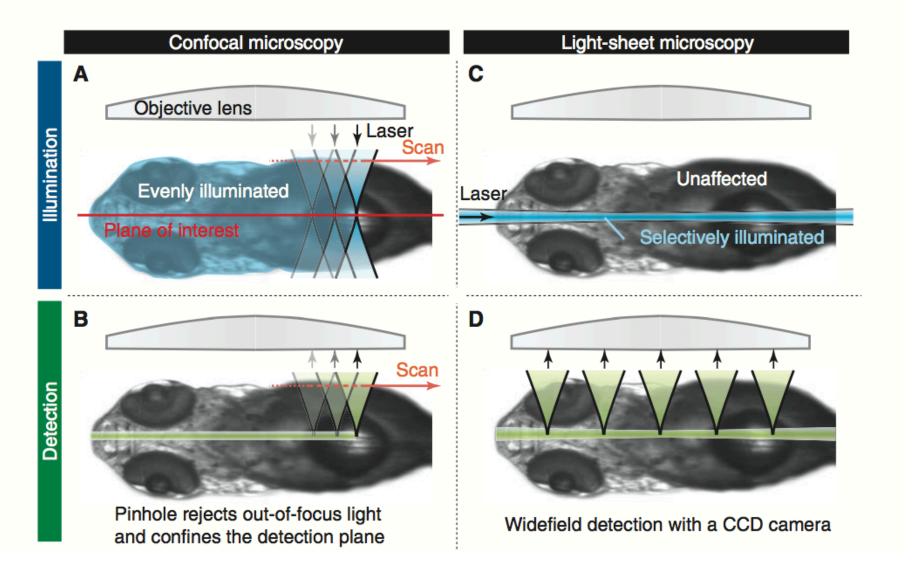
Sample hooked in place & suspended from motorised stage

Selective Plane Illumination

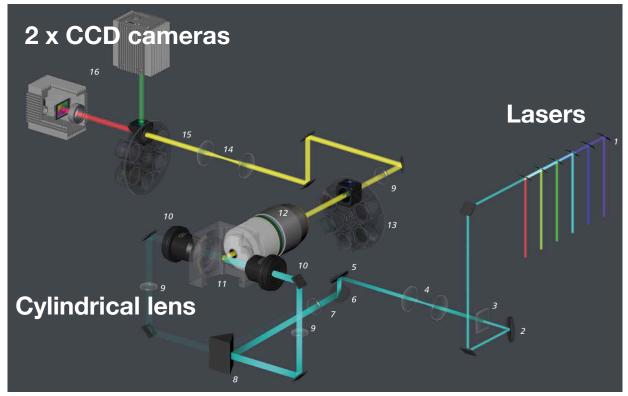


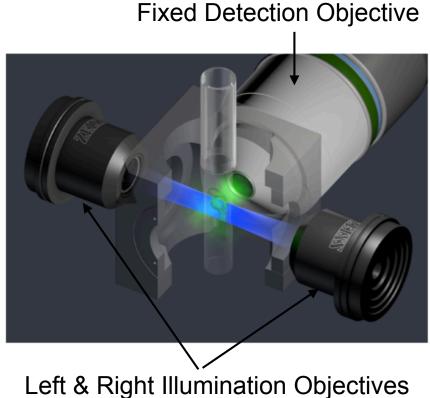
- 1) Single plane of sample illuminated by laser lightsheet
- 2) Fluorescent Emission detected by orthogonally positioned lens
- 3) Each plane of sample is sequentially exposed to the lightsheet
 - = 3D volume of the sample imaged

Faster & less damaging



Dual-Side Illumination



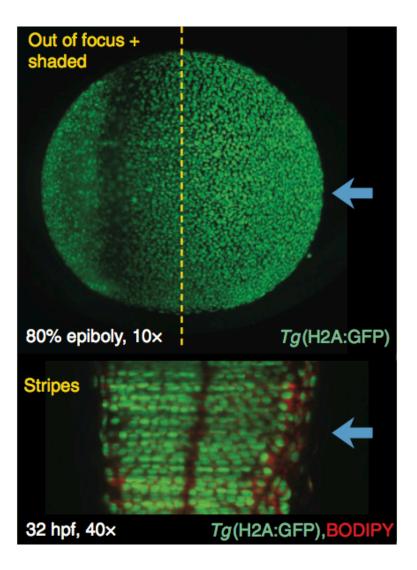




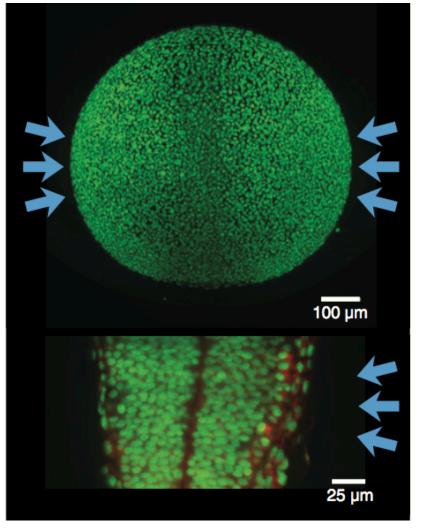
5x/0.16 EC Plan-Neofluar; WD = 5.6mm 20x/1.0 W Plan Apo; WD = 2.4mm (water immersion) 63x/1.0 W Plan Apo; WD = 2.1mm (water immersion)

Illumination issues

Single-side Collimated illumination



Dual-side Multidirectional illumination



Illumination issues

Without Pivot scanner

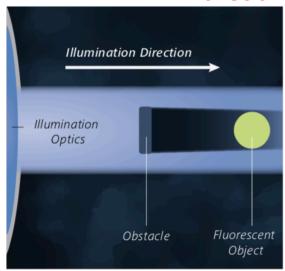
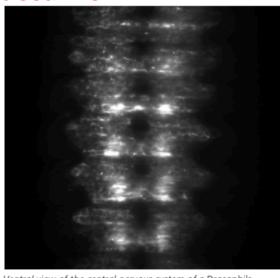
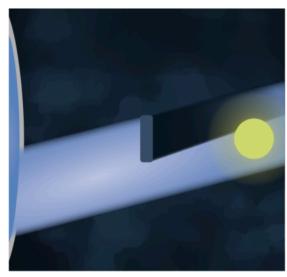


Figure 1



Ventral view of the central nervous system of a Drosophila melanogaster embryo.

With Pivot scanner



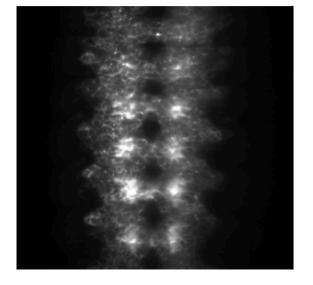
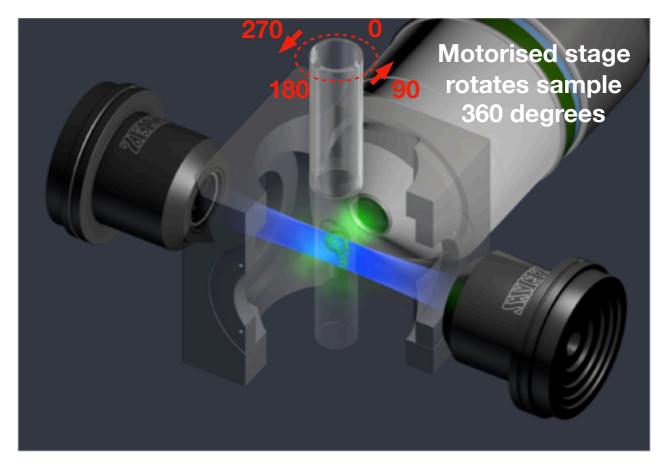


Figure 2

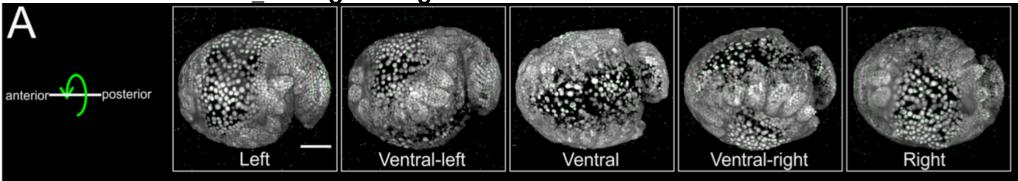
Multi-view Imaging

Sample rotation



- The sample can easily be imaged from multiple view angles
- Post-processing is required to form a single data set
- This is improves axial resolution especially important for large samples

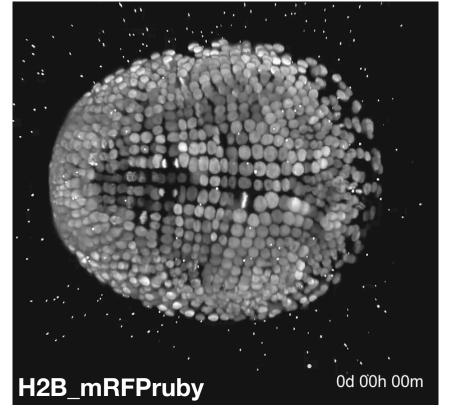
5 views_45 degree angles





Parhyale hawaiensis

All 5 views merged into 1 data-data-set

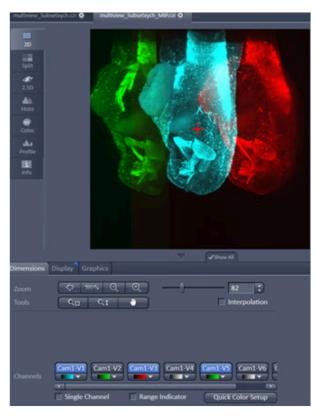


Wolff et al., (2018) Elife: 7

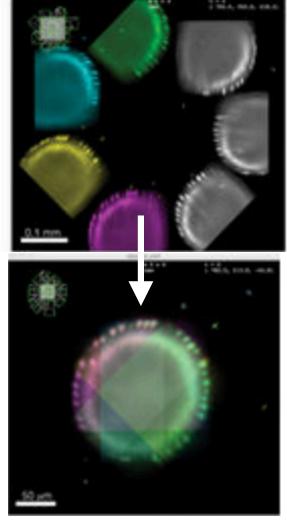
- ~0.2 um diamete fluorescent "fiducial" beads are embedded in the agarose
- By matching up these points, each of the z-stack volumes can be transformed to the same coordinate space & merged
- Beads = "microspheres"
 - Blue (365/430 nm)
 - Green (505/515 nm)
 - Orange (560/580 nm)
 - Red (575/600 nm)
 - Dark red (660/680 nm)

Multi-view Reconstruction

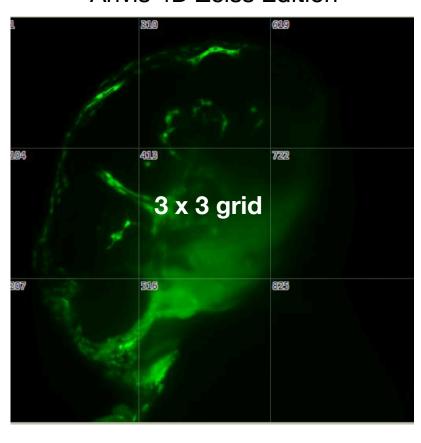
Zen Zeiss Software



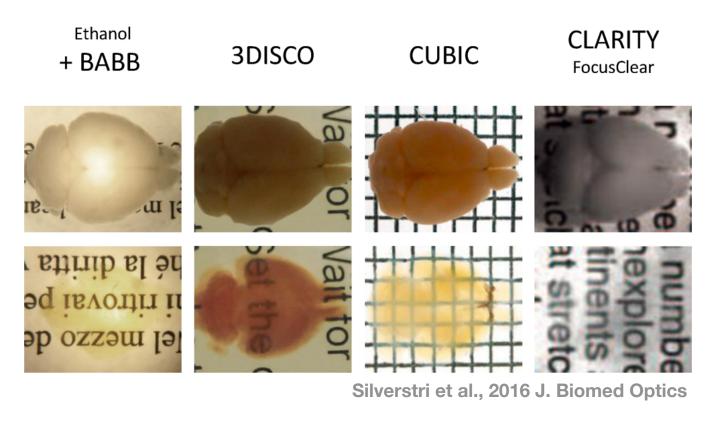
Mutliview Reconstruction Preibisch (MDC Berlin)



TilingArivis 4D Zeiss Edition



Large Fixed Samples will require clearing



- Multiple Methods exist -e.g. DISCO, PEGASOS, CLARITY, FLUO CLEAR
- Requires empirical testing to ID the best for each sample type
- Protocols can takes days/weeks depending on the sample

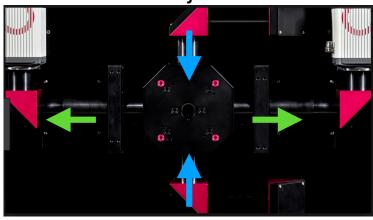
Commercial Versions

Zeiss Z1 Lightsheet (released 2013) **Zeiss Lightsheet 7** (2020)



Luxendo (Bruker) Lightsheet released 2015

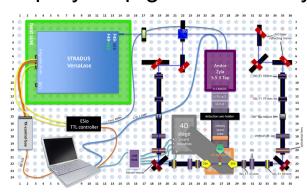
2 x detection objectives



Bespoke Versions

OpenSPIM.org

Step-by-step guide to build your own

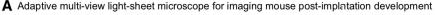


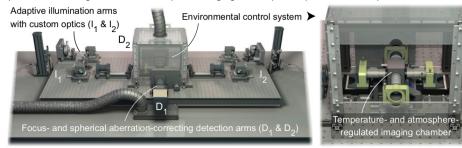
Pitrone et al. 2013 Girstmair et al., 2016

\$50k

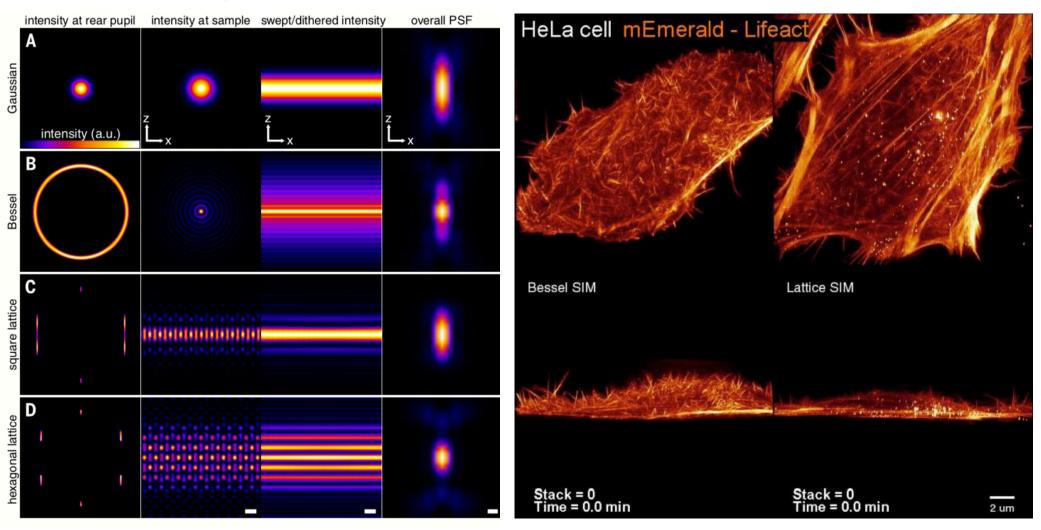
Adaptive multiview microscope

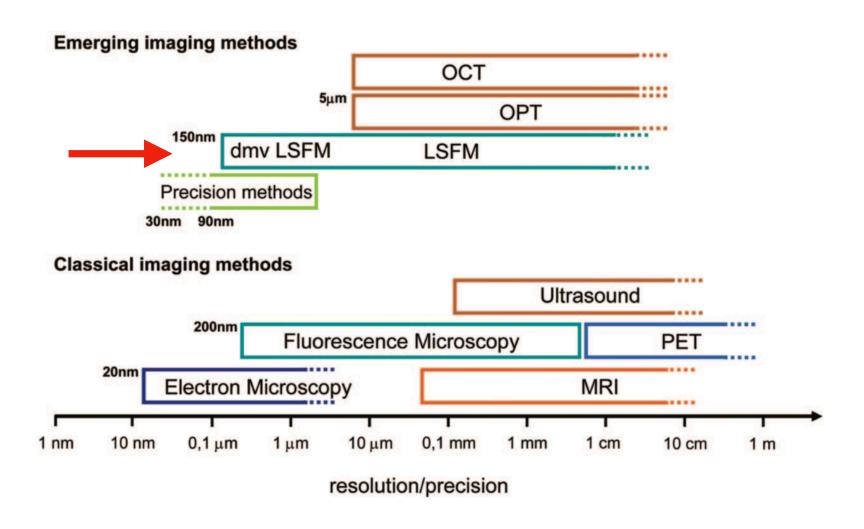
Keller lab (Janelia Research Campus)



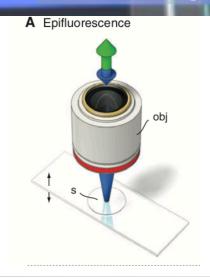


High Resolution LS Microscopy Lattice Lightsheet

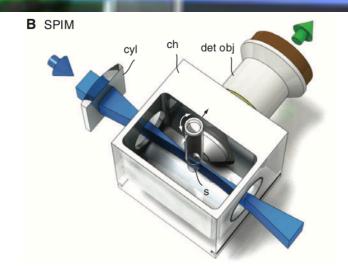




dmv LSFM = deconvolve multiview lightsheet fluorescence microscopy







Whole depth of sample illuminated

One focal plane illuminated at a time

Pin-hole reduces out of focus light

No pin-hole (deconvolution post-processing maybe required)

Easy to mount samples

Sample mounting requires optimisation

Slow acquisition (mins/z-stack)

Fast acquisition (secs/z-stack)

Single view imaging

Multiview imaging

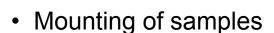
Additional modules exist (FRAP/Semi-Super-resolution/Laser ablation)

Requires bespoke equipment/microscope

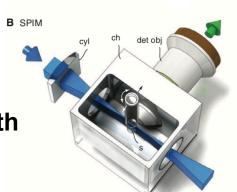
Challenges

Fixed samples

 Optimisation of clearing with IHC/ Fluorescent marker



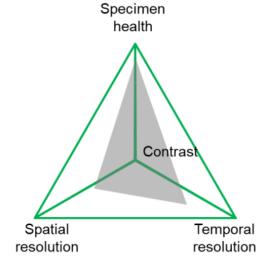
 Reconstruction of image volumes from multi-view angles



Live imaging

Mounting of live samples

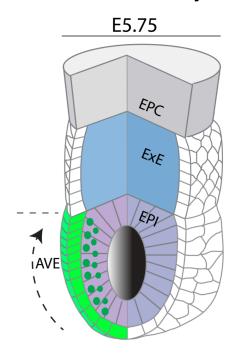
Optimising imaging parameters



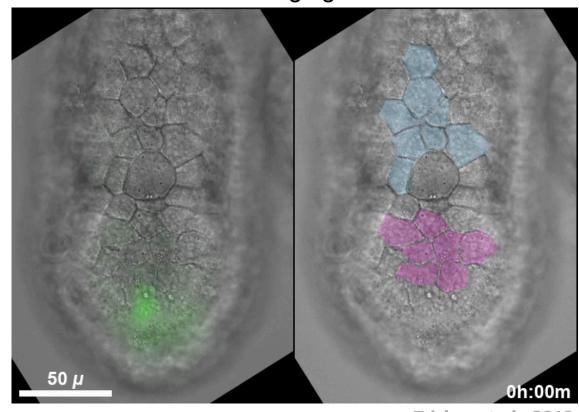
Post-processing large amounts of data
 -TBs of data!

Ligthsheet case study

Mouse embryo



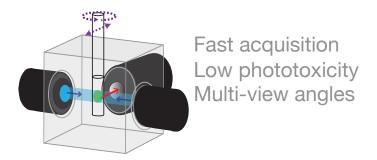
Confocal imaging & DIC



Trichas et al., 2012

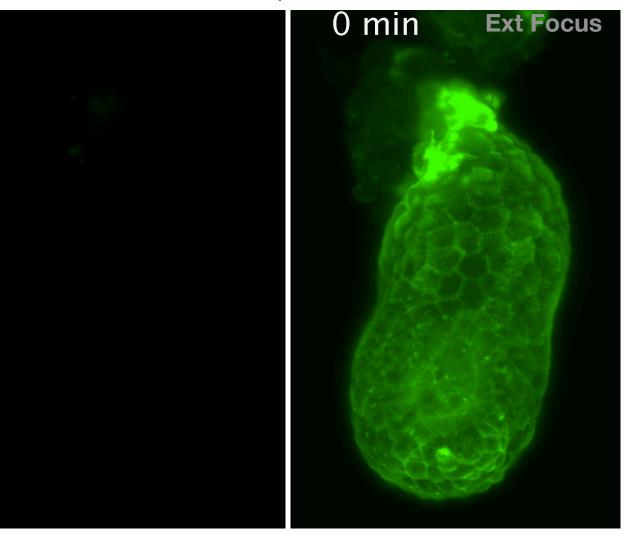
- Q- How are Cell Movements Co-ordinated Tissue-wide in VE?
- AVE Migration takes place over 3-5 hours
- E5.5 embryos are highly light sensitive
- Conventional imaging could only capture a sub-set of the embryo

Ligthsheet Imaging Zeiss Z1 Lightsheet



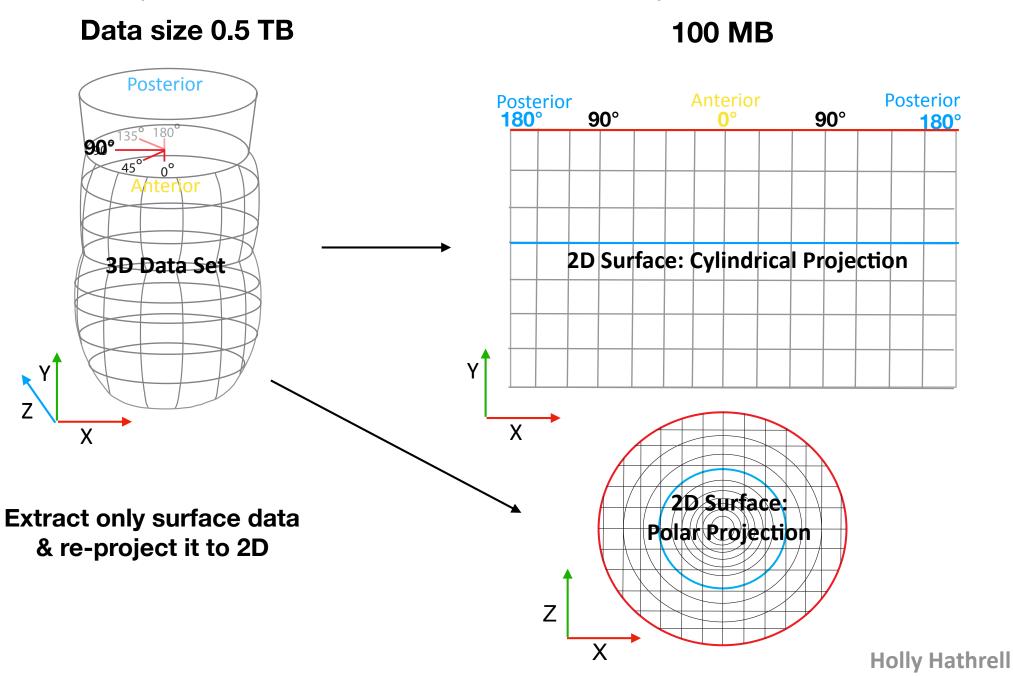
2 full z-stacks (2um step) 2 x view angles (0,180) Every 5 mins 10 hours

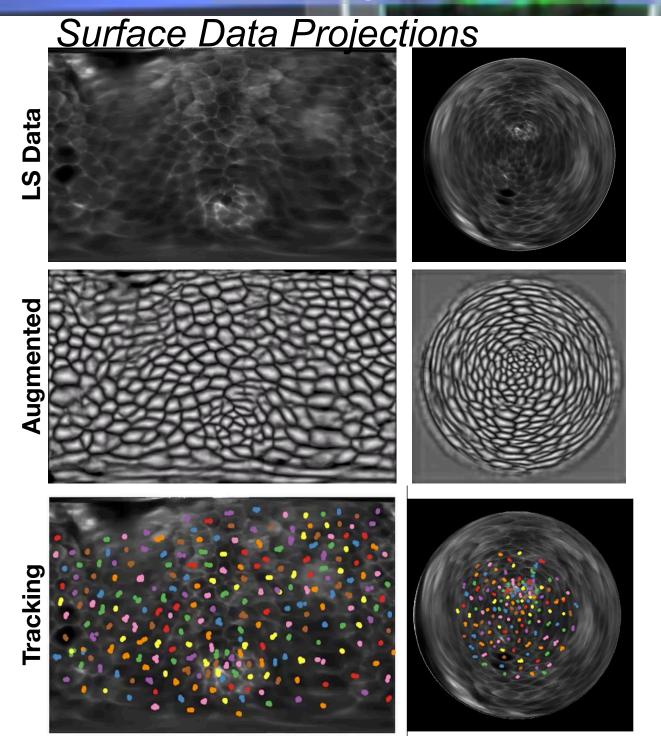
Timelapse Datasets



Lightsheet imaging enabled visualisation of all cells in a single embryo Challenge = Large Data Sets

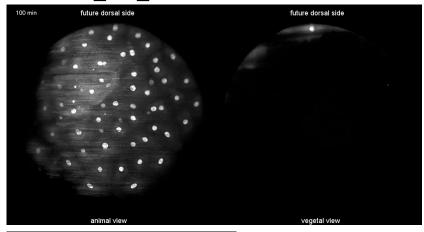
Can we present VE SPIM data in a more visually accessible manner?

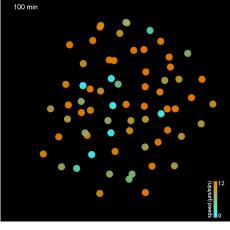


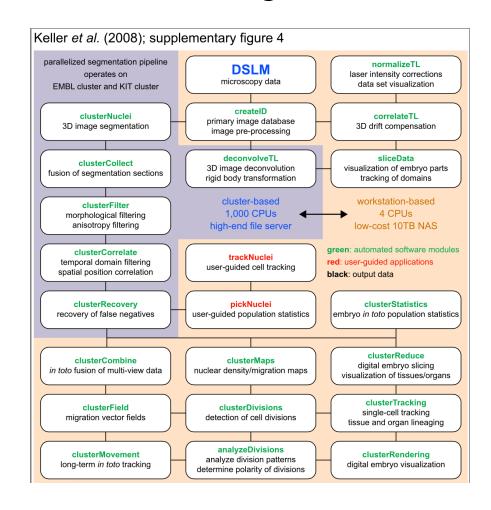


Extracting Quantitative Data - Cell tracking

Histone_2A_GFP







Automated tracking software

Imaris MaMut - Massive Multi-view tracking

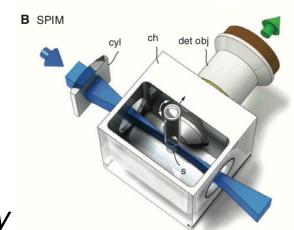
Arivis RACE -Real-time accurate cell-shape extractor

Summary

Selective plane fluorescence imaging

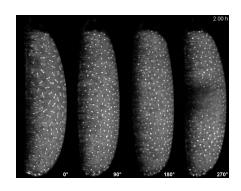
Optically sections samples

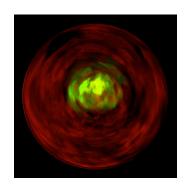
Combines fast acquisition & low photo-toxicity

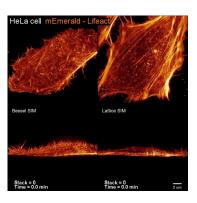


Enables multiview imaging

An emerging technology - we are in the early generations of commercial microscopes







Any Questions?