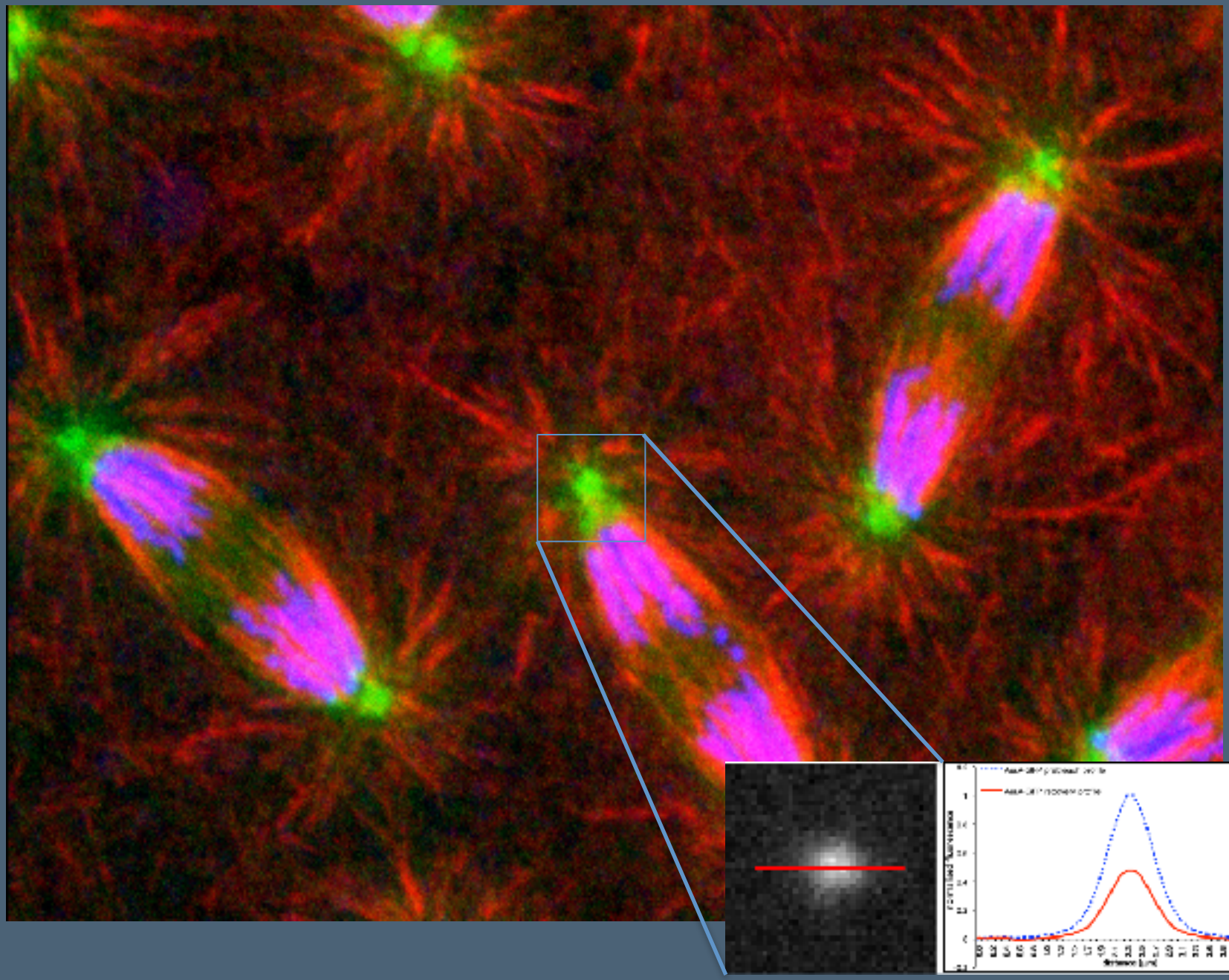
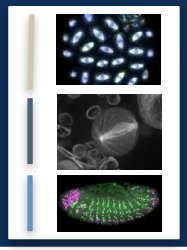
dunn school
bioimaging facility

Confocal Microscopy

(Increasing contrast and resolution using optical sectioning)
Lecture 7

November 2019



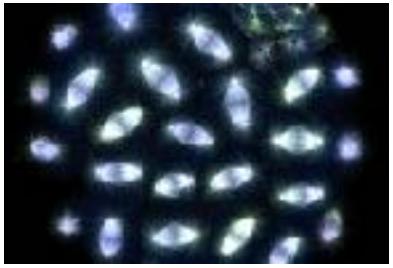


3 Flavours of Microscope

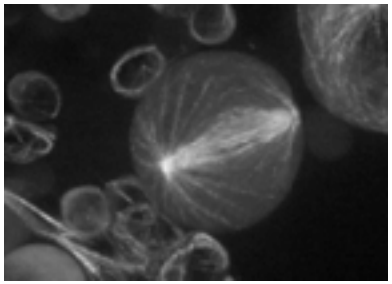
Problem:
Out of Focus
Light

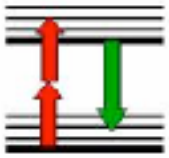
Confocal

Laser
Scanning

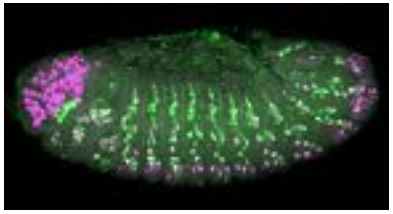


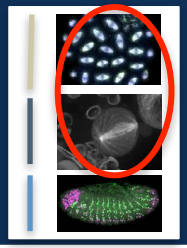
Spinning
disc





2-Photon





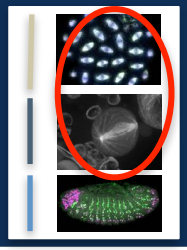
A short History of Confocal

Confocal “concept’ patented by Marvin Minsky in 19

Eggar and Petran developed “spinning disc” confocal in late 1960s

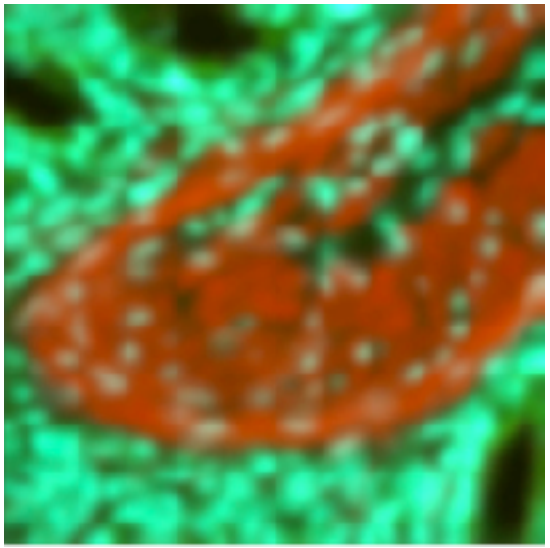
Brakenhoff, Stelzer developed “stage” scanning confocal in late 1970

White, Amos and Wilson developed the MRC500 point scanning confocal
-Marketed commercially in 1987



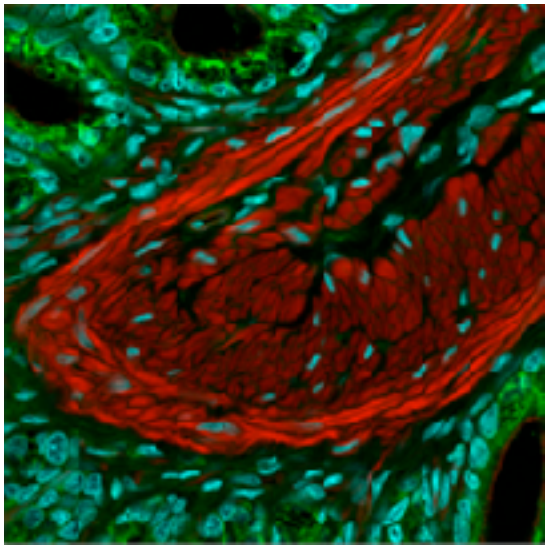
Comparison Widefield Vs Confocal

Widefield

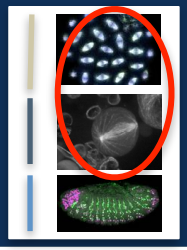


Out of focus light ‘blurs’ image

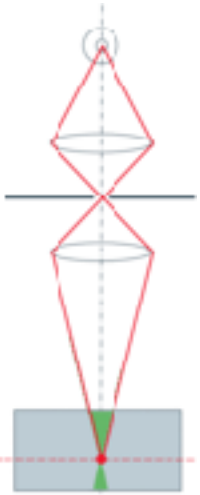
Confocal



Out of focus light is blocked

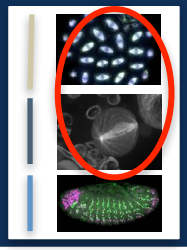


Principle of Confocal Microscopes

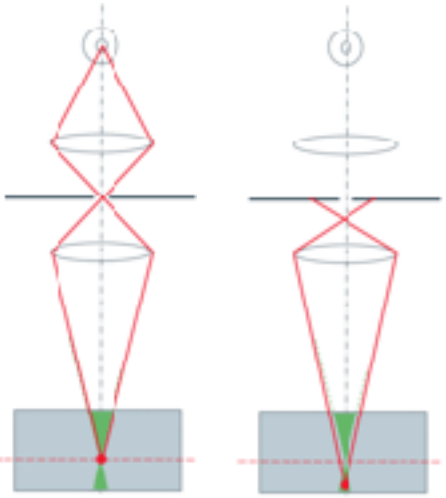


Pinhole diaphragm in the Conjugated focal plane = CONFOCAL

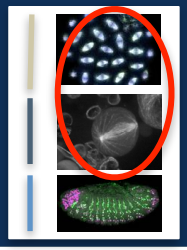
in focus light (from the optical section) passes through the pinhole and into the detector



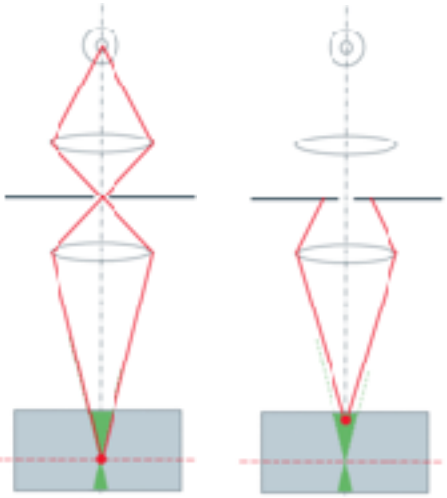
Pinhole – blocks out-of-focus light



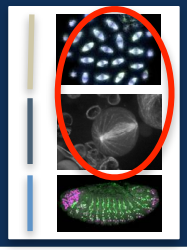
light from below the optical section crosses infront of the pinhole and doesn’t pass through the pinhole



Pinhole – blocks out-of-focus light



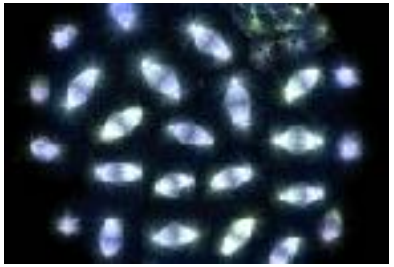
light from above the optical section also doesn’t pass through the pinhole aperture



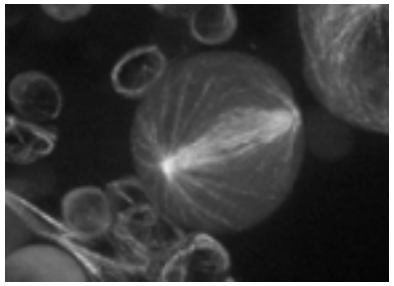
Confocal Microscopes

Confocal

Laser Scanning



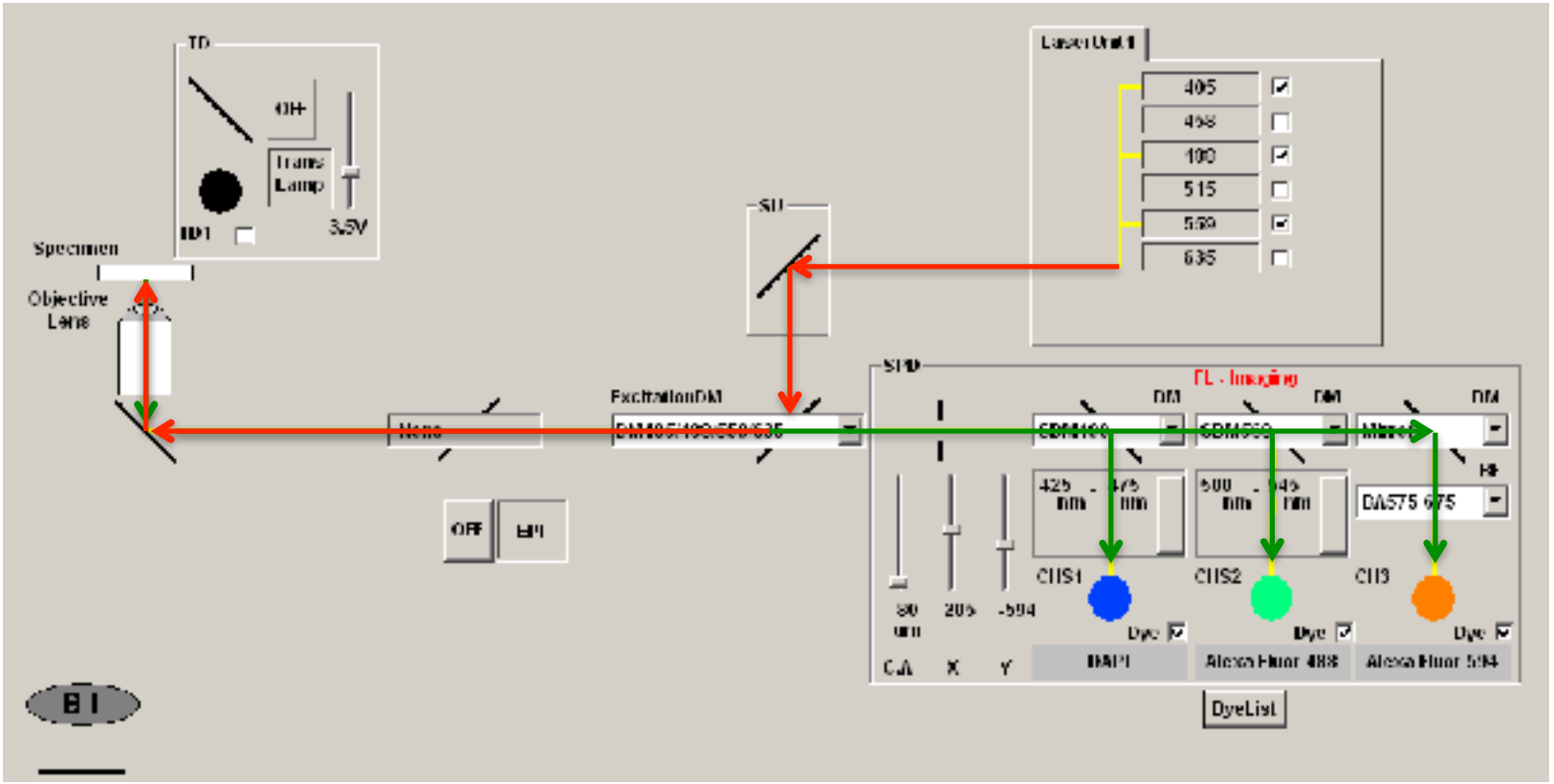
Spinning disc



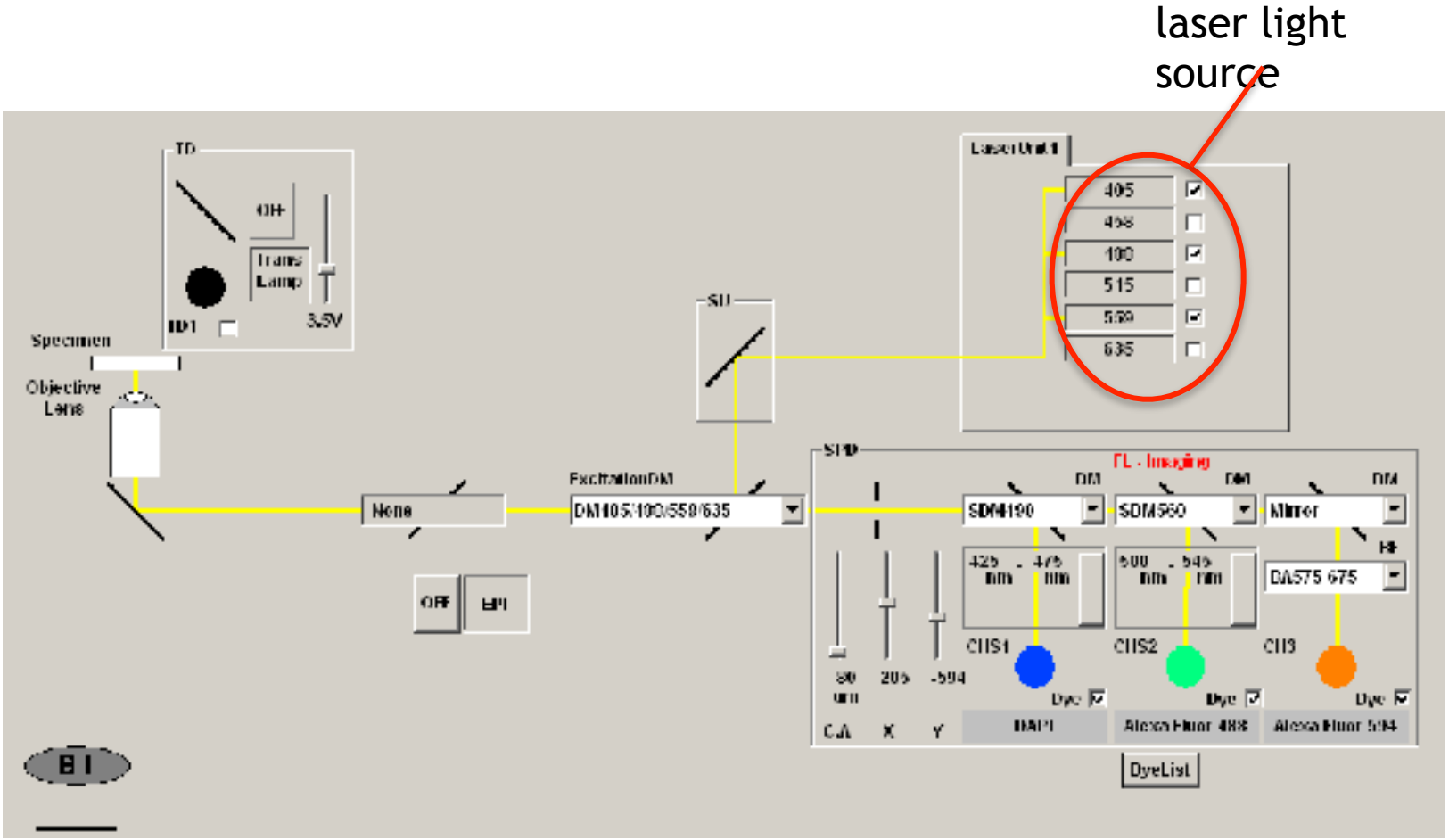
Laser Scanning Confocal Microscope

Laser Scanning Confocals are great to get ‘pretty’ images

Laser Scanning Confocal - components



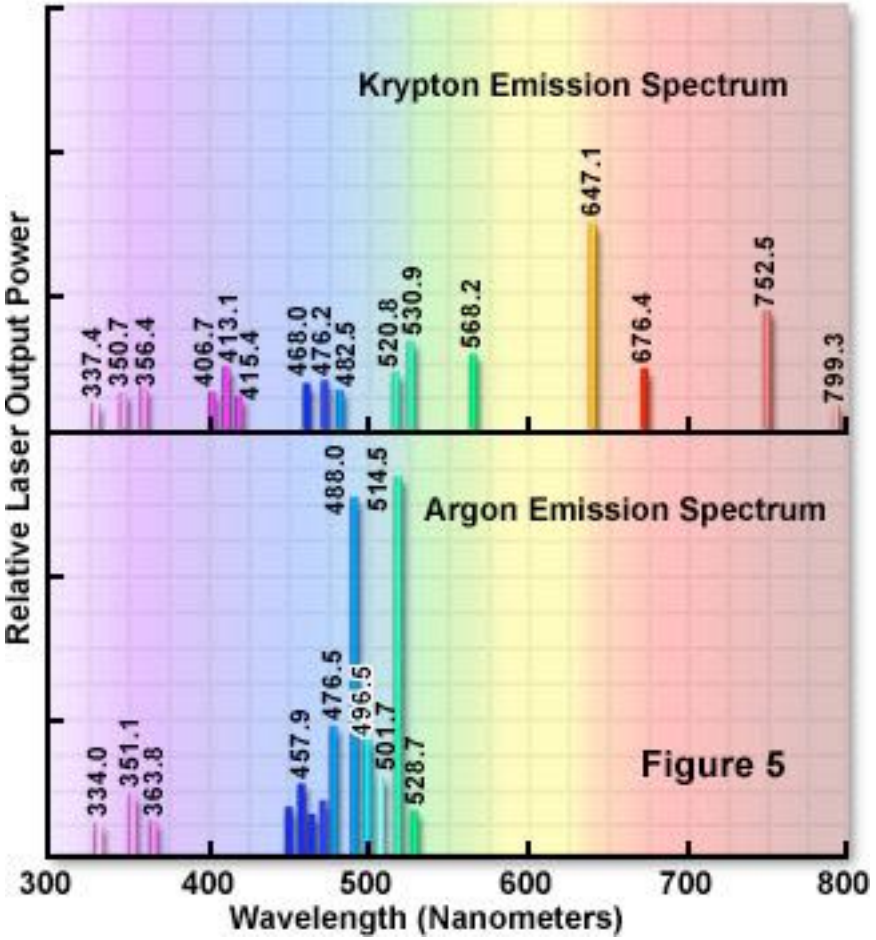
Laser Light Source



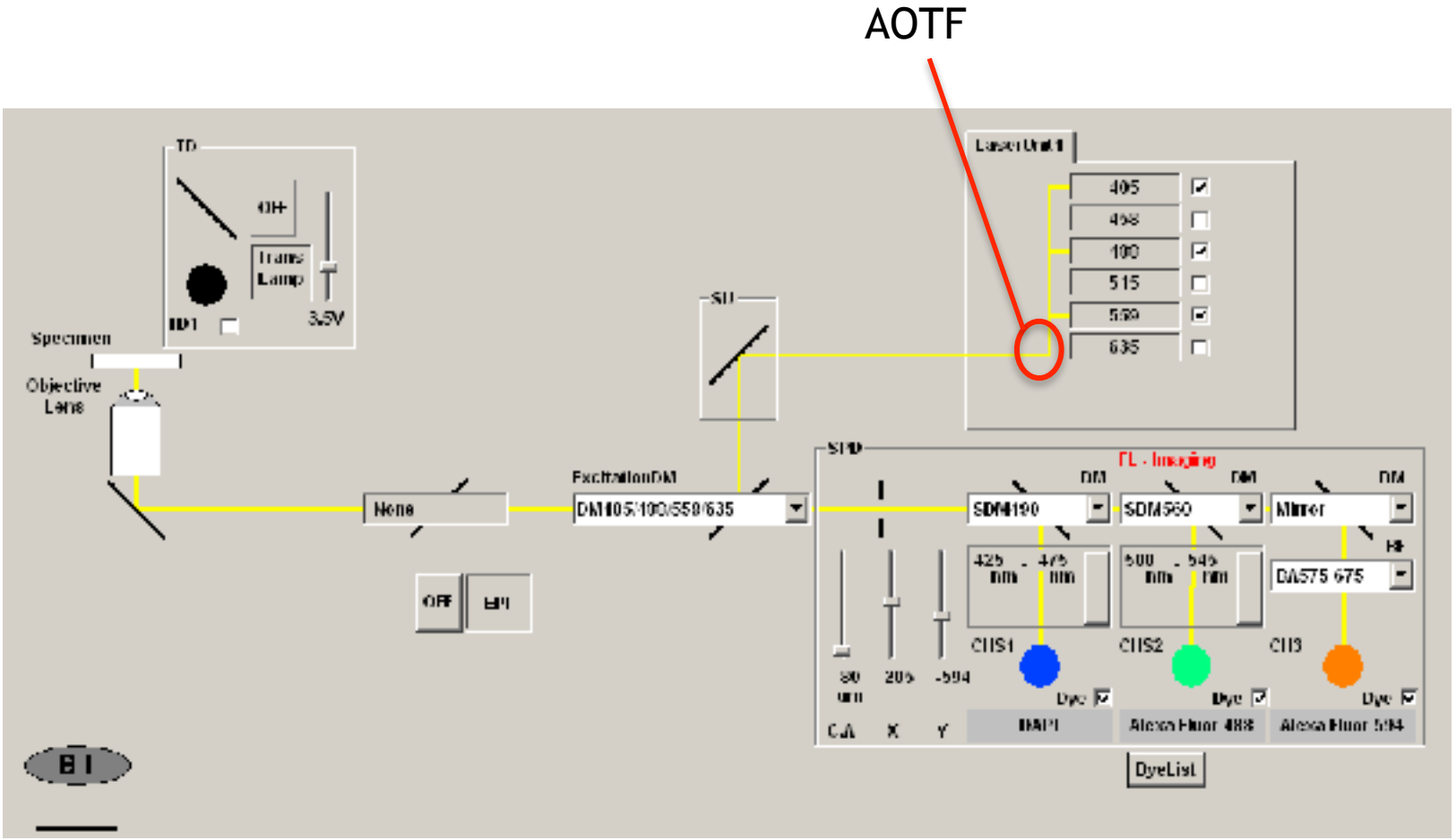
Laser Light Source

enables tighter control of fluorophores excited

Laser Emission Spectra

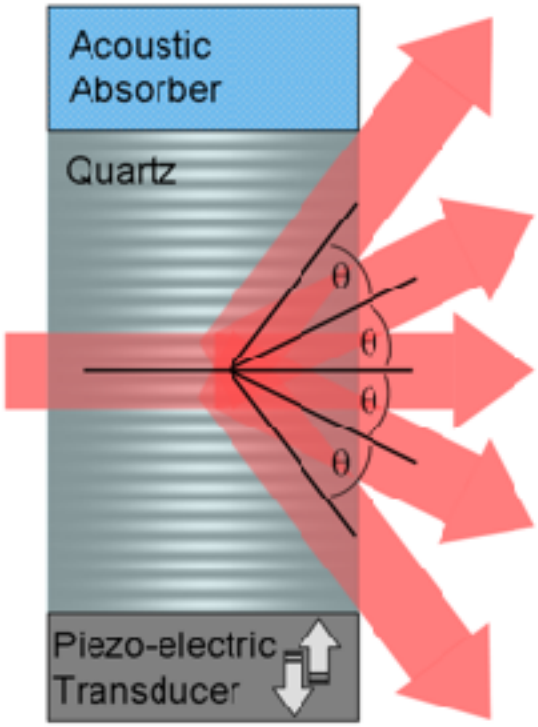


AOTF
Acousto-Optic Tunable Filter



AOTF
Acousto-Optic Tunable Filter

THEORY

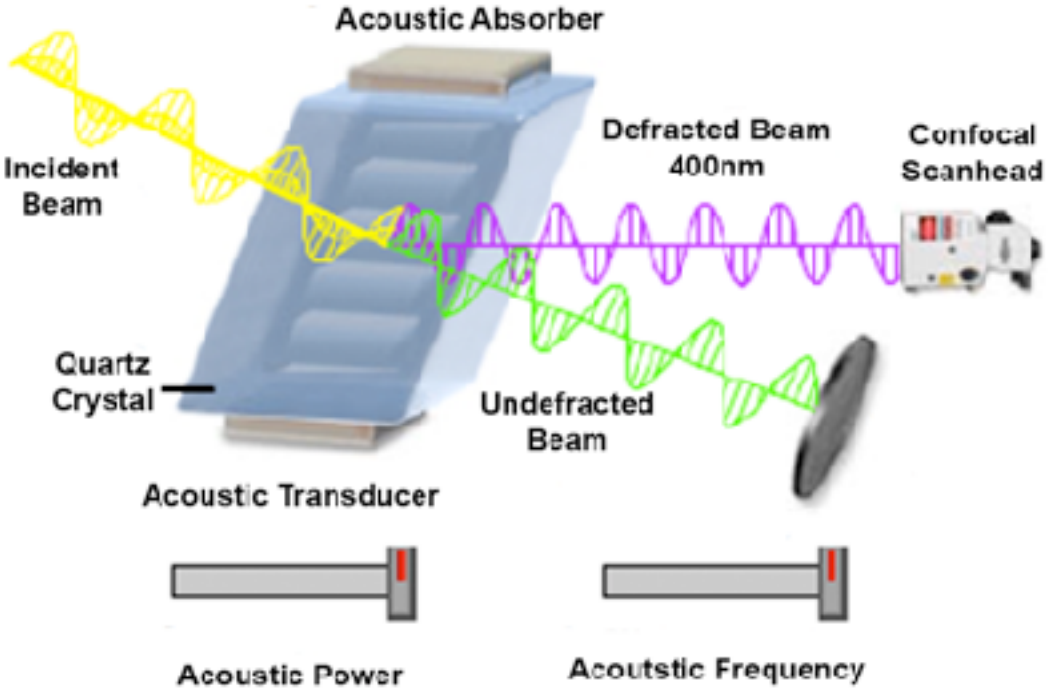


acousto-optic effect:

Acoustic wave excited within the quartz gives rise to variations in the refractive index

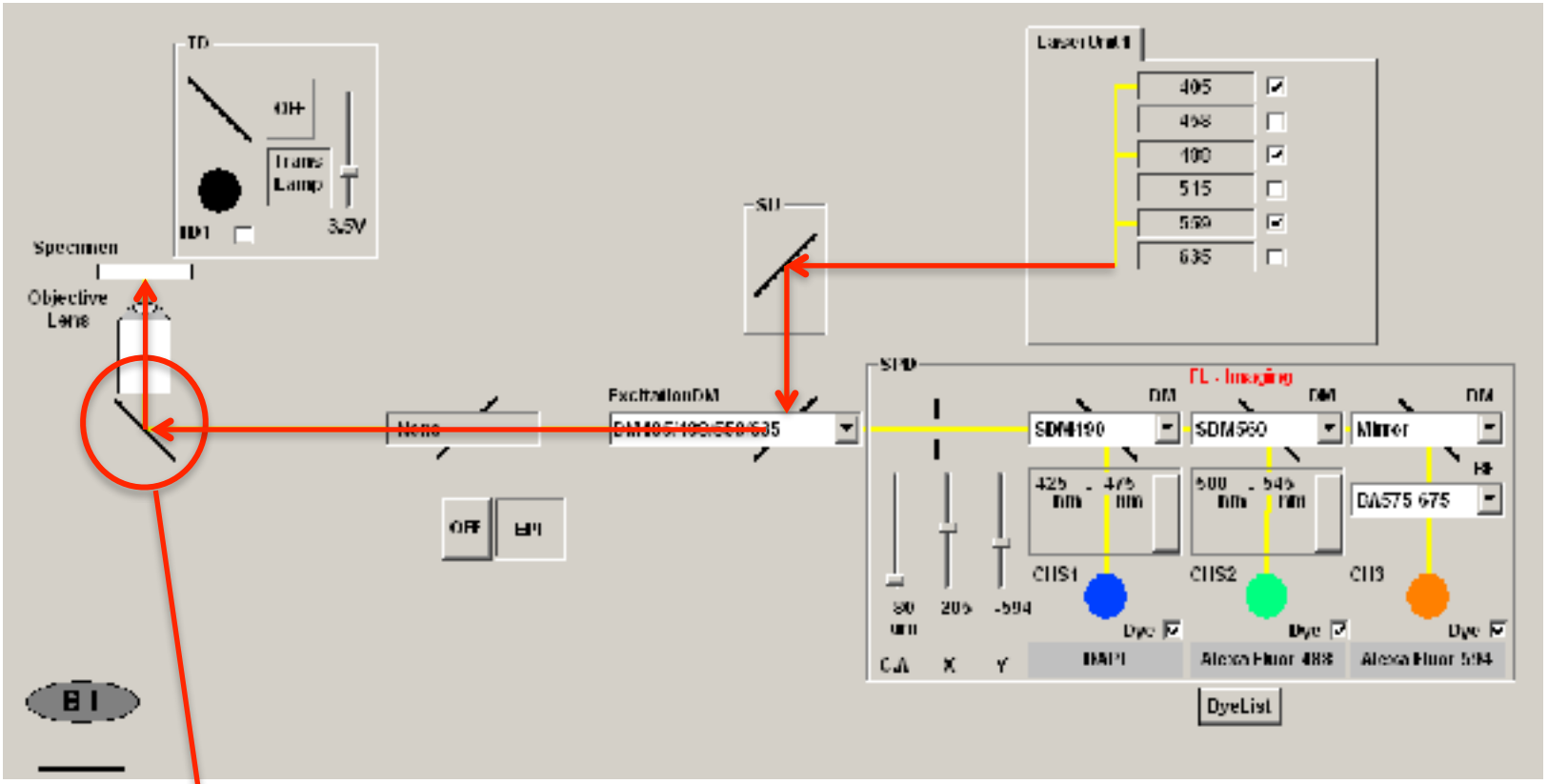
The wavelength of the diffracted light is dependent on the acoustic frequency in the quartz. By tuning the frequency of the acoustic wave, the desired wavelength of the optical wave can be diffracted acousto-optically.

AOTF
Acousto-Optic Tunable Filter



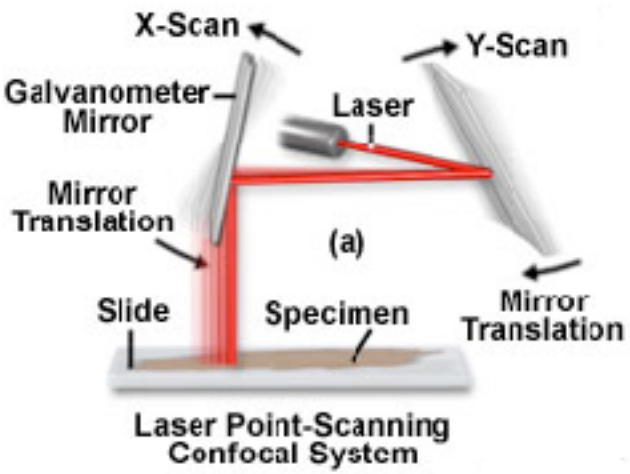
Quick On/Off of lasers
Very fast changes between excitation

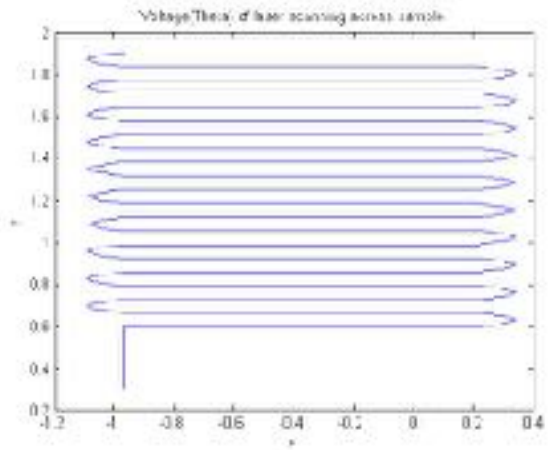
Galvo Scanning Mirrors



Galvo Scanning Mirrors

Galvo Scanning Mirrors

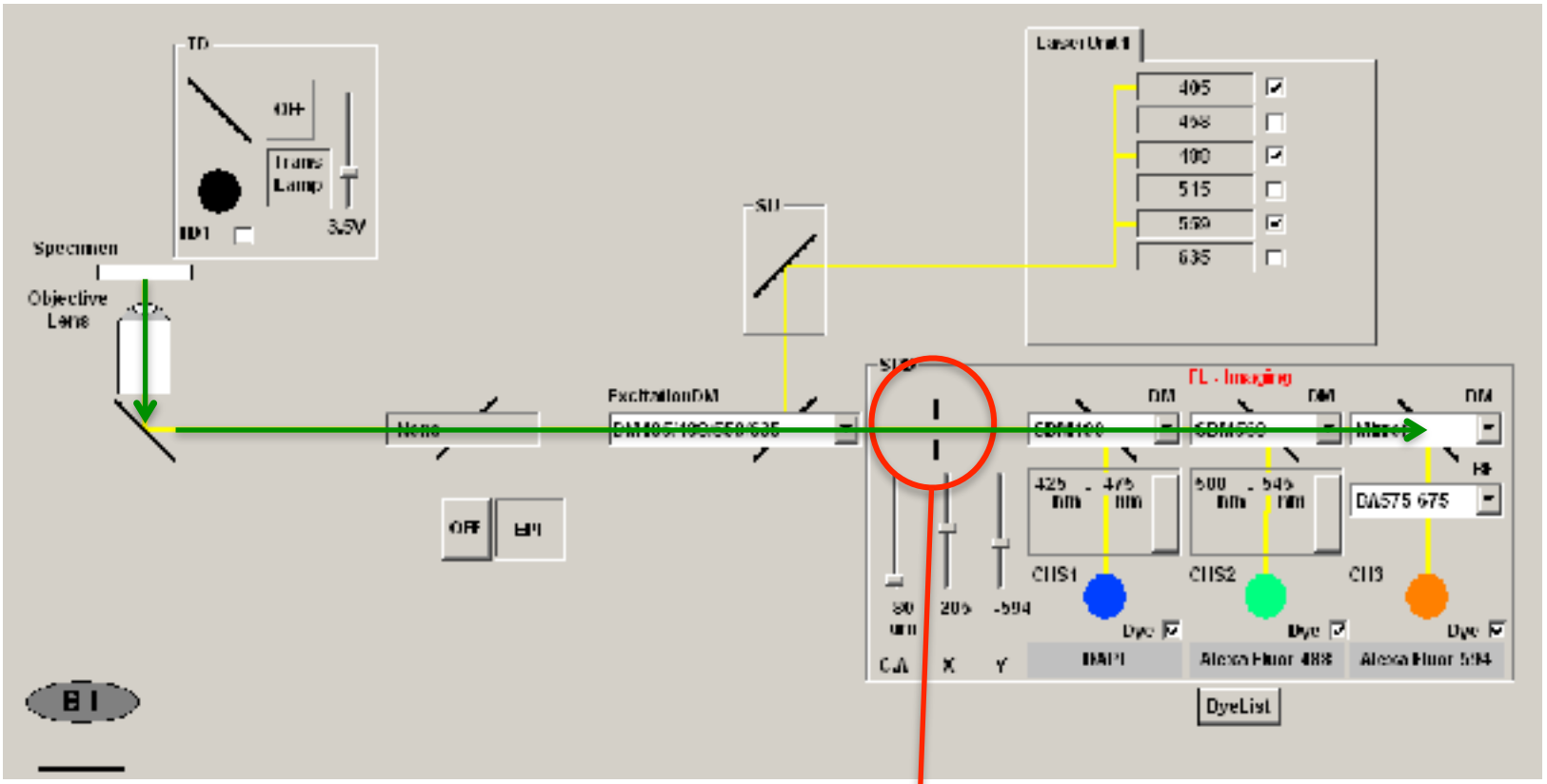




Sample excited at one point at a time

Adjustable Pinhole

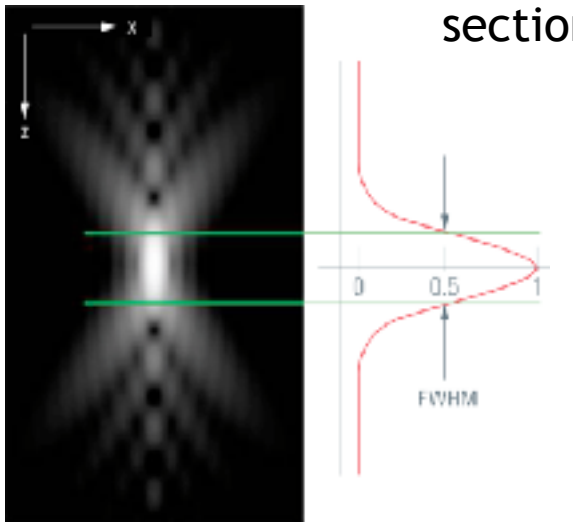
AOTF



pinhole

THEORY

Pinhole – Optical Sectioning



FWHM=Full Width Half-Maximum

Shorter the wavelength the thinner the optical section

Diameter of the pinhole: Smaller pinhole thinner optical section

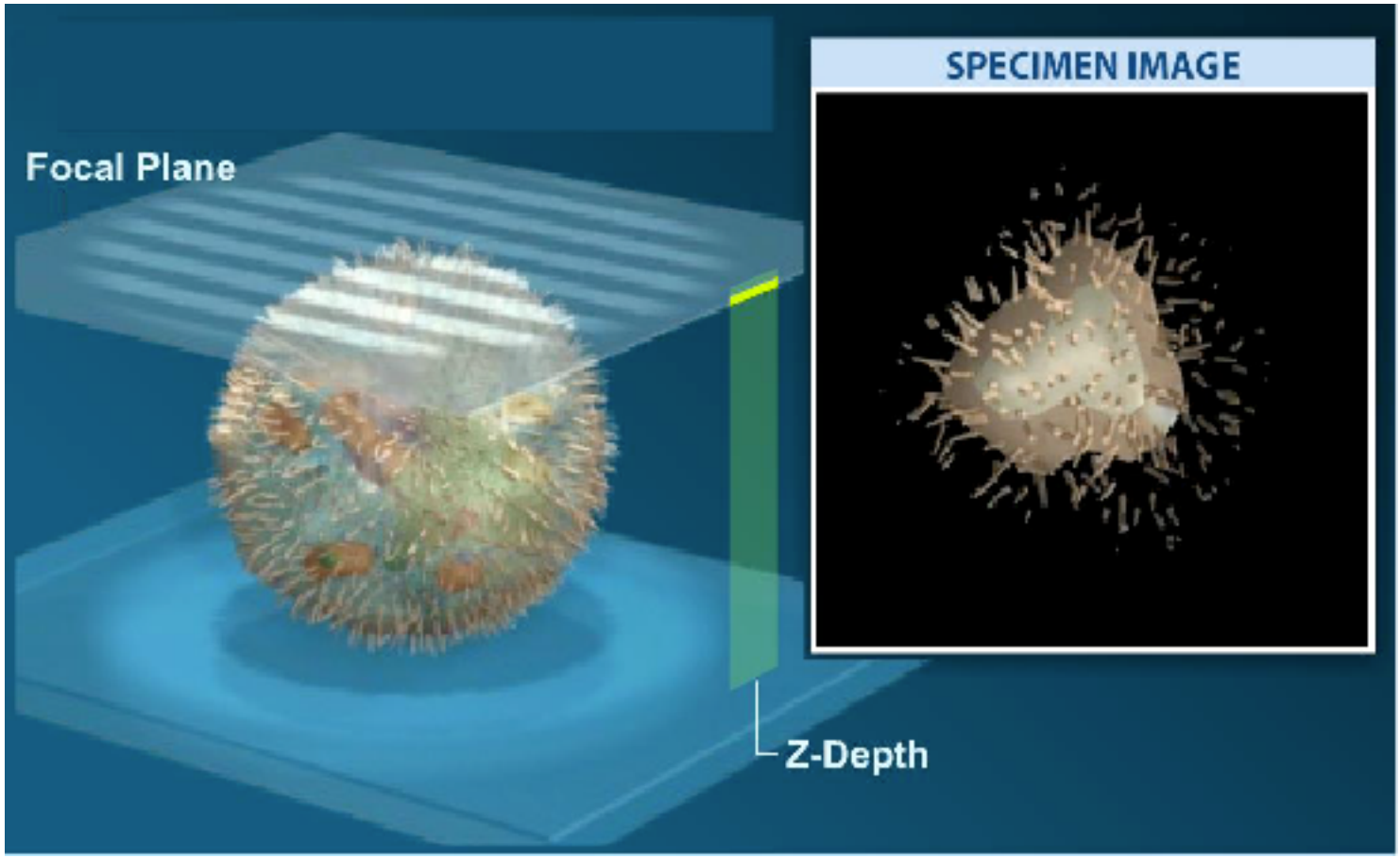
$$FWHM_{axial} = \sqrt{\left(\frac{\lambda_{exc} \cdot n}{NA^2}\right)^2 + \left(\frac{n \cdot \sqrt{2} \cdot PH}{NA}\right)^2}$$

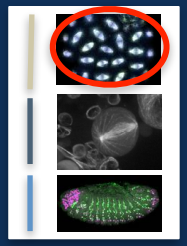
The higher the NA. the thinner the section

Weak signal > open pinhole > more light but thicker

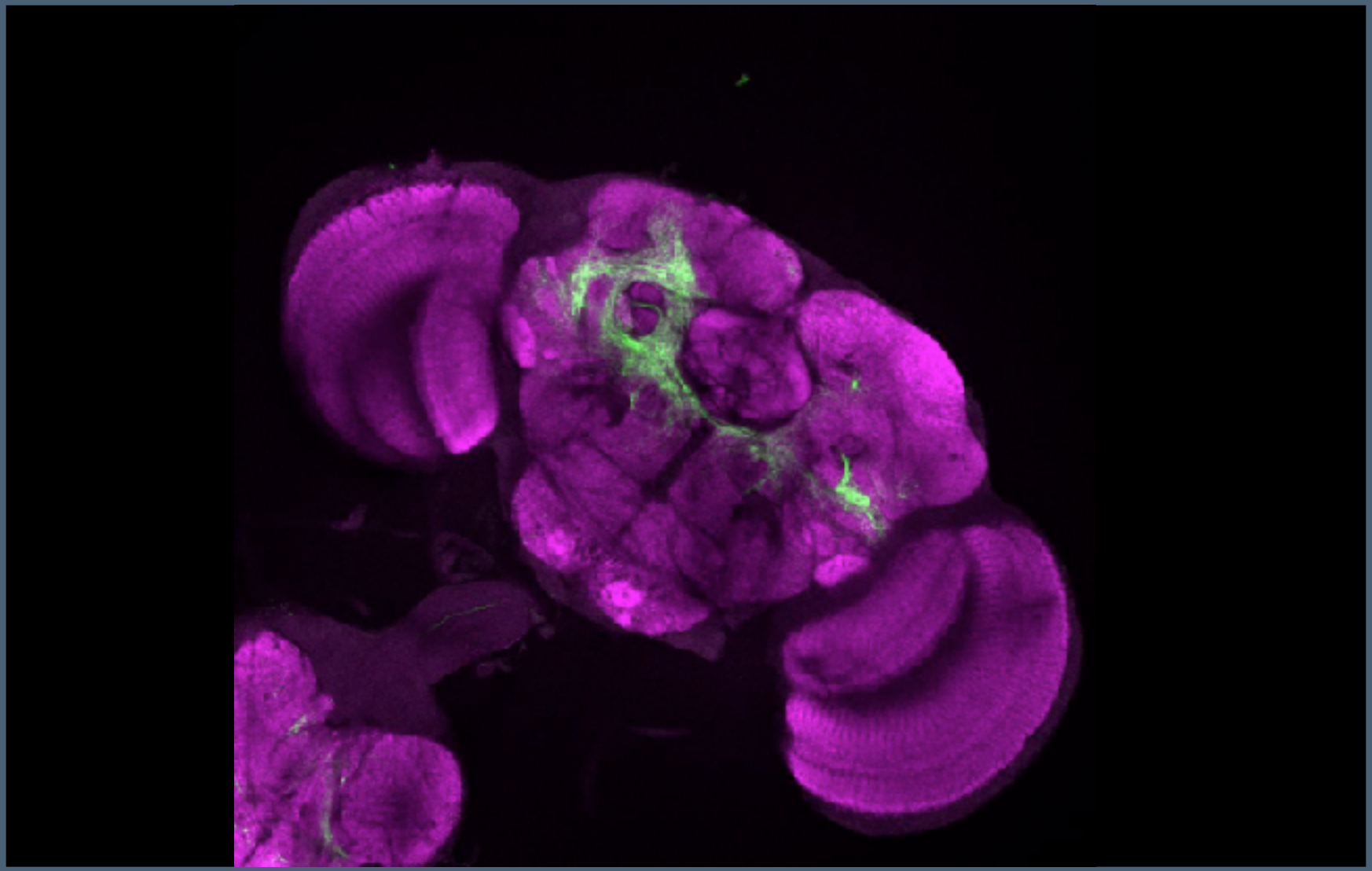
Confocal enables 3D reconstruction

Optical section



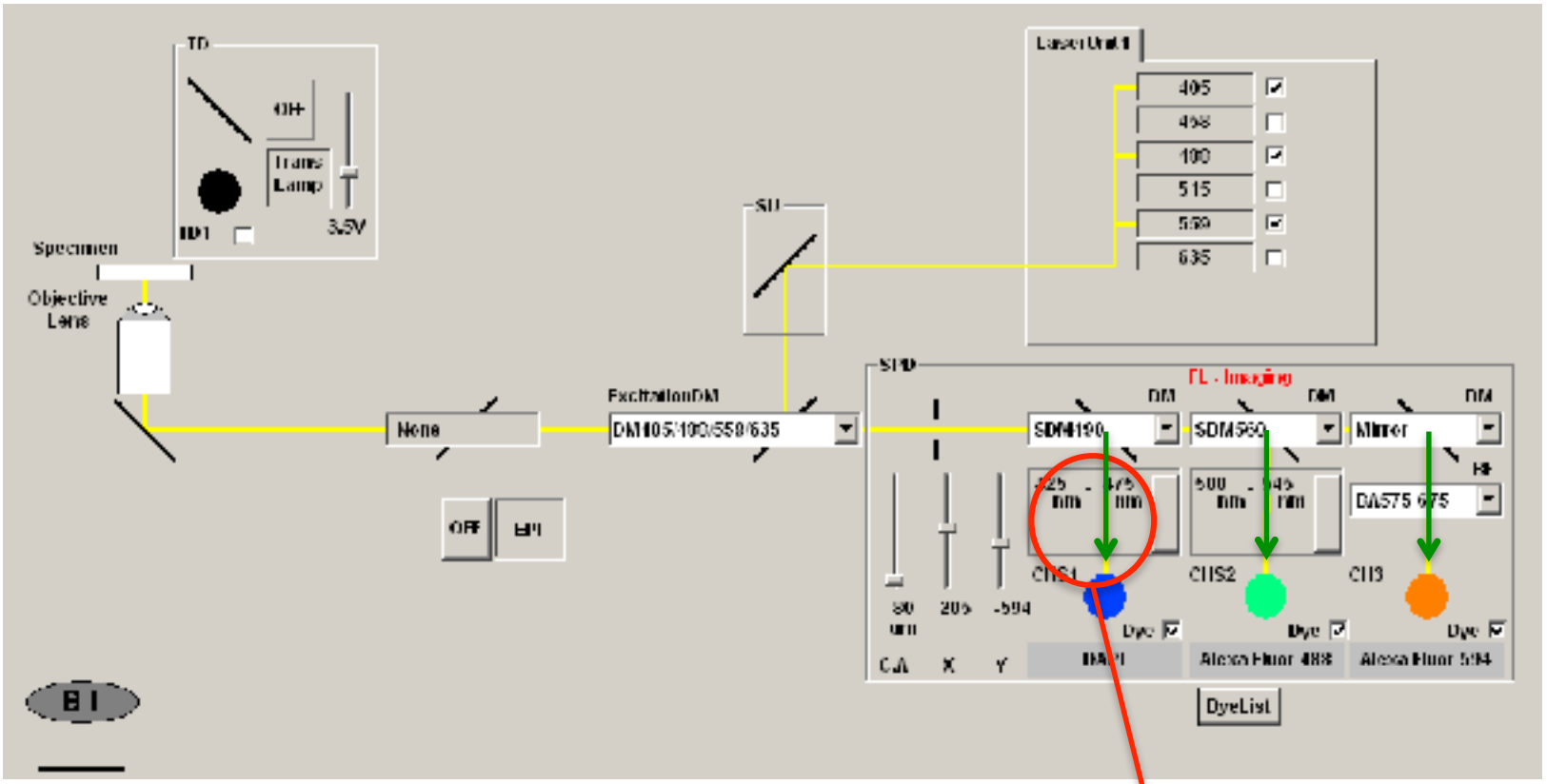


Confocal enables 3D reconstruction

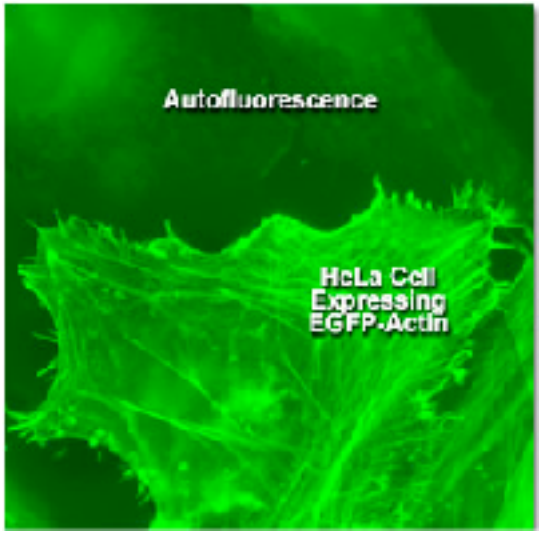


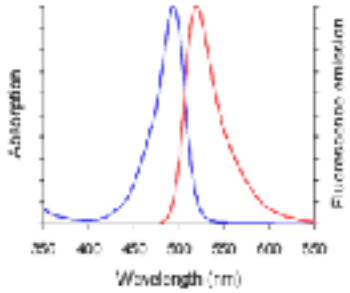
Adult Drosophila head (C. Rezeval Goodwin Lab)

Variable Detector Slit



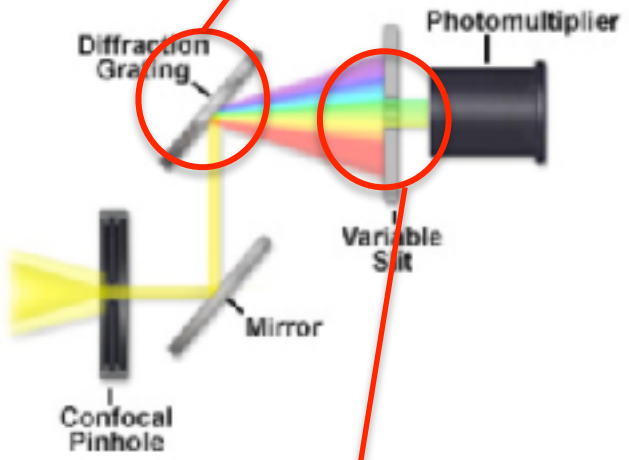
Spectral Unmixing





Light emitted from fluorophore as a spectrum

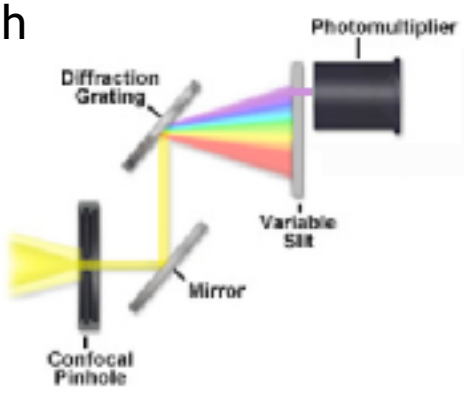
Defraction grating separates wavelengths over physical area

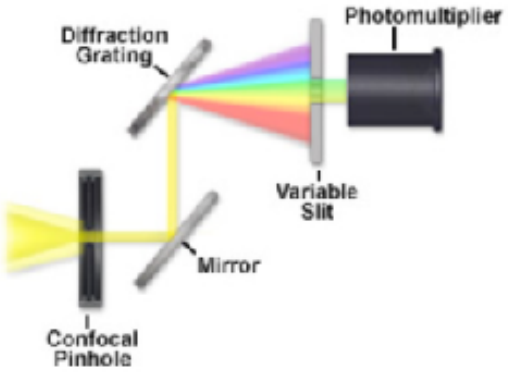


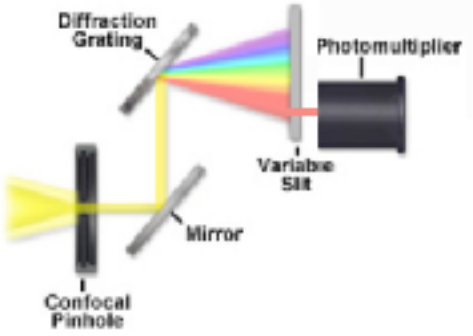
Variable slit lets through only certain wavelengths

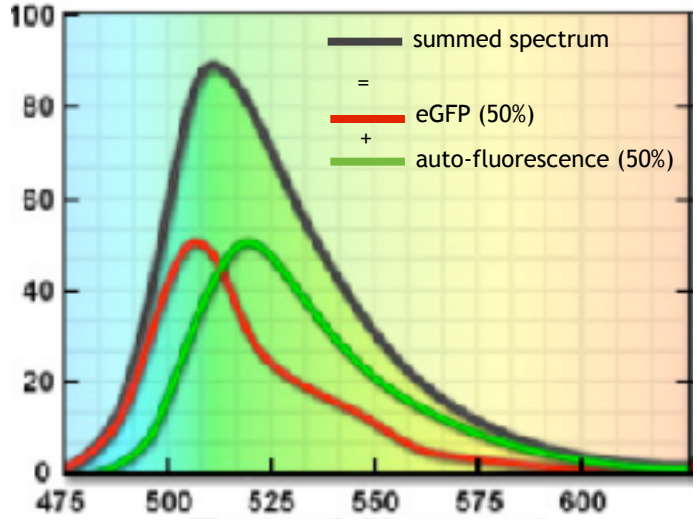
Spectral Unmixing

At each pixel:



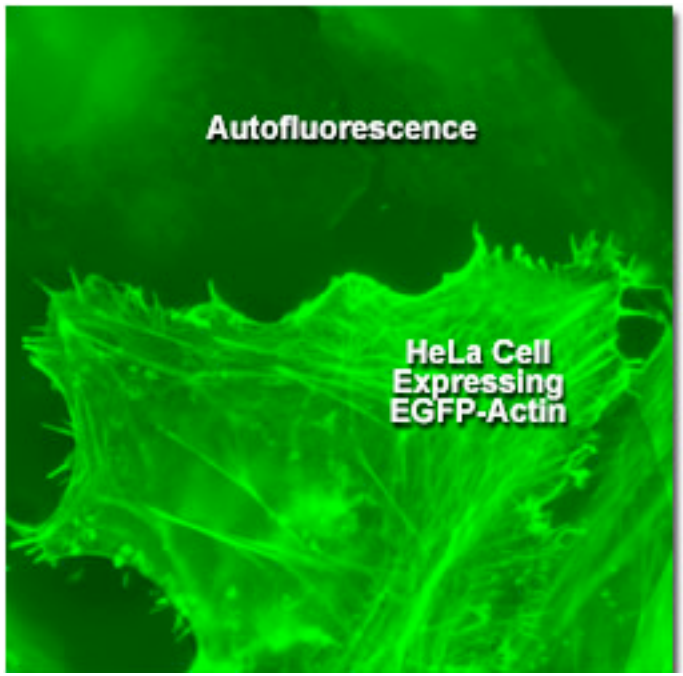


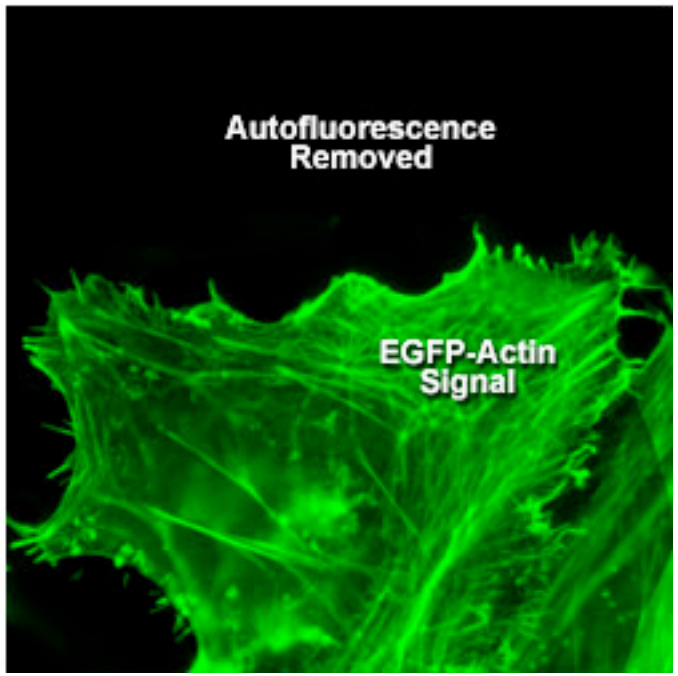




Match the summed spectrum with all possible summed combinations from a library
At each pixel you therefore know the proportion of each fluorophore present

Spectral Unmixing removal of



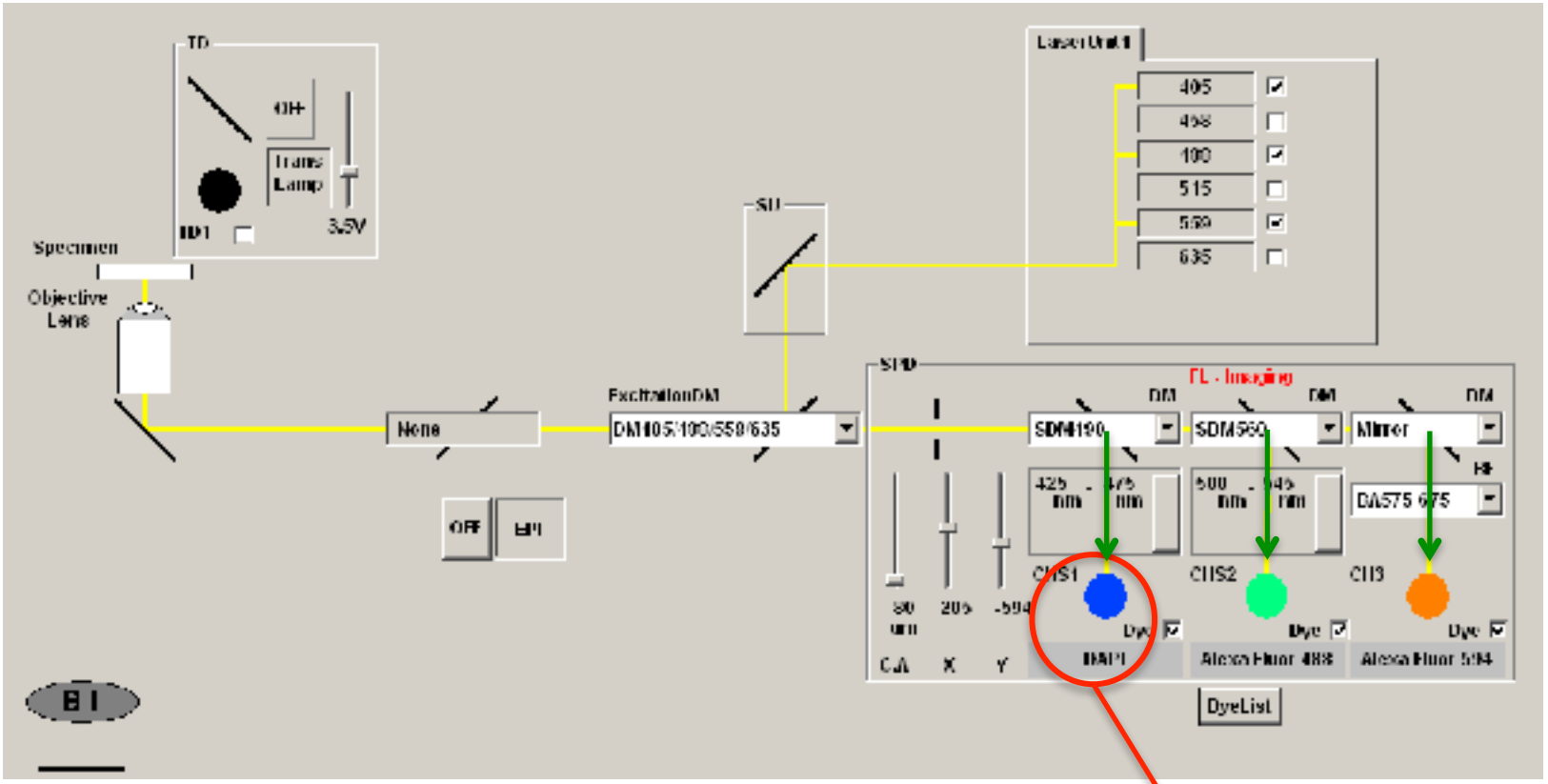


At each pixel:

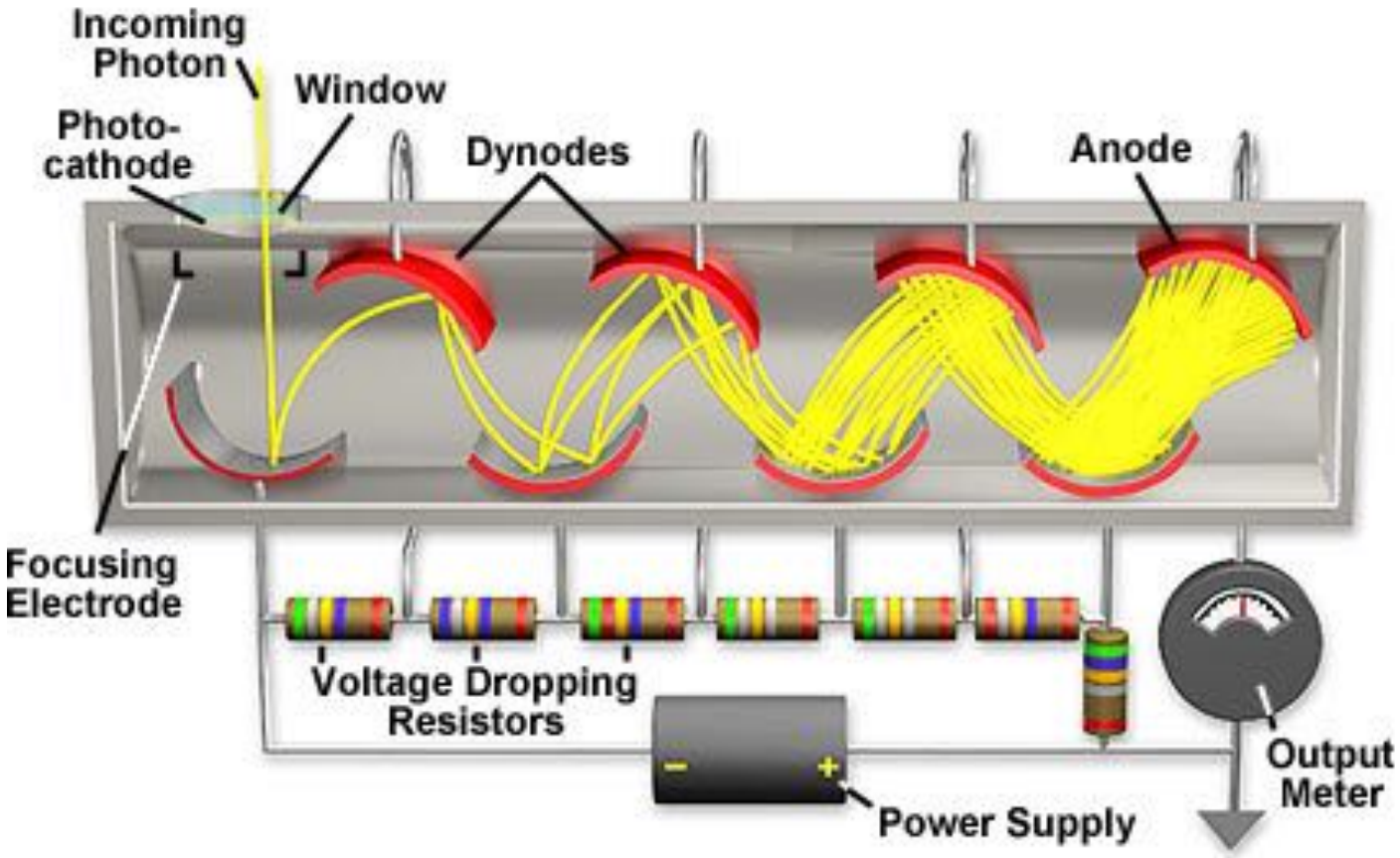
Calculate the proportion of the pixel is due to autofluorescence.

Subtract the autofluorescence from the ‘true’ GFP value.

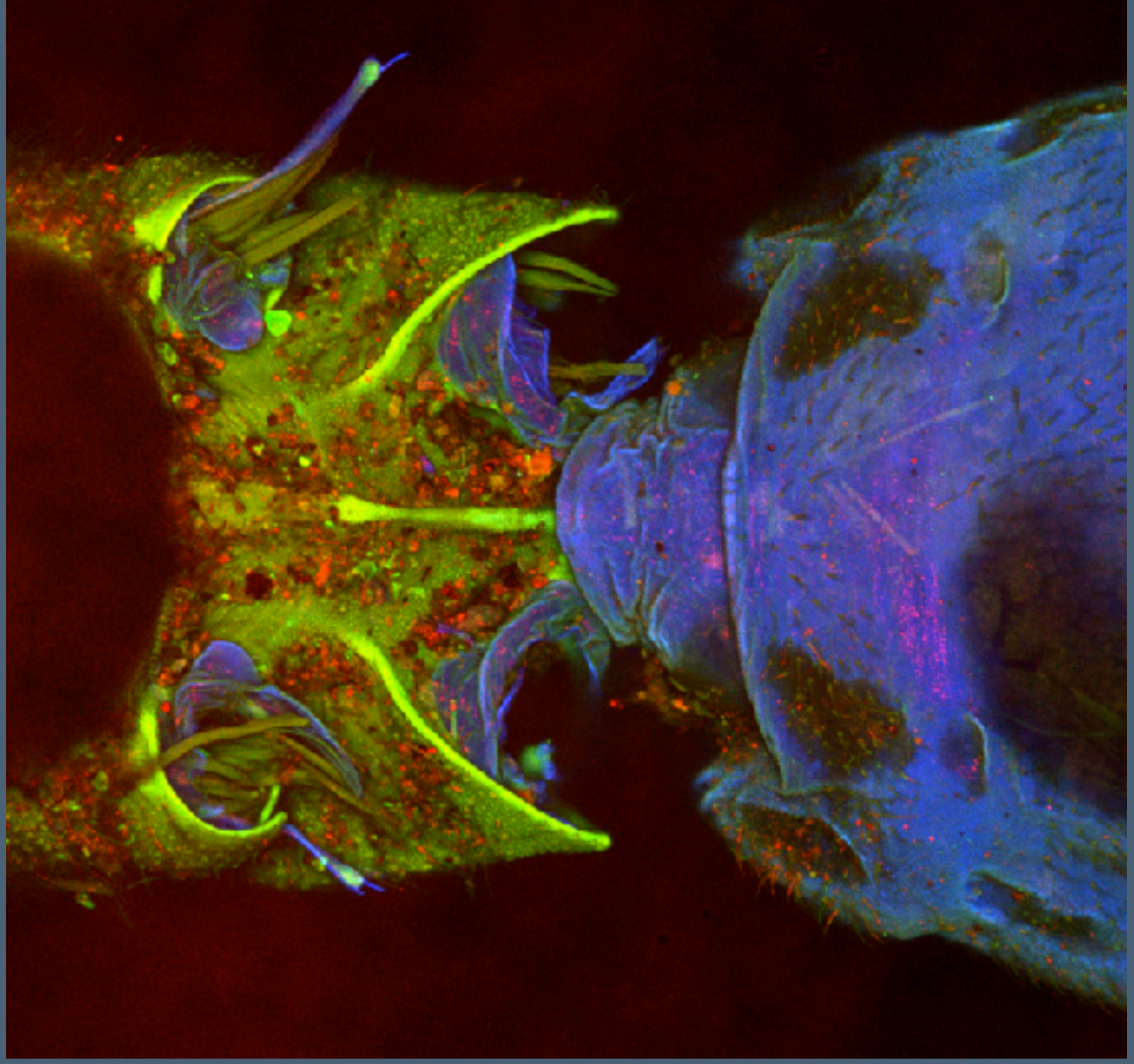
PMT – Photon Multiplier Tube



PMT – Photon Multiplier Tube

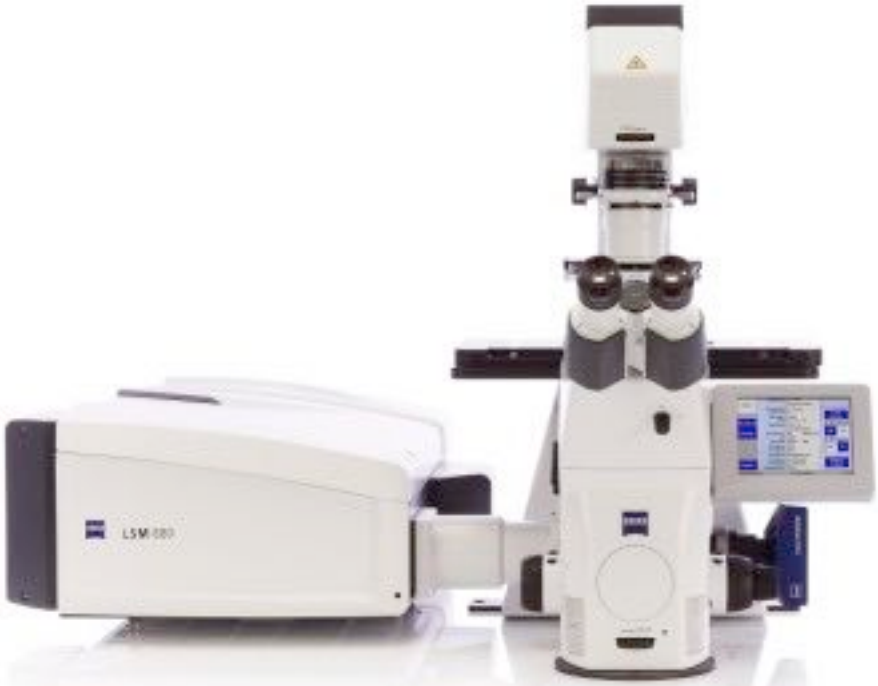


Very Low Noise
Huge Signal Amplification



insect autofluorescence

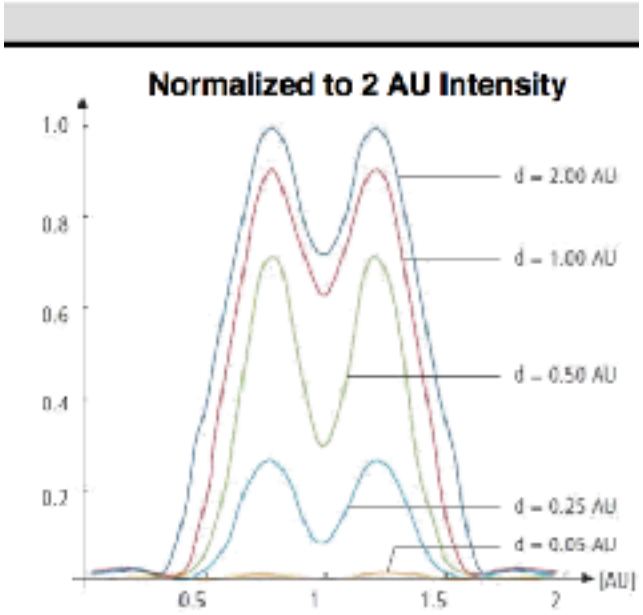
‘Airy-Scan’ technology



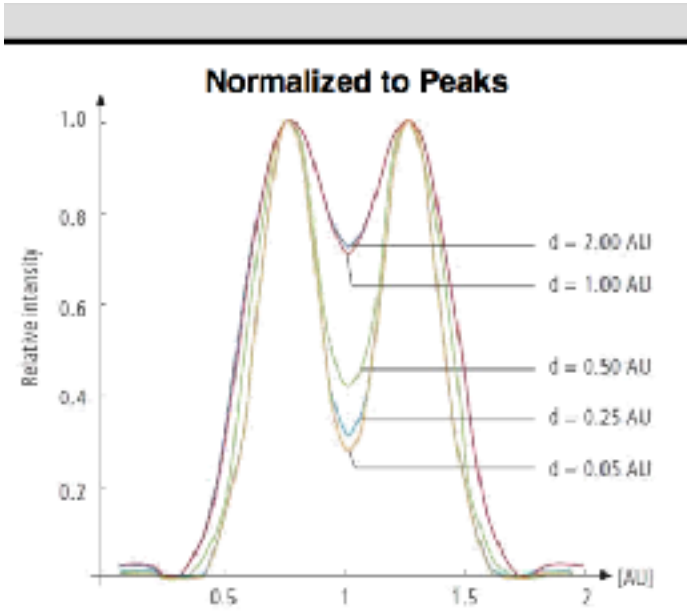
THEORY

‘Airy-Scan’ technology

Small Pinhole, signal loss but resolution gain..



However, constricting the pinhole actually yields a drastic reduction in signal below 1 AU

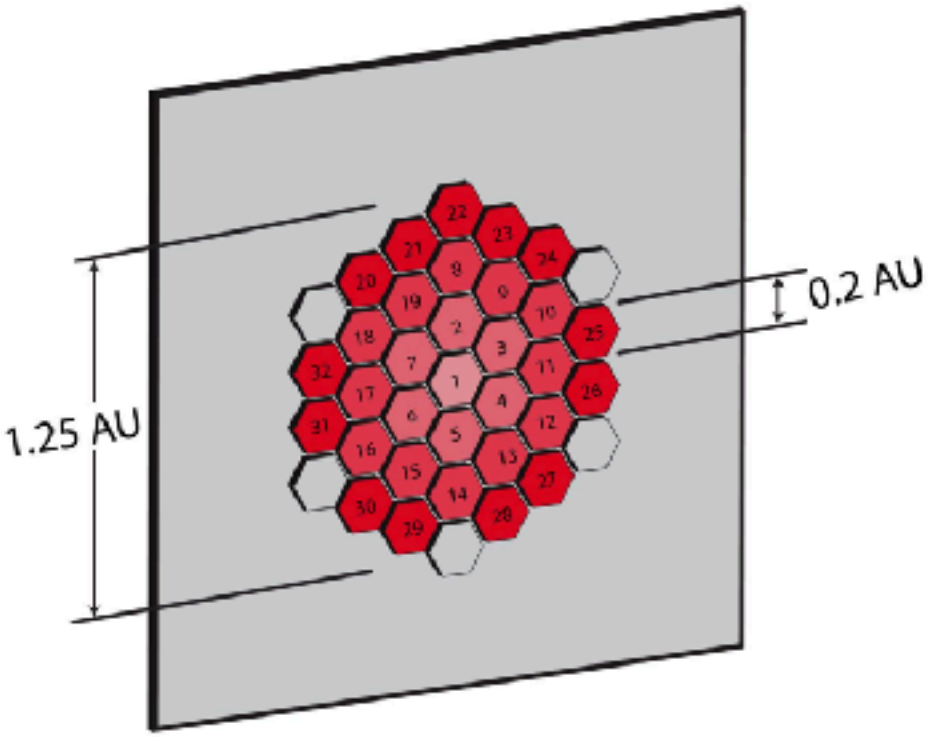


Small pinhole diameters lead to improved resolution steadily until about 0.2 AU, results in deeper dips between two objects

THEORY

‘Airy-Scan’ technology

let through all the emitted light
capture 0.2AU on each detector

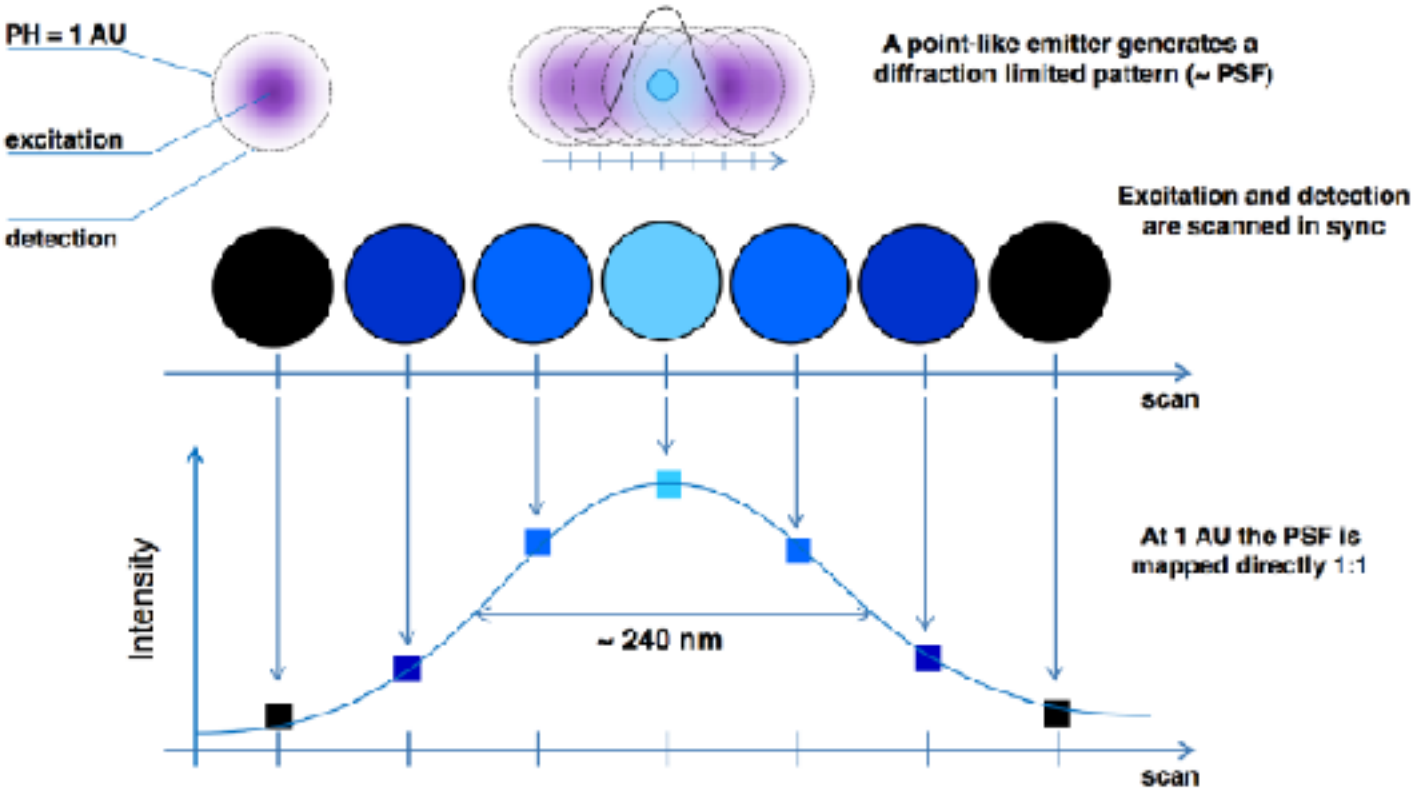


- 32 GaAsP detectors in hexagonal lattice
- Each detector approximately 0.2 AU in diameter
- Total detection area approximately 1.25 AU in diameter
- **Simultaneous improvement in resolution and signal**

THEORY

‘Airy-Scan’ technology

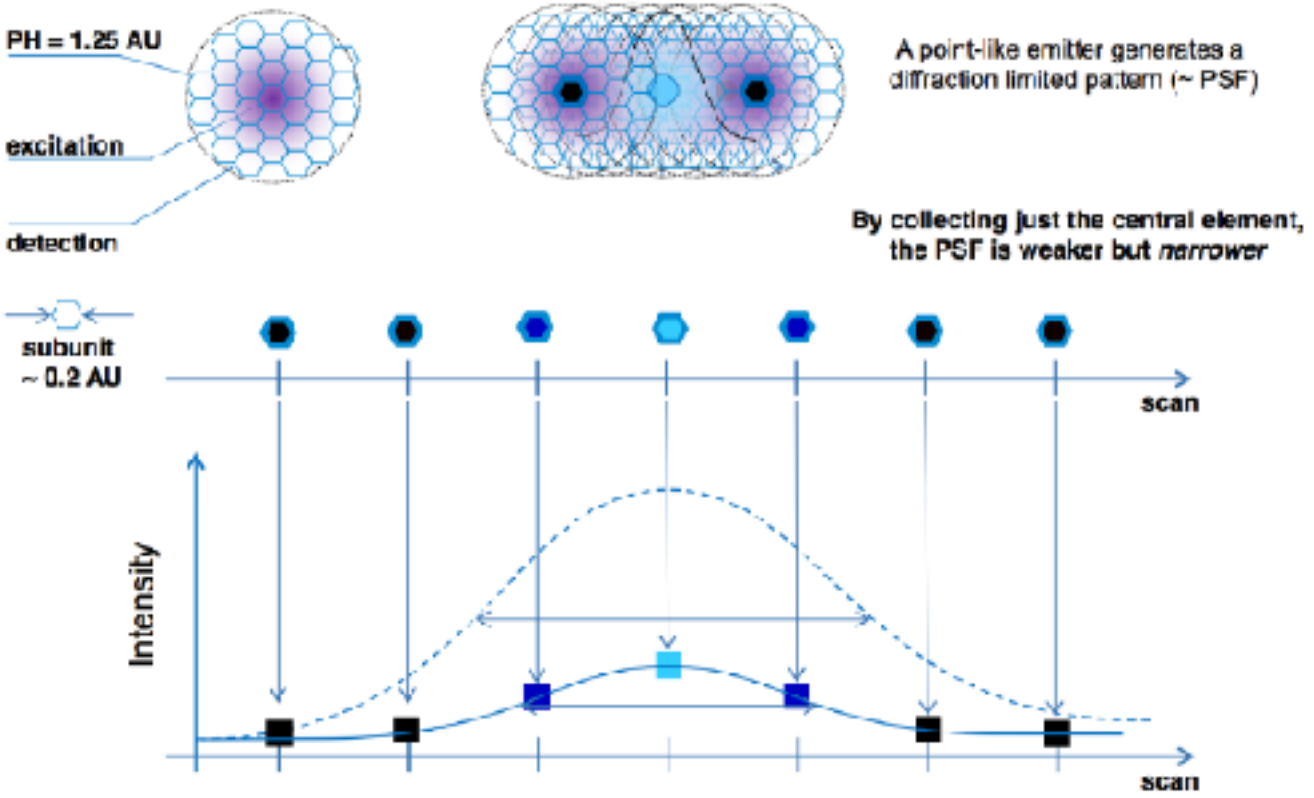
point of light scanned with 1AU ‘standard’ detector



THEORY

‘Airy-Scan’ technology

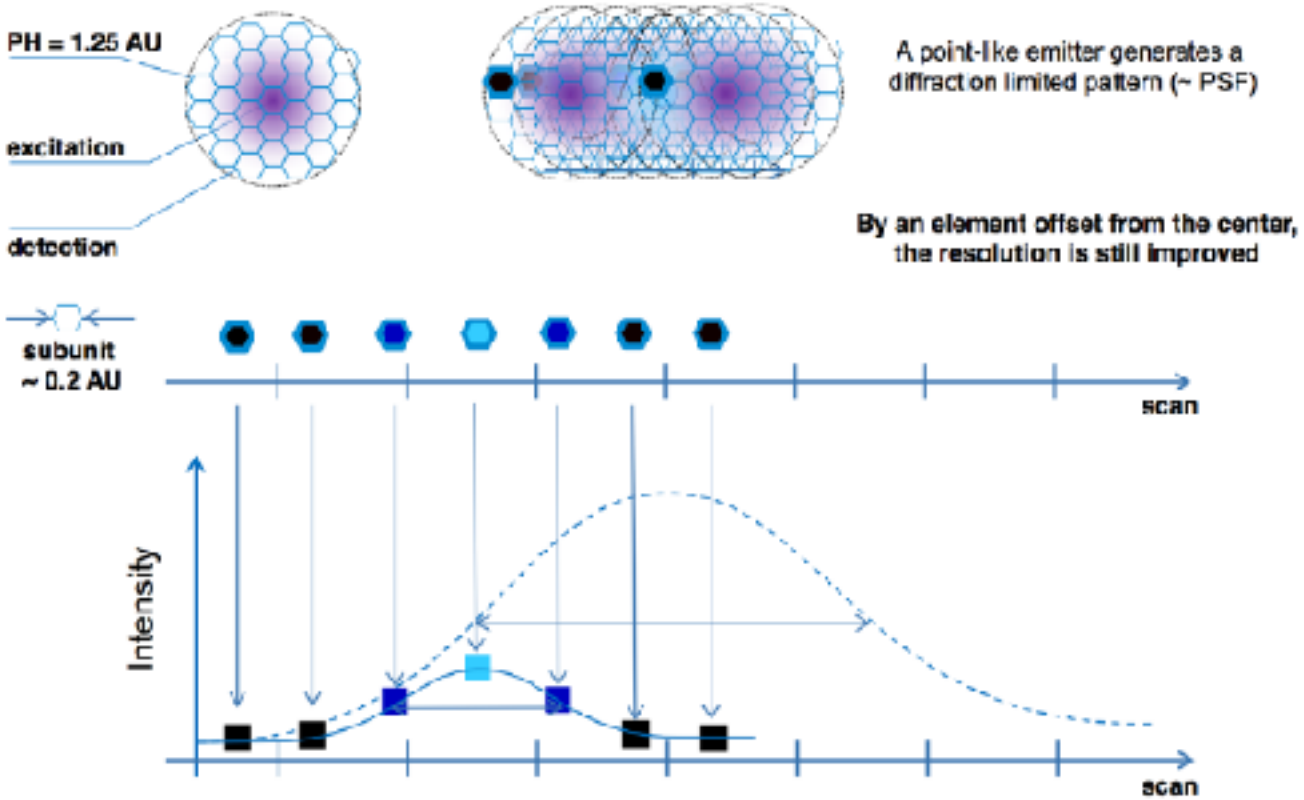
point of light scanned with 0.2AU ‘Airyscan’ detector
>increased resolution



THEORY

‘Airy-Scan’ technology

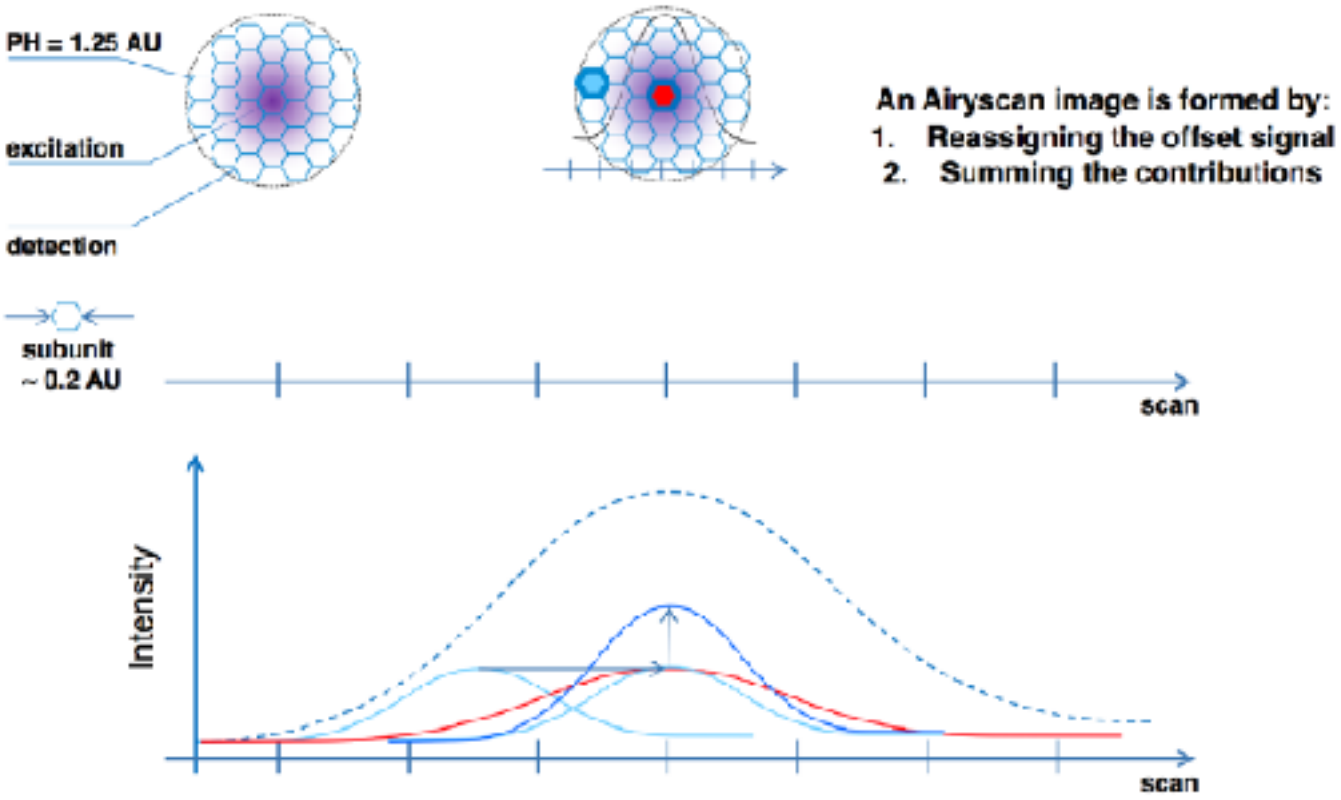
each 0.2AU ‘Airyscan’ detector provides
>increased resolution



THEORY

‘Airy-Scan’ technology

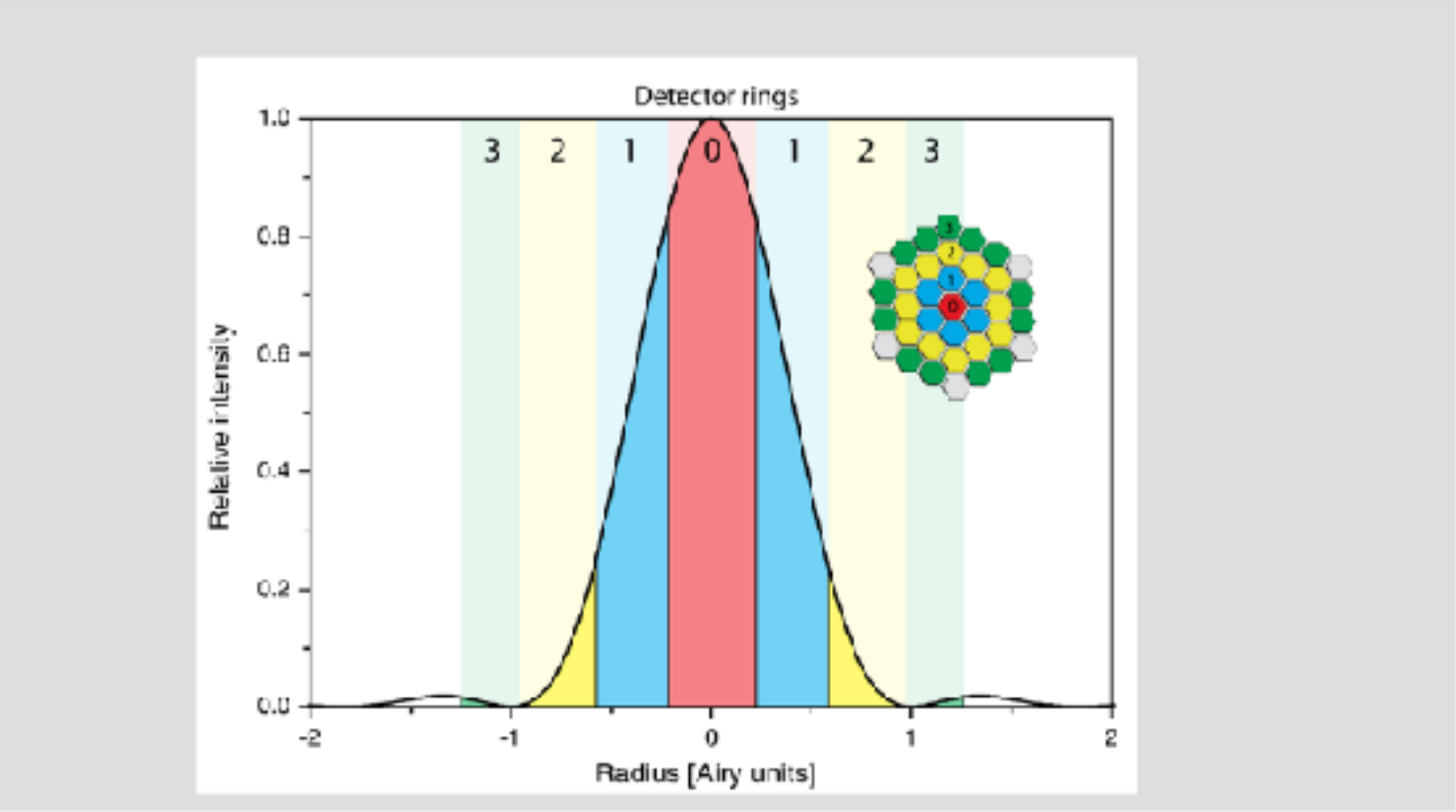
each 0.2AU ‘Airyscan’ detector info
is reassigned and summed

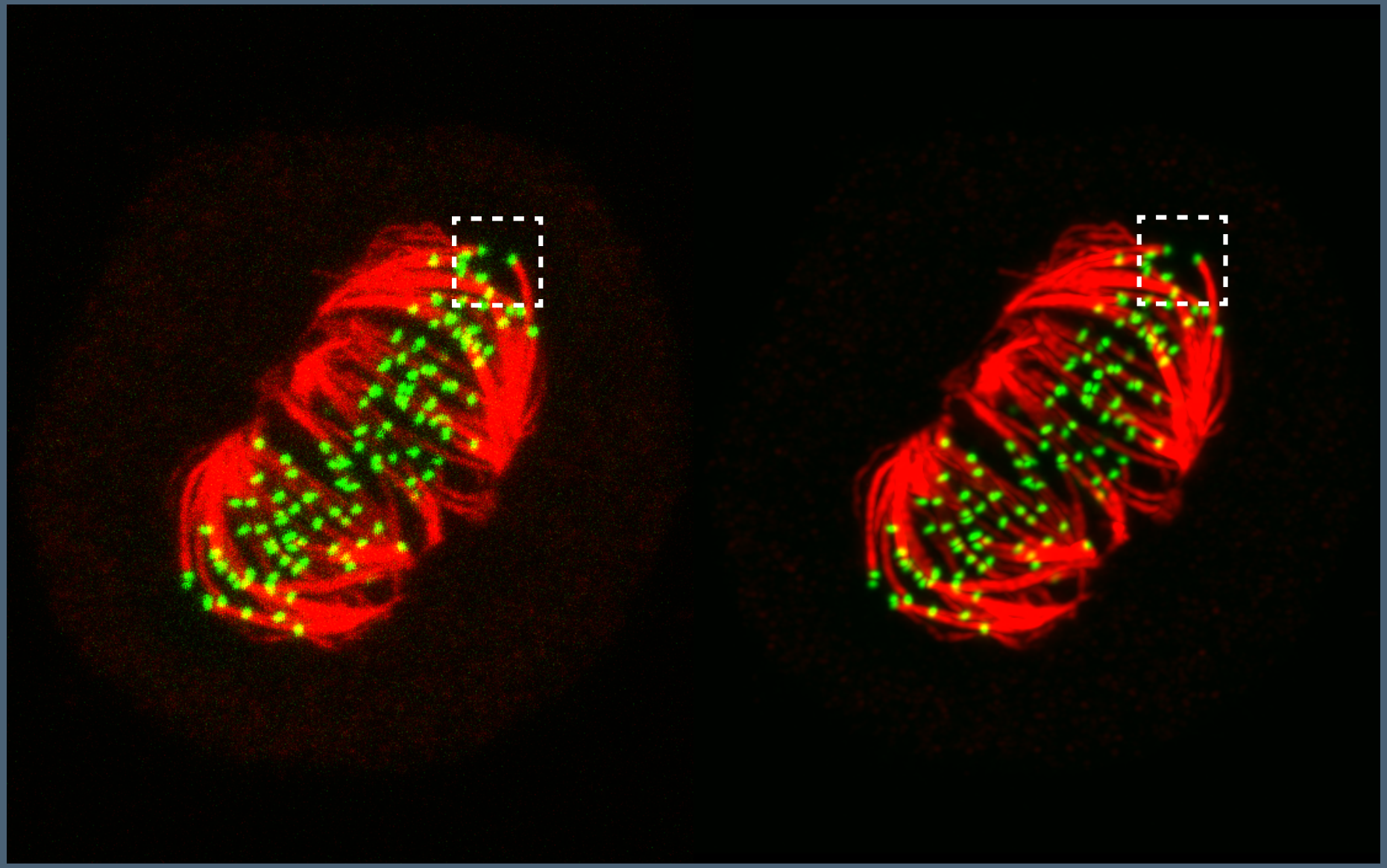


THEORY

‘Airy-Scan’ technology

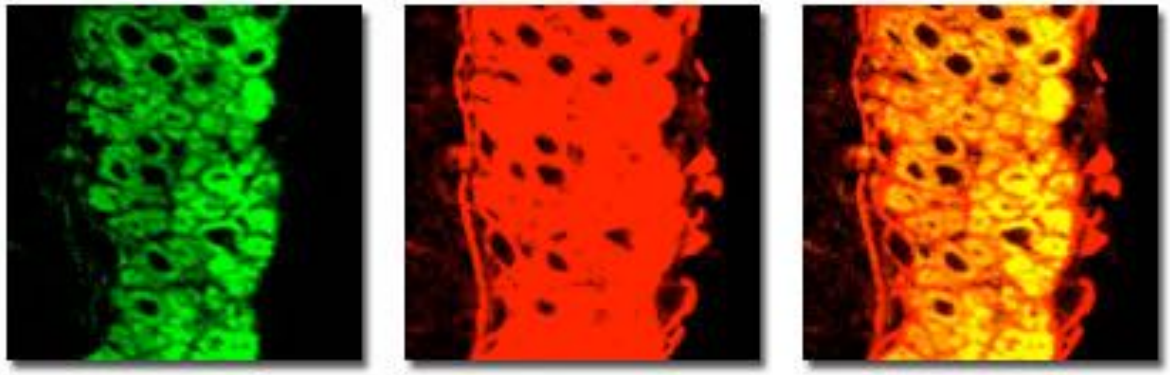
effective PSF is now smaller.. > increased resolution (1.4x - 1.7x)



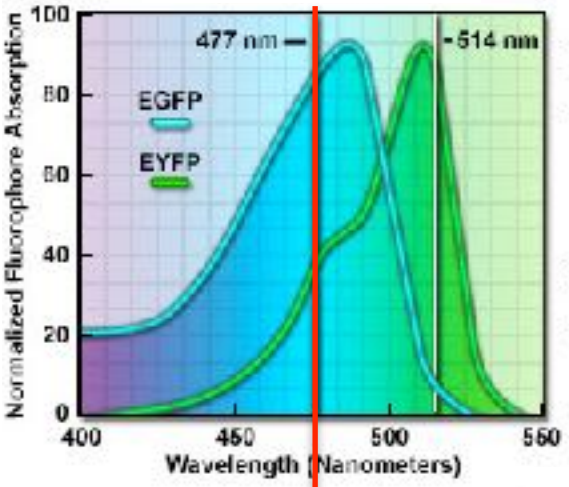


kinetochores (James Banecroft, Gruneberg Lab)

‘bleed-through’

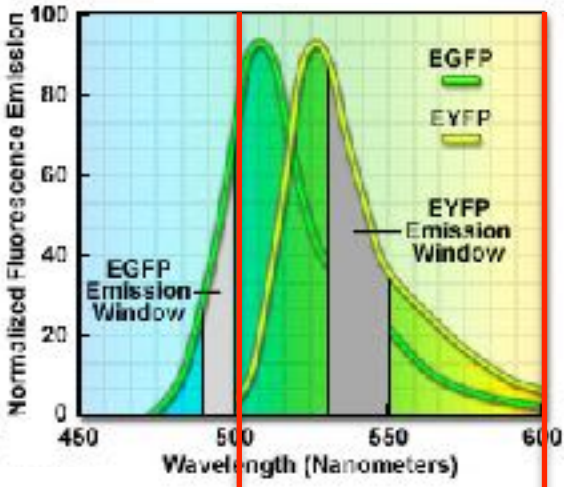


Absorption spectral profiles



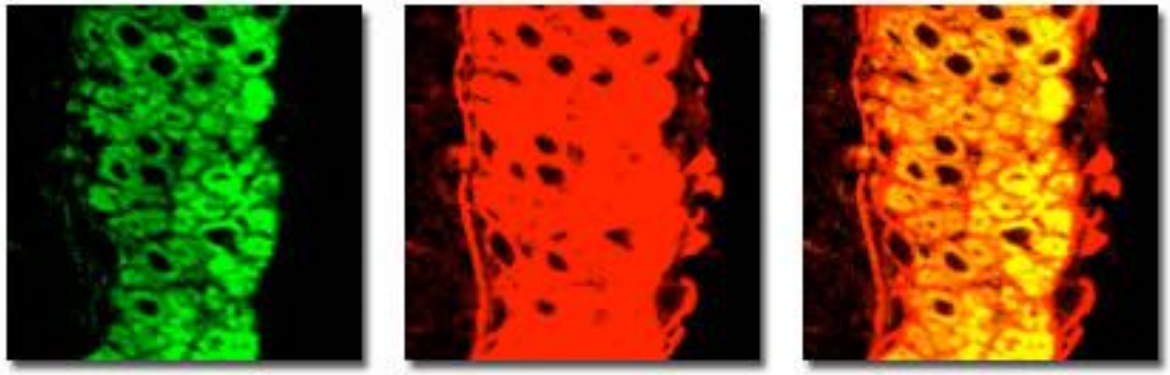
Excite at 477nm

Emission spectral profiles

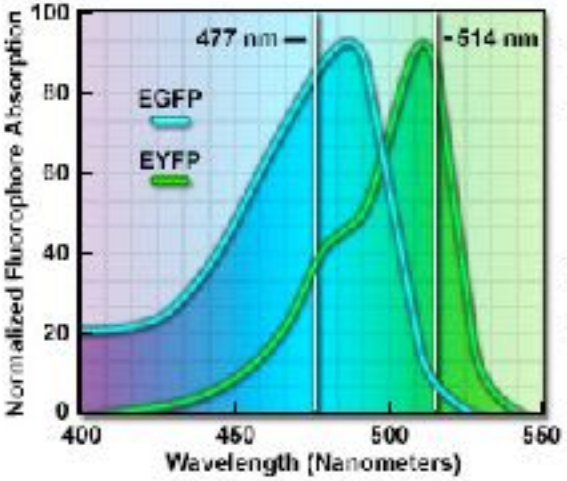


overlapping emission

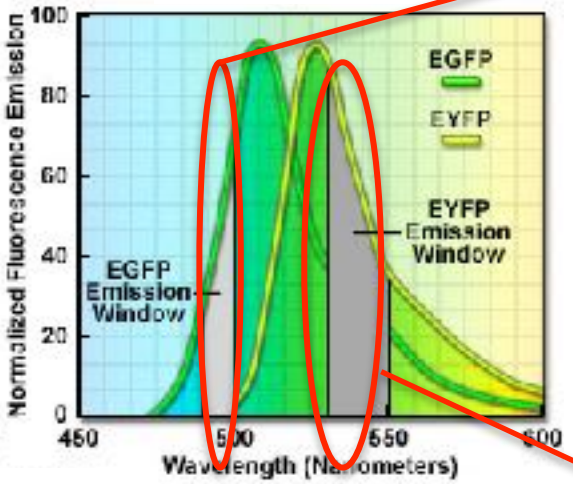
minimising‘bleed-through’ Variable Slits

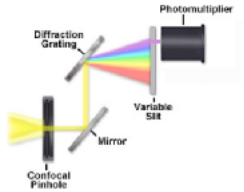


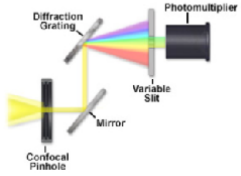
Absorption spectral profiles



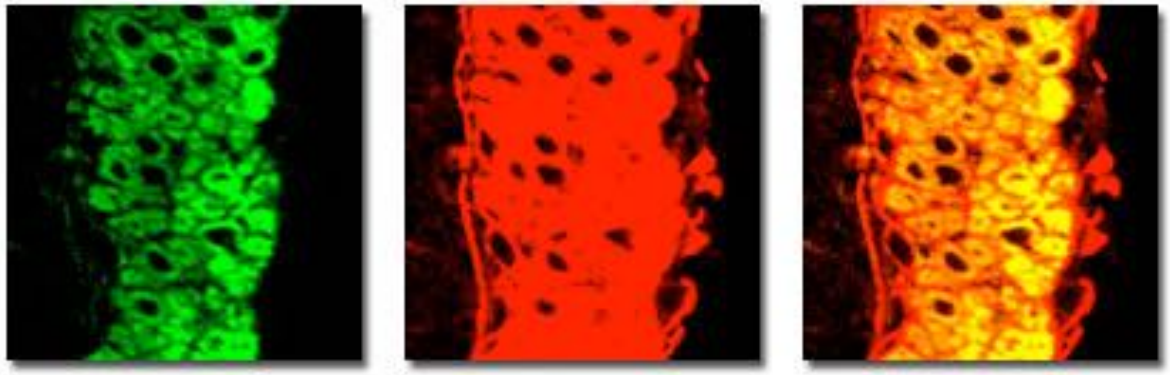
Emission spectral profiles



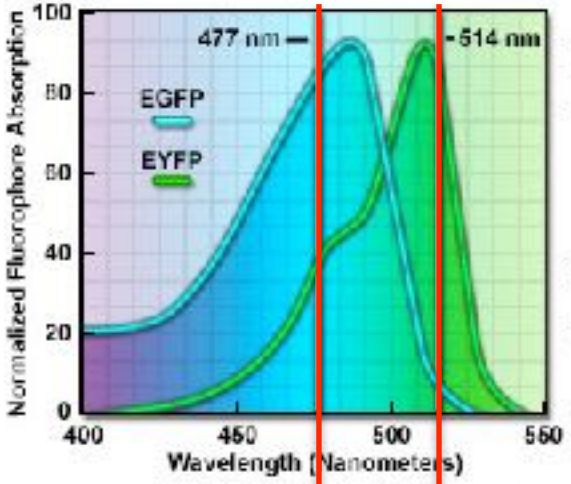




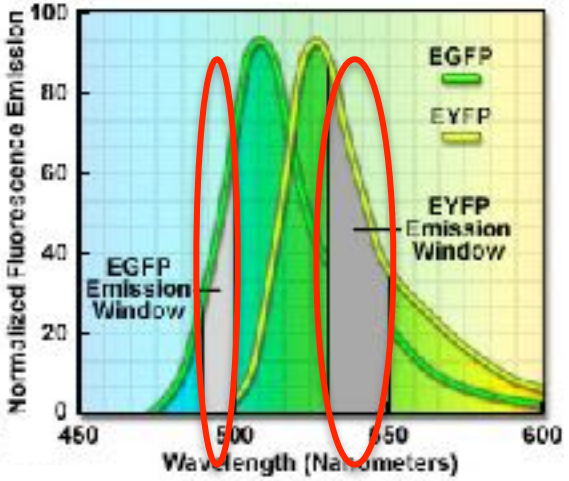
minimising‘bleed-through’ Sequential Scanning



Absorption spectral profiles



Emission spectral profiles

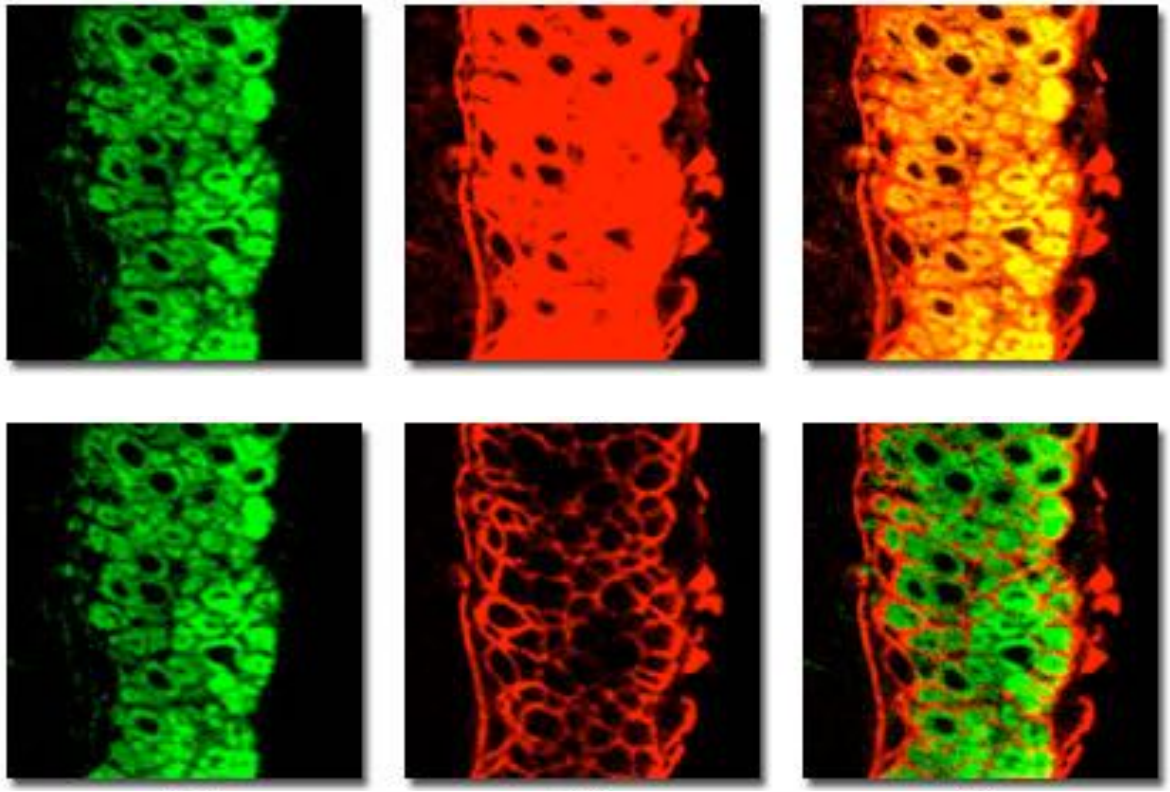


Excite at 477nm

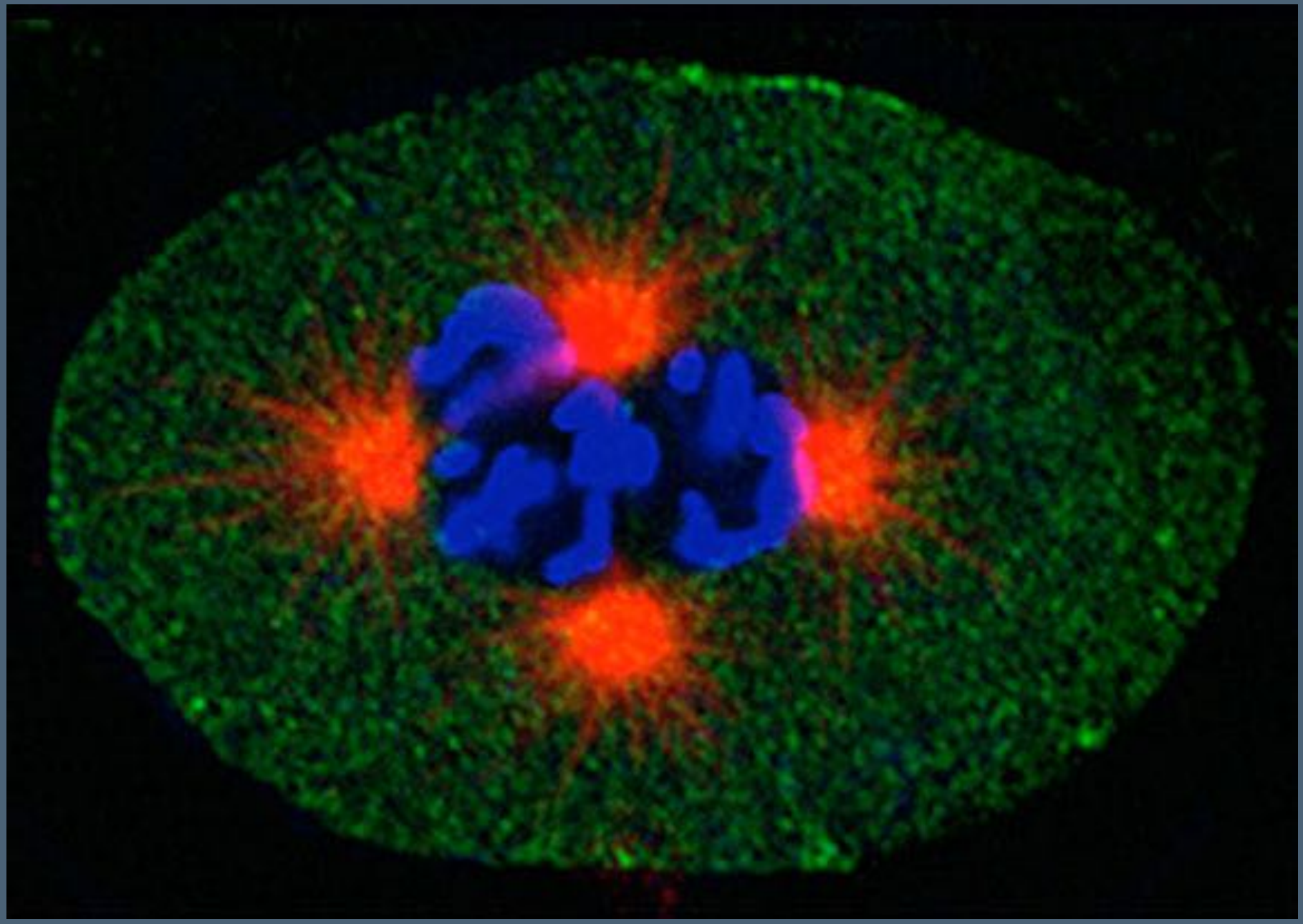
Excite at 514nm

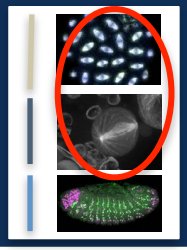
Temporal separation

minimising ‘bleed-through’

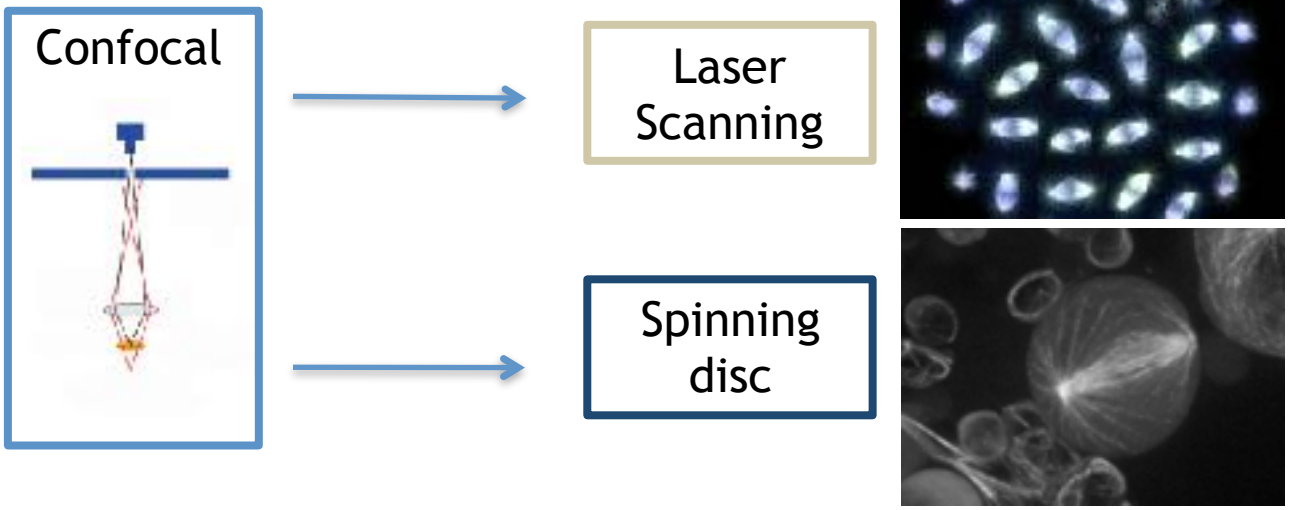


Adjust detector slit widths
Use sequential scanning

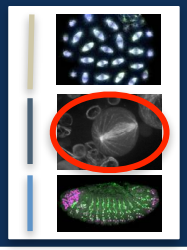




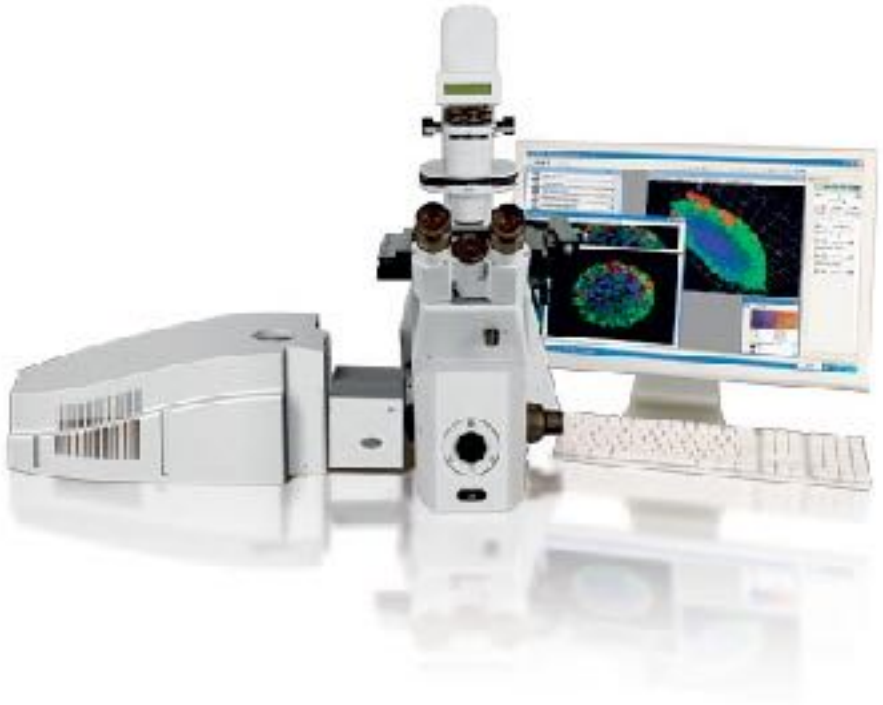
Confocal Microscopes



Both are confocals

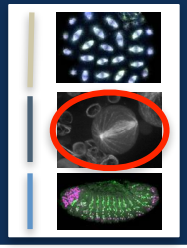


Spinning Disc Confocal

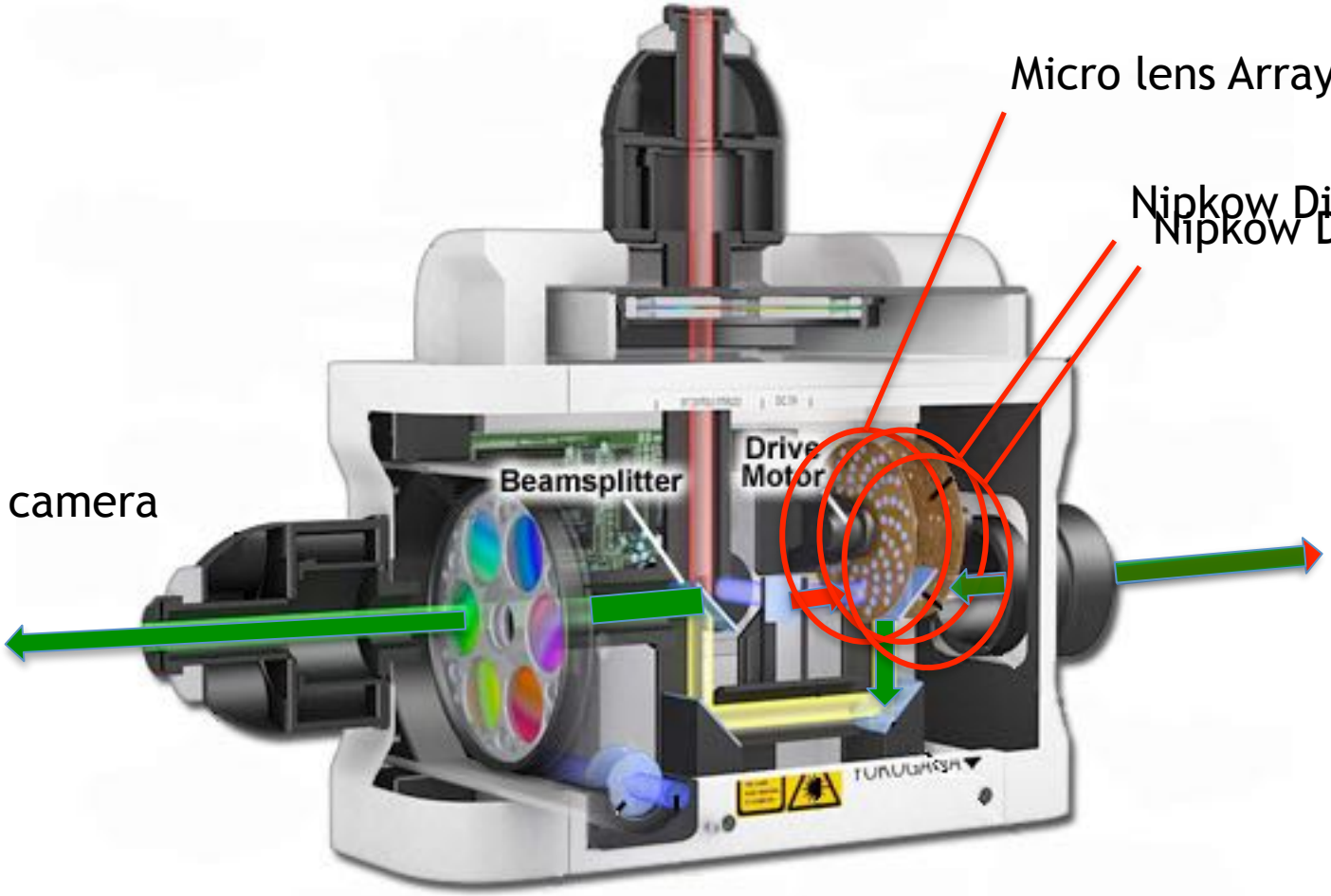


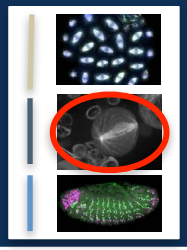
Great for live cell imaging

Can collect many images per second

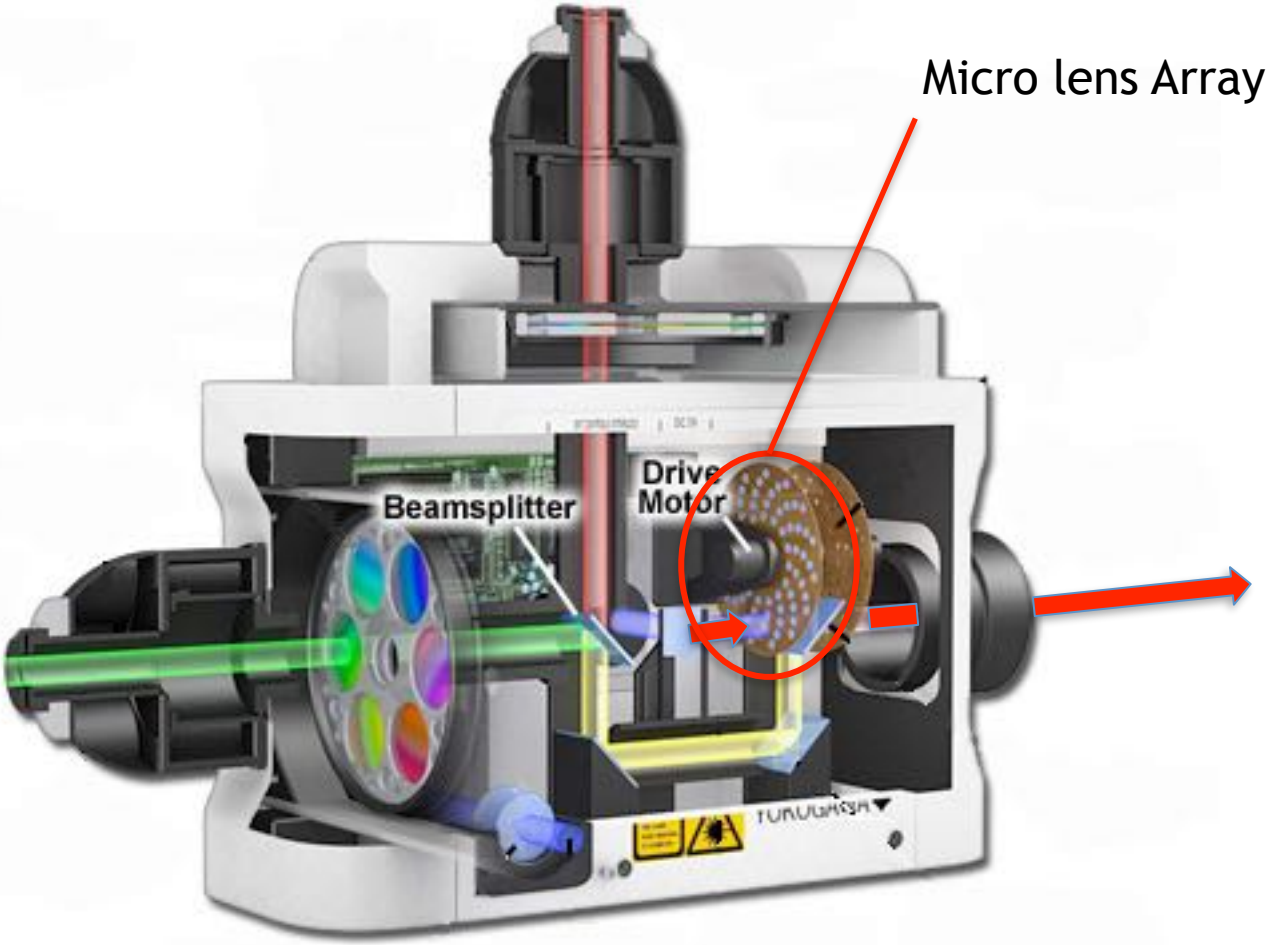


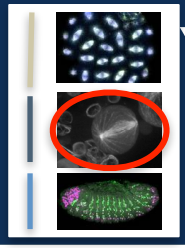
Yokogawa CSU-X1 Spinning Disc



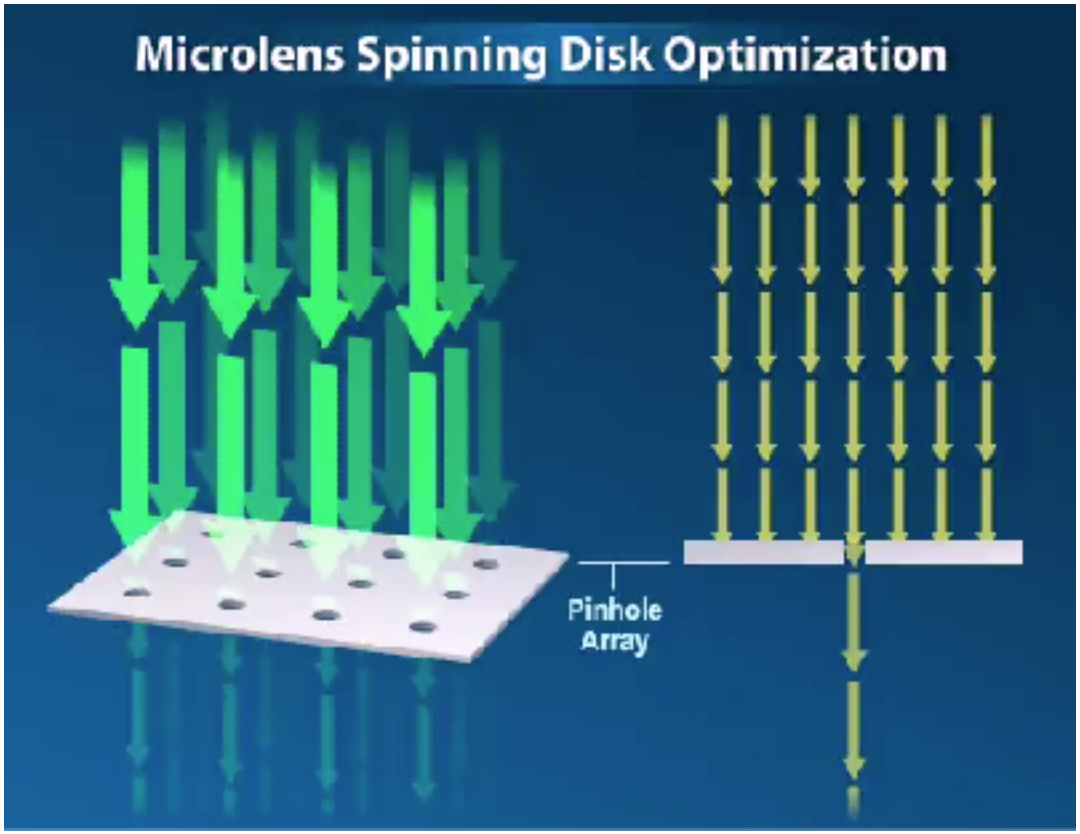


Yokogawa CSU-X1 Spinning Disc





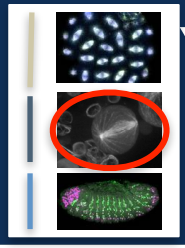
Yokogawa Spinning Disc Confocal Microlenses



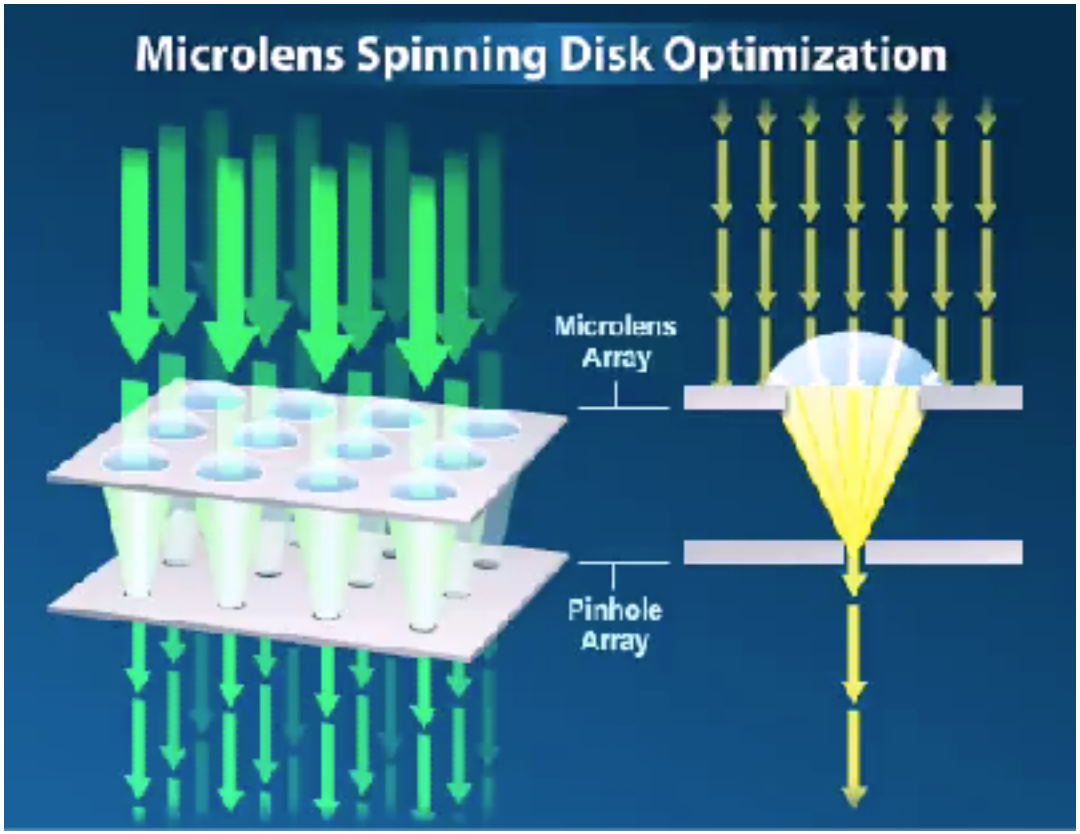
just a pinhole array –
Optimised for ‘cofocality’
and ‘crosstalk’

too much light is blocked
from reaching the specimen

Only 4% light passes through disc

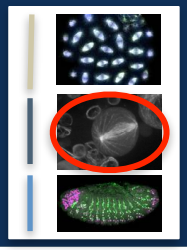


Yokogawa Spinning Disc Confocal Microlenses

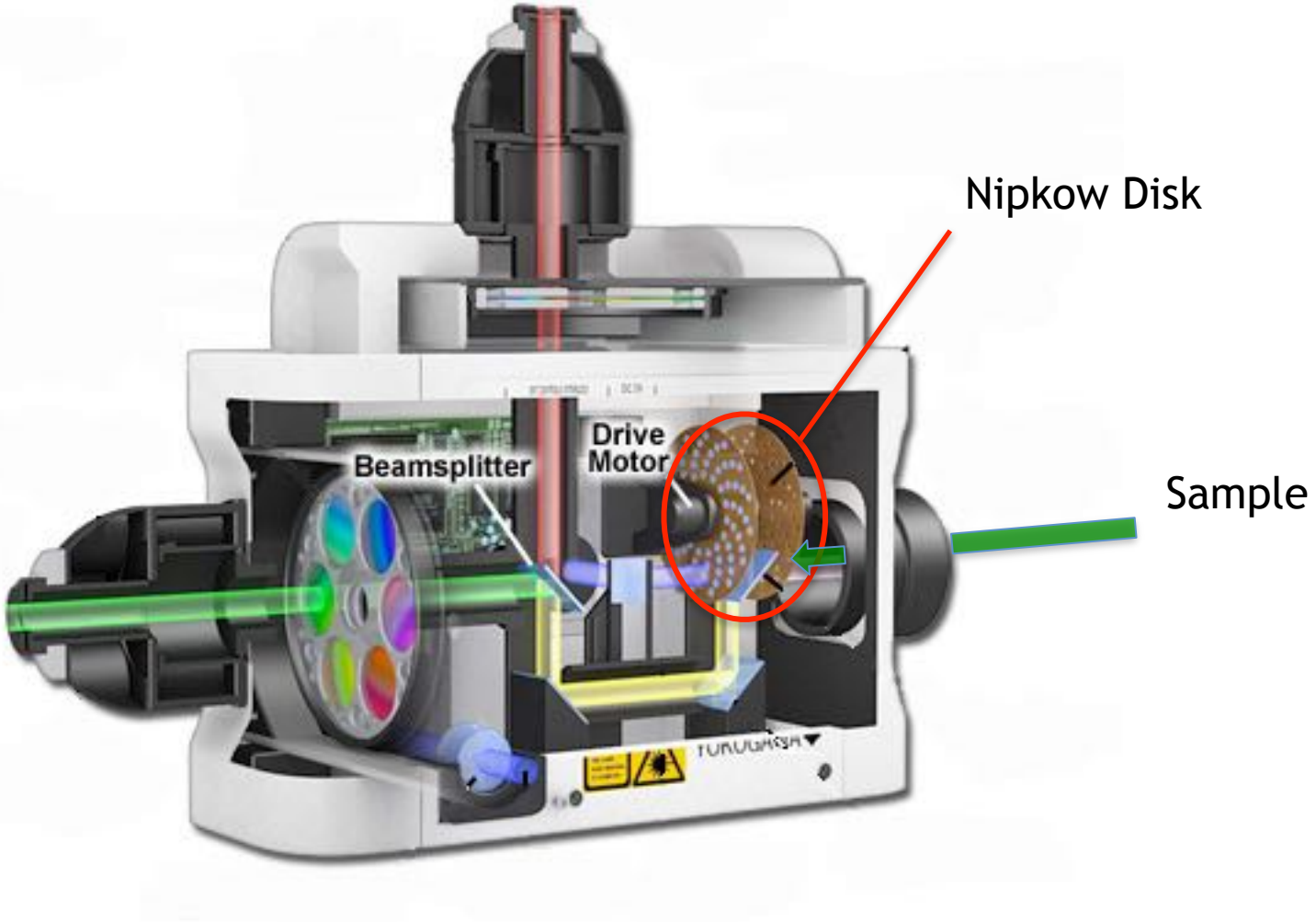


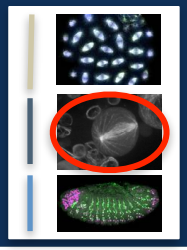
micro-lens array increase the light reaching the specimen

Typically 56% light passes through disc

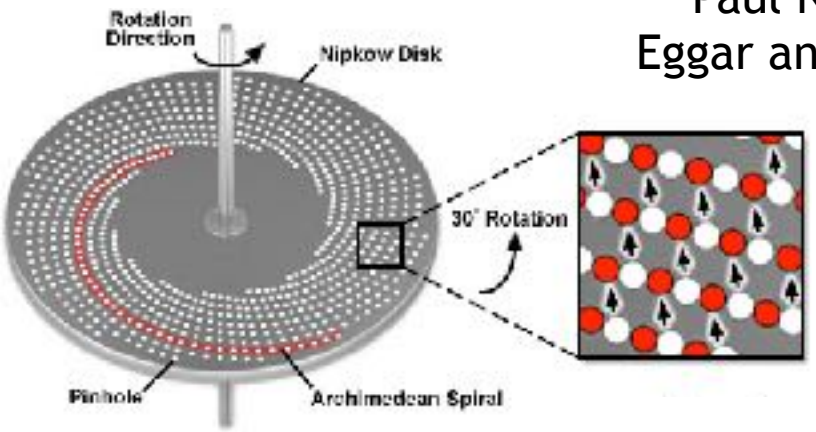


Yokogawa CSU-X1 Spinning Disc

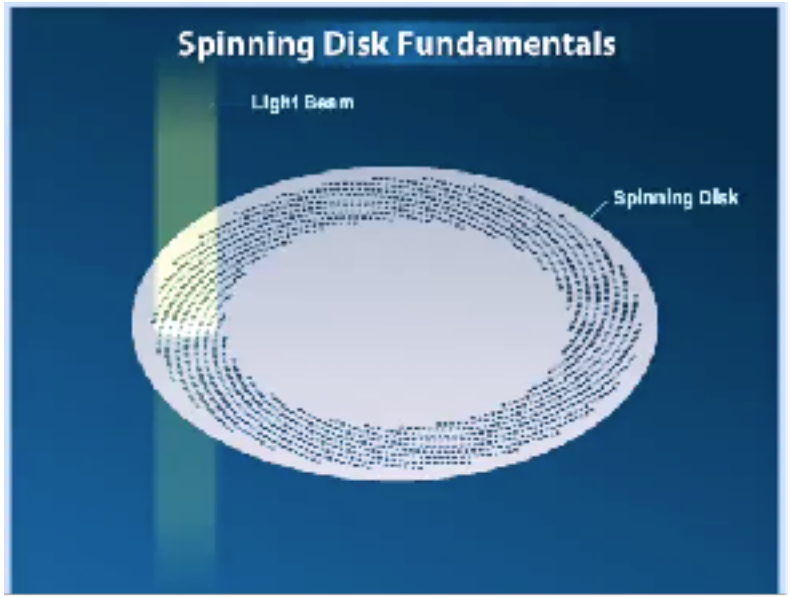




The Nipkow Disk

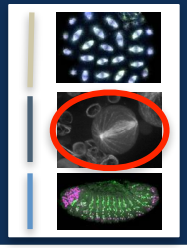


Paul Nipkow, 1884
Eggar and Petran, 1967

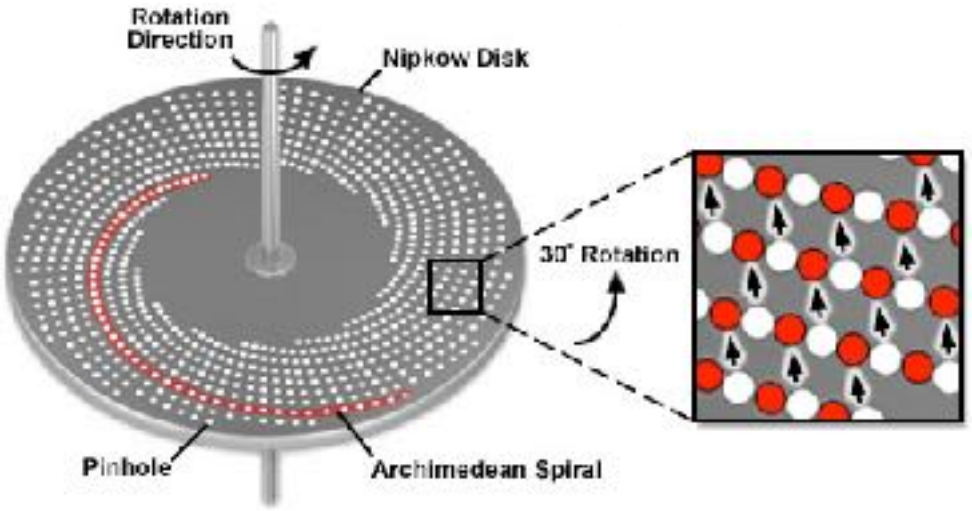


Approx. 1000 pinholes

Single frame created with each 30-degree of rotation of disc (12 frames per rotation)

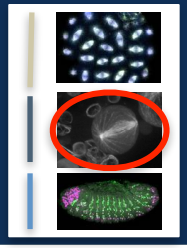


The Nipkow Disk

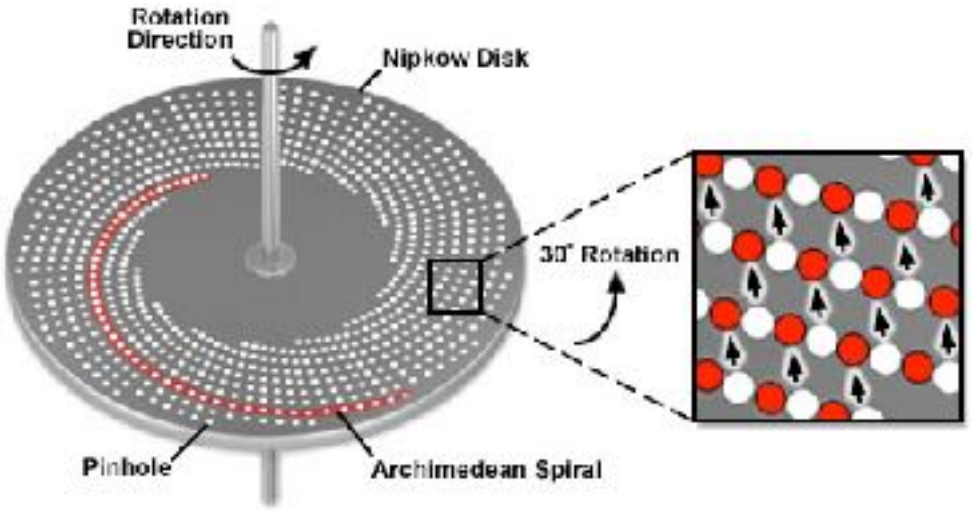


Larger pinholes - brighter image, but less “confocal”

Pinholes fixed size: Typically = 50um

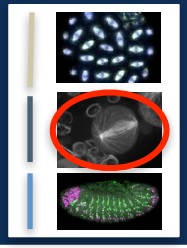


The Nipkow Disk

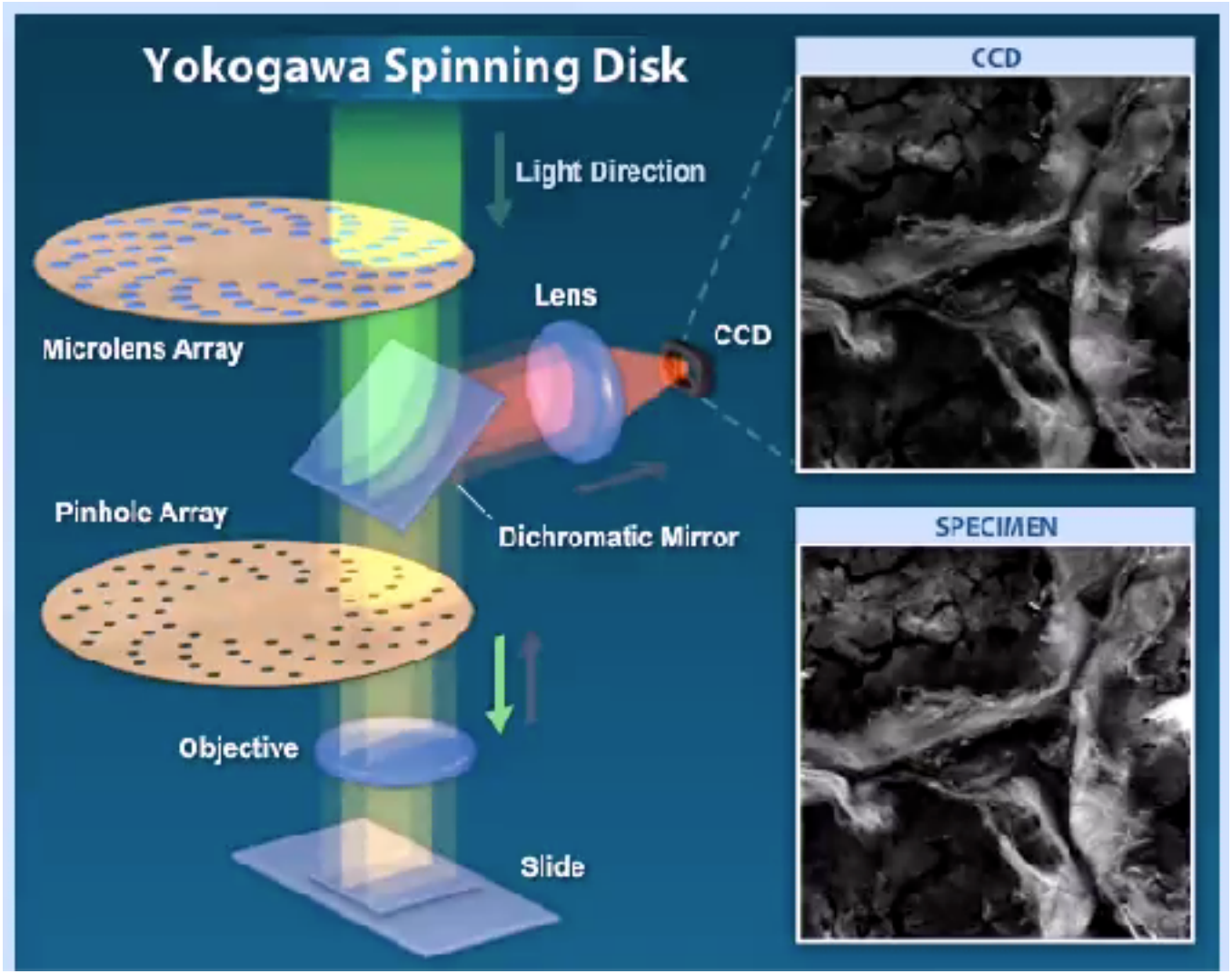


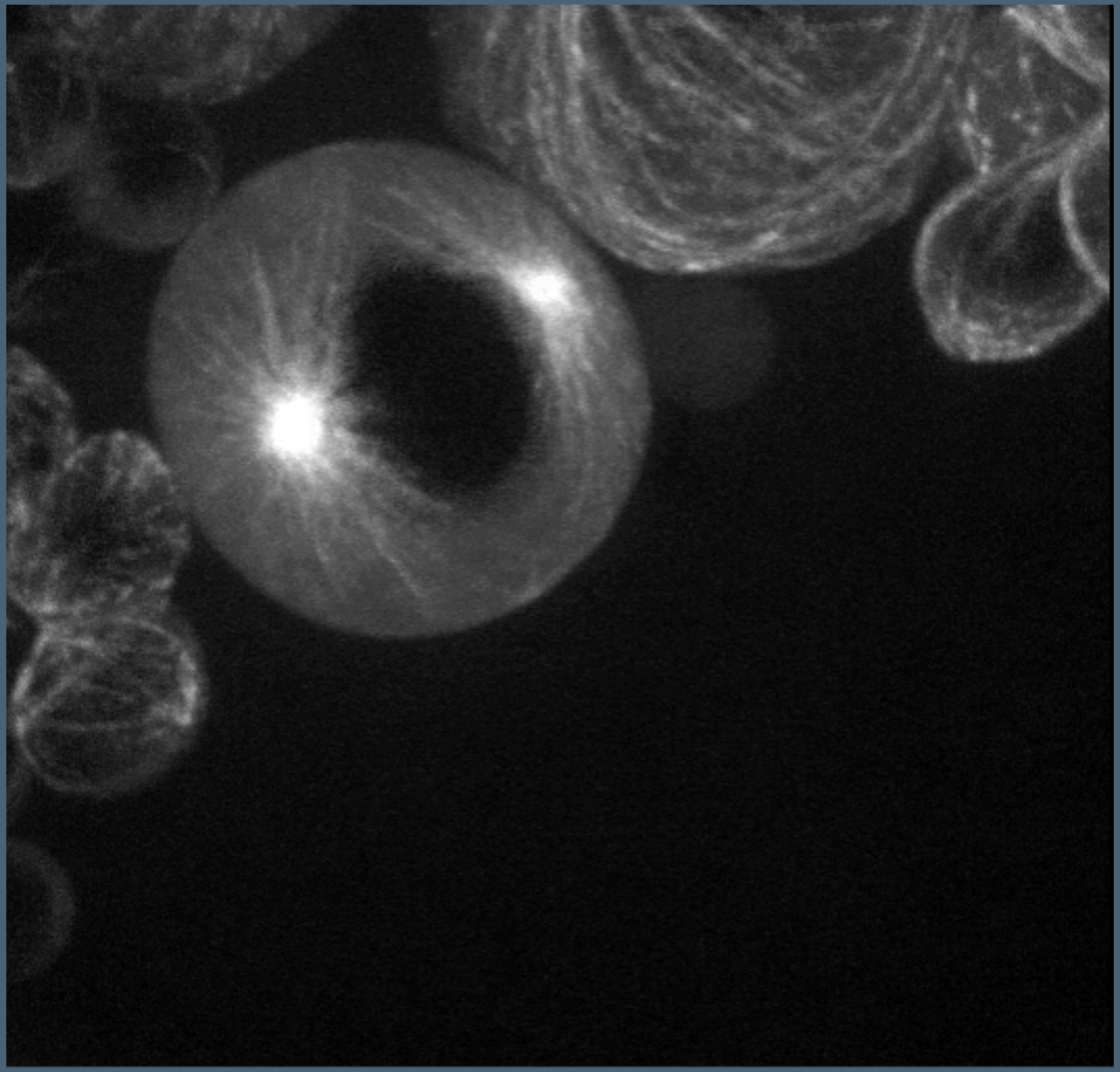
Constant Battle:
Smaller spacing - more light gets through, but "crosstalk"

Pinhole Spacing Typically = 2.5um apart

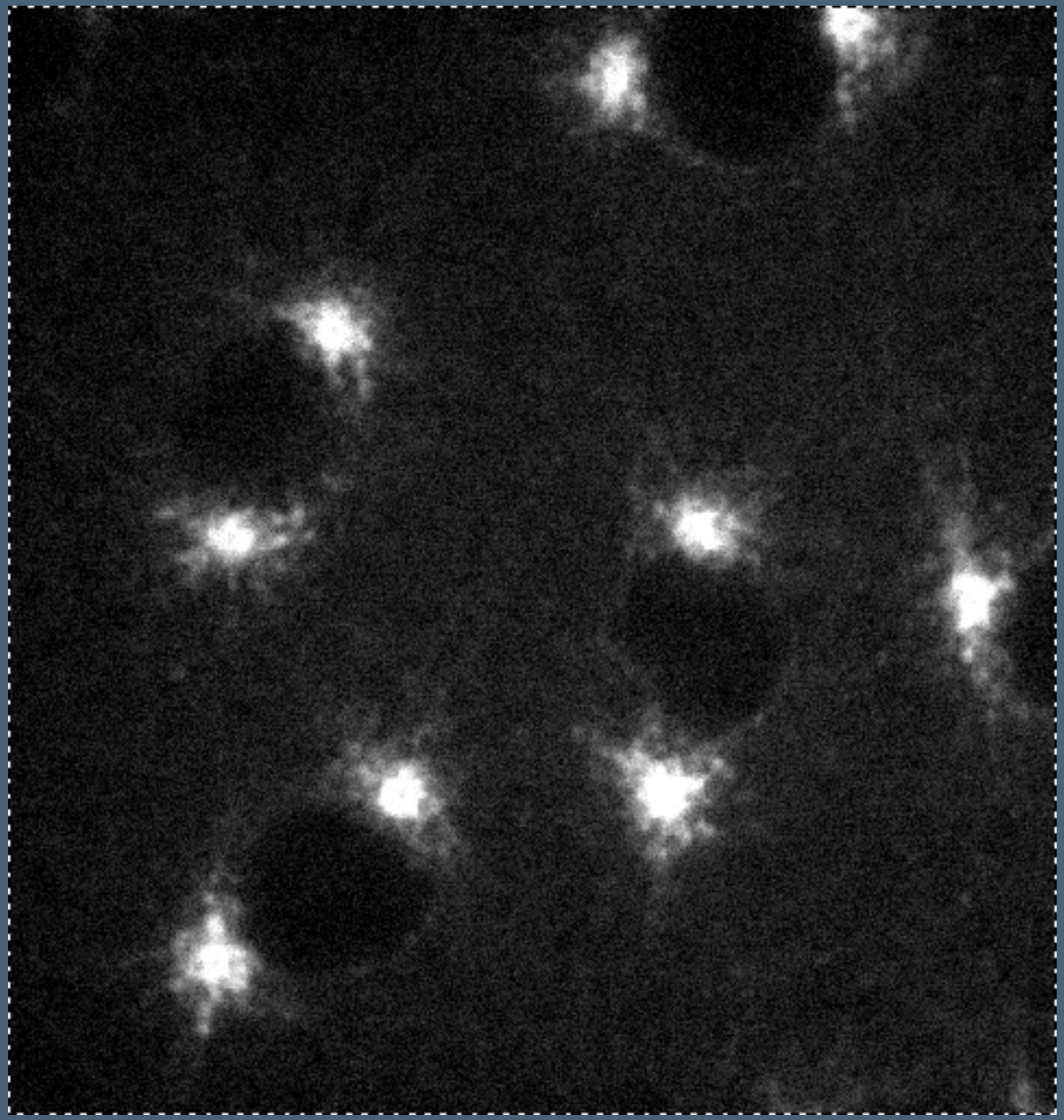


Yokogawa Spinning Disc Confocal

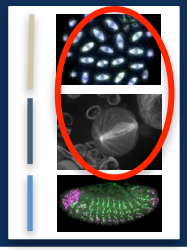




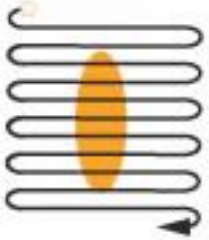
Cell division in brain stem cells (neuroblasts), Raf

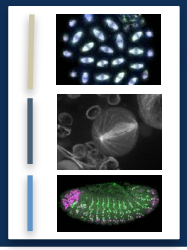


MT binding protein in *Drosophila* embryo, Raff L



Point Scanning Vs Spinning Disc

	Point Scanning	Spinning Disc
		
Speed	Slow (secs)	Fast (msecs)
Sensitivity	OK	OK
Flexibility	Good	Poor
Bleaching	Poor	Good
Pretty Pictures	Unbeatable!	Pretty damn good!
Pretty Movies	Good – if process slow	Unbeatable!

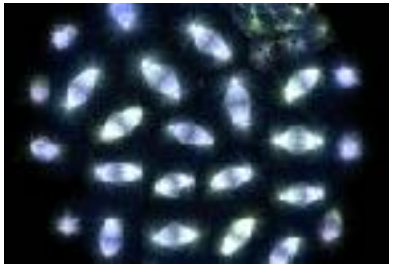


3 Flavours of Microscope

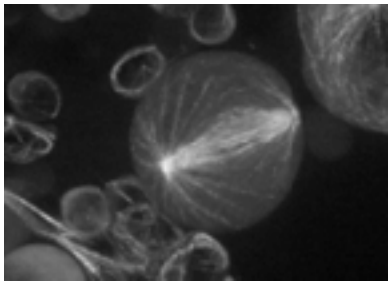
Problem:
Out of Focus
Light

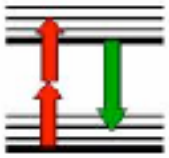
Confocal

Laser
Scanning

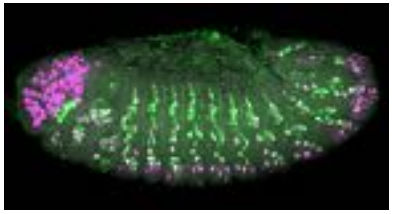


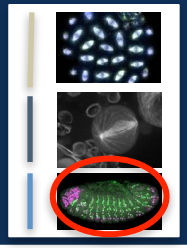
Spinning
disc





2-Photon



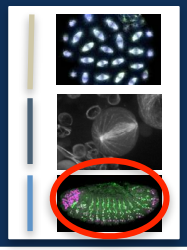


2-photon Microscope

Not a ‘confocal’

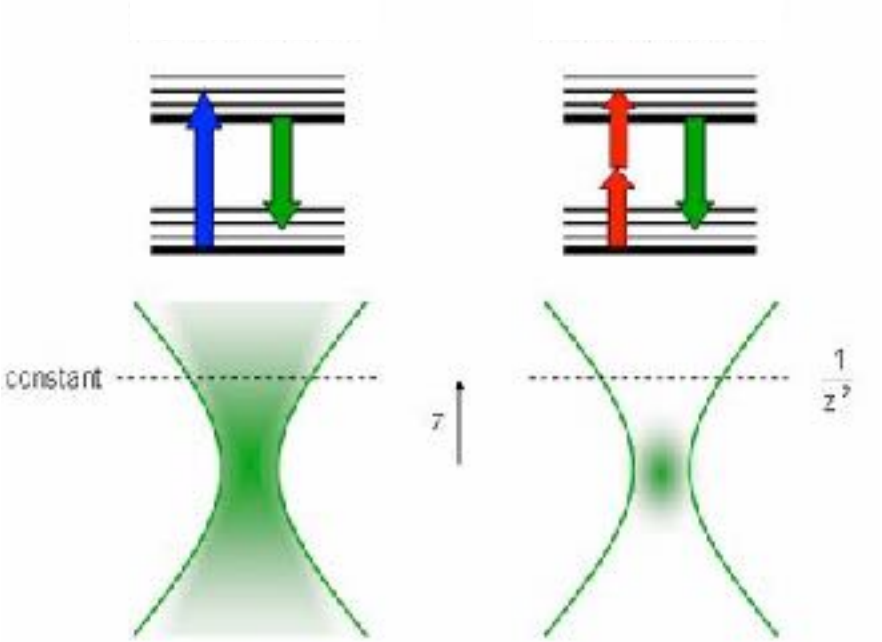
for imaging deeper into thick specimens

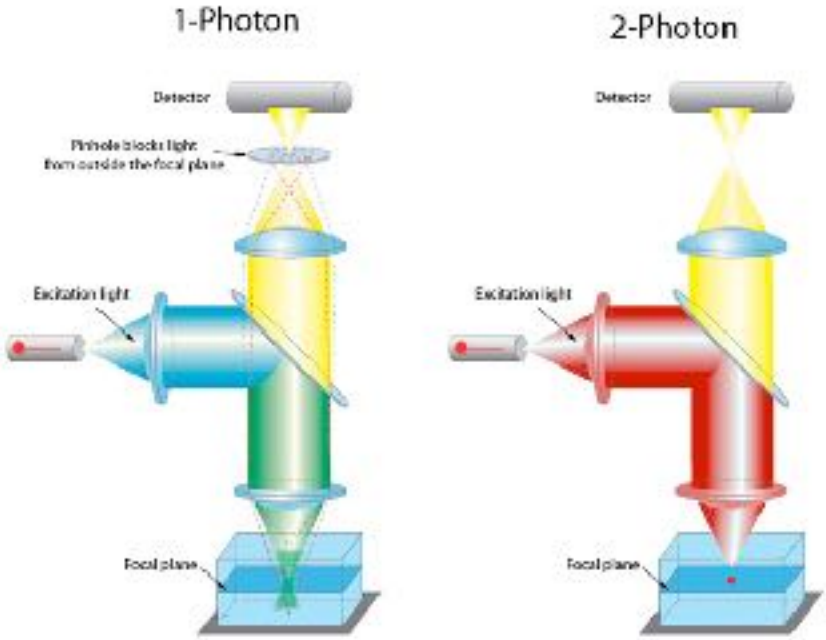
less damaging to biological samples



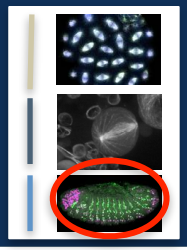
Confocal Vs 2-photon

1 Photon Excitation 2 Photon Excitation



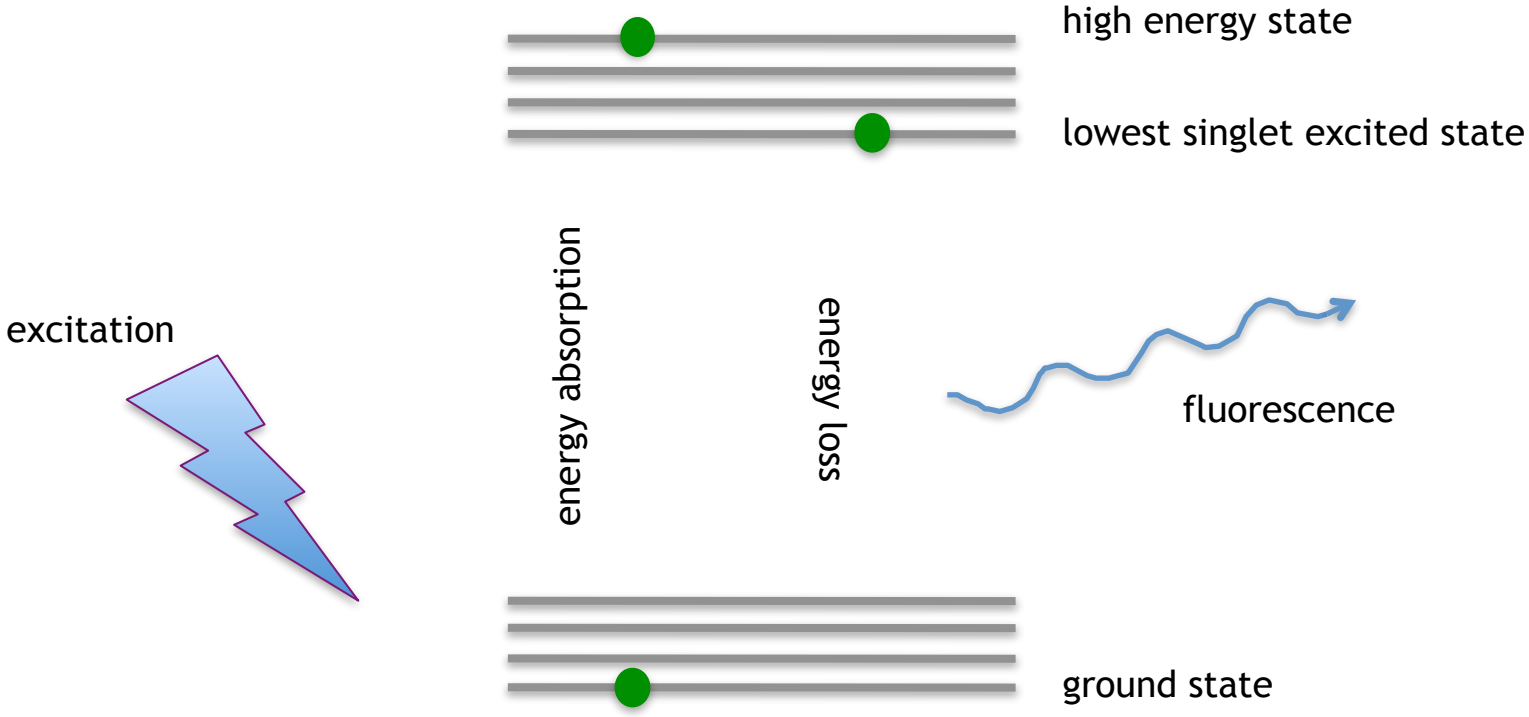


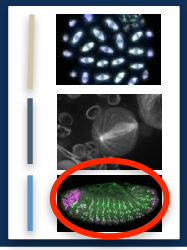
There is no out of focus



THEORY

1 Photon Excitation





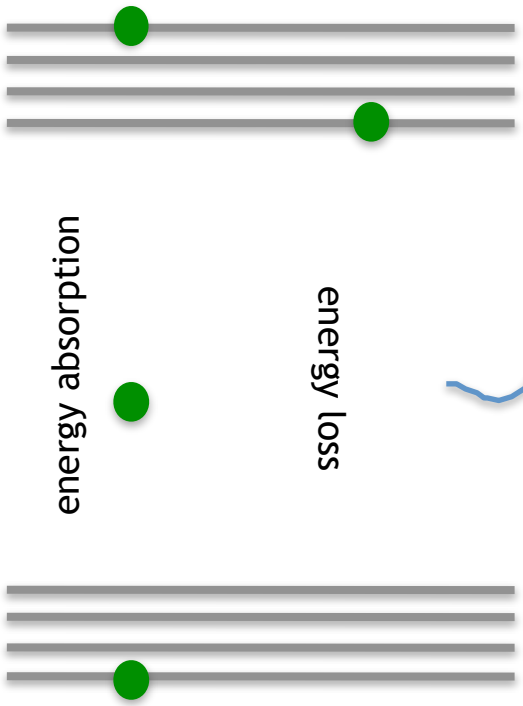
THEORY

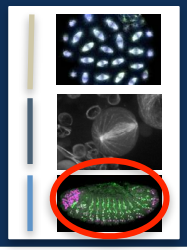
2 Photon Excitation

Almost simultaneous

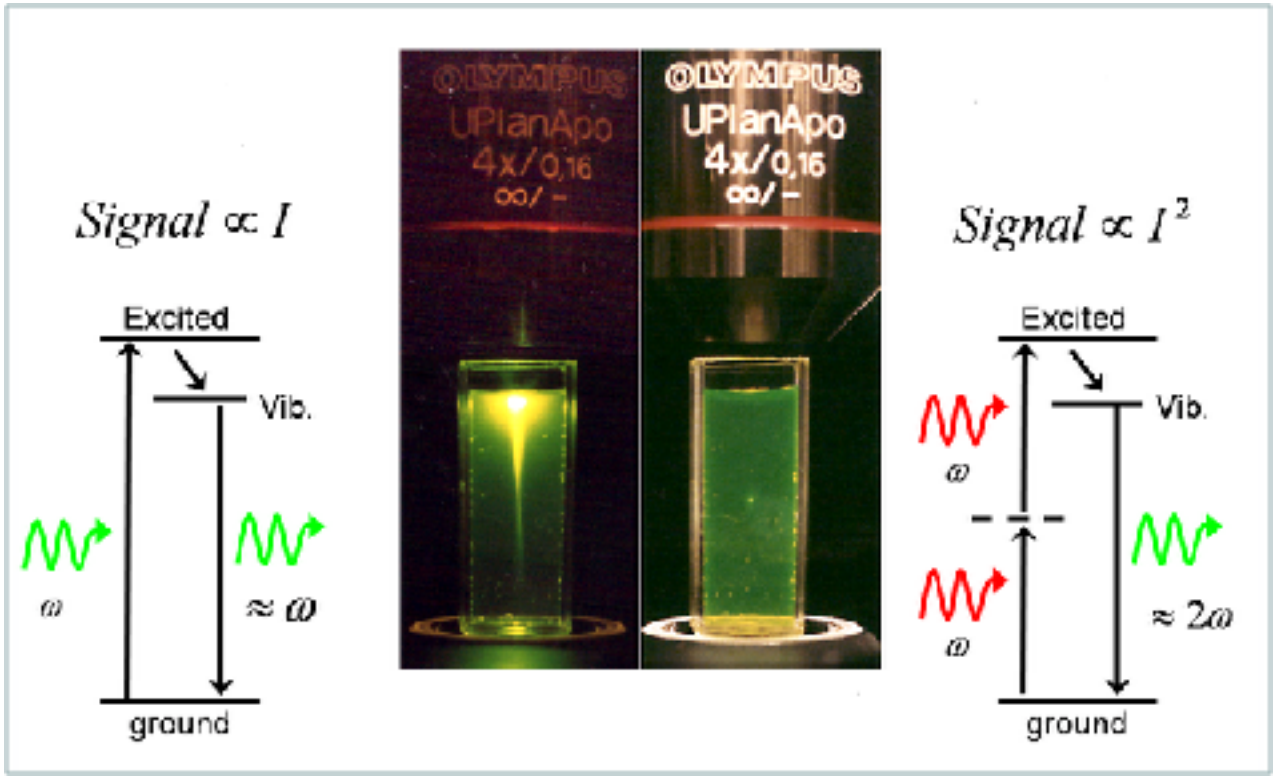
2nd low energy (IR~700nm) pulsed excitation

Low energy (IR ~700nm) Pulsed excitation



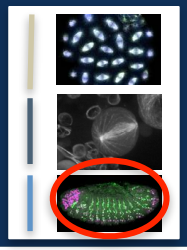


Principle of 2-photon

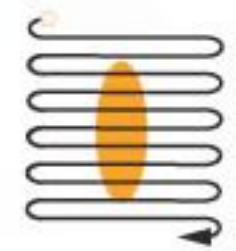


Near simultaneous, two photon event highly unlikely, only really possible a focal point

Tightly focused excitation



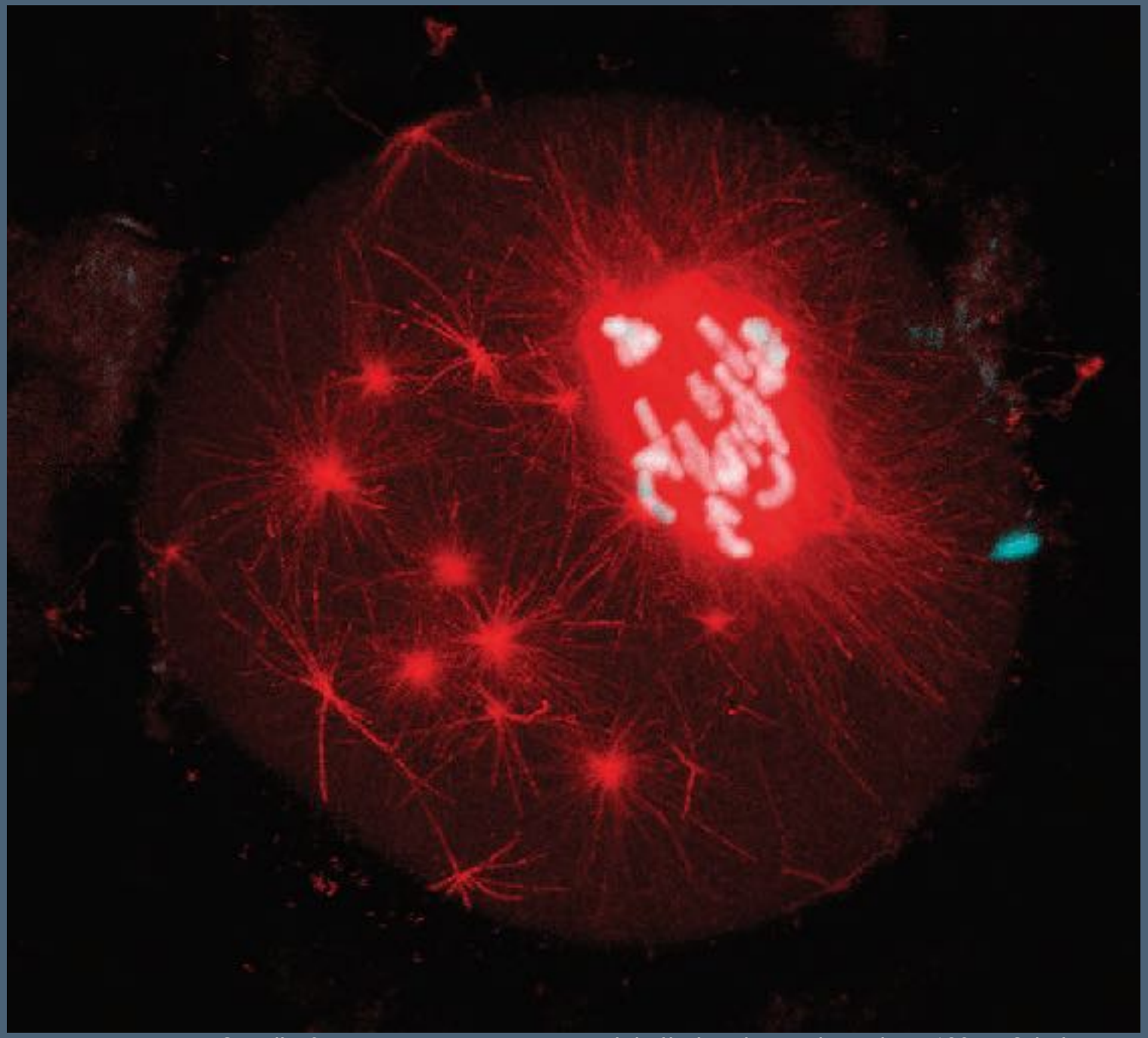
2-photon Microscope



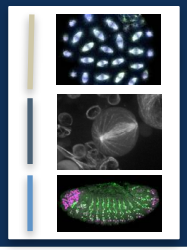
Pulsed excitation laser is then scanned across the sample.

Longer wavelengths are scattered to a lesser degree than shorter ones, and penetrate deeper into the sample.

In addition, these lower-energy photons are less likely to cause damage outside the focal volume.



Spindle formation in mouse ooctye, labelled with Hoechst, Alexa 680. M Schuh. EMBL, Heidelberg, G



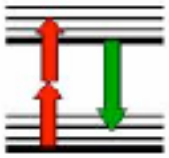
3 Flavours of Microscope

Problem:
Out of Focus
Light

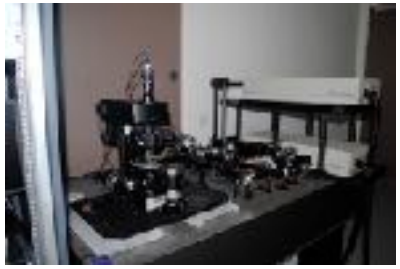
Confocal

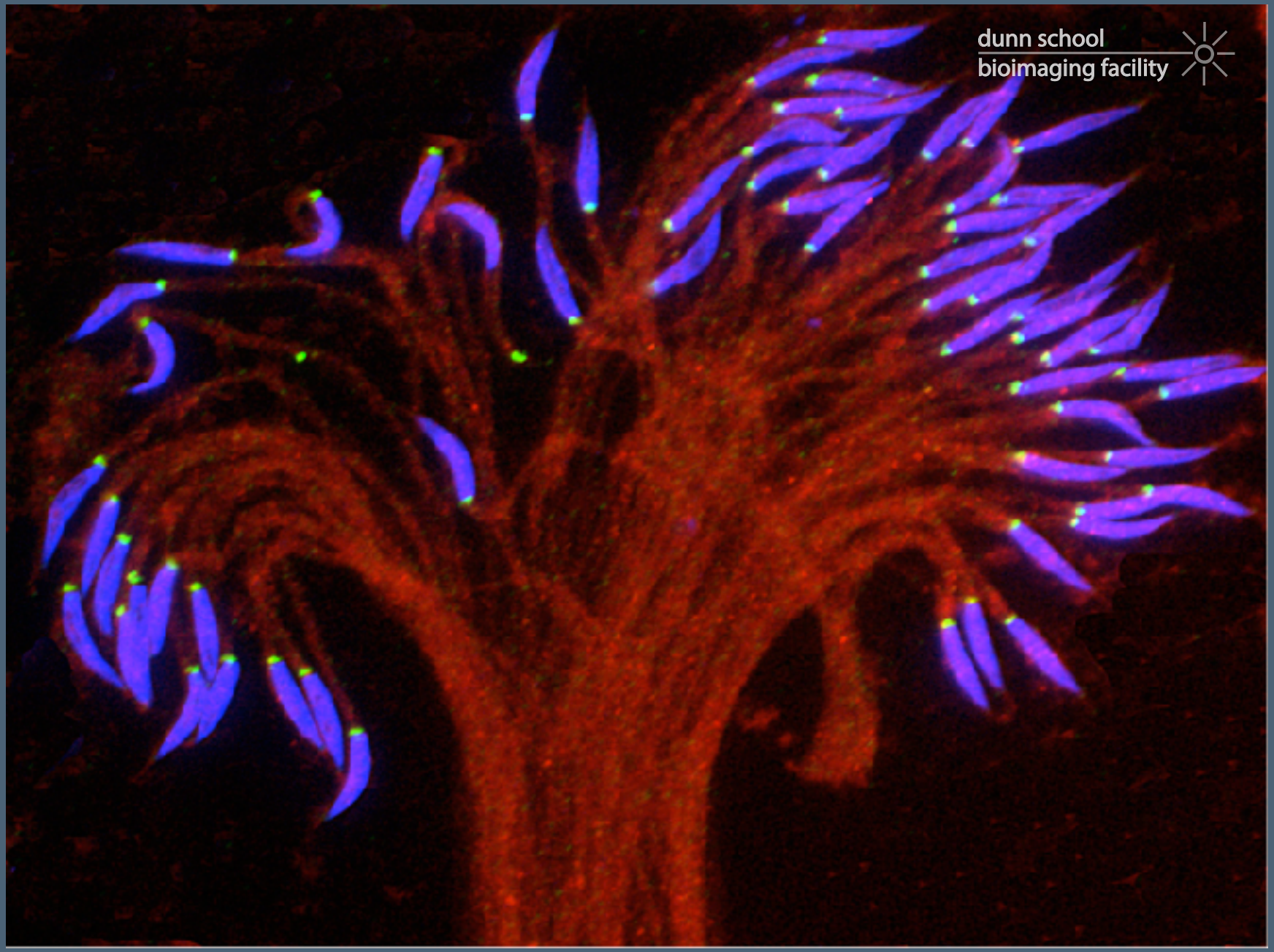
Laser
Scanning

Spinning
disc



2-Photon




dunn school
bioimaging facility