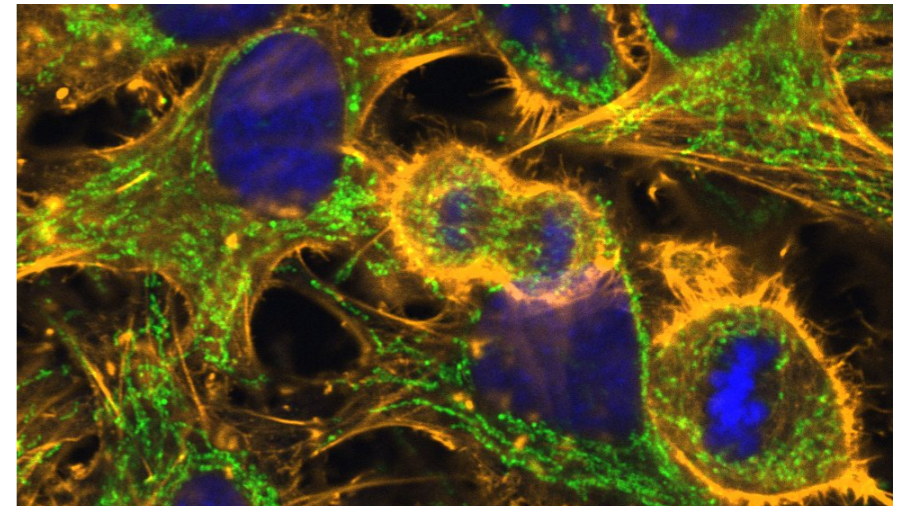
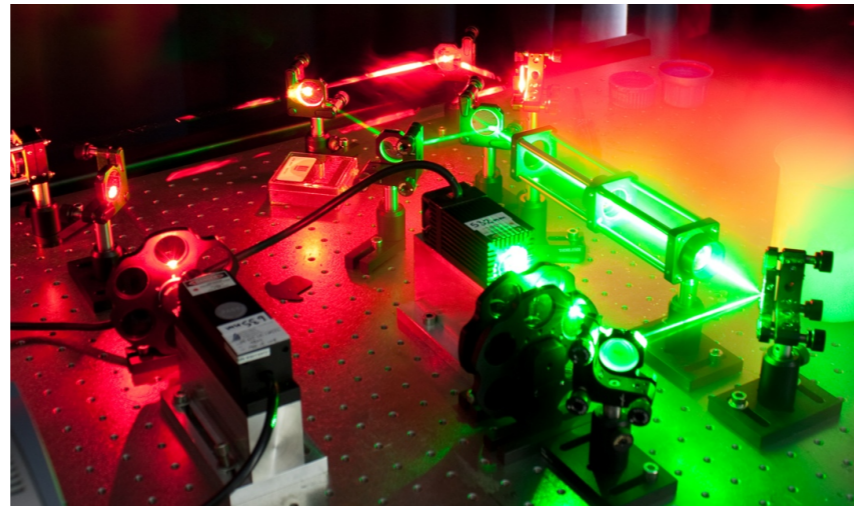
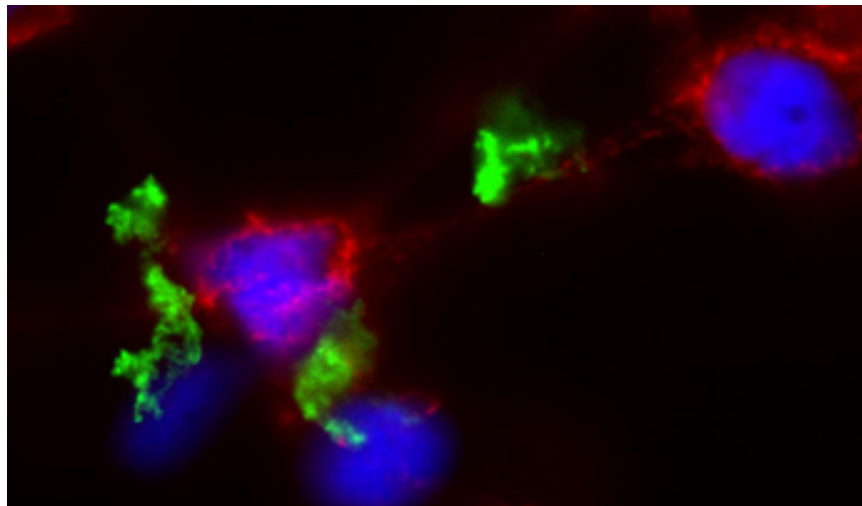


# Understanding and Applying Fluorescence Microscopy



**Carina Mónico**

Micron assistant manager

## Part I:

What is fluorescence?

Why fluorescence?

Fundamental problem in fluorescence microscopy

Components of the fluorescence microscope: dichroic mirror

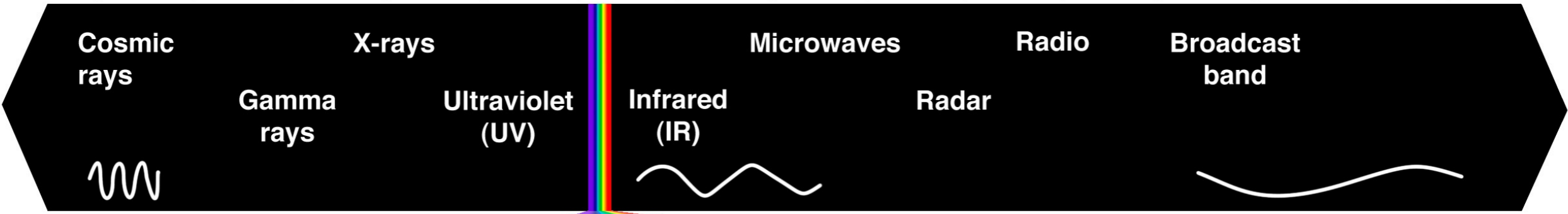
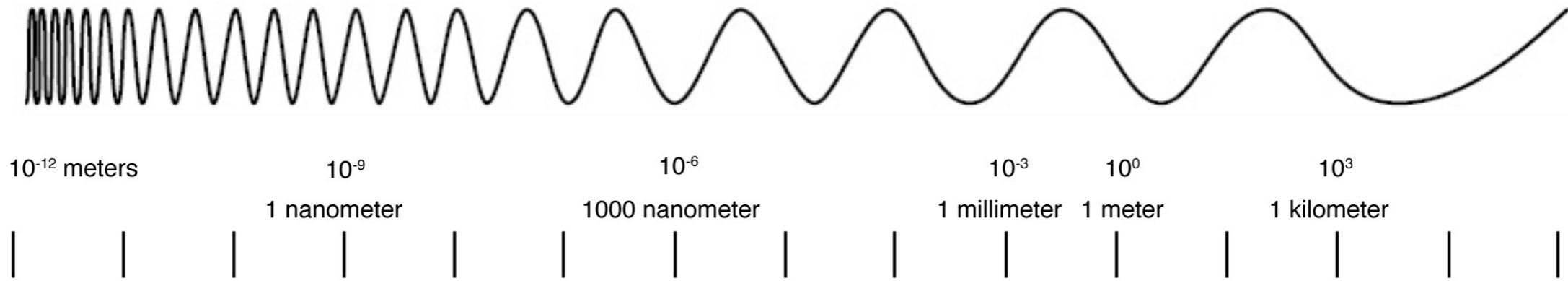
Fluorescent light sources

**Part II:** Tips on sample preparation - Fixed samples

**Part III:** Point Spread Function and Optical Transfer Function



# Light: the electromagnetic spectrum



Short Wavelengths

Long Wavelengths

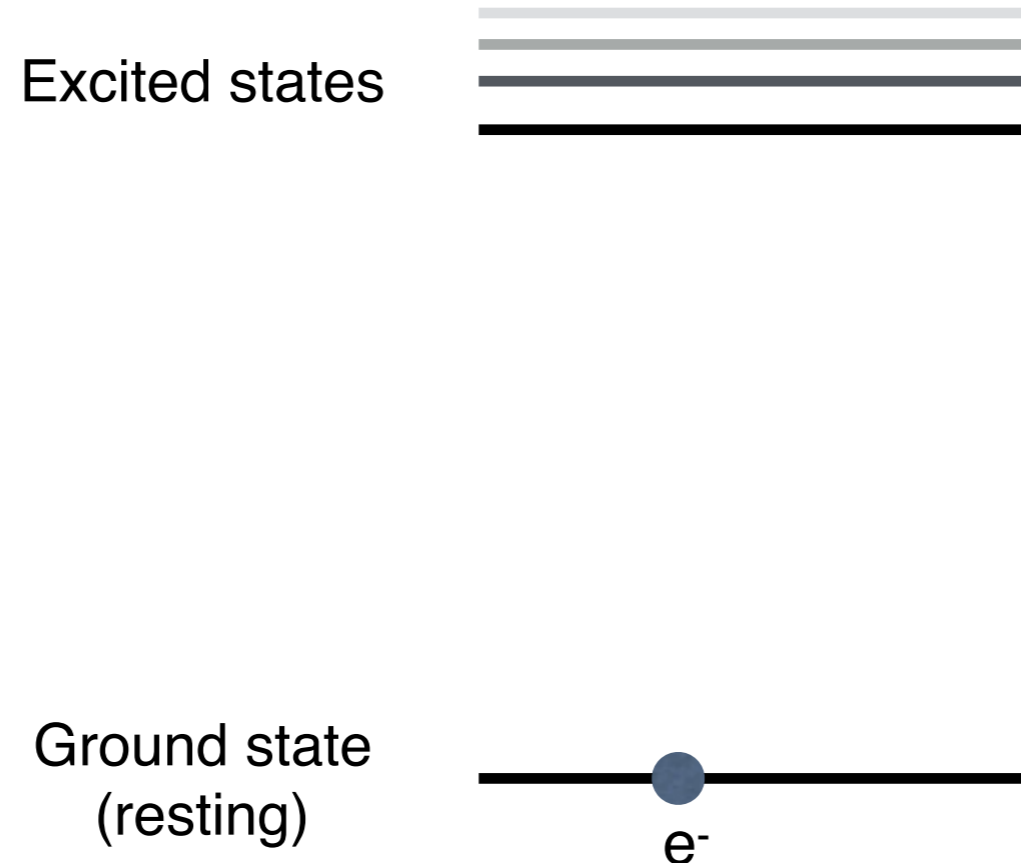


400 nanometers      500 nanometers      600 nanometers      700 nanometers

380 – 700 nm visible to the human eye

# What is Fluorescence?

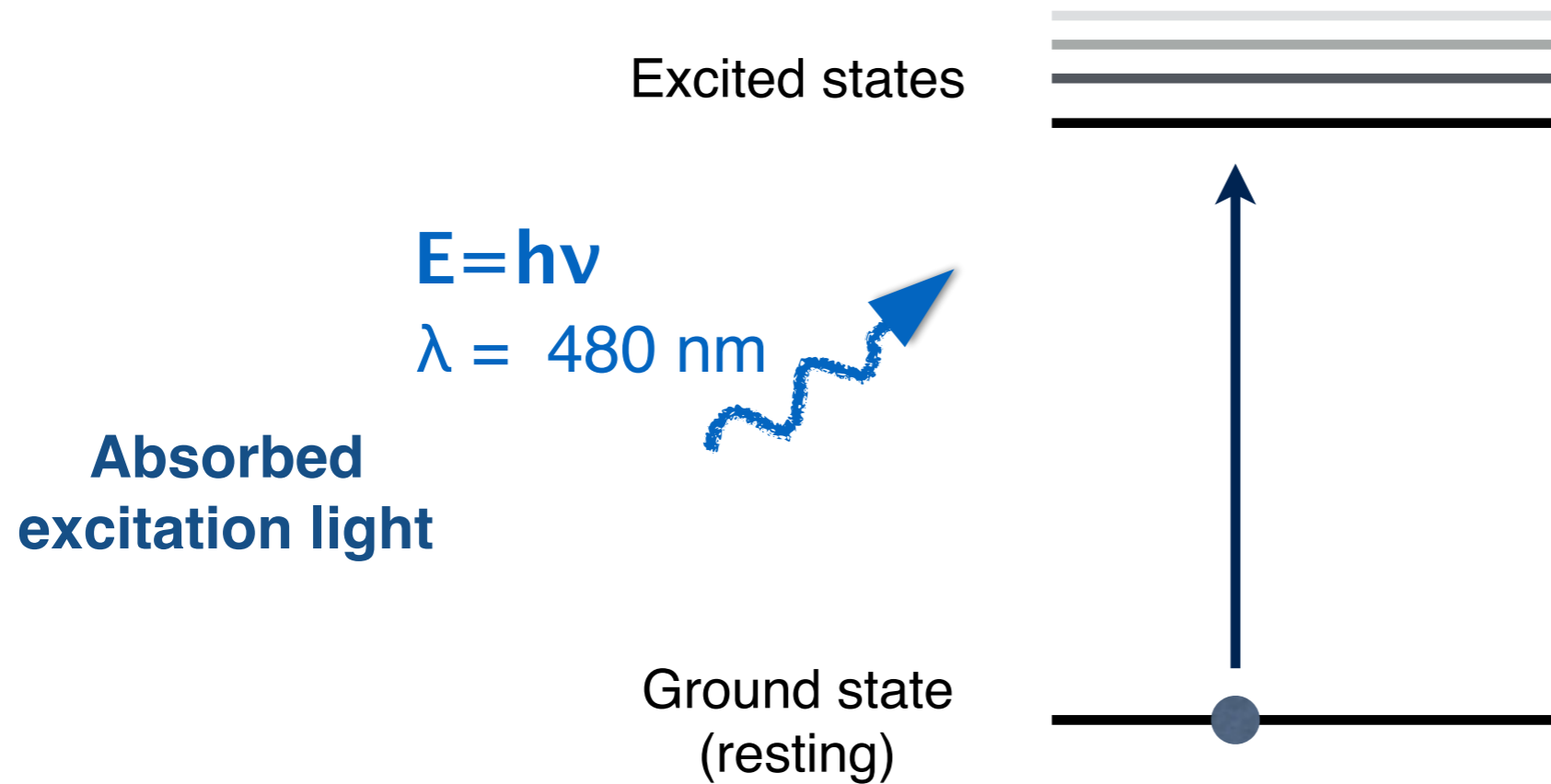
**Fluorescence** is the emission of light by a molecule that has absorbed light



Molecules have discrete levels of energy

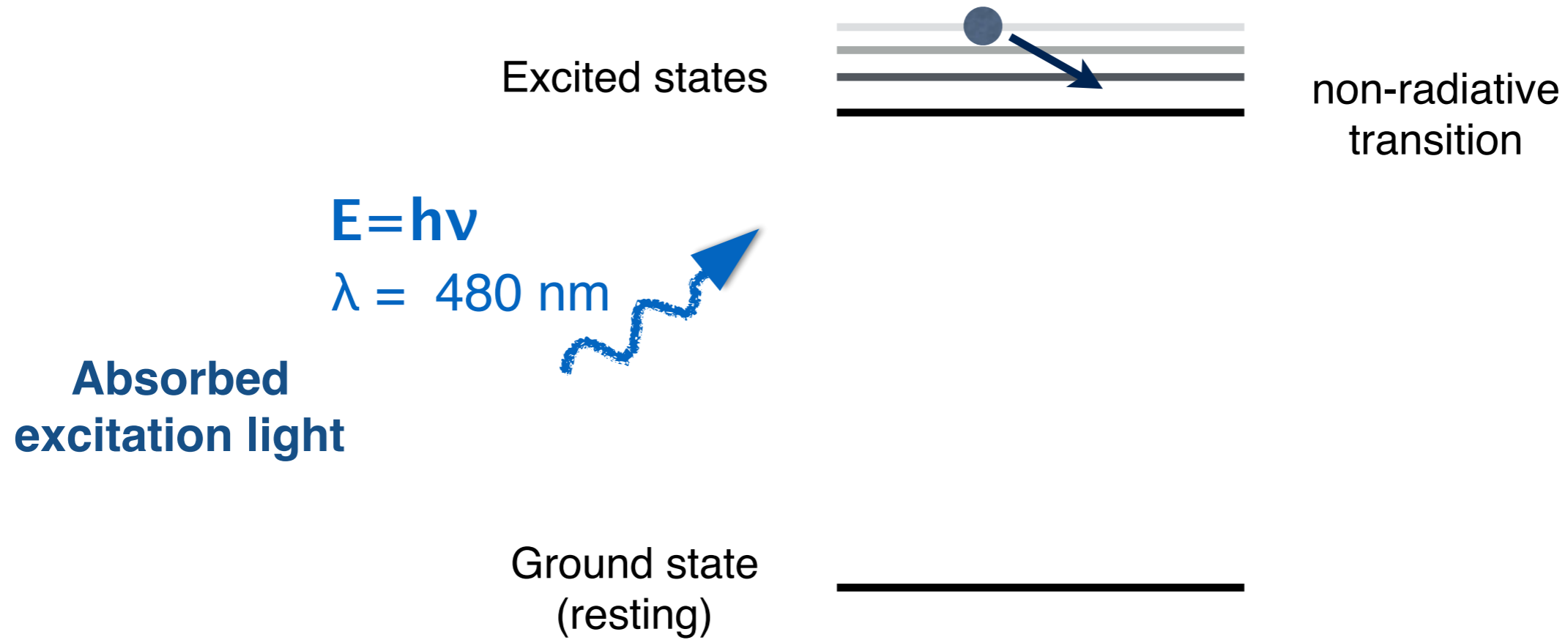
# What is Fluorescence?

**Fluorescence** is the emission of light by a molecule that has absorbed light



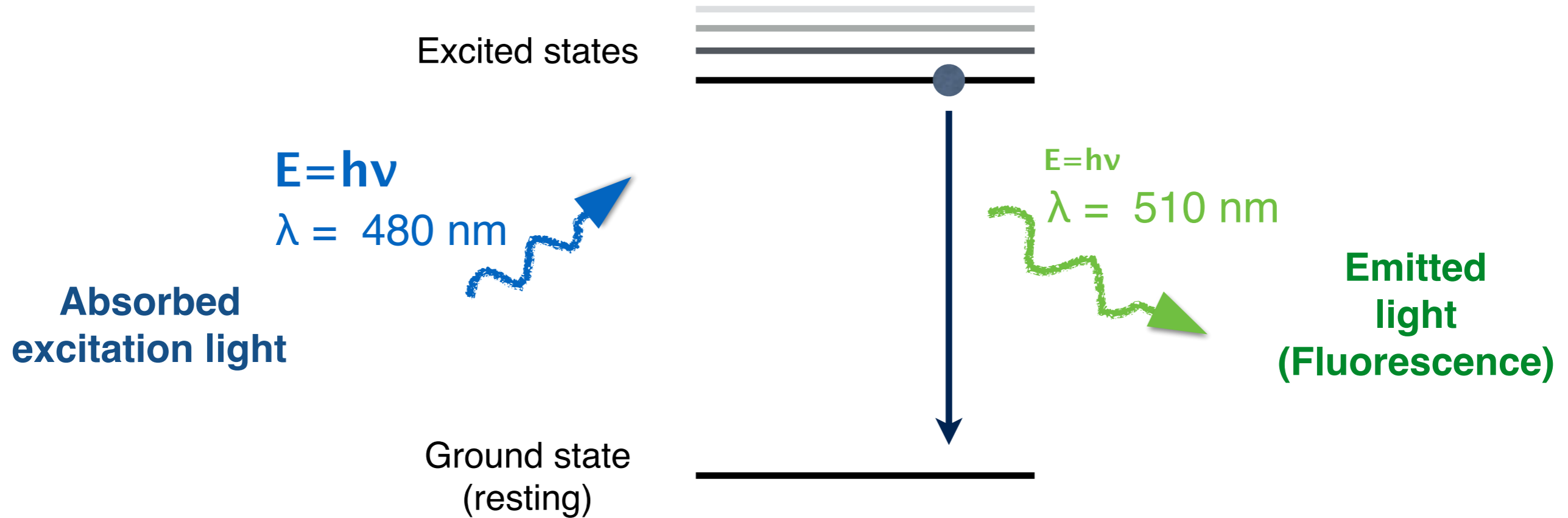
A photon is the energy unit for light to interact with matter

# What is Fluorescence?





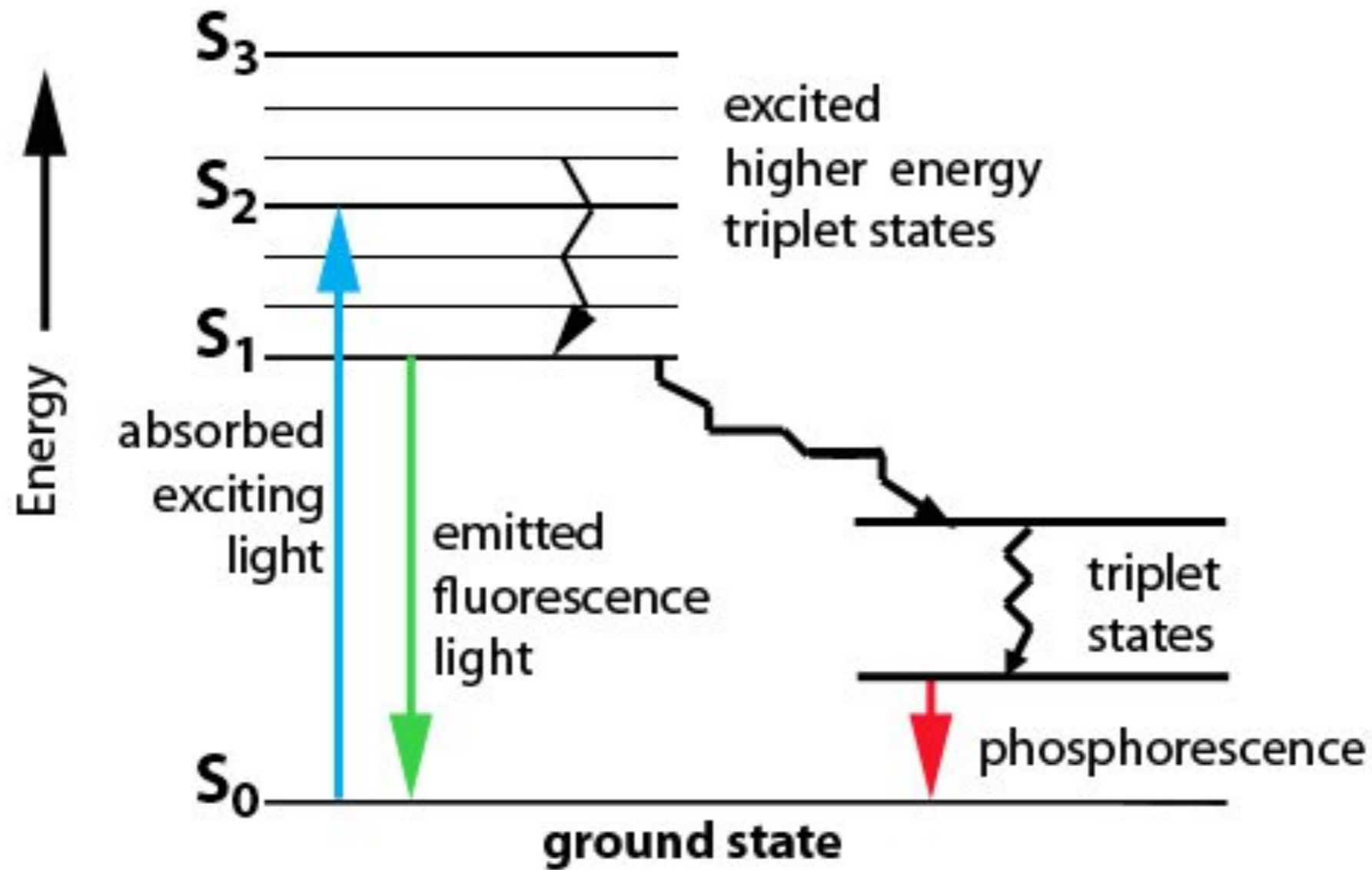
# What is Fluorescence?



**Fluorescence** has higher wavelength than absorbed light

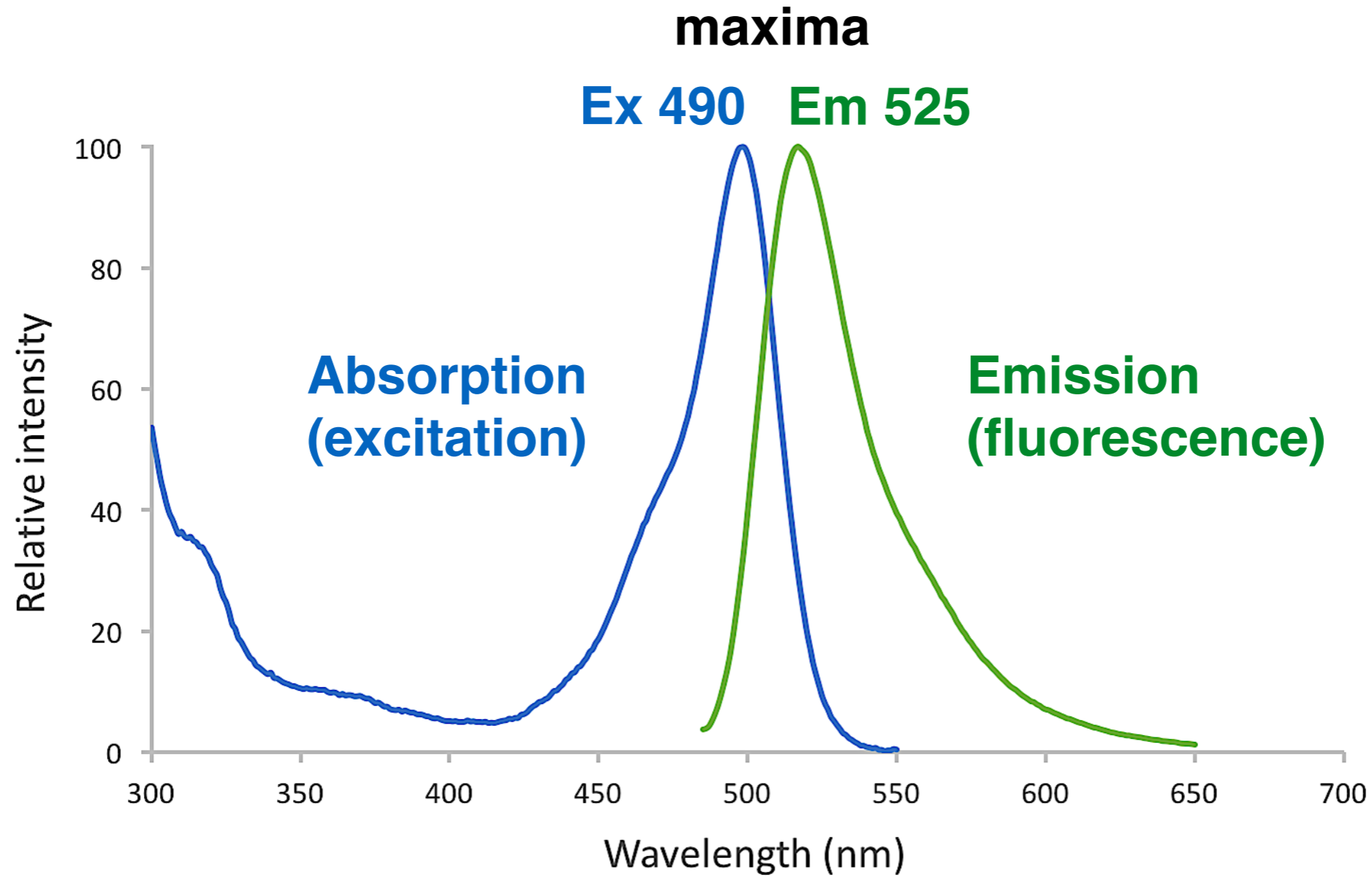
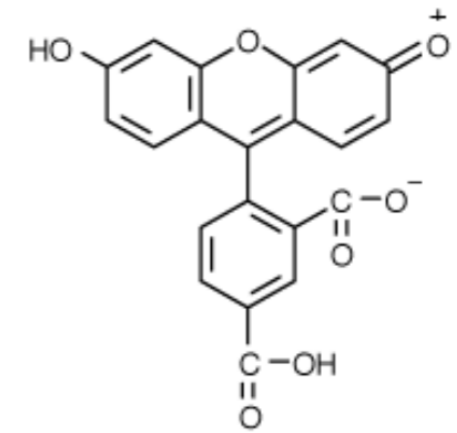
The full picture is represented on the Jablonski diagram...

→ Lecture 6



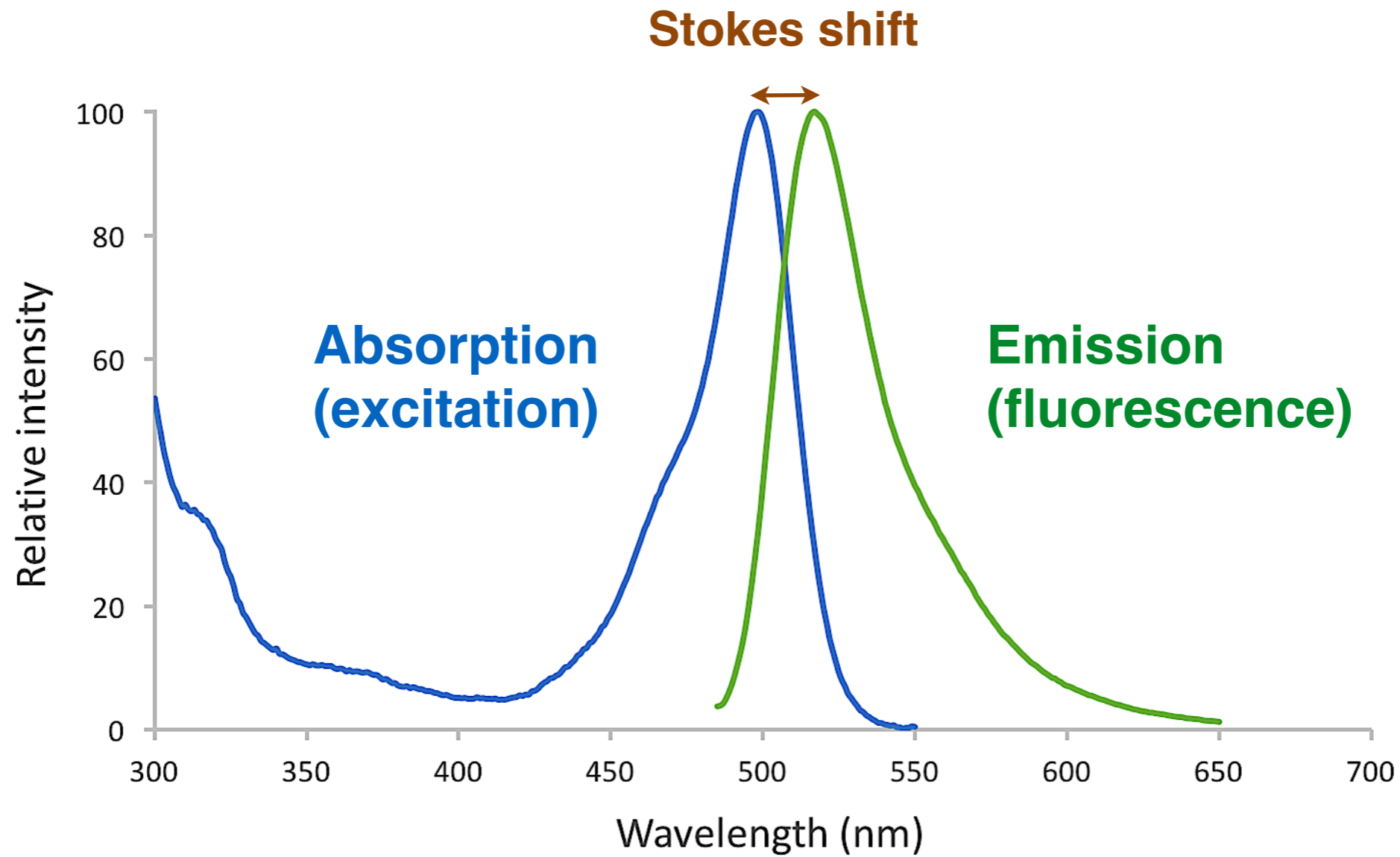
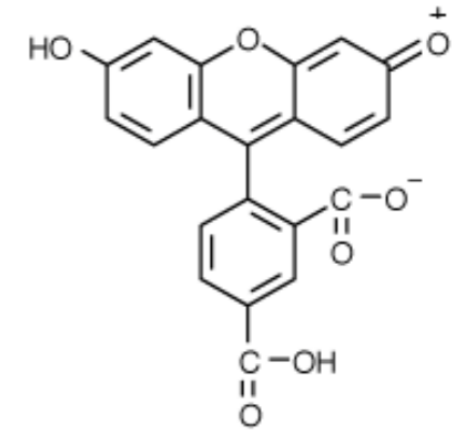
# Fluorescence Spectra

Fluorescein (FITC)



# Fluorescence Spectra

Fluorescein (FITC)





## Genetically encoded fluorescent proteins

- GFP, YFP, mCherry

## Organic dyes

- Alexa, ATTO, Fluorescein, DAPI, Cyanine (Cy3, Cy5)
- Fluorescent labelled antibodies (immunofluorescence)

## Inorganic dyes

- Quantum Dots

## Endogenous species

- Elastin, collagen, metabolic coenzymes (NADH, FAD)

# Why Fluorescence?

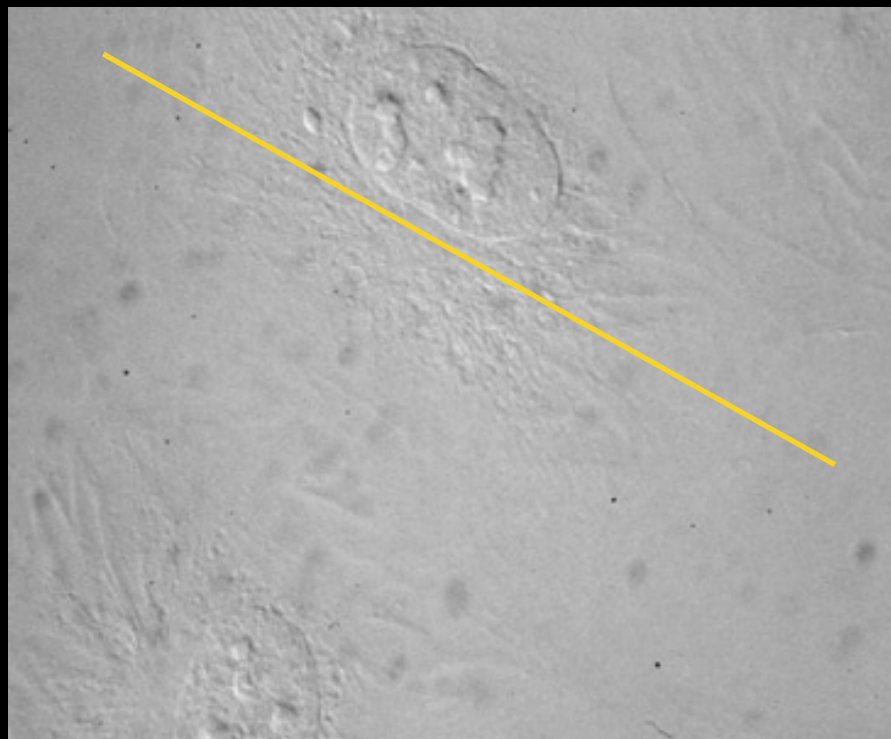


Chris Teren: <https://www.youtube.com/watch?v=PhclTQ3g0s8>

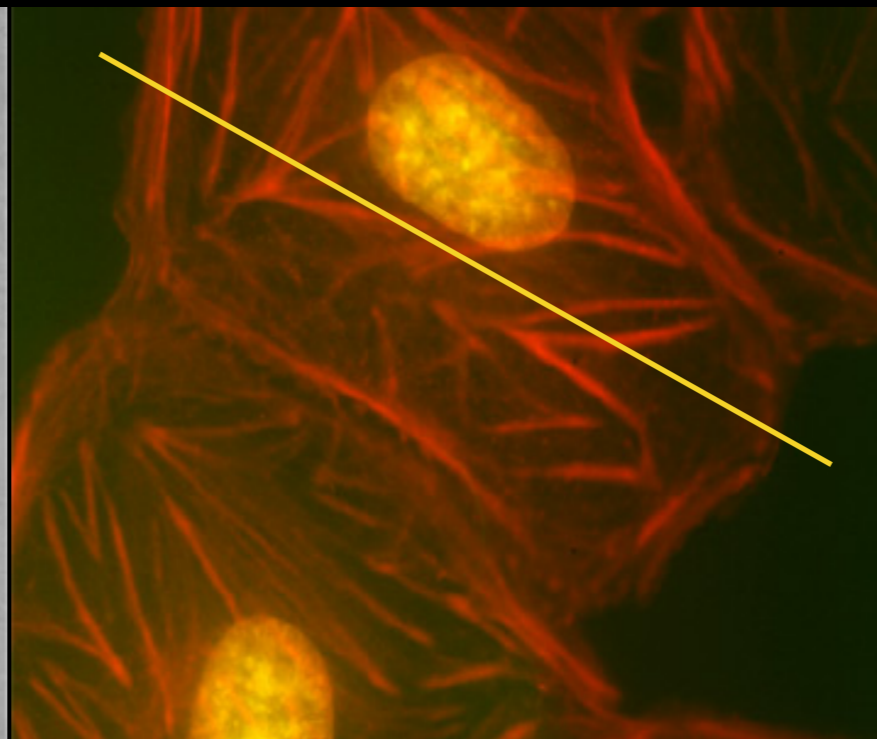
**CONTRAST**

# Why Fluorescence?

- Weak signal against dark background is easier to measure
- High signal to background - contrast



bright field (DIC)



fluorescence



Intensity profile

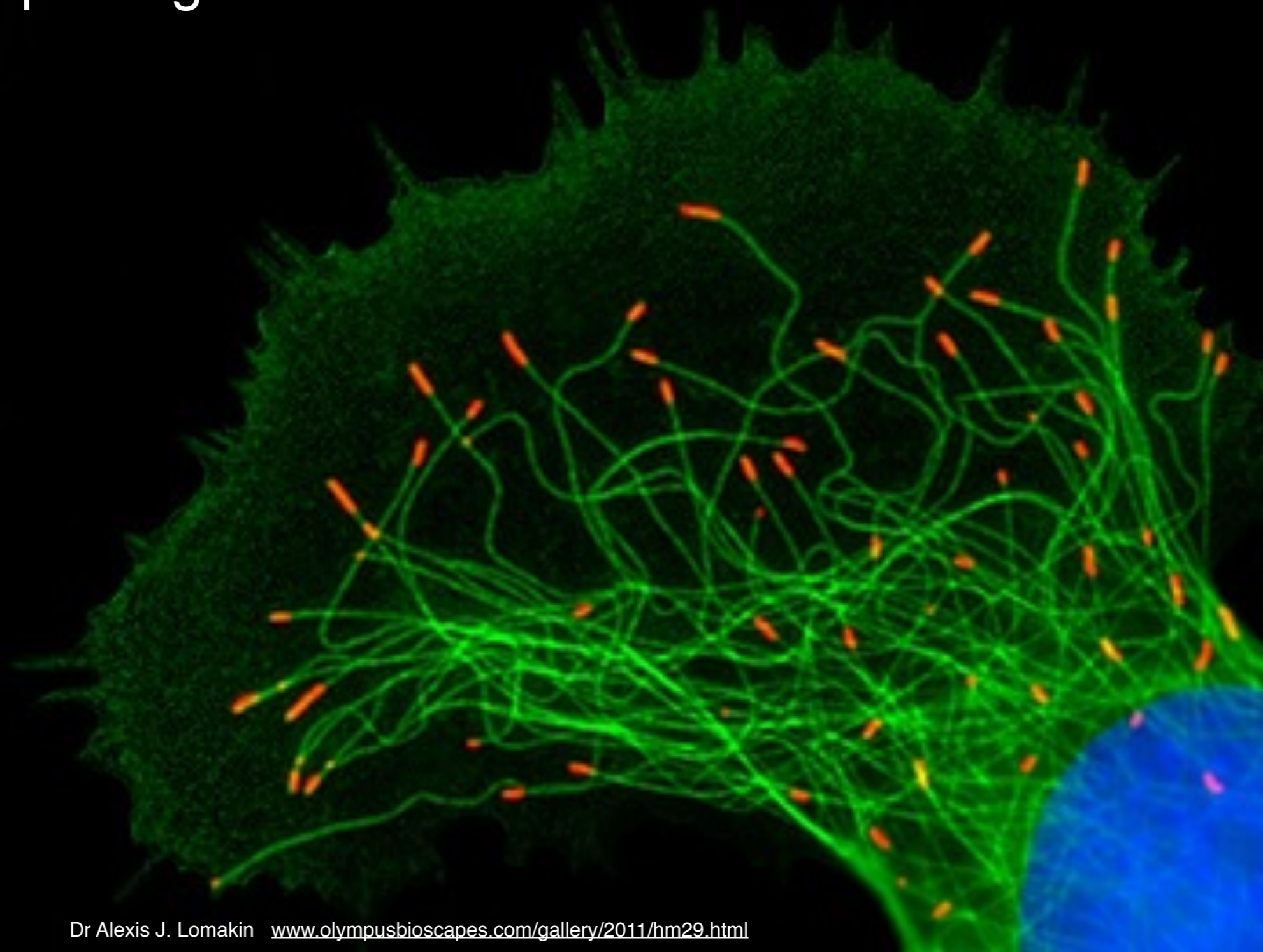
# Why Fluorescence?

- Selective labeling
- Ease of multiplexing
- Quantitative

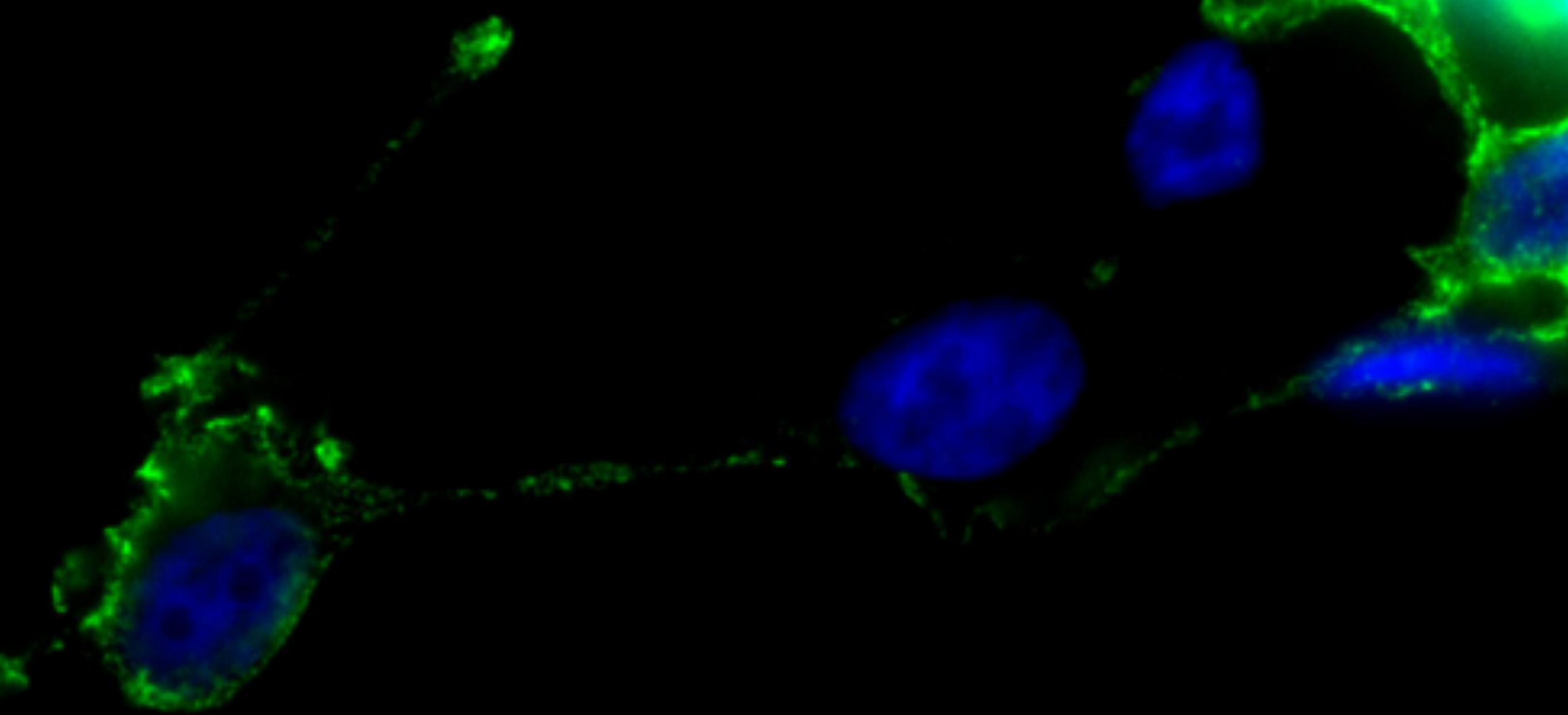
**Microtubules**

**Microtubule Plus ends**

**Nucleus**







Why is the background black In a fluorescent image ....?

**Widefield  
deconvolution**

**Confocal**

**TIRF**

**FCS**

**dSTORM**

**PALM**

**Multi-photon**

**STED**

**3D-SIM**

# Fundamental problem in fluorescence microscopy

**STRONG  
illumination**

vs.

**WEAK  
fluorescence signal**



produce high-efficient illumination of the specimen



capture weak fluorescence emission

illumination  
EXCITATION

EMISSION  
fluorescence

Dichroic mirror

# Dichroic mirror - at the heart of fluorescence microscopy

Dichroic mirrors are made by coating a glass substrate with a series of optical coatings

Incident Light

“unwanted” wavelengths

Reflected Light



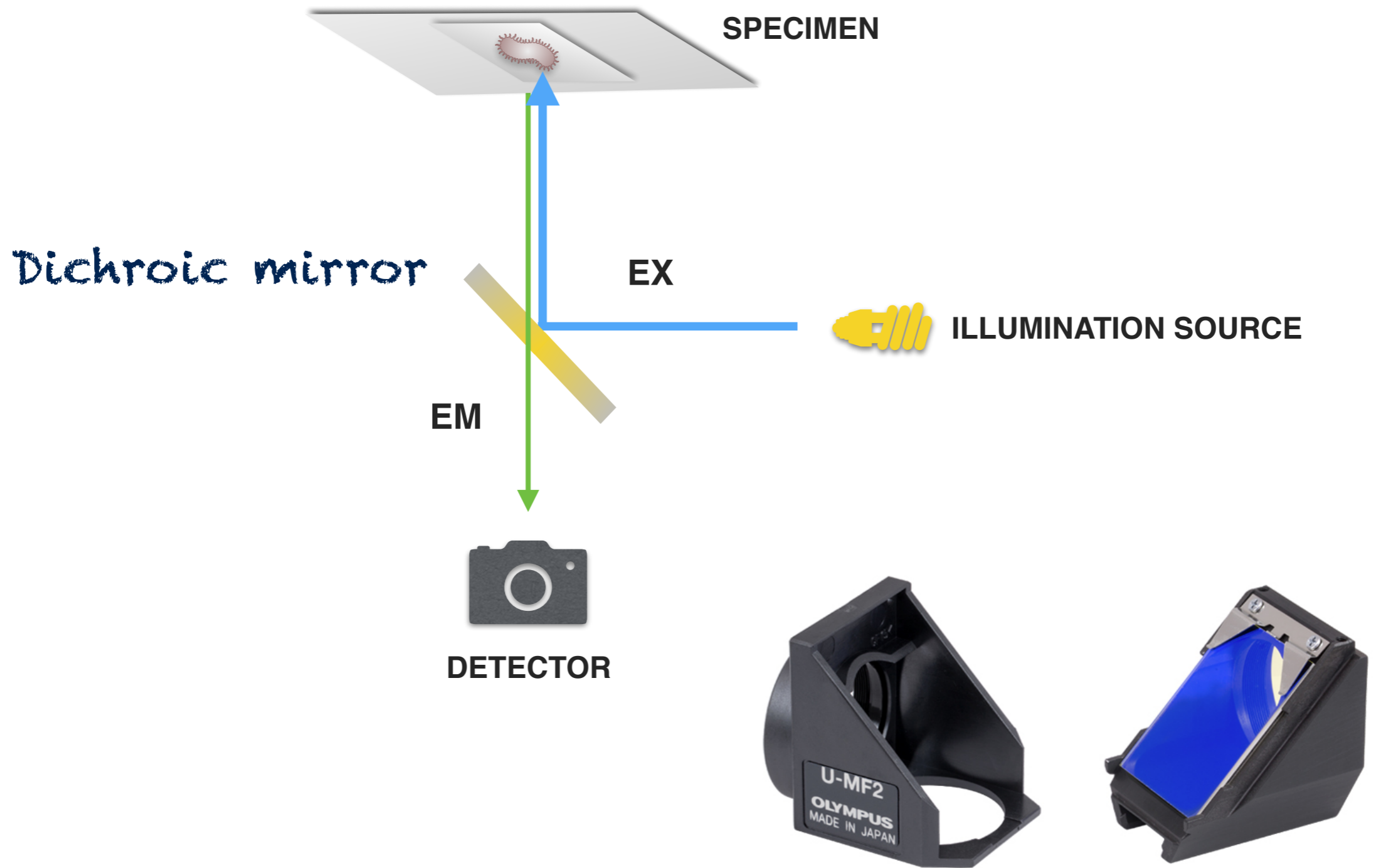
Transmitted Light

“wanted” fluorescence





# Dichroic mirror - at the heart of fluorescence microscopy

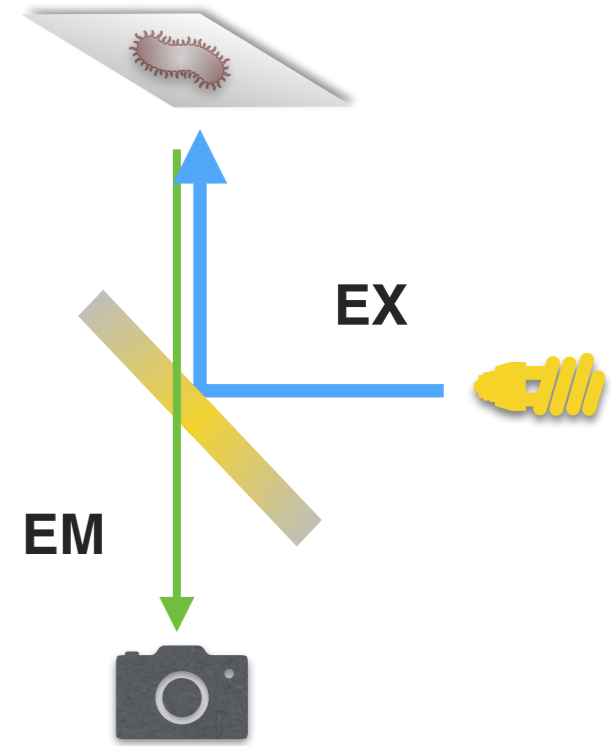
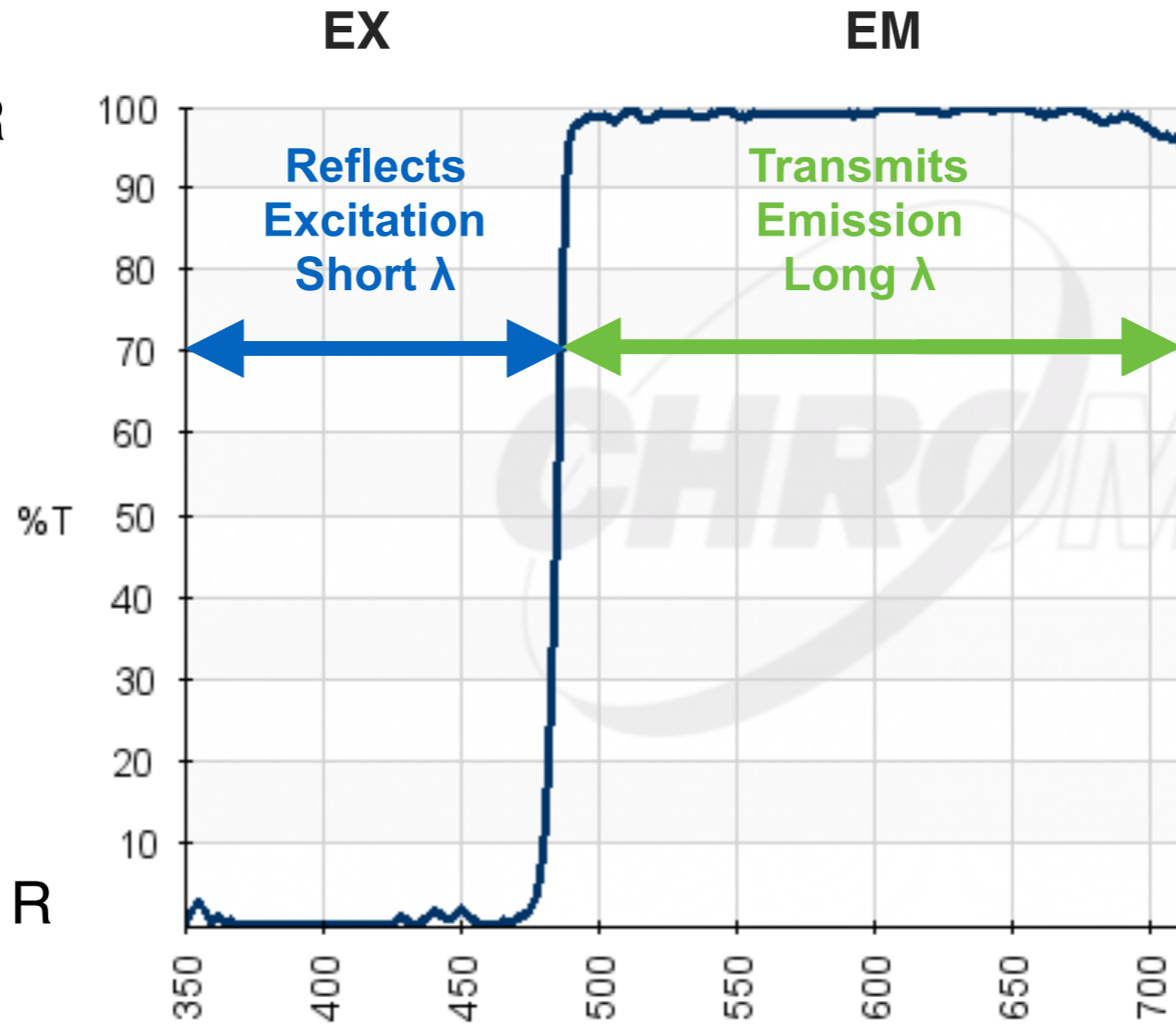


# Dichroic mirror - Spectral properties

Dichroic

100% T = 0% R

0% T = 100% R

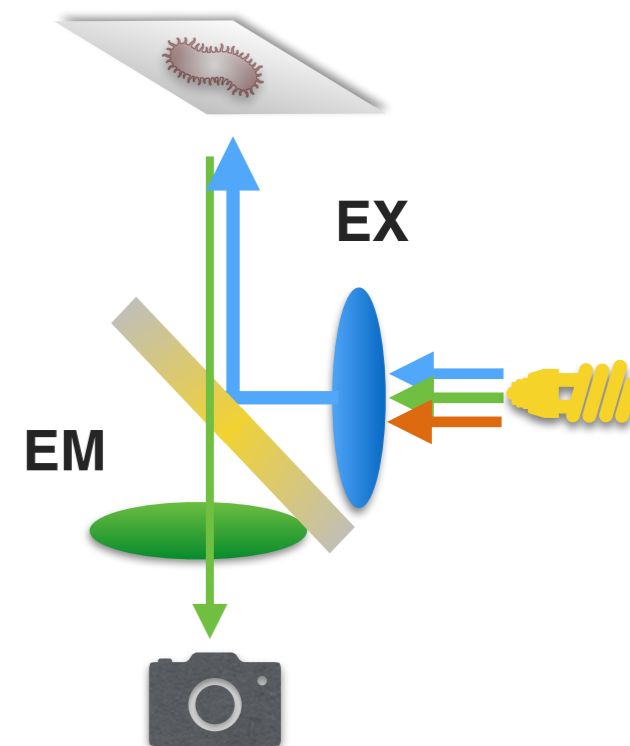
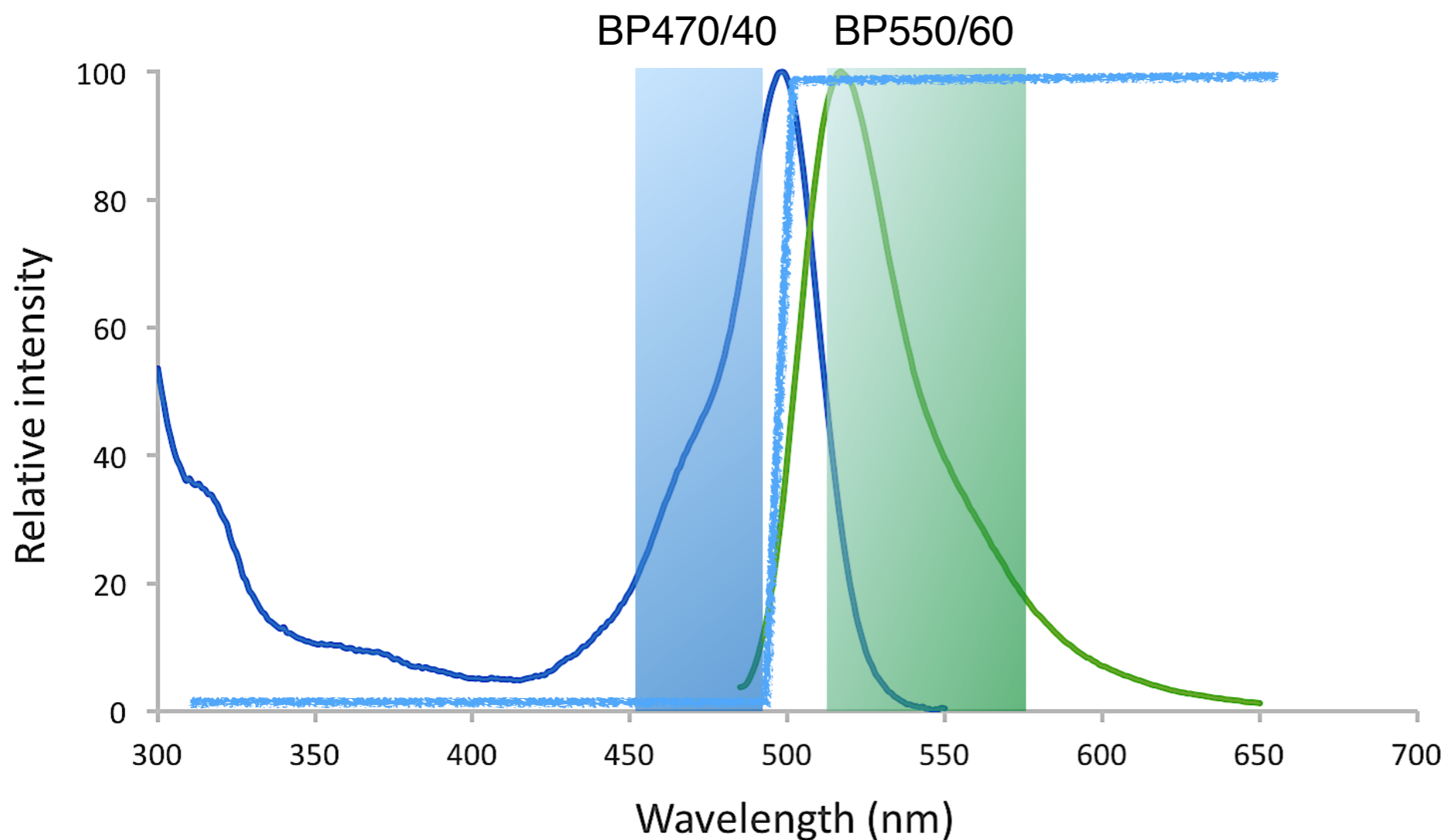
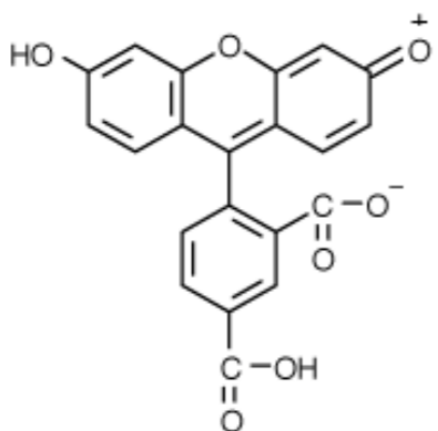


Separates excitation light from emission light

# Dichroic, excitation and emission filters

... related to dye spectrum

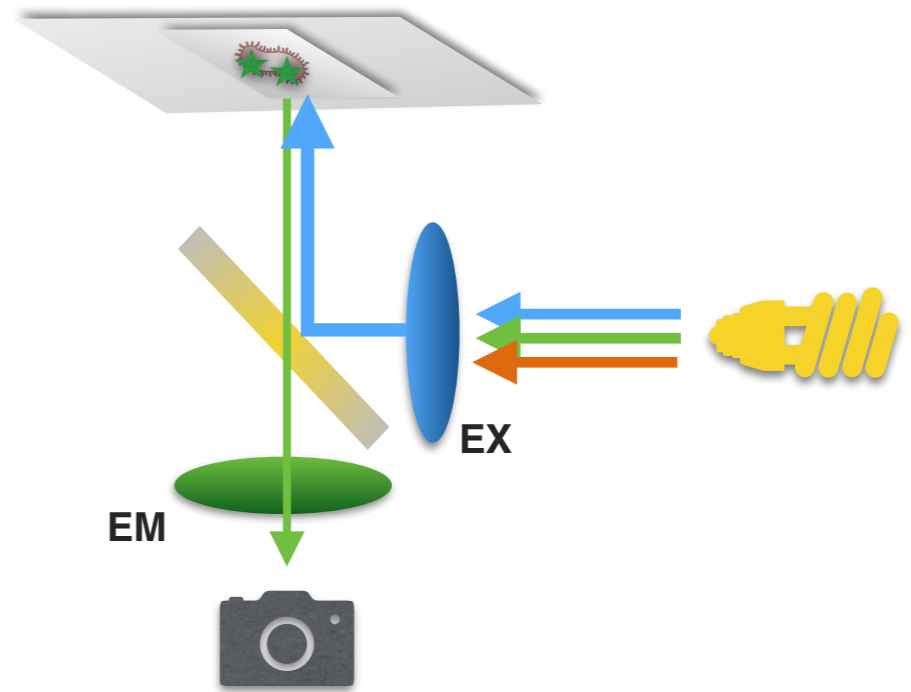
Fluorescein (FITC)



# What about multiplexing...?



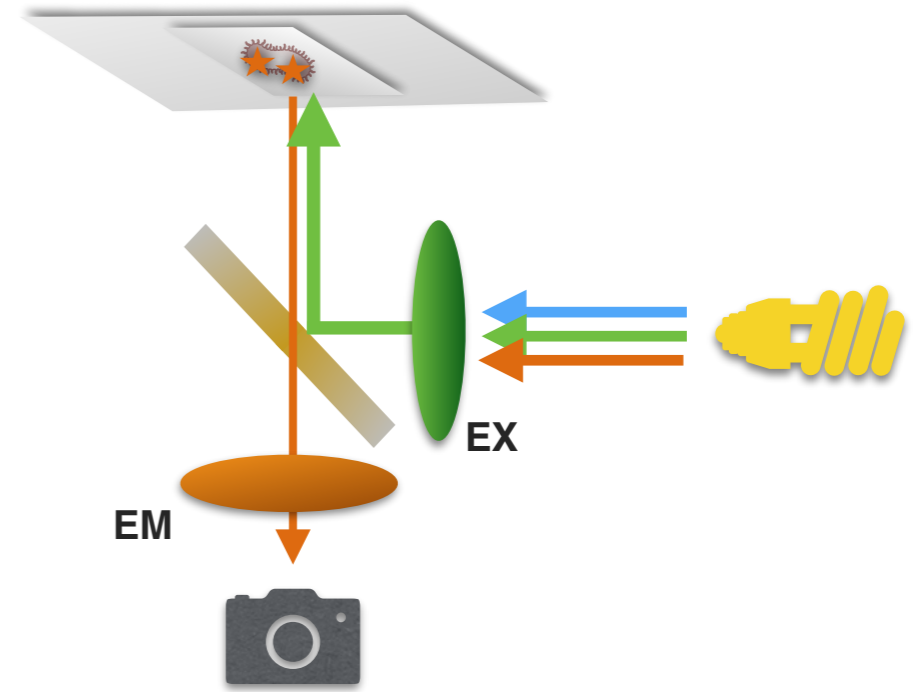
Fluorescein (FITC), GFP



# What about multiplexing...?



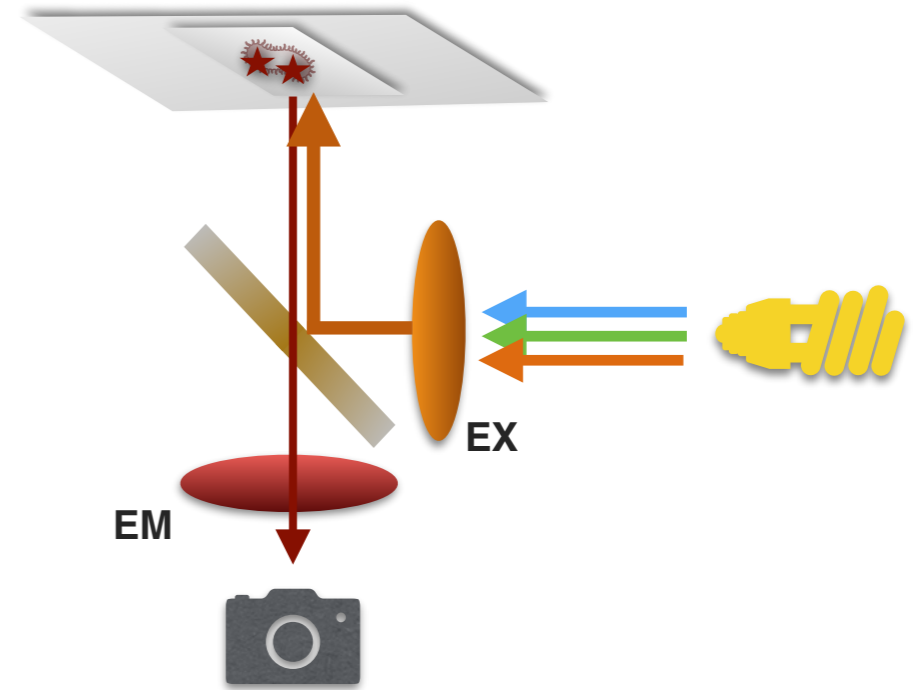
Rhodamine (TRITC)



# What about multiplexing...?



mCherry, Cy5

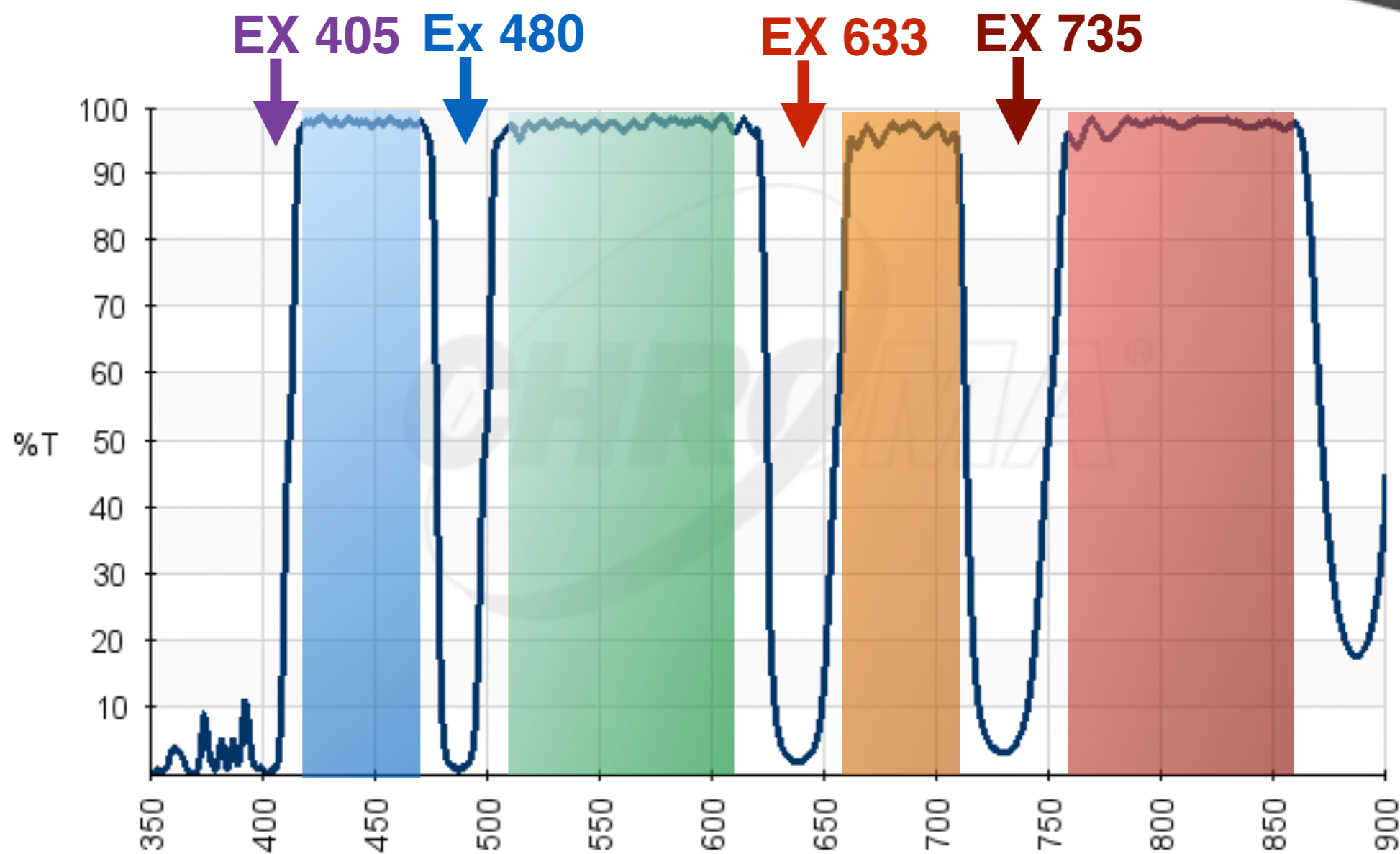




# What about multiplexing...?



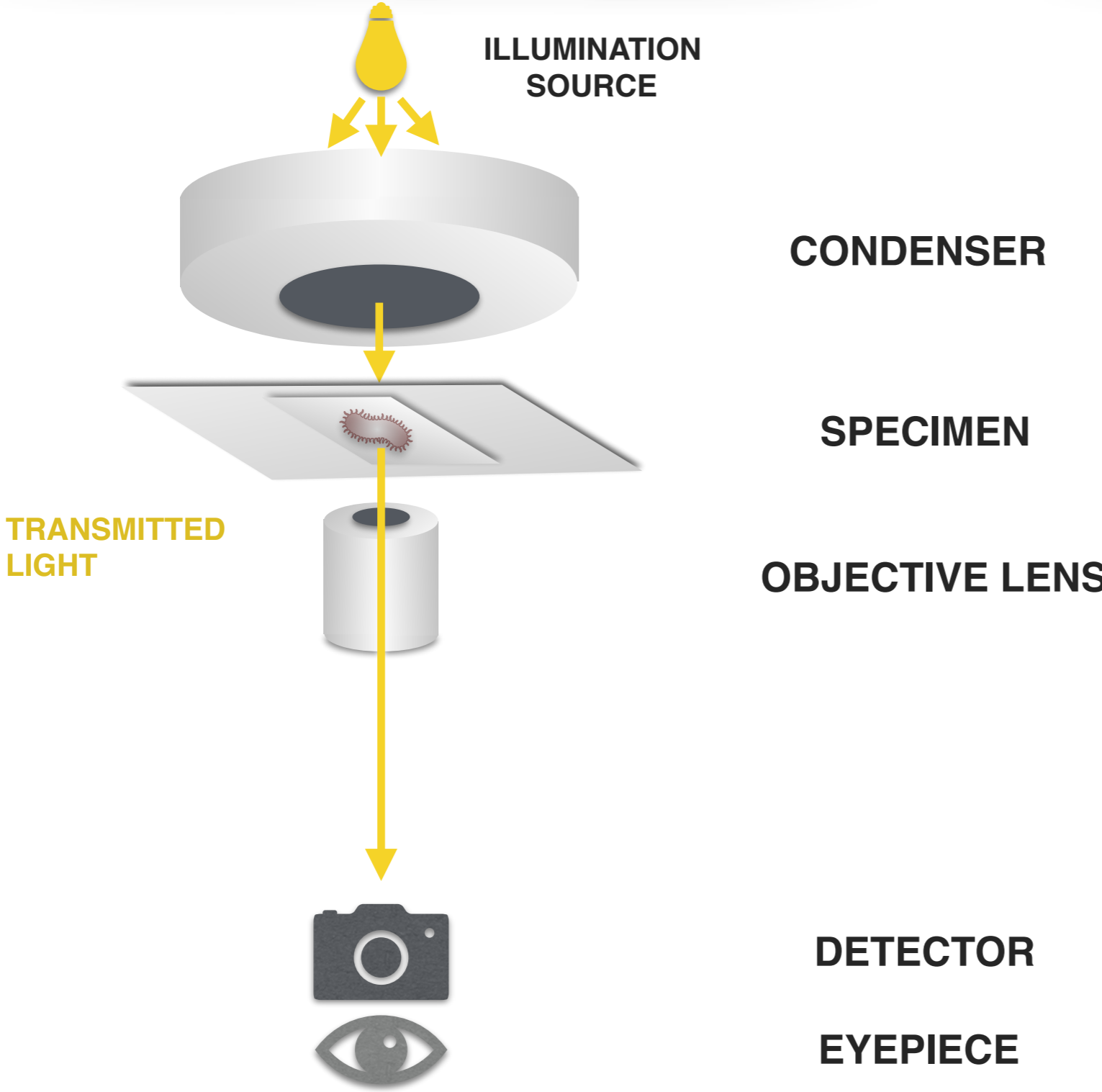
Polichroic



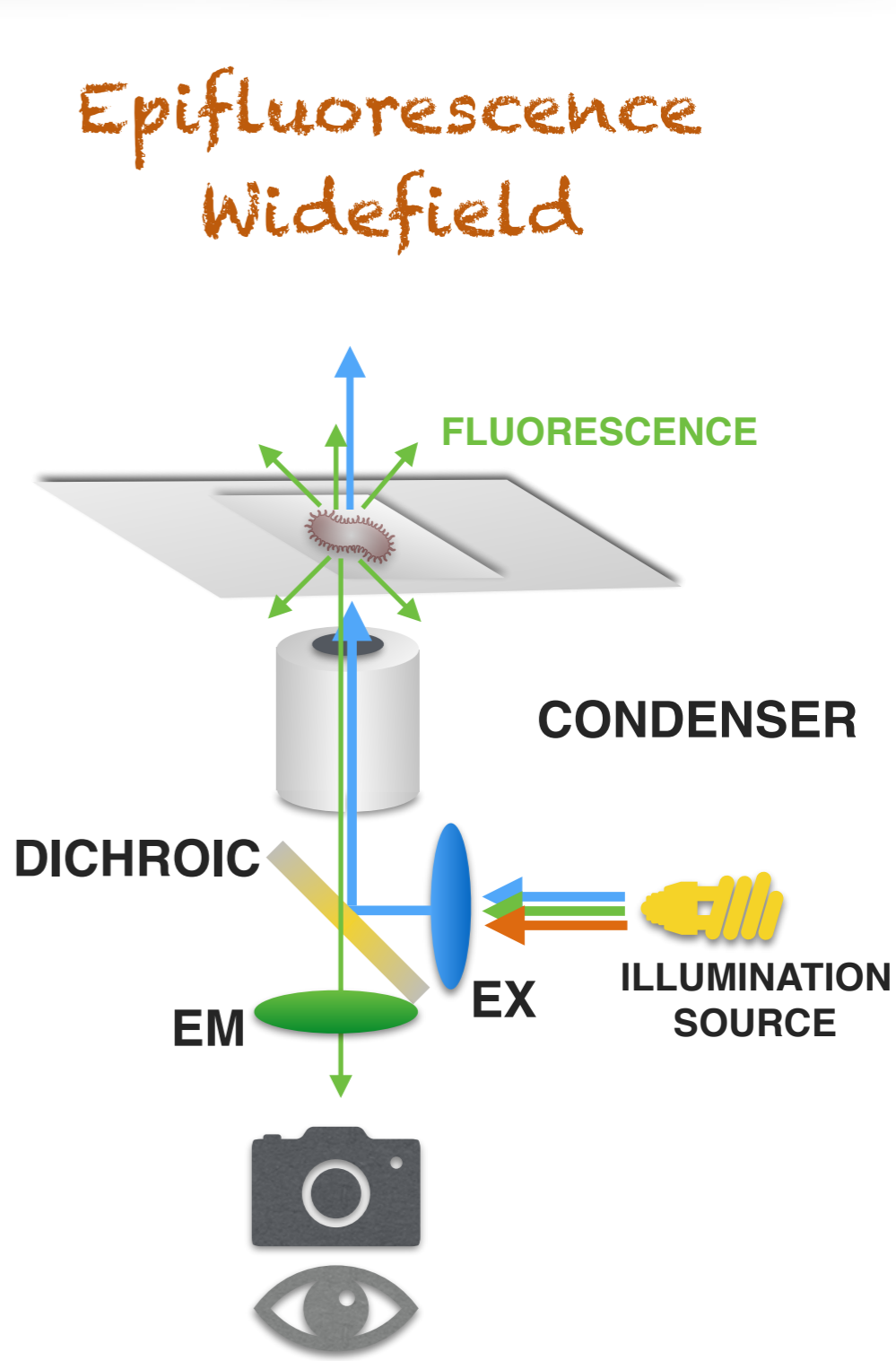


# Components of a microscope: Brightfield vs. Fluorescence

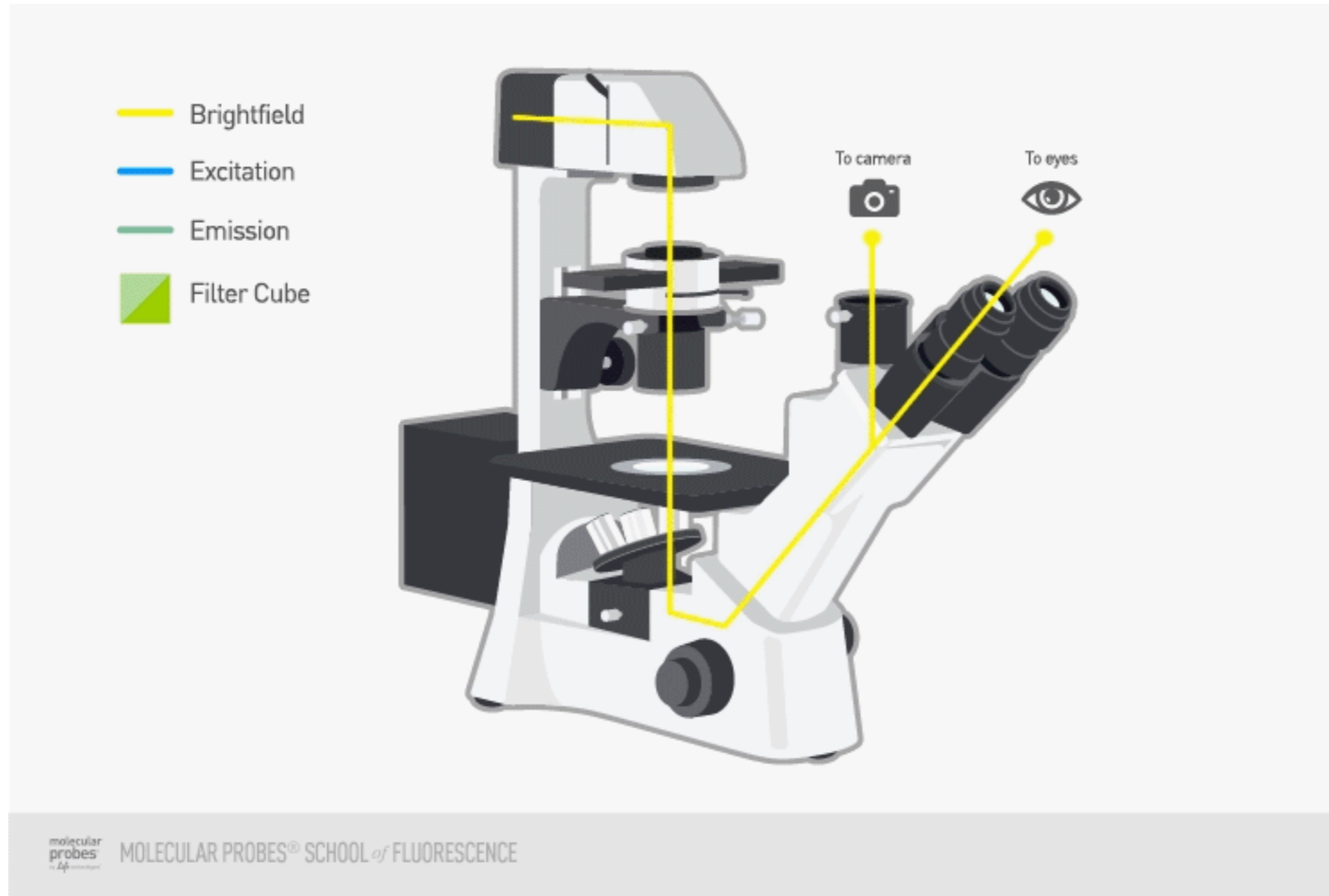
## Transmitted Light (Brightfield)



## Reflected Light (Fluorescence)

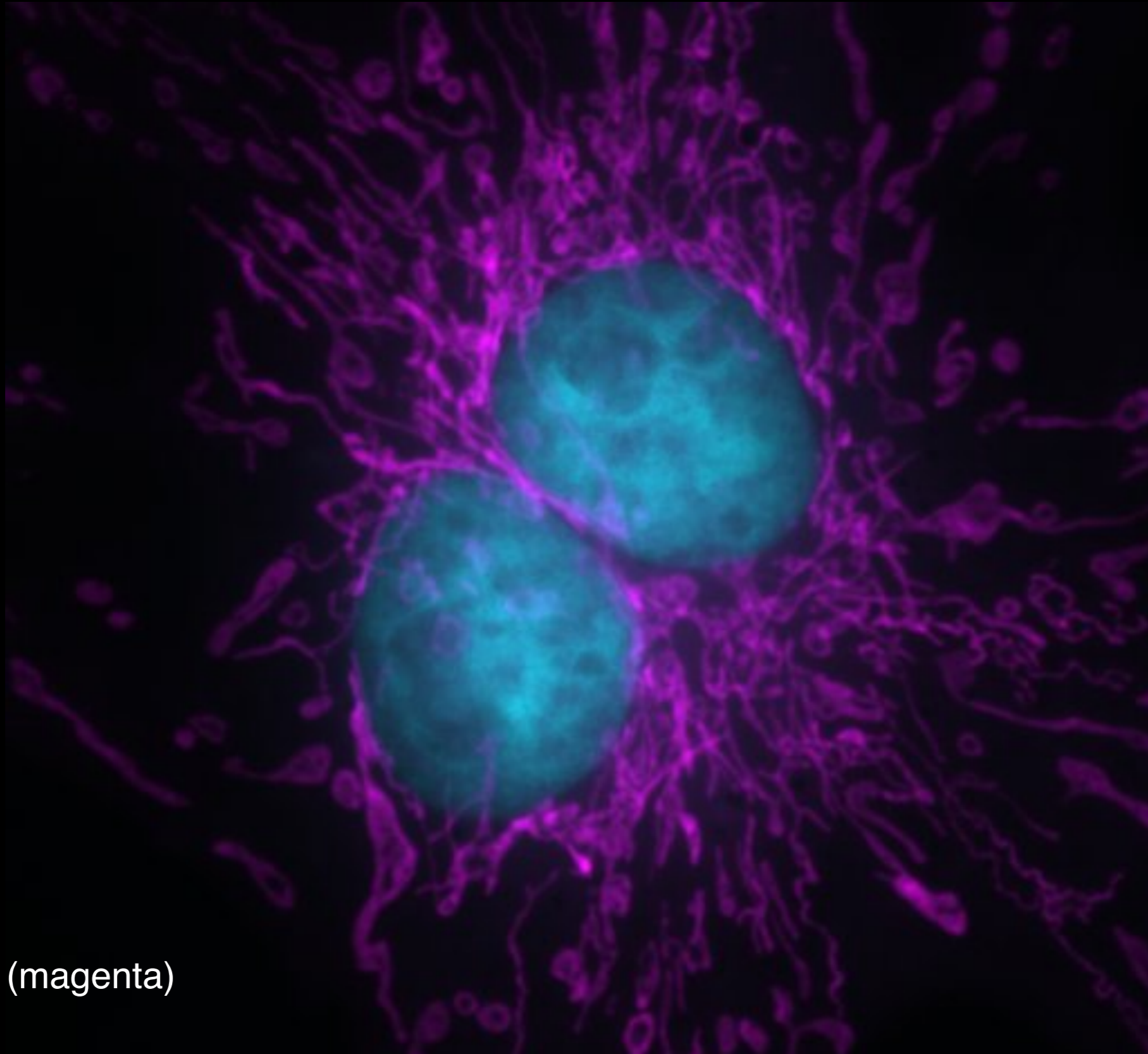


# Epifluorescence vs Brightfield light paths (inverted)



# Widefield Fluorescence Microscopy

The whole field of view is collected at once



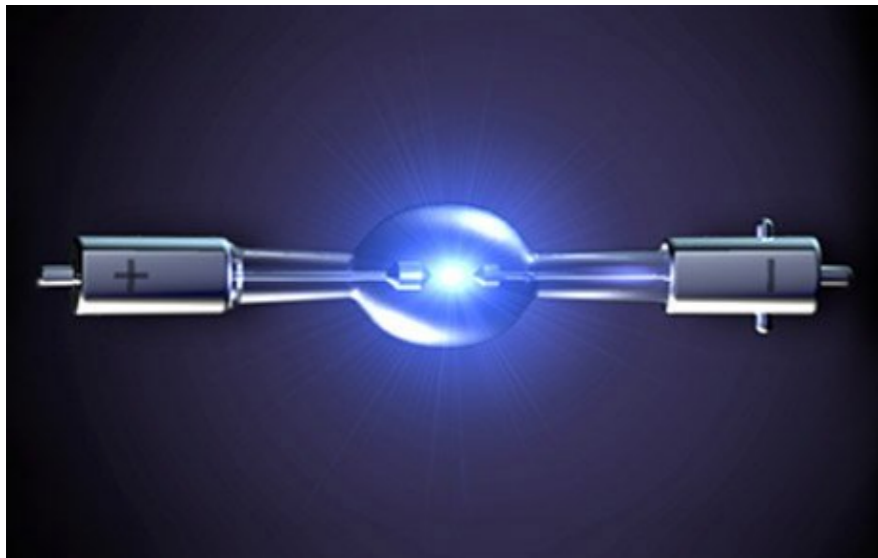
BPAE cells  
Mitotracker Red (magenta)  
DAPI (cyan)

# Illumination sources for widefield fluorescence microscopy

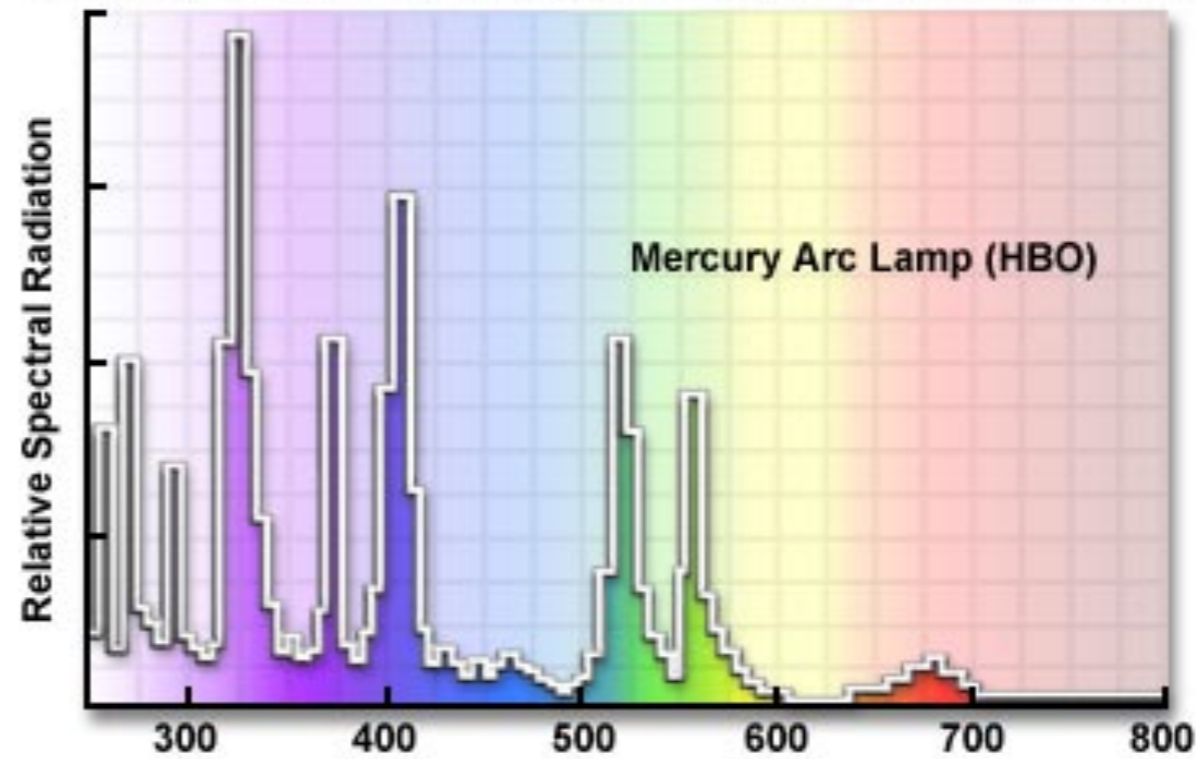
## Widefield fluorescence

### Arc Lamp Mercury

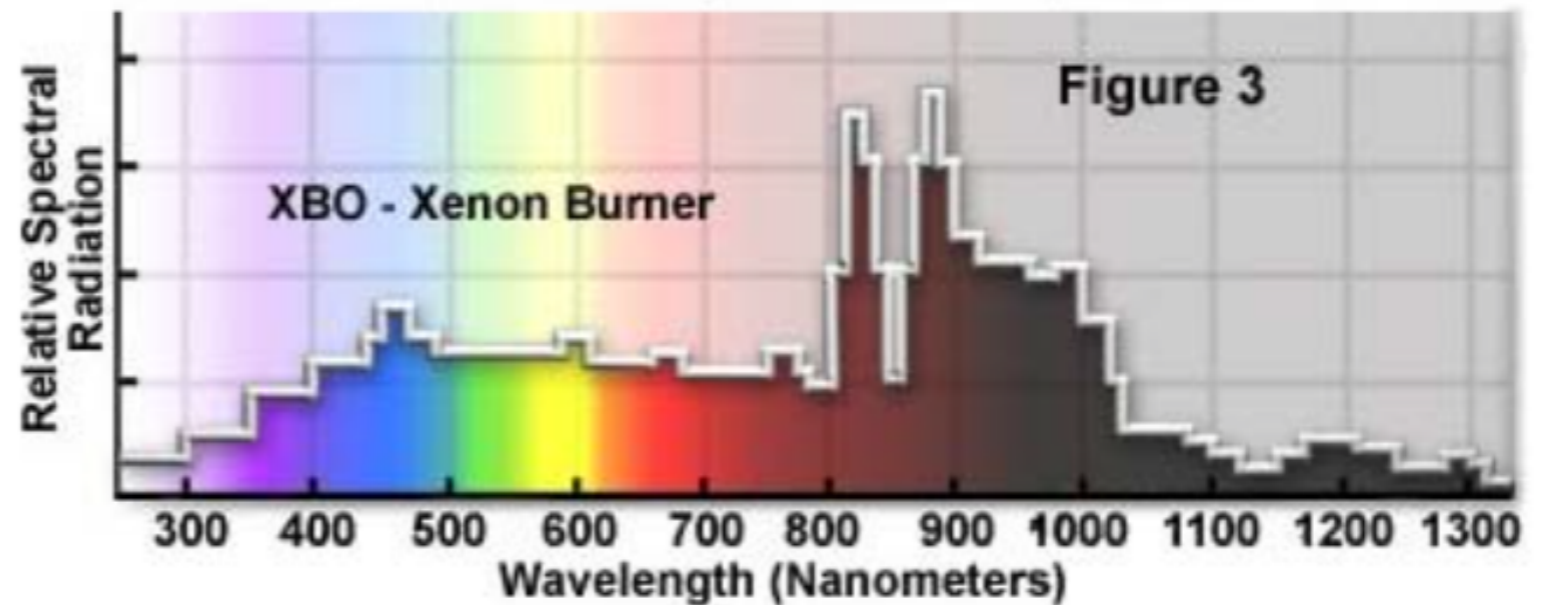
- 200h
- hazardous
- *out of use*



Mercury Arc Lamp UV and Visible Emission Spectrum



Xenon Arc Lamp Emission Spectrum



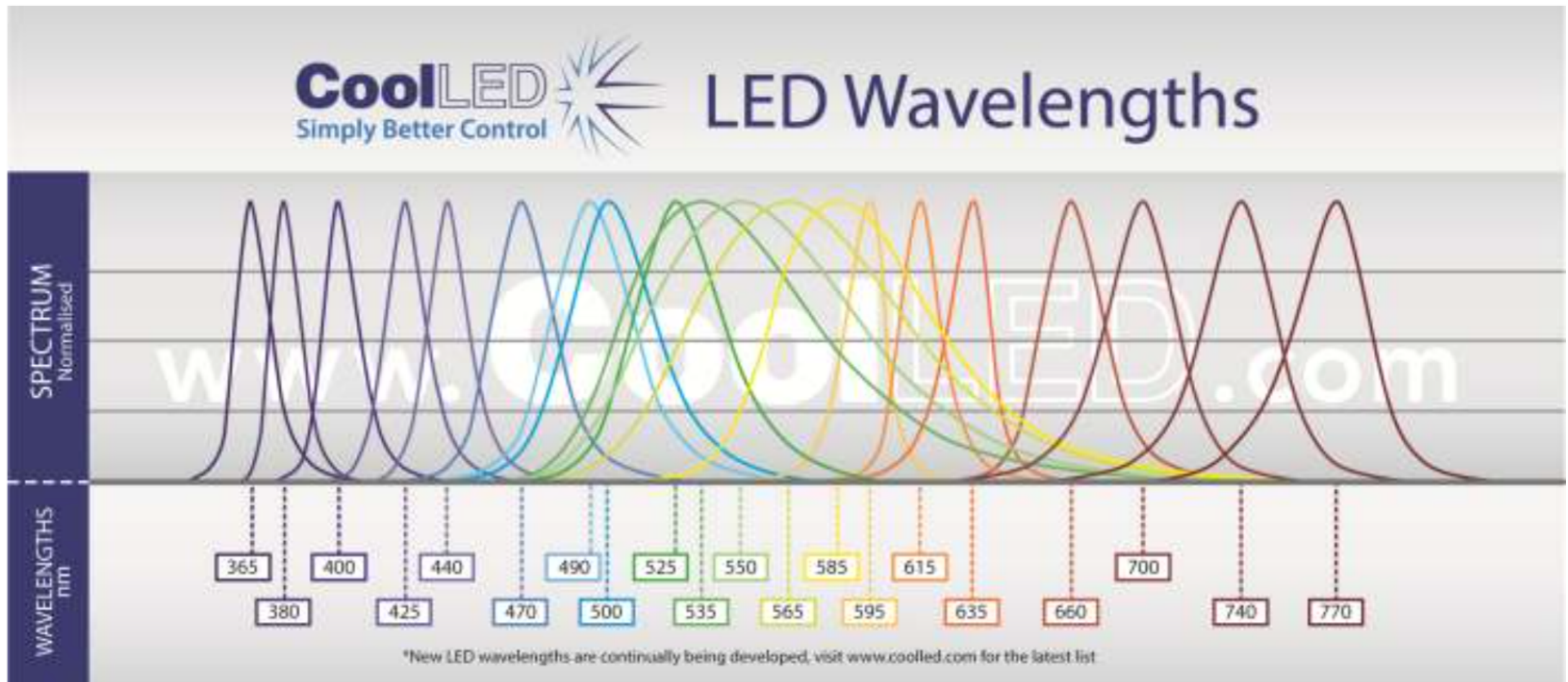
simultaneous excitation of multiple fluorophores over a wide wavelength range



# Illumination sources for widefield fluorescence microscopy

## State of the art for widefield fluorescence

## LEDs Light Emitting Diodes



<http://www.cooled.com/product-detail/led-wavelengths/>

- Wide range of lines available
- 25,000 h

# Illumination sources for fluorescence microscopy

Confocal

2-photon

TIRF

Super-resolution

Only discrete lines!

lines Alexa dye

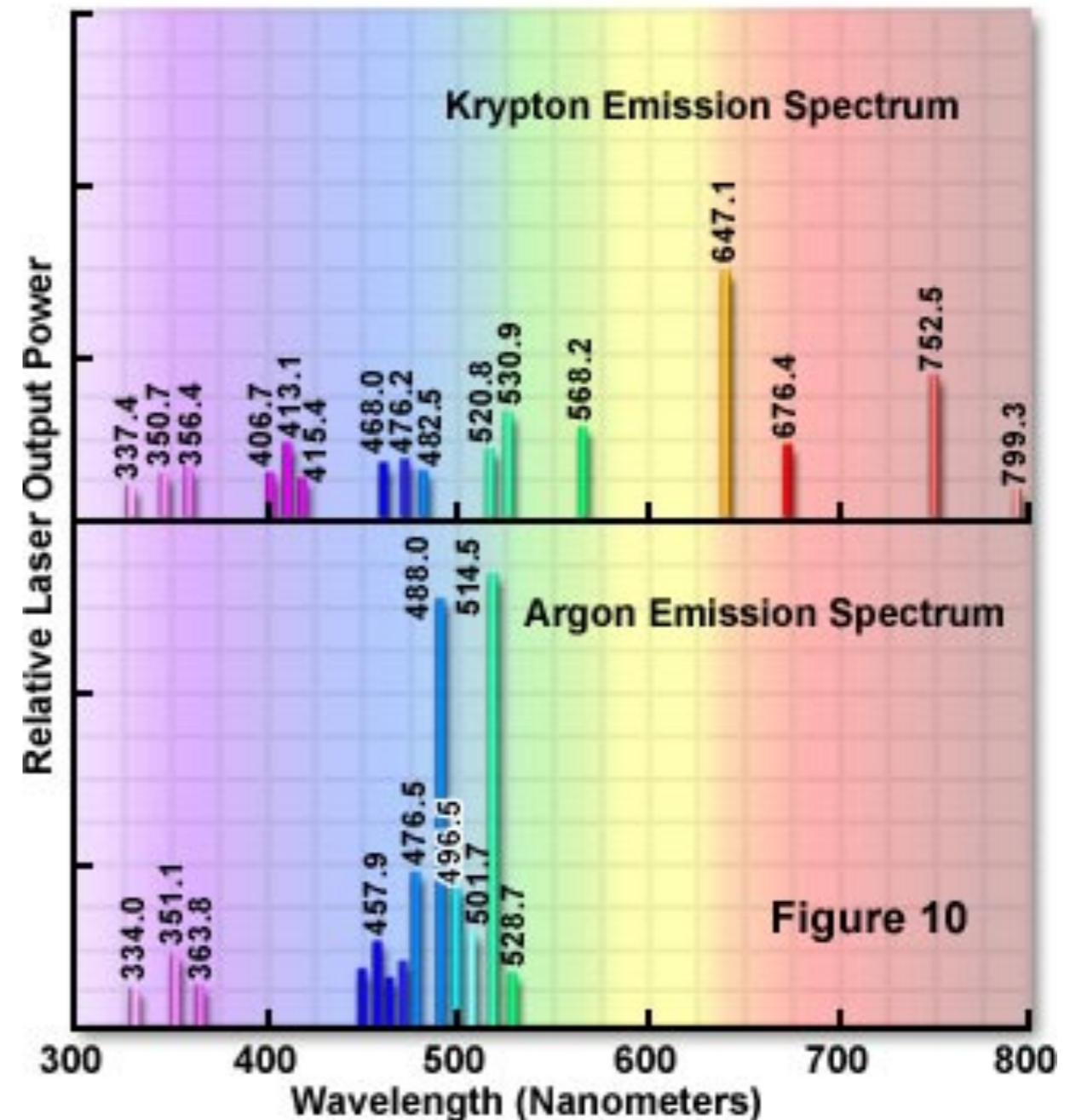
405	405	440
440	430	540
488	488	515
514	514	540
561	568	605
633	633	645

Narrow beams of highly monochromatic, coherent and collimated light

## Lasers

(light amplification by stimulated emission of radiation)

Laser Illumination Source Emission Spectra

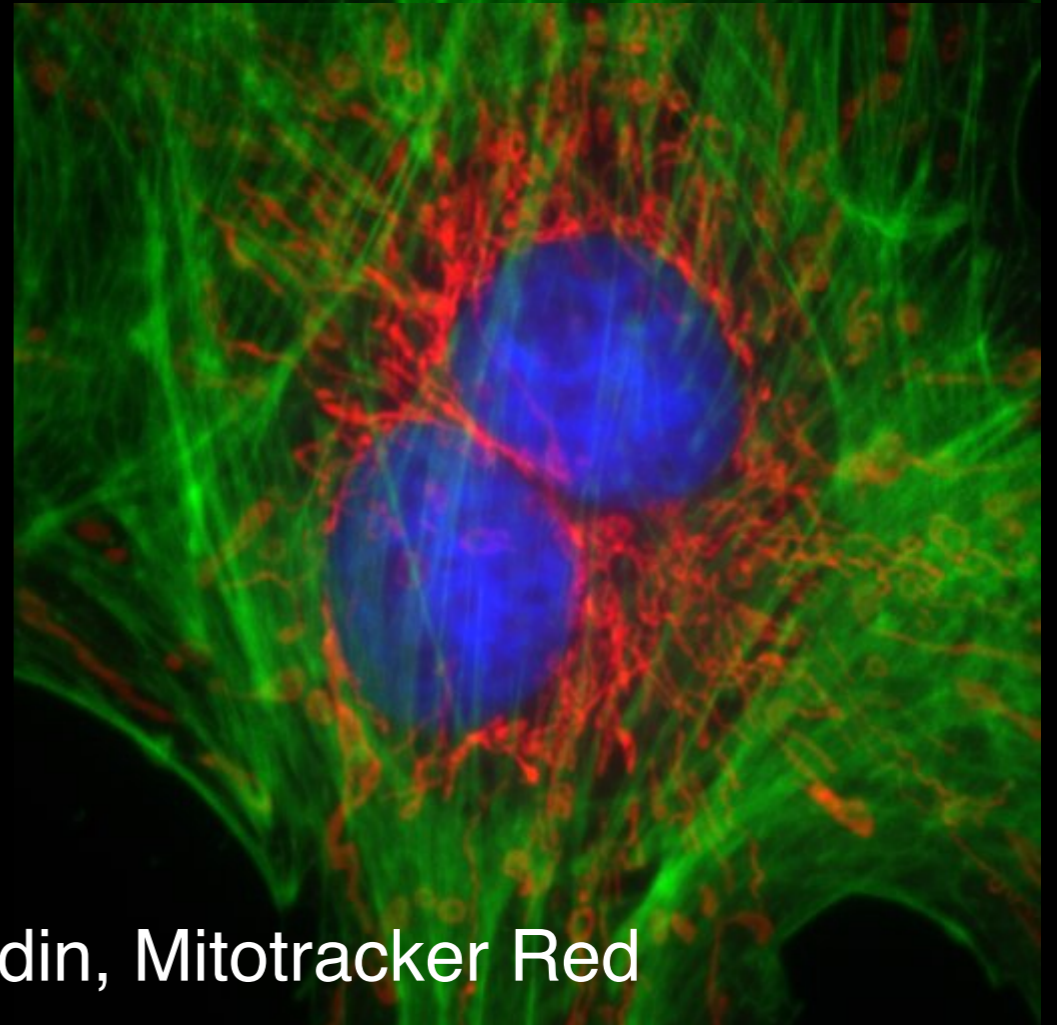
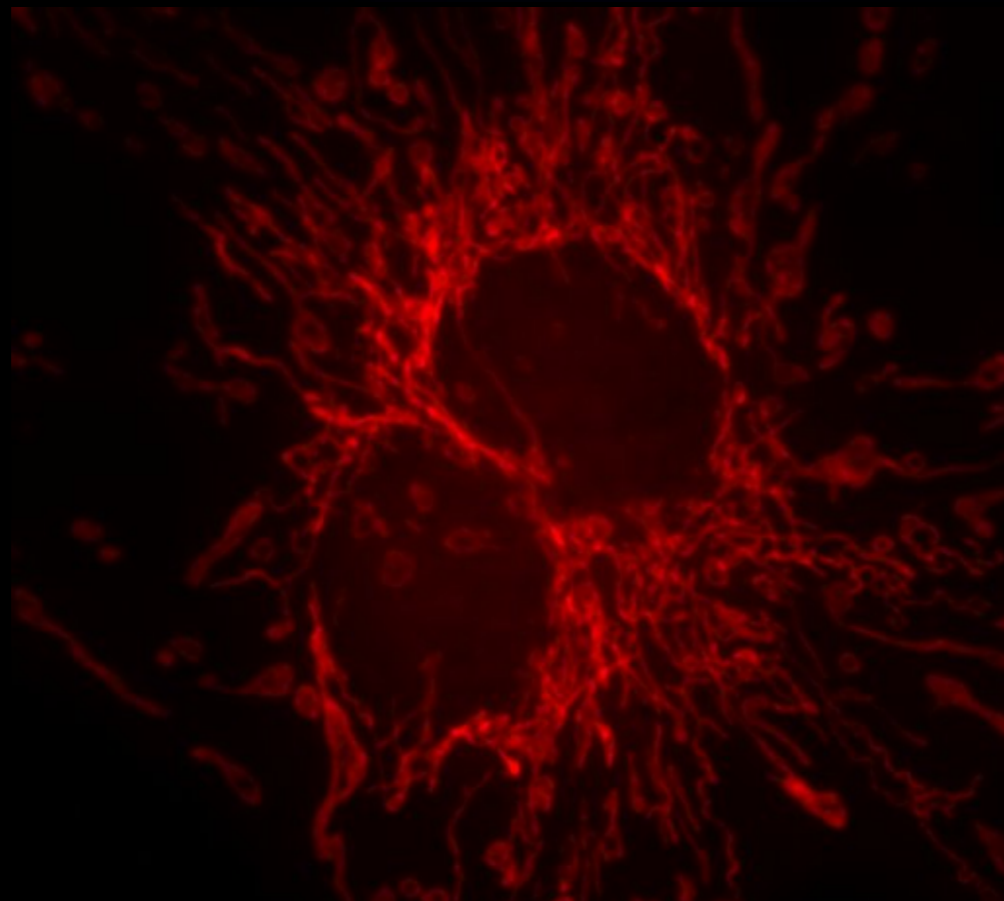
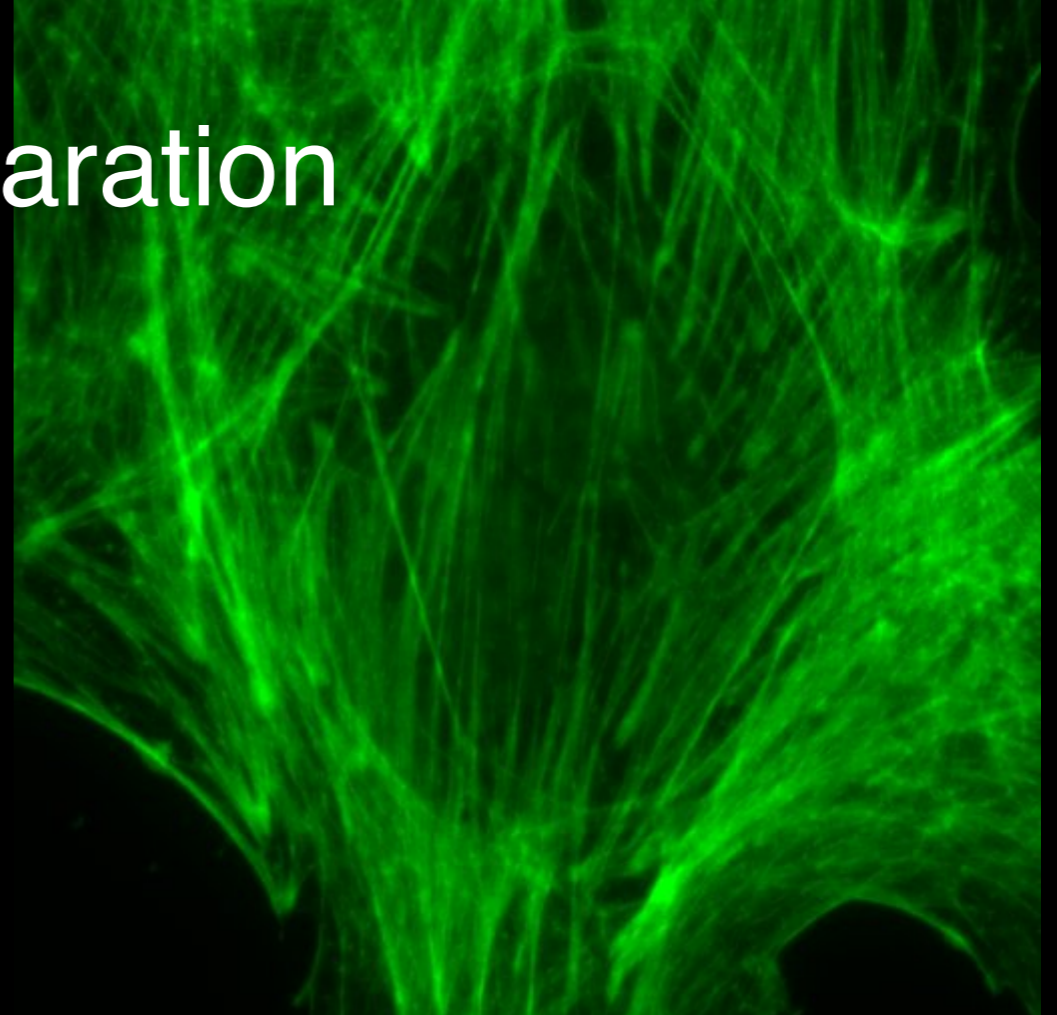
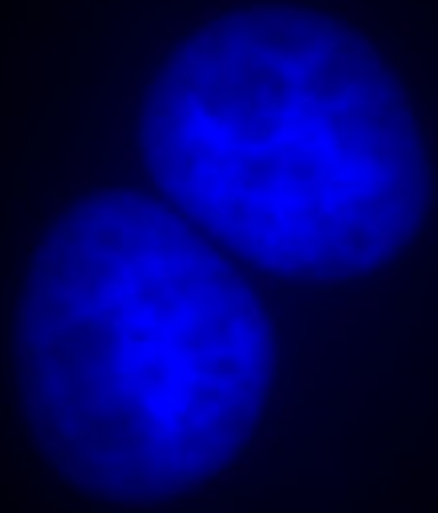




## Part 2 Tips on sample preparation

Fixed samples

(*in vivo* lecture 9)



BPAE cells stained with DAPI, Alexa 488 phalloidin, Mitotracker Red

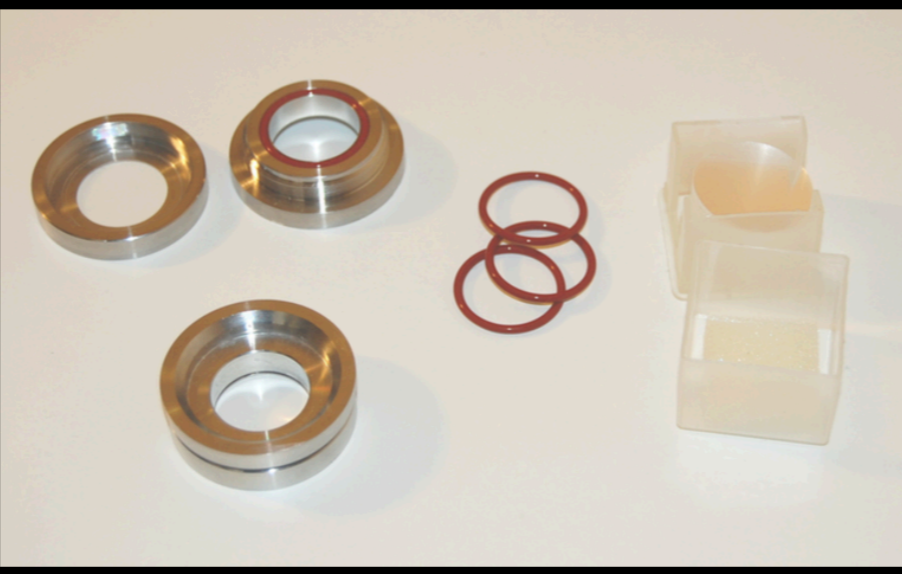
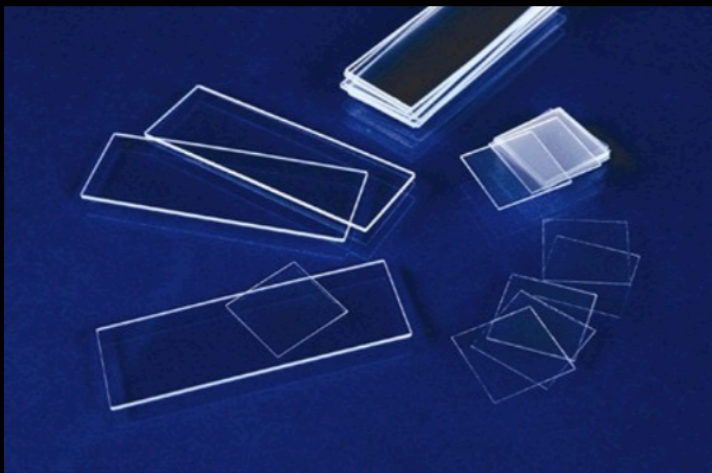


# Why work with fixed material?

1. Convenience / Throughput
2. Widely applicable molecular labeling:  
Immunofluorescence  
FISH
3. Ease of multiplexing bright stable labels

# Immobilising the specimen

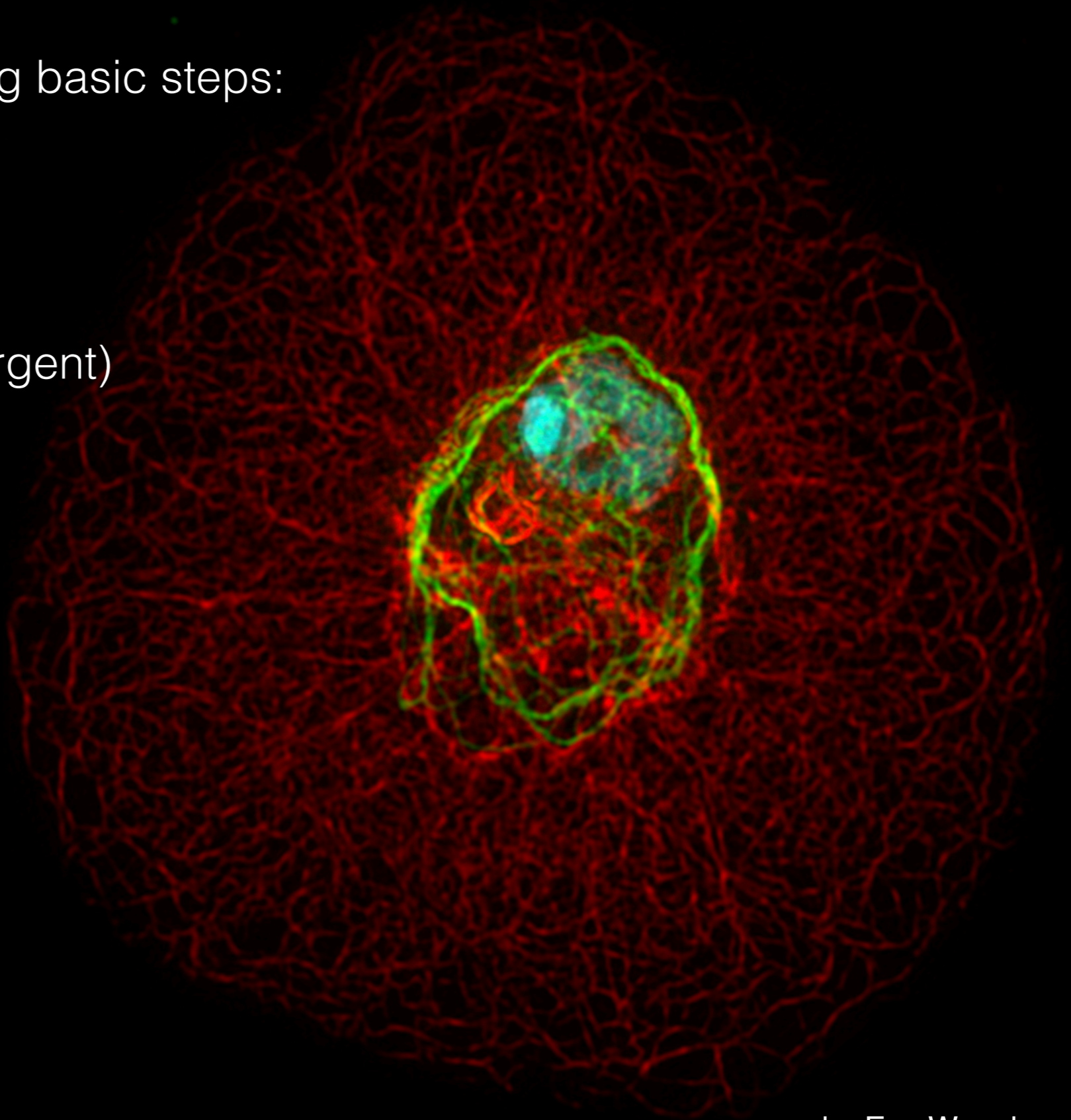
*Sample holder must be suitable for imaging*



# Typical Immunocytochemistry protocol

Most are variants of the following basic steps:

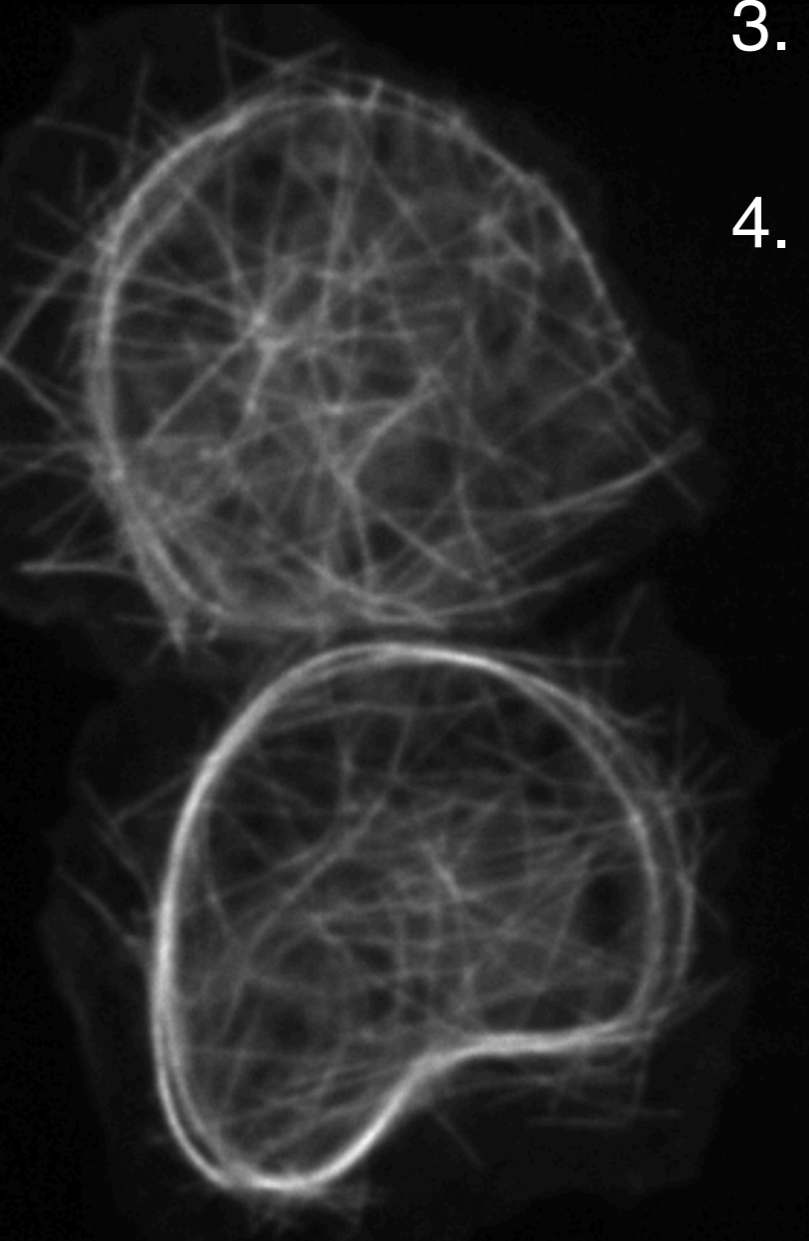
1. Fixation (e.g. PFA)
2. Permeabilisation (e.g. detergent)
3. Washes (e.g. PBS)
4. Blocking (e.g. serum)
5. 1<sup>o</sup> antibody
6. Washes (e.g. PBS)
7. 2<sup>o</sup> antibody
8. Washes (e.g. PBS + H<sub>2</sub>O)
9. Mounting (e.g. Vectashield)





# Fixation: preservation of cells or tissue in a life-like state

1. Preserve structural features
2. Uniform fixation throughout the sample
3. Enable dye labeling
4. Reduce background fluorescence



Microtubules in *Drosophila* macrophages

Left :

Live cells expressing Jupiter-GFP

Right:

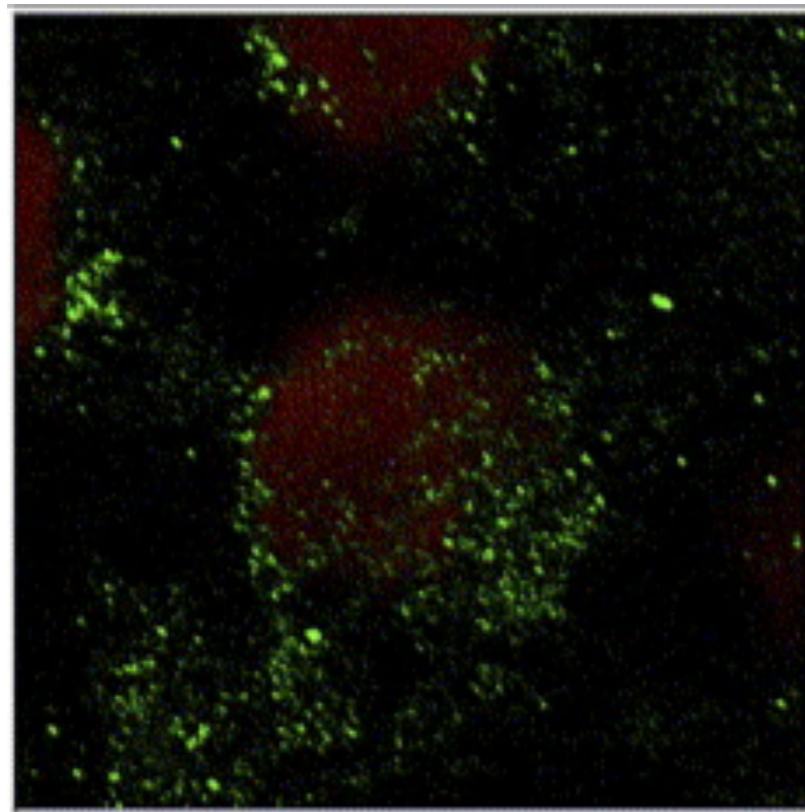
PFA fixed cell stained with anti-tubulin antibody and Alexa Fluor 488



# Types of Fixation

## Denaturing fixation:

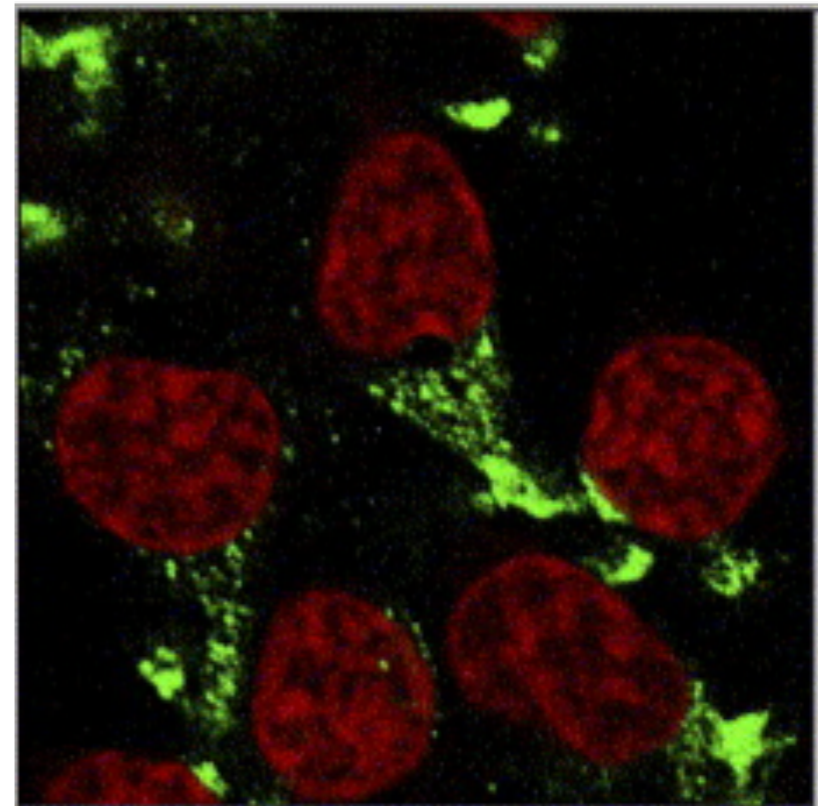
Cold methanol



destroys 3D protein structure  
dissolves lipids into micelles

## Cross-linking fixation:

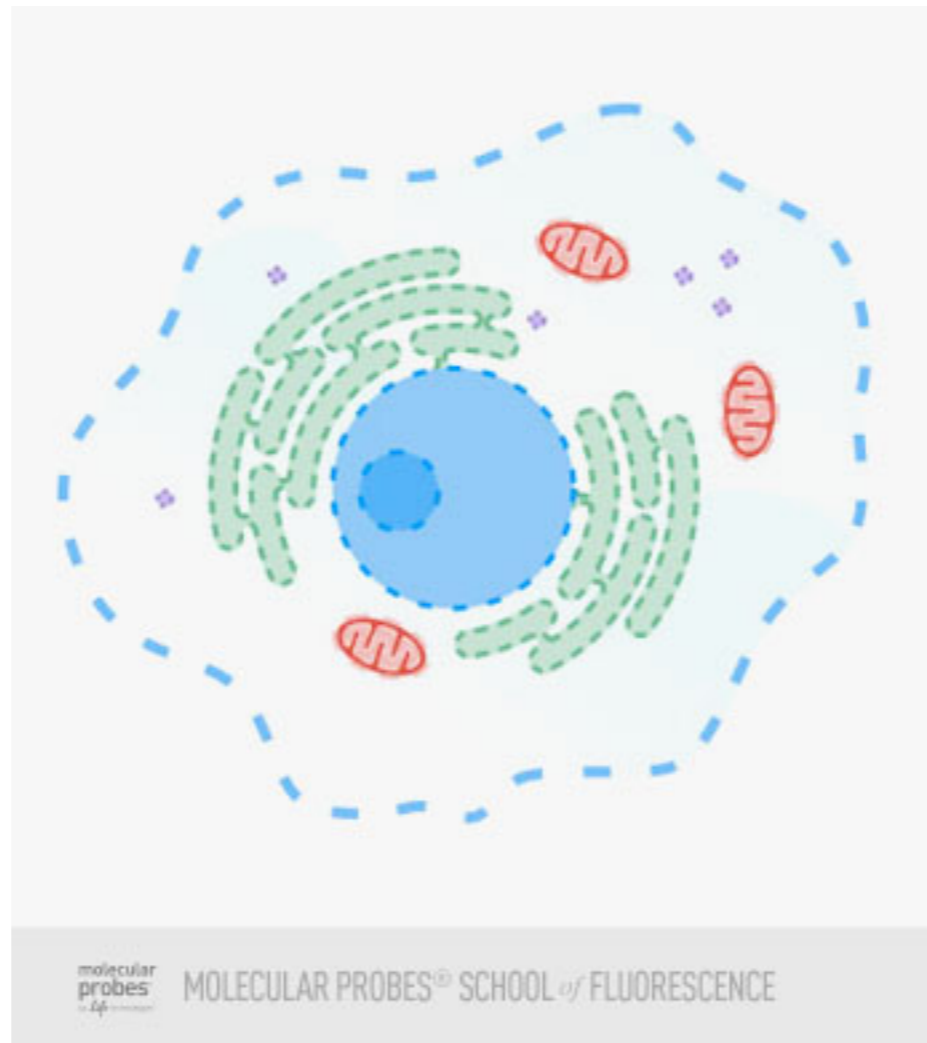
Formaldehyde (PFA)



binds to proteins and some lipids,  
but not RNA, DNA or most sugars

Sometimes a combination of both is necessary ...

## Removal of some lipids with detergents



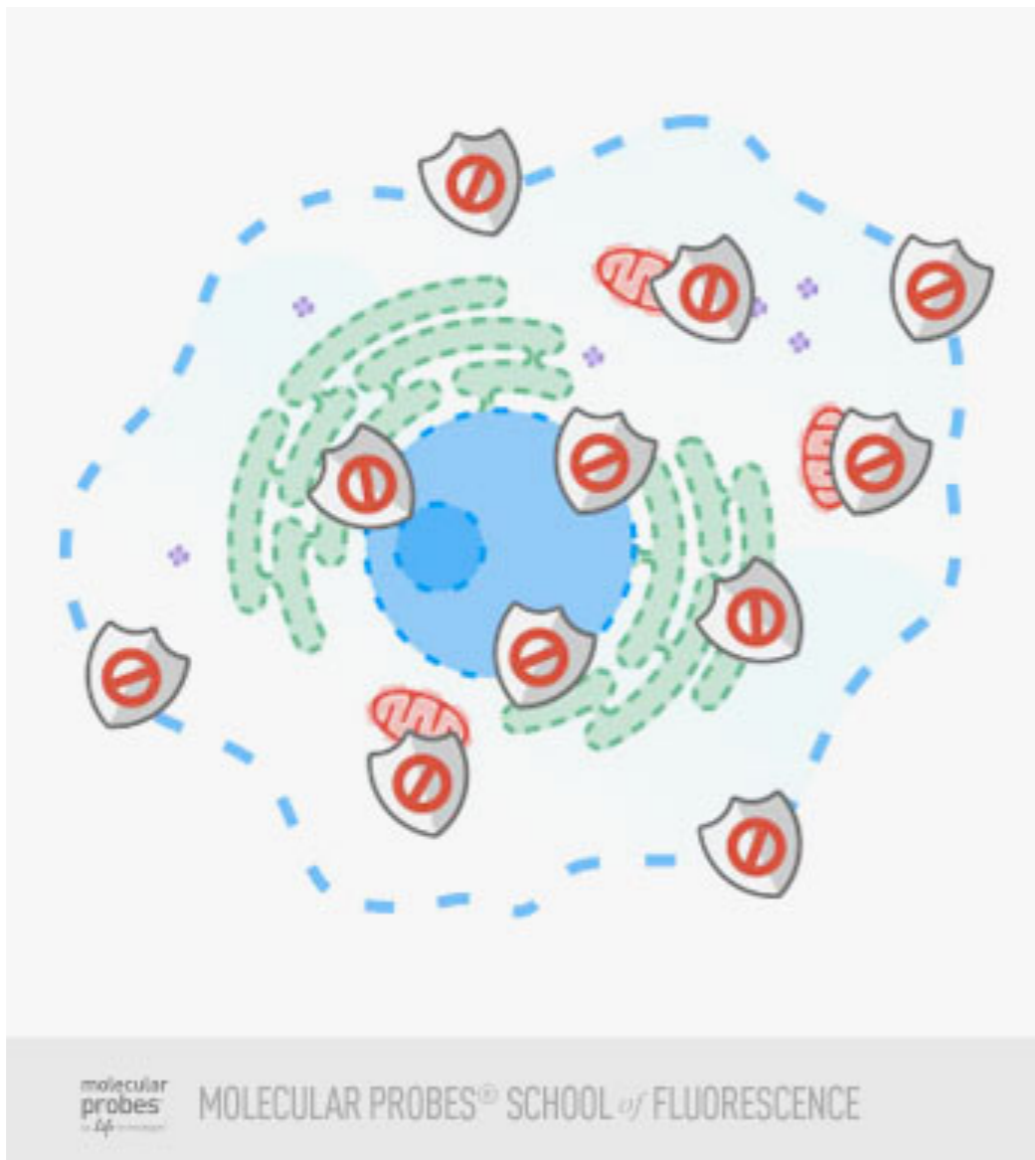
**Tween 20**

**Triton X-100**

Goal: to allow large labels (antibodies) to penetrate fixed cells/tissue

## Reduction of nonspecific staining

done with a solution containing excess of protein



**Bovine Serum Albumin (BSA )**

**Casein (or non-fat dry milk)**

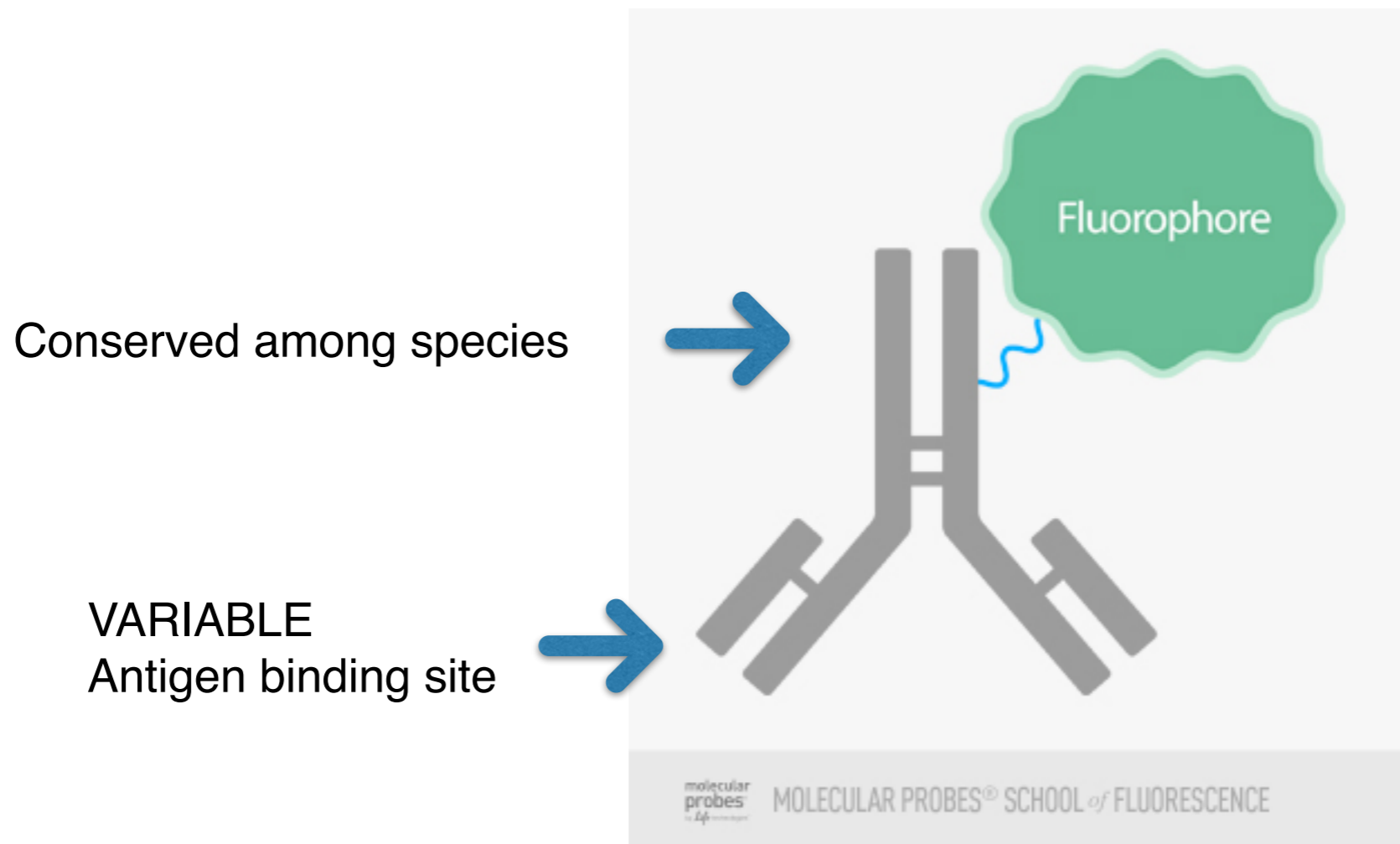
blocking proteins prevent low-affinity antibody interactions elsewhere in the sample



# Immunolabeling (antibodies)

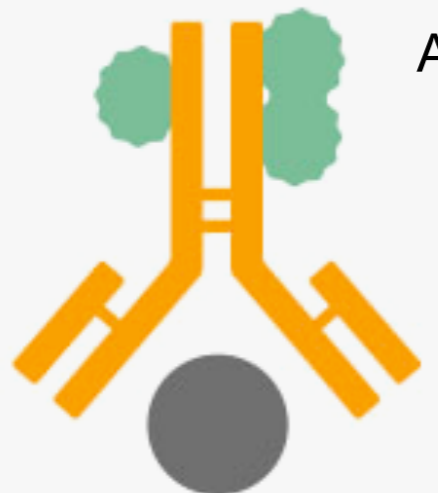
**Antibody** (large Y-shaped protein called immunoglobulin) produced by the immune system, found in the blood or other body fluids of **vertebrates**.

The **antibody** recognises unique parts of the foreign target called an **antigen**.



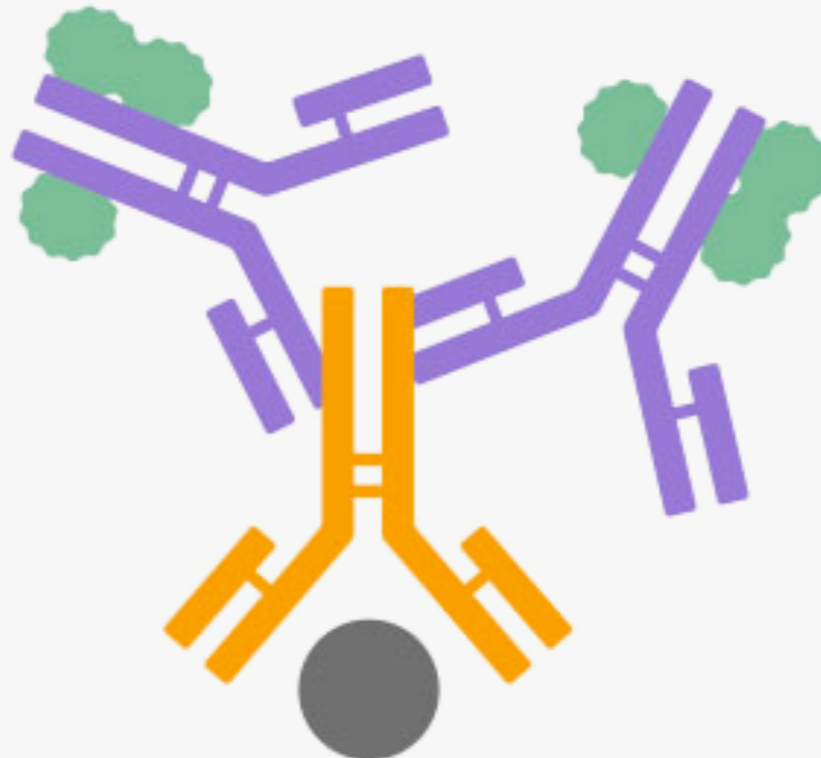
# Immunolabeling (antibodies)

Direct immunofluorescence



Alexa 488

Indirect immunofluorescence or secondary detection



mouse anti-tubulin



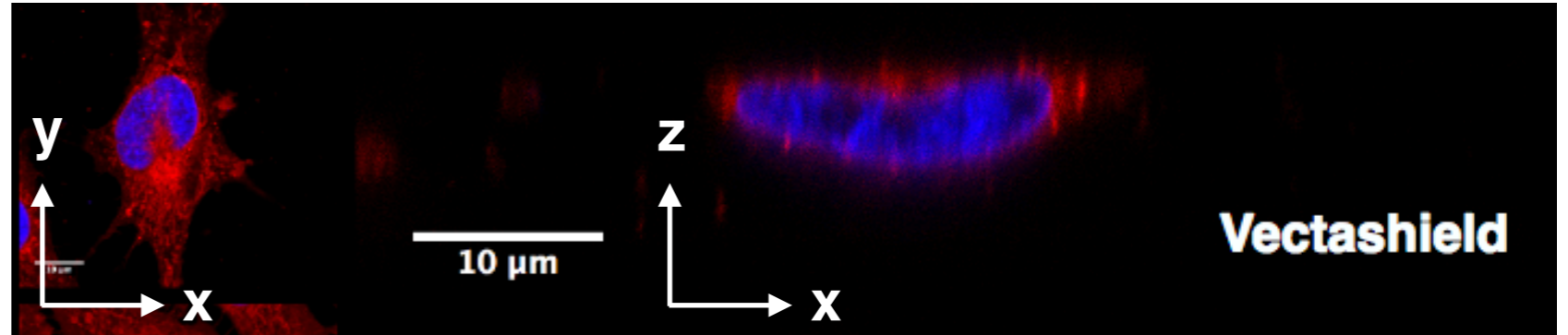
goat anti-mouse



tubulin

# Mounting

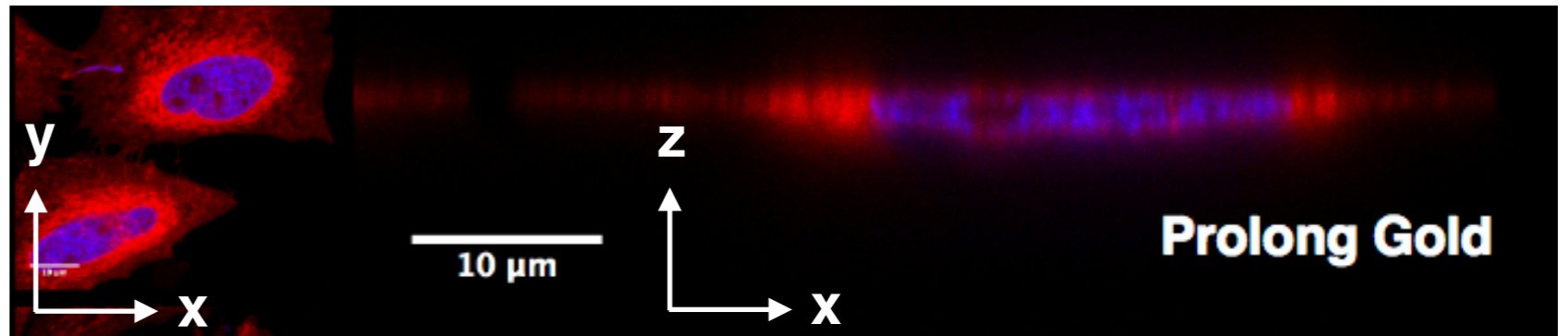
- Non-hardening



- Short-term storage (days to few weeks)

- **Dabco, Glicerol, Vectashield** (antifading agent, but does not work with FarRed dyes)

- Hardening



- Long term storage (months)

- It can flatten the cell if polymerises too fast

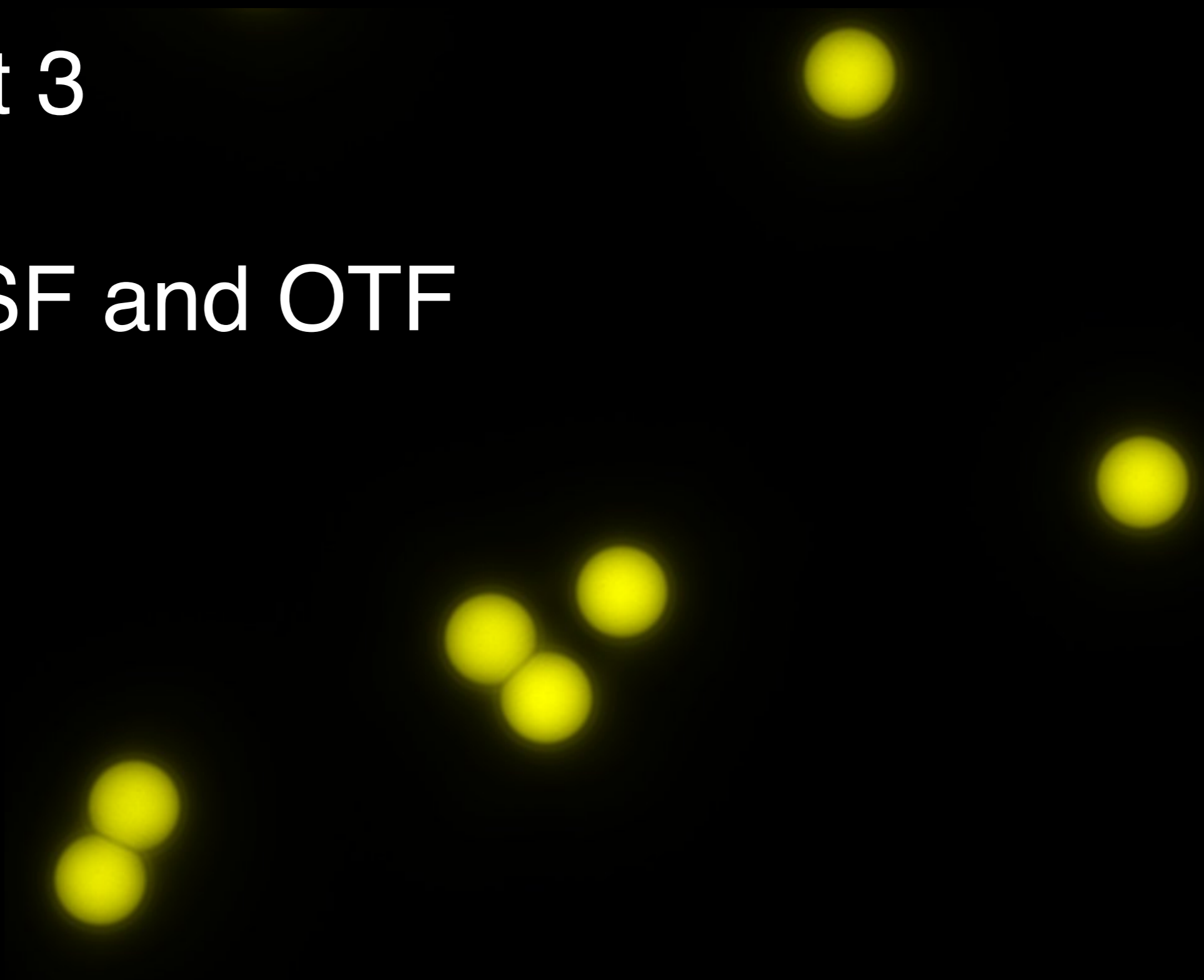
- **Prolong Gold, Prolong Glass, Vectashield harset**

# Experimental controls: the key for reliable results

- No primary or secondary antibody (autofluorescence)
- Incubate with secondary but not primary antibody
- Check cross-talk between dyes and microscope filterset
- Test specificity in knock-out /knock-down cells

# Part 3

## PSF and OTF



## Point Spread Function

**PSF is a measure of the microscope response to a point source of light**

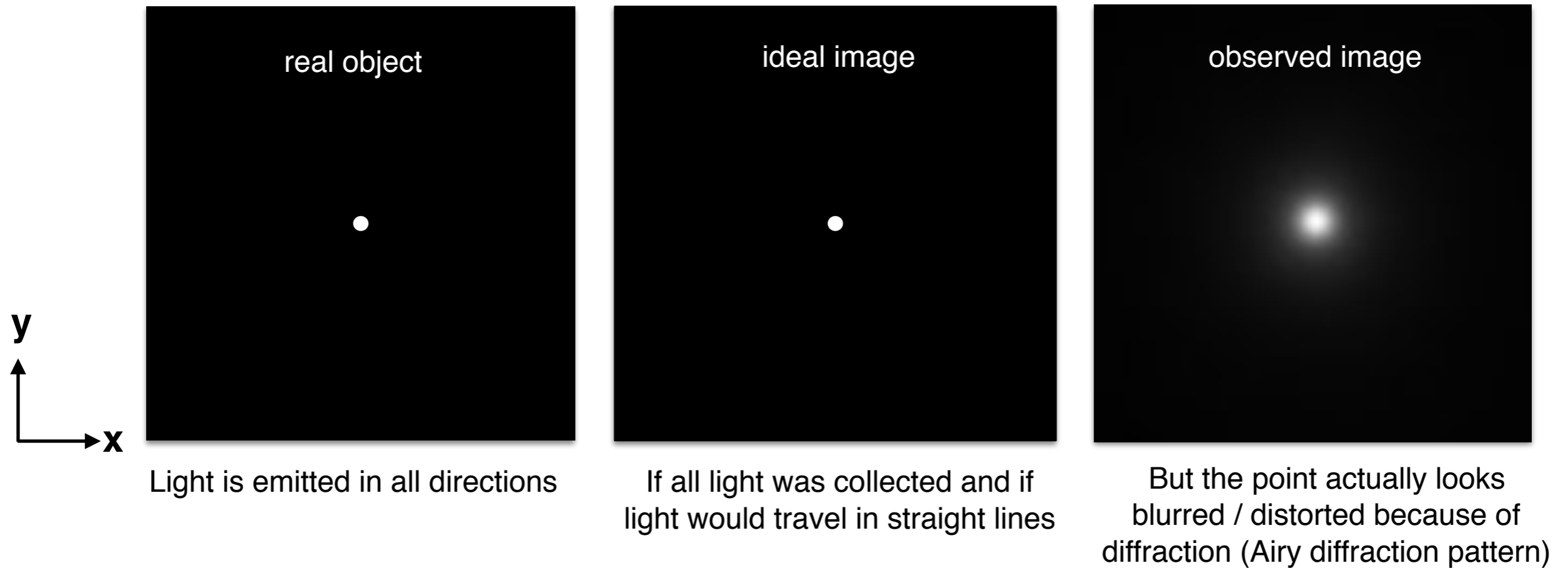
Why bother?

- microscope performance
- spherical aberrations
- x, y, z info
- image quality
- alignment
- optical resolution

# PSF (Point Spread Function) in fluorescence

## Point Spread Function

How does light spread out from a single point?



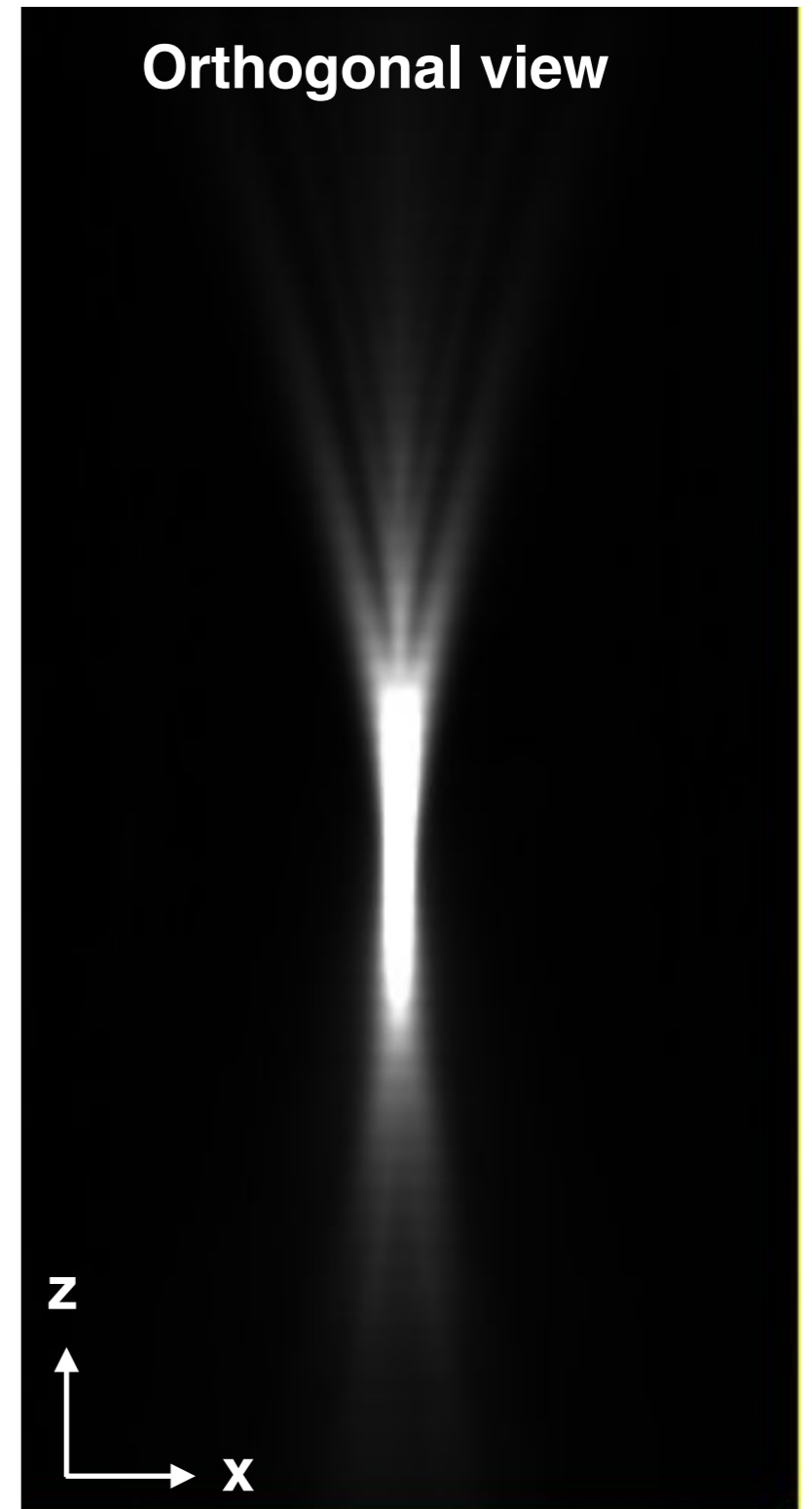
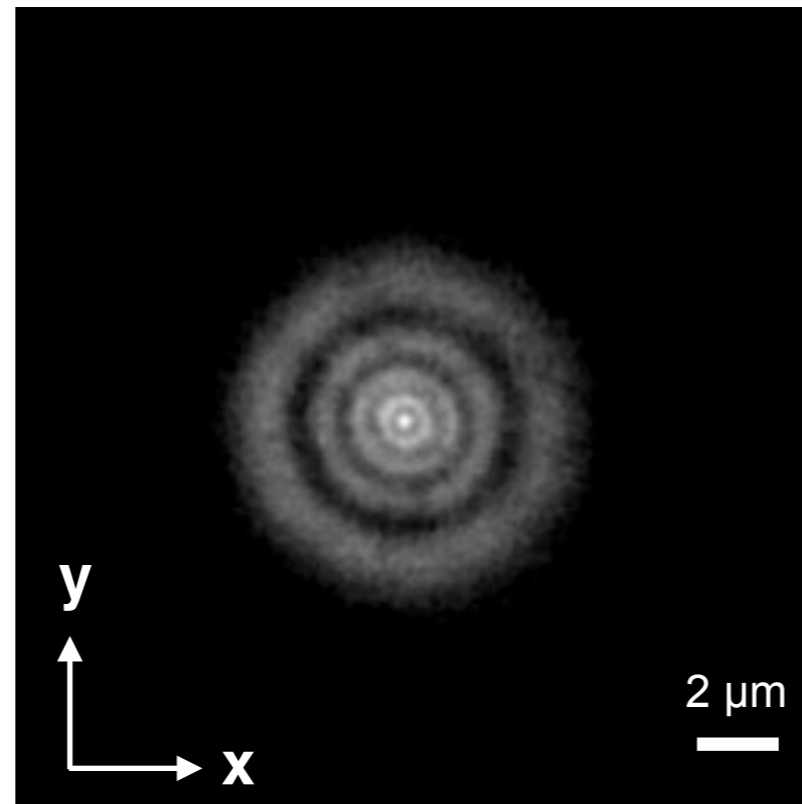
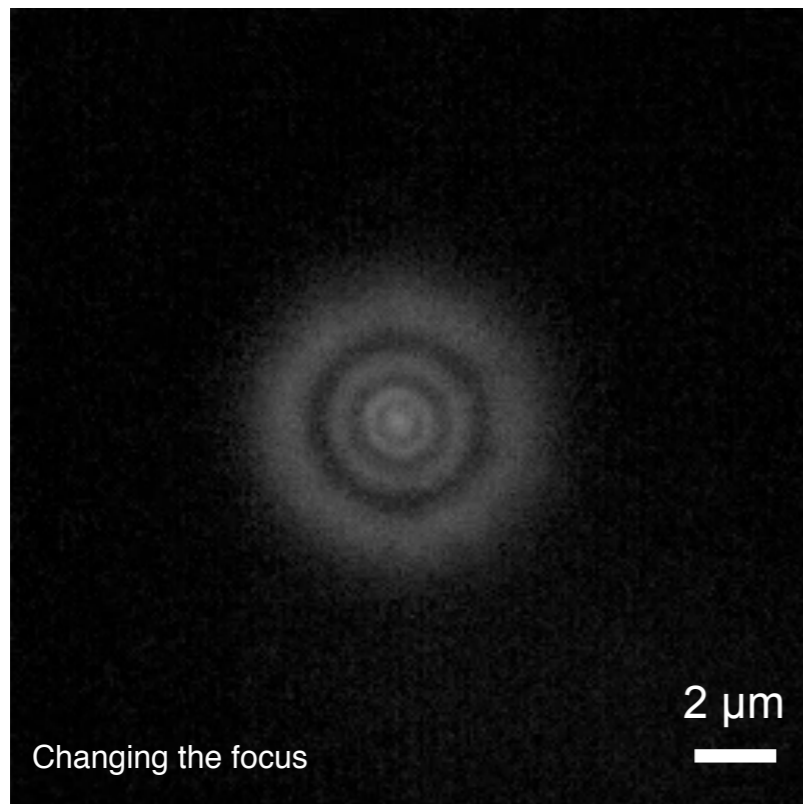
**Fluorescent bead, single dye, or a fluorescent protein as a point source of light**



# PSF (Point Spread Function) in fluorescence

## PSF

red fluorescent 170 nm bead



## Airy disk diffraction pattern

(concentric rings)

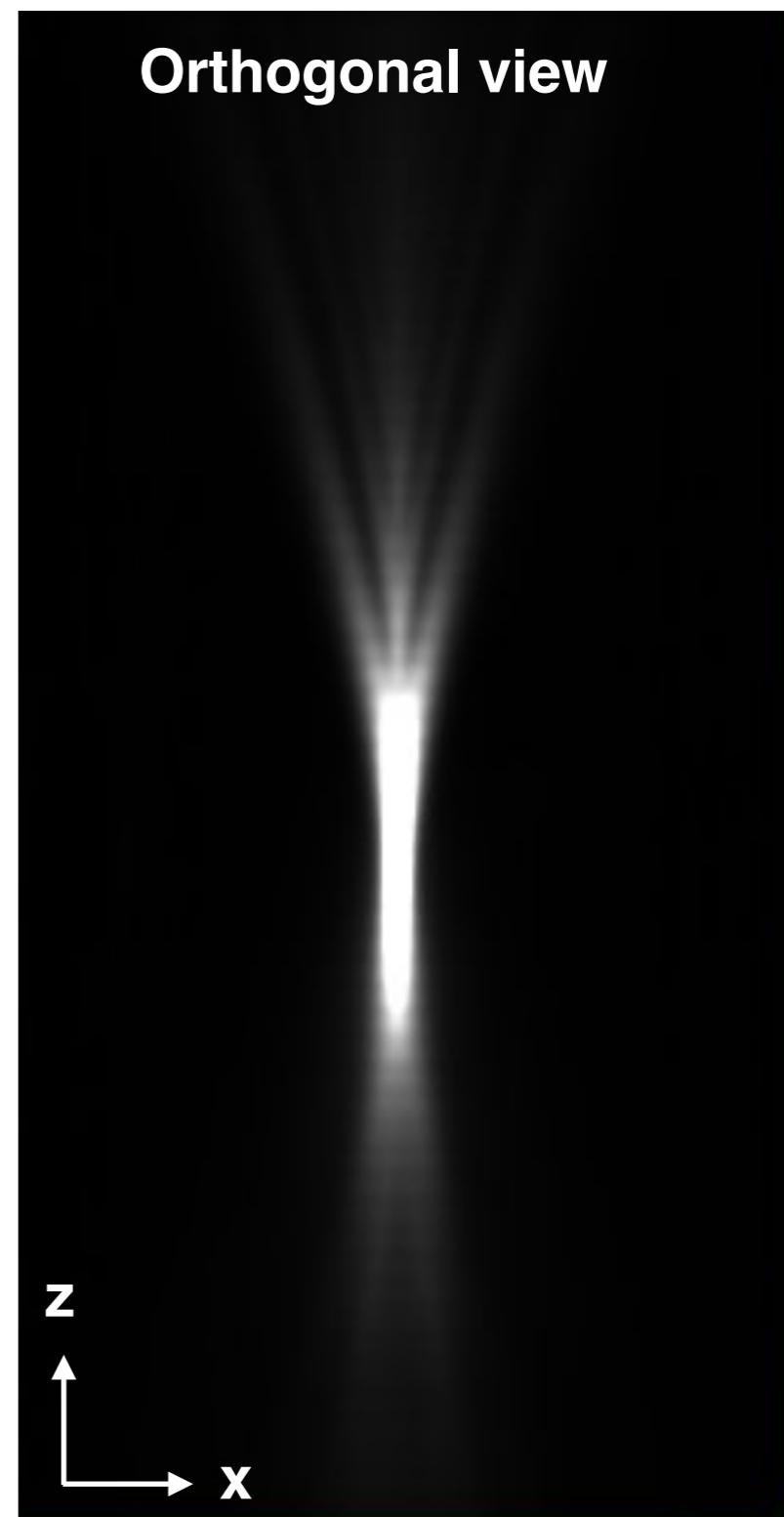
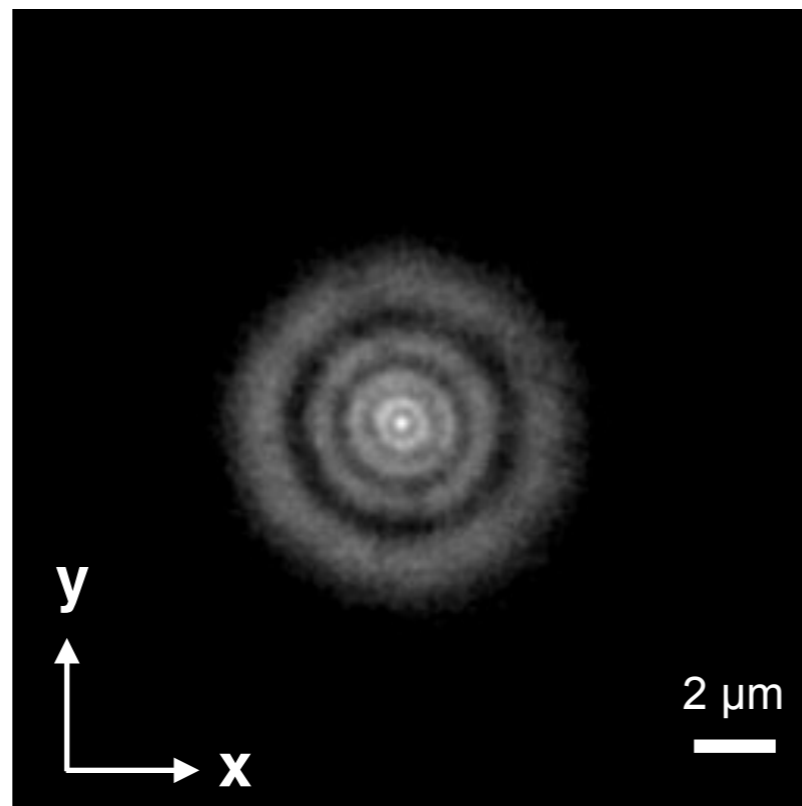
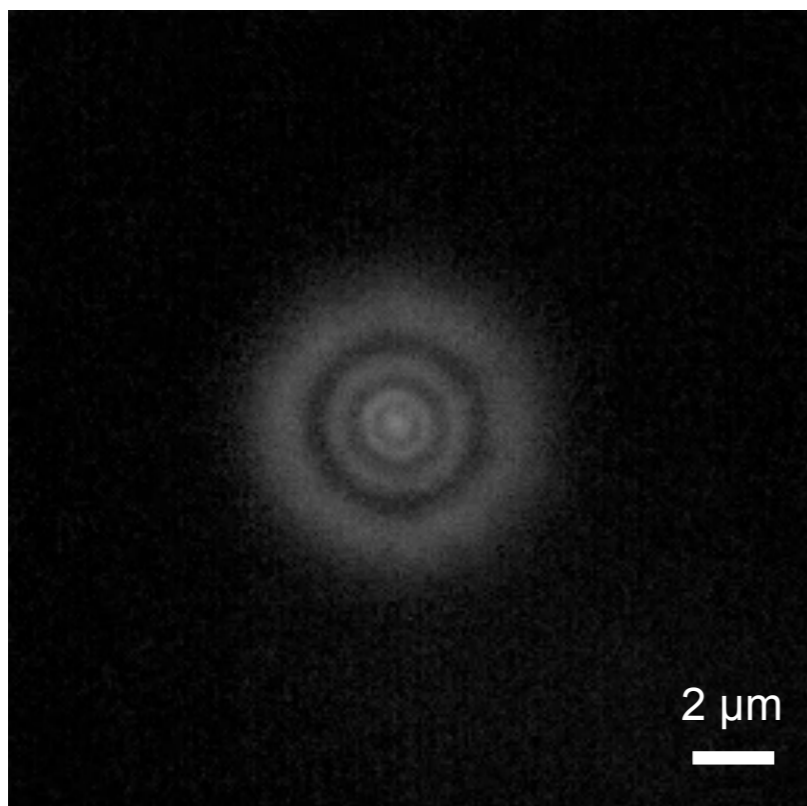
Light waves emitted from a point source are not focused into an infinitely small point by the objective

They converge together and interfere in the image plane

# PSF is the 3D image of a point-like object under the microscope

## PSF

red fluorescent 100 nm bead



## What can we observe?

- Blur is broader in z than xy  *RESOLUTION*
- How symmetric is the distribution

ALIGNMENT, SPHERICAL ABERRATIONS, MISMATCH REFRACTIVE INDEX

# Why blurred and how is the Airy diffraction pattern generated?

Objective lens

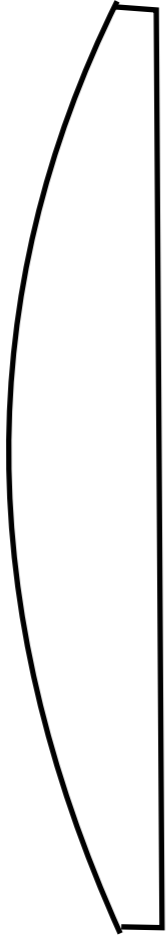
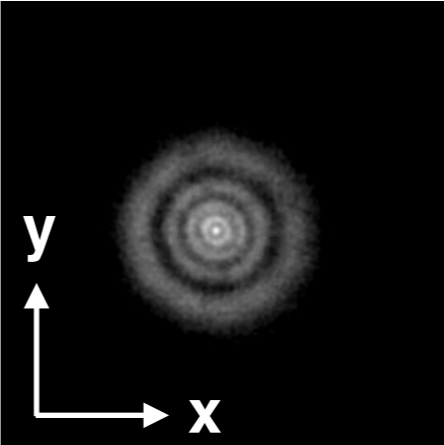
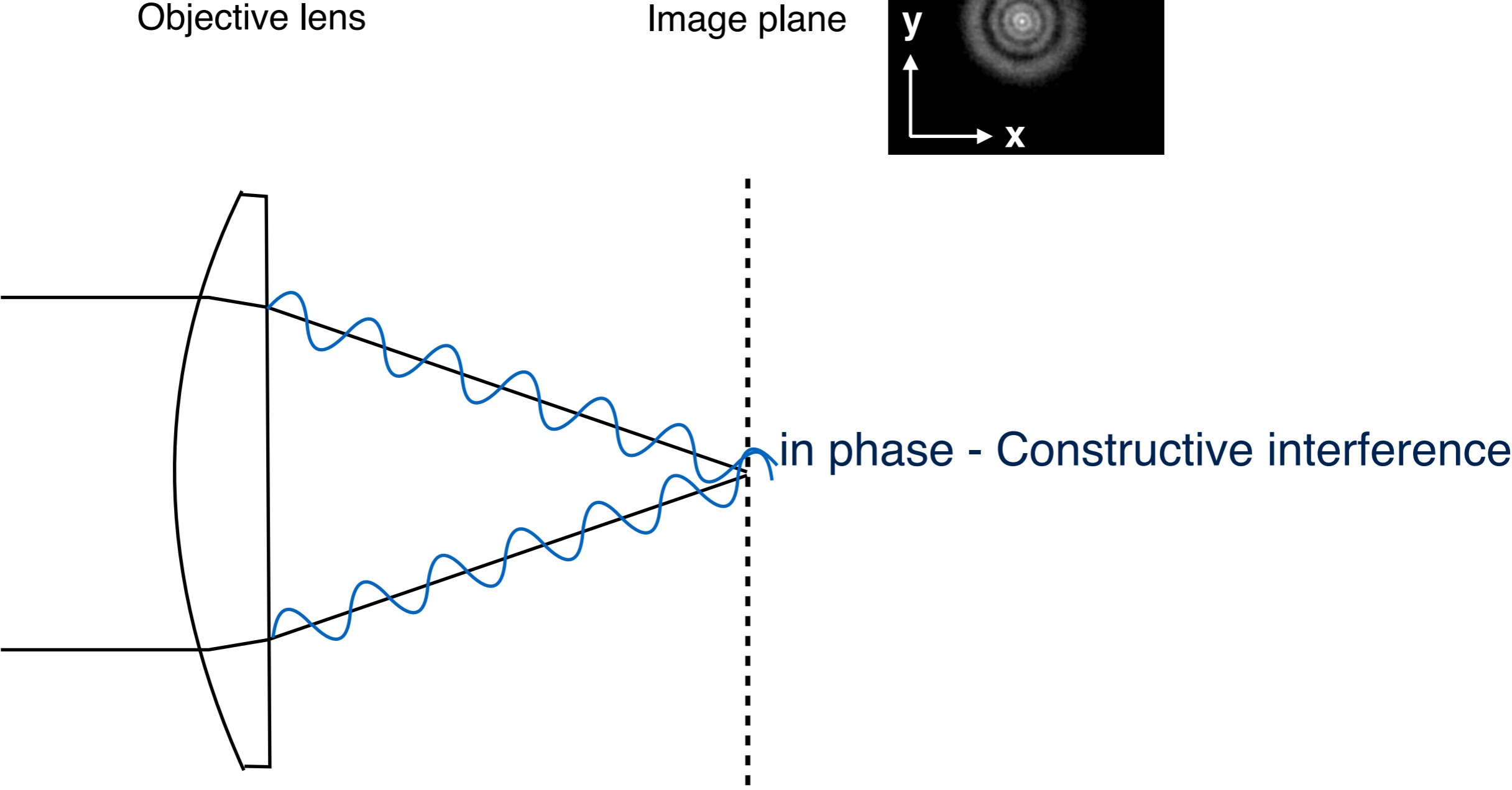


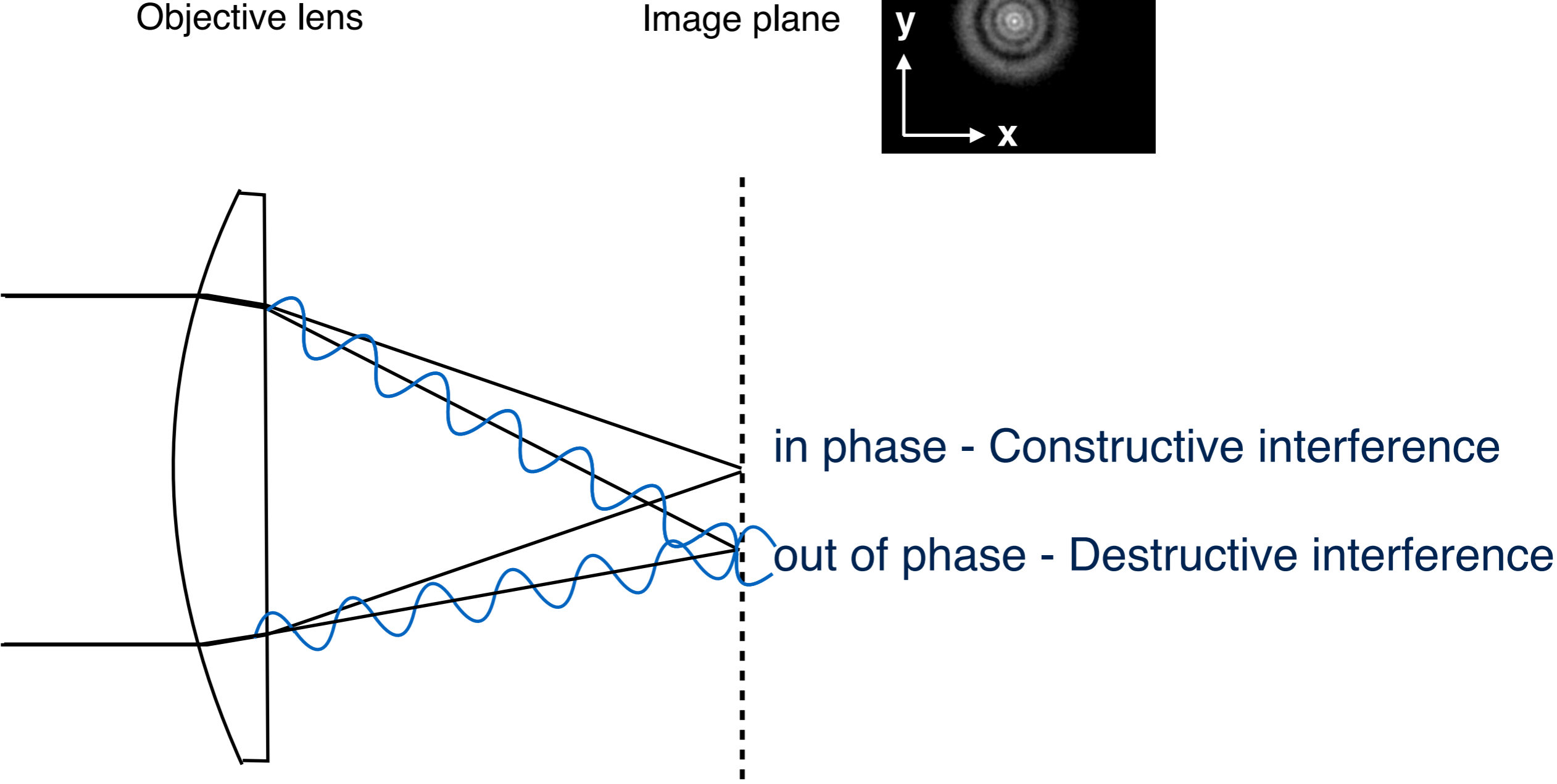
Image plane



# Why blurred and how is the Airy diffraction pattern generated?

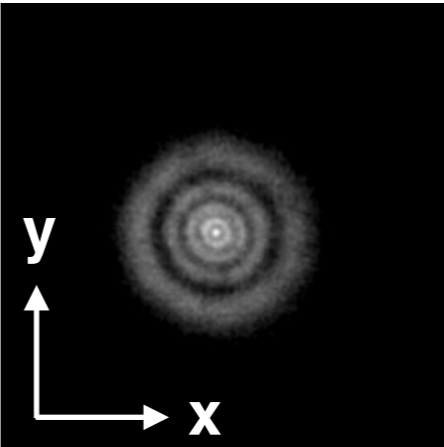
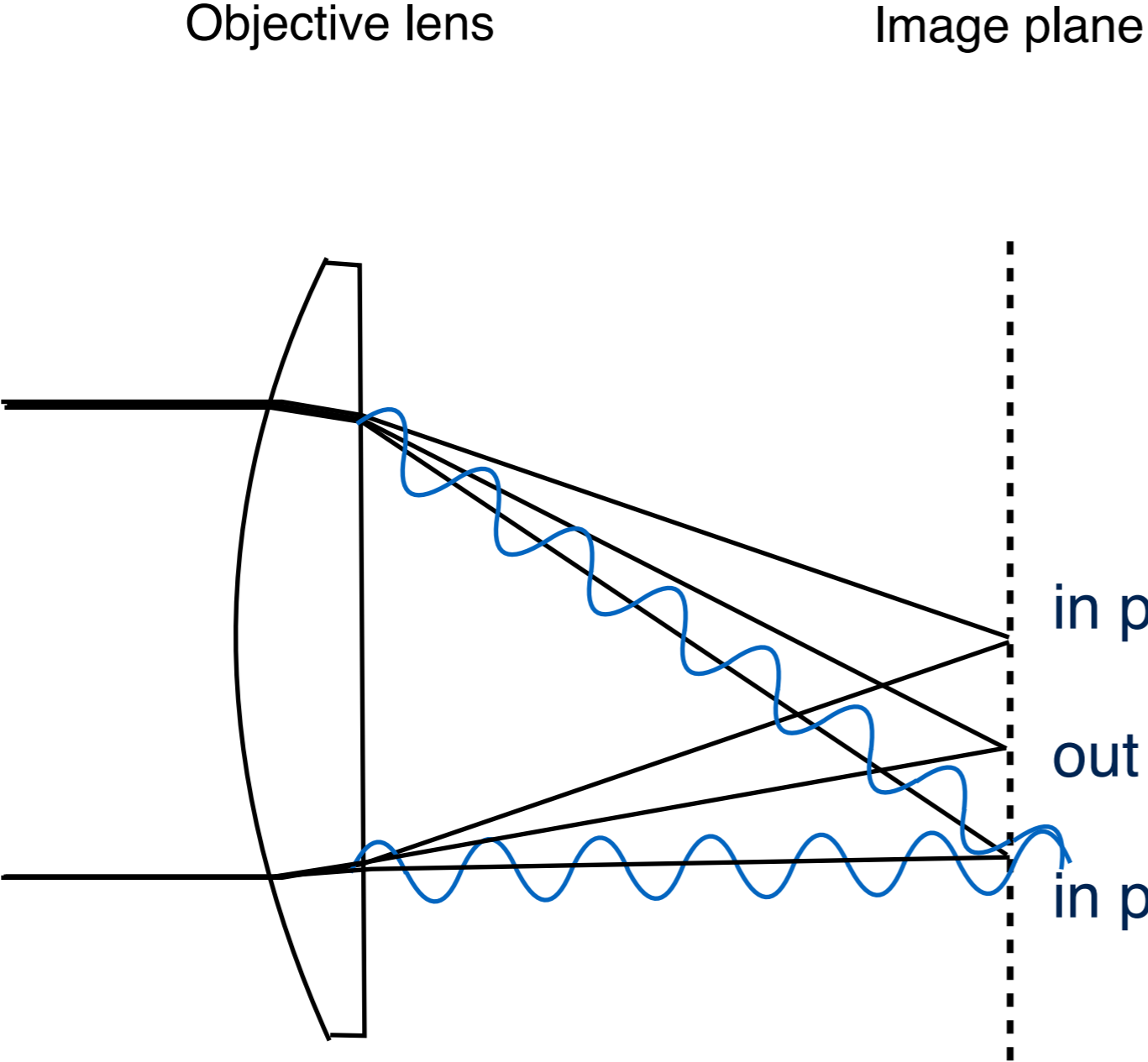


# Why blurred and how is the Airy diffraction pattern generated?





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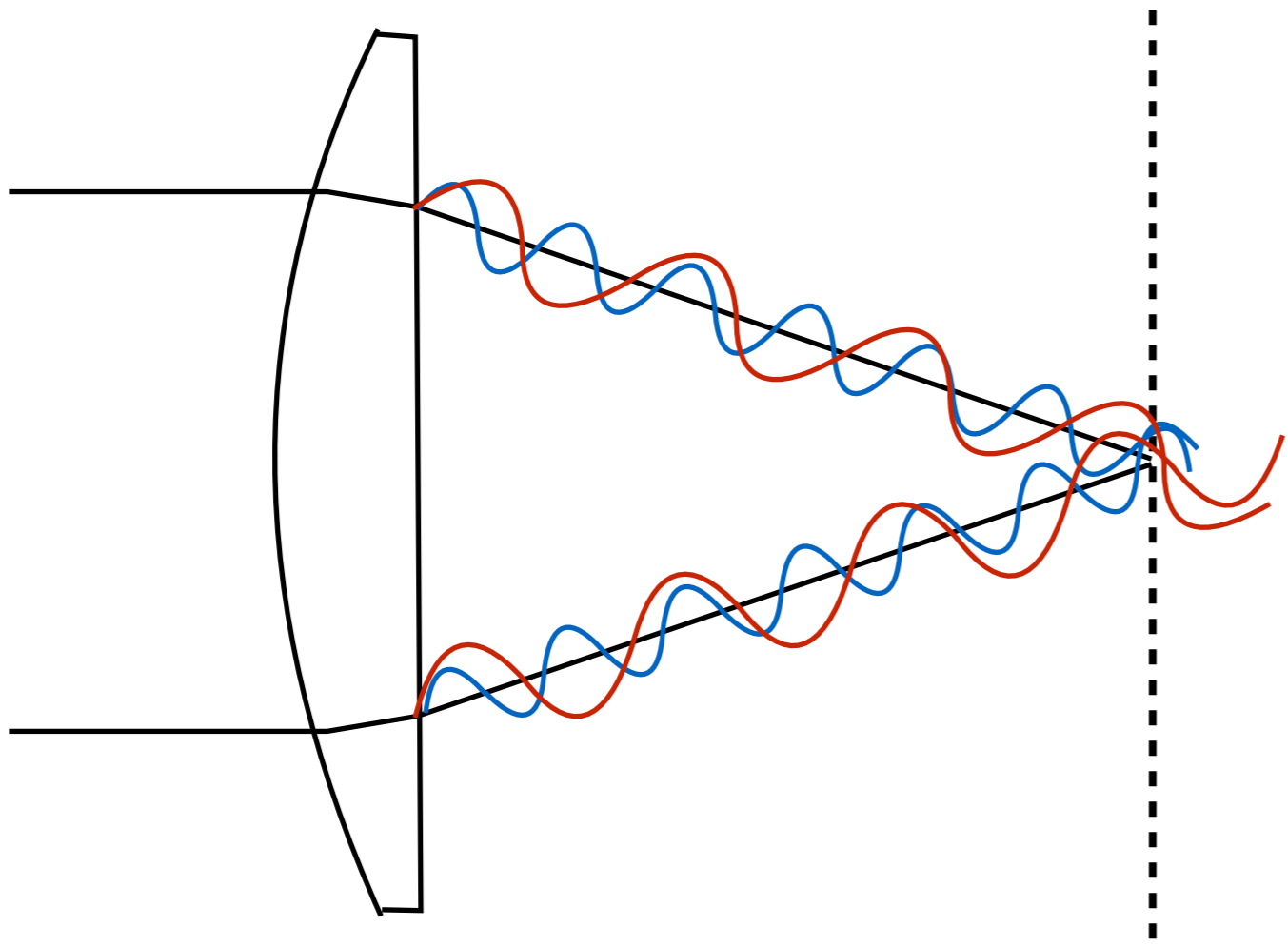
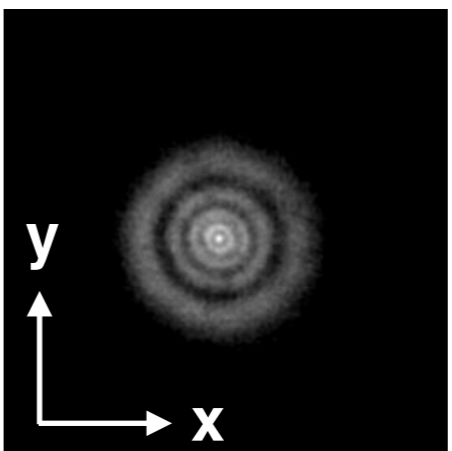


in phase - Constructive interference  
out of phase - Destructive interference  
in phase - Constructive interference

# What does depend on...?

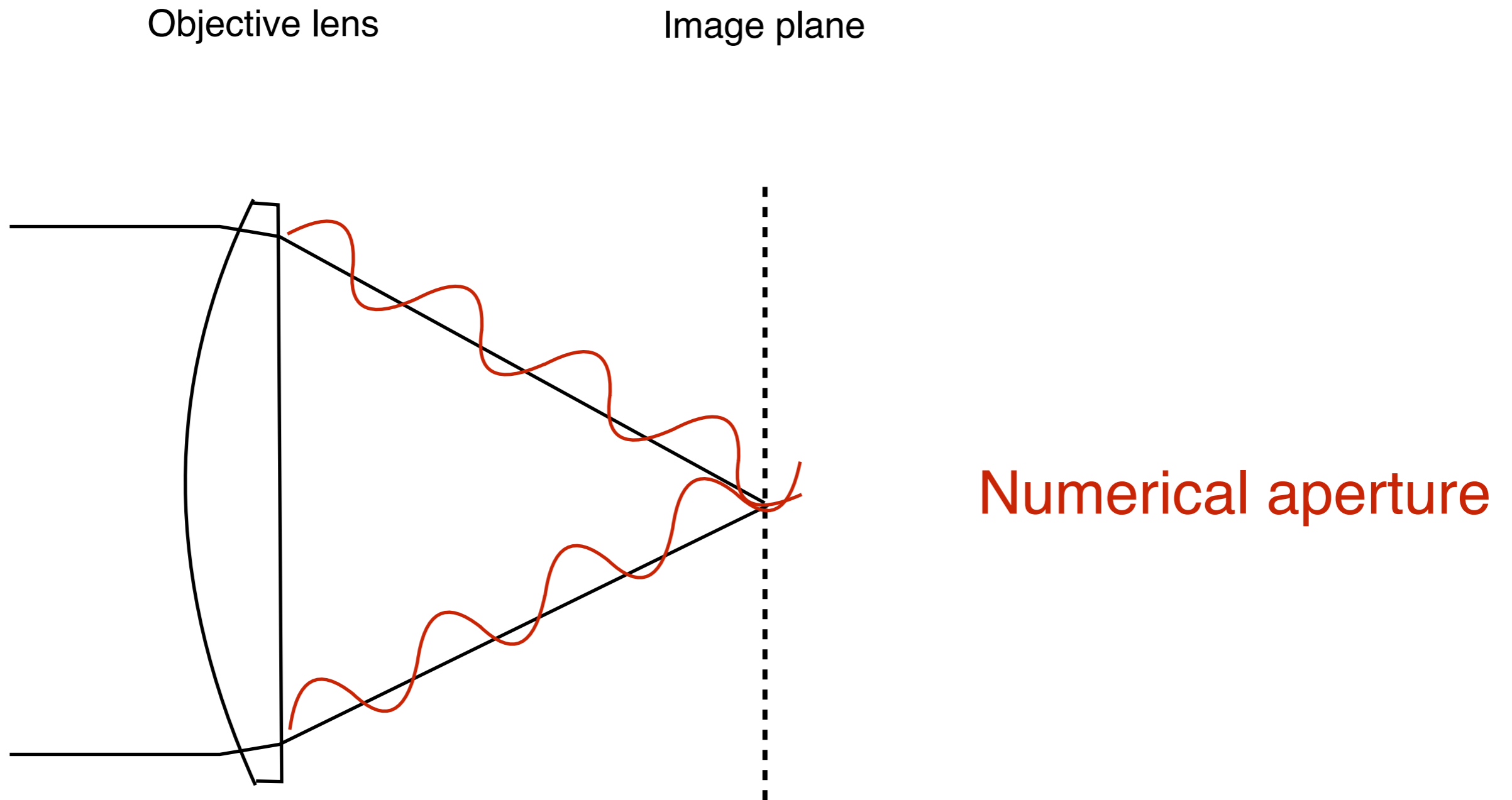
Objective lens

Image plane



wavelength

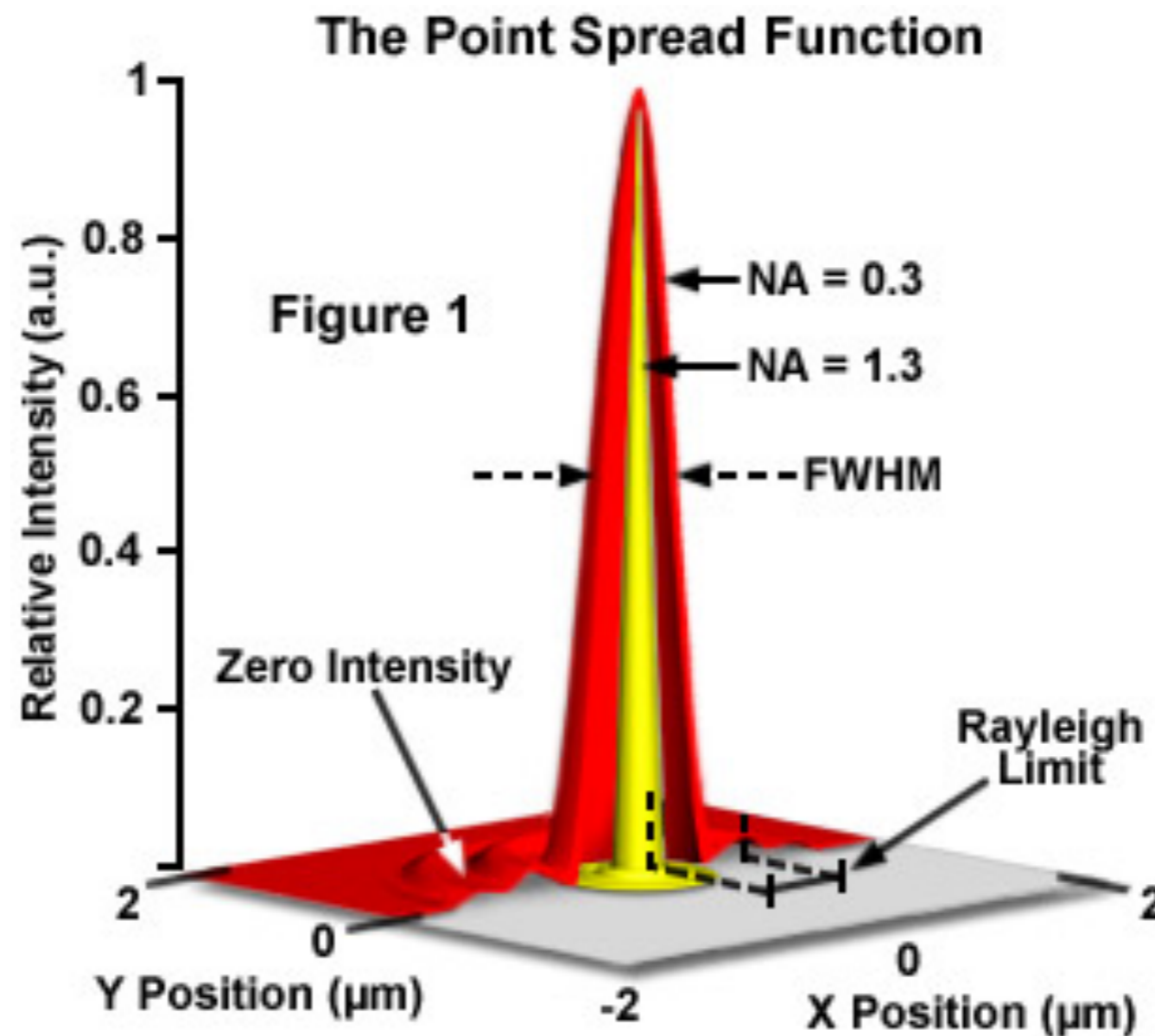
# What does depend on...?



Higher numerical aperture, less distortion, higher resolution

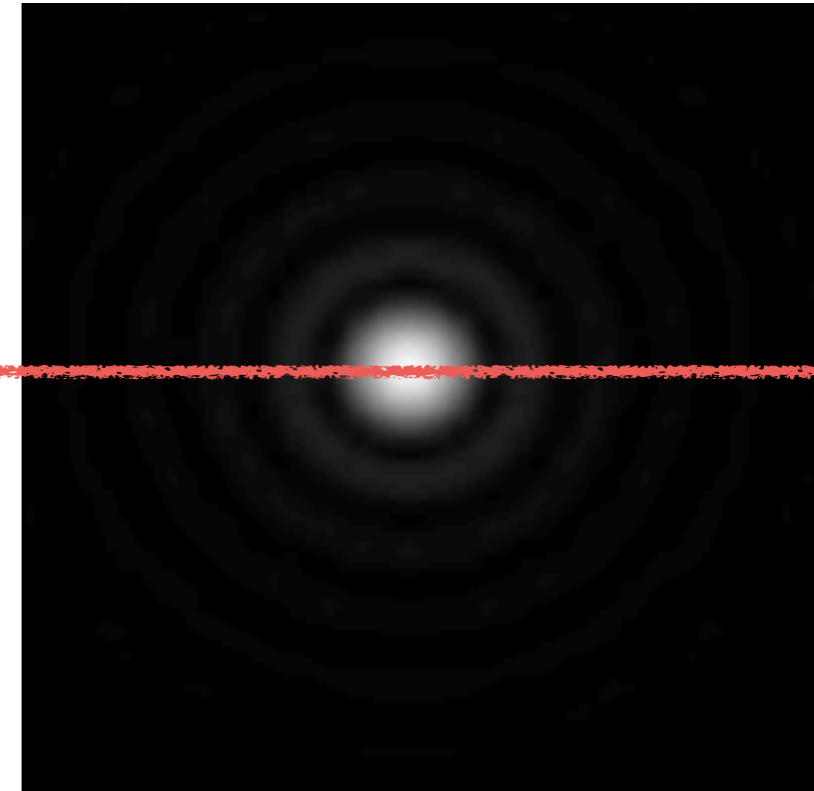
# What does depend on...?

Numerical aperture



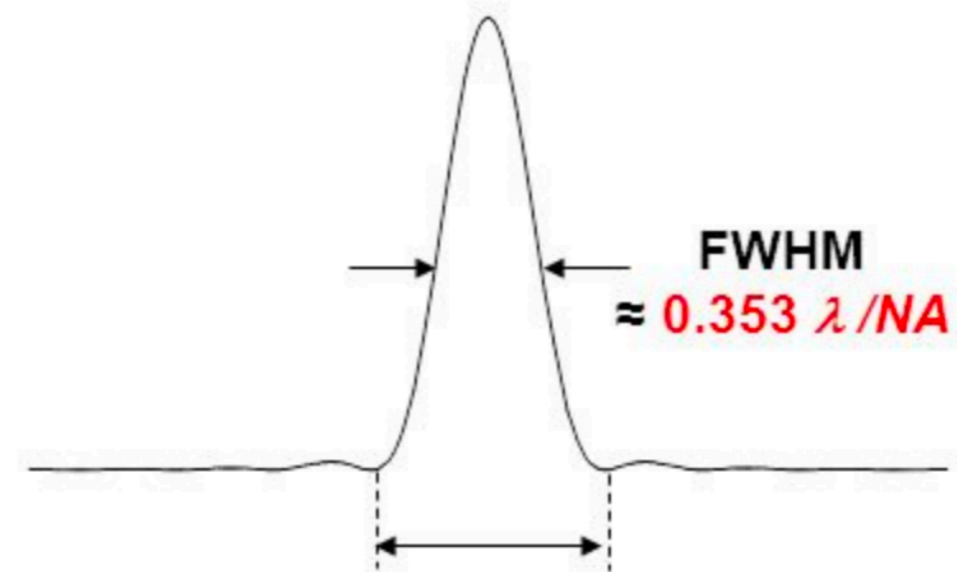
Higher numerical aperture, less distortion, higher resolution

# PSF is a way to measure resolution



As the Full Width at Half Max (FWHM) of the PSF

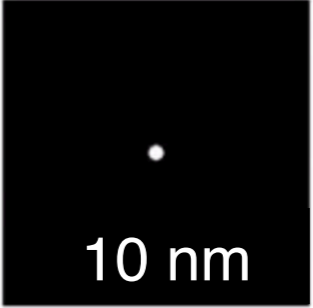
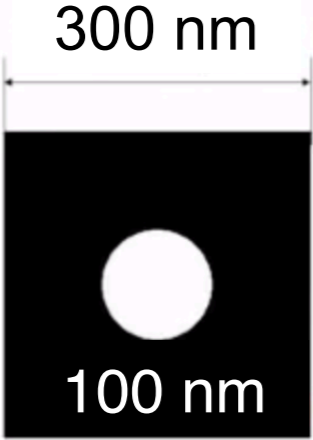
As the diameter of the Airy disk (first dark ring of the PSF) = "Rayleigh criterion"



Airy disk diameter  
 $\approx 0.61 \lambda / NA$



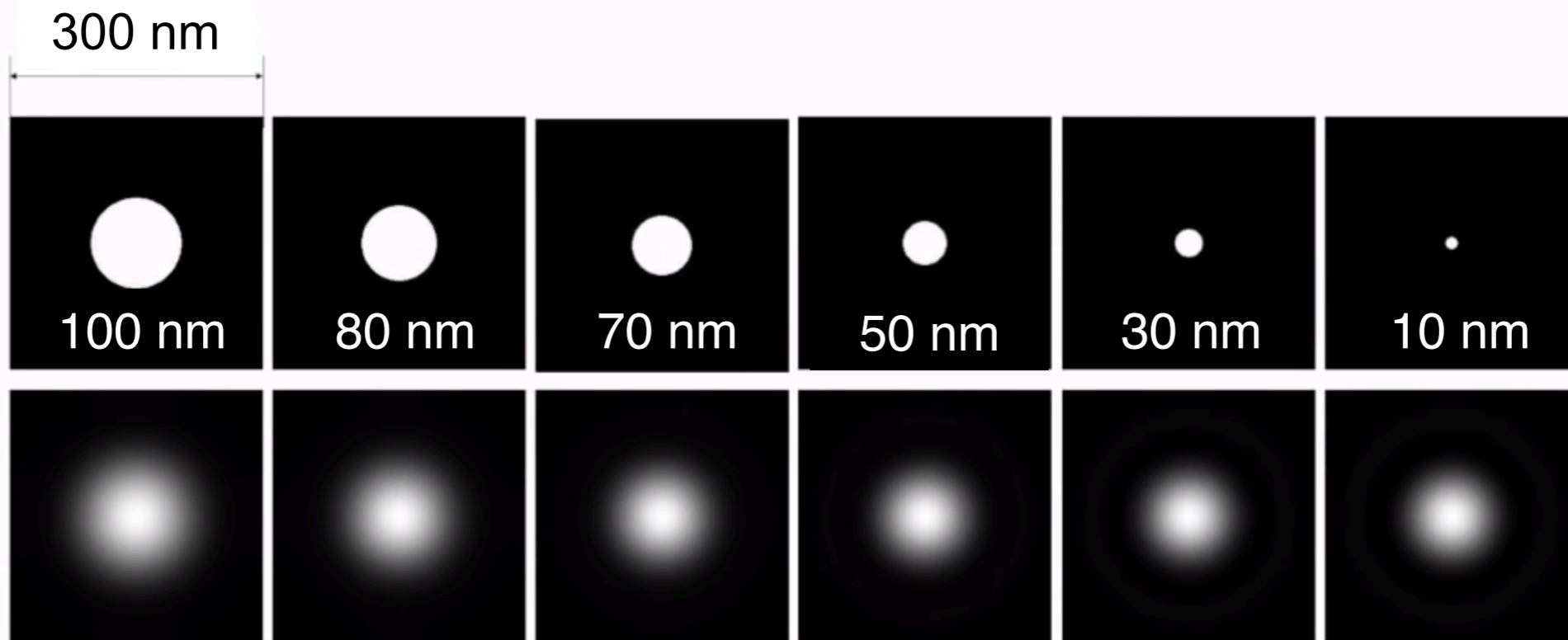
# How is the PSF of a small object?



1.4NA objective

$\lambda = 0.48 \mu\text{m}$

# How is the PSF of a small object?



1.4NA objective

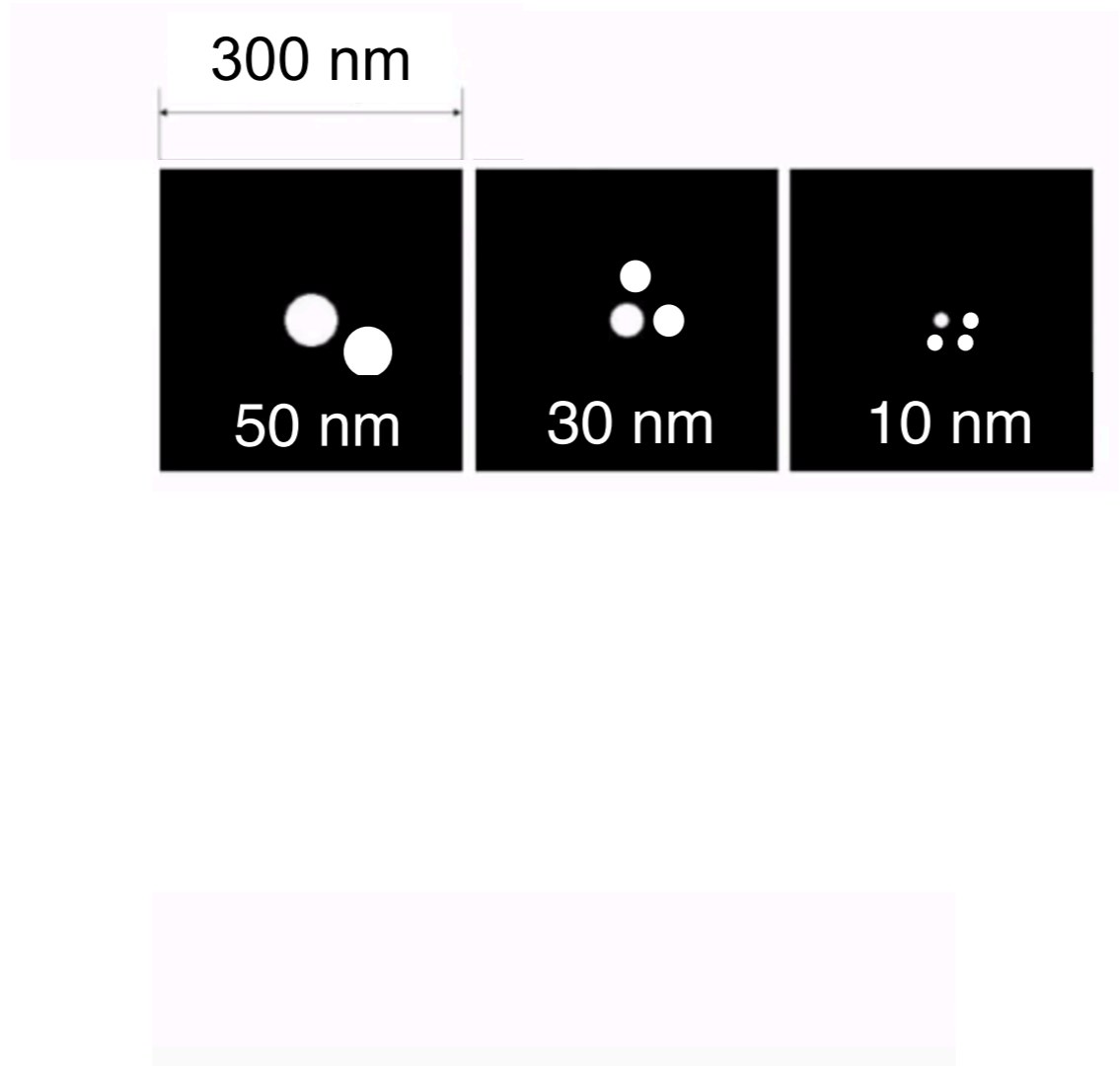
$\lambda = 0.48 \mu\text{m}$

$$d = \frac{\lambda}{2NA}$$

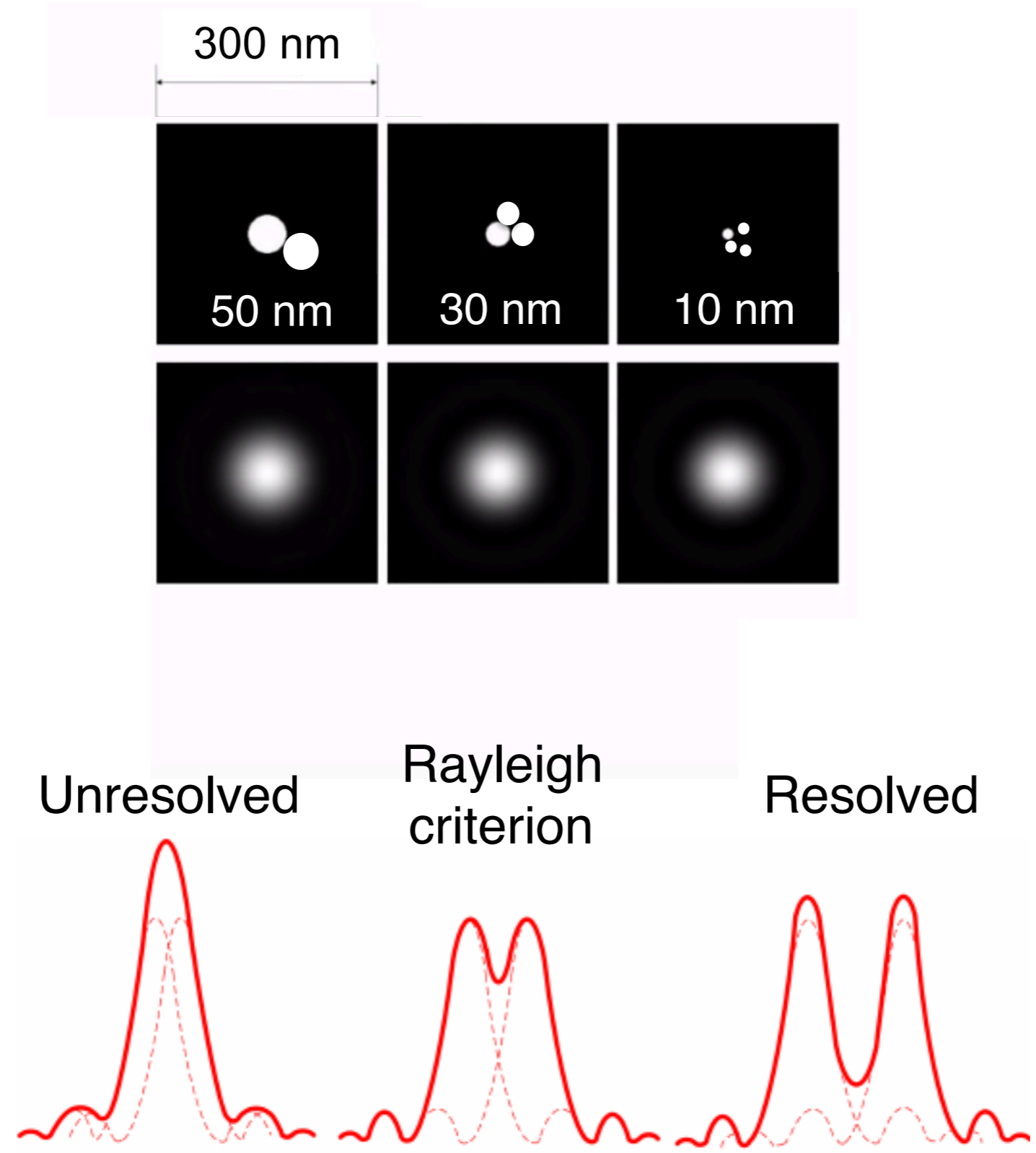
$\sim 170\text{nm}$

Abbe's diffraction limit

# How is the PSF of many small objects?



# PSF of many small objects

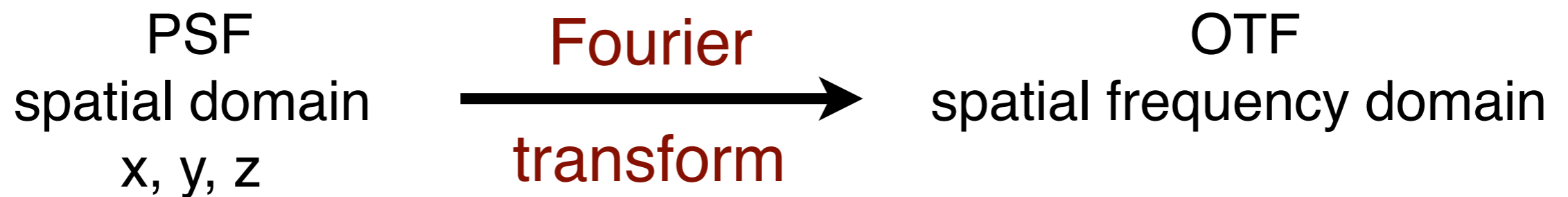


# OTF (Optical transfer function)

Used in widefield-deconvolution and Super-resolution (SIM)

OTF is the **Fourier transform** of PSF

FT algorithm computes  
a signal into its  
**frequency** domain



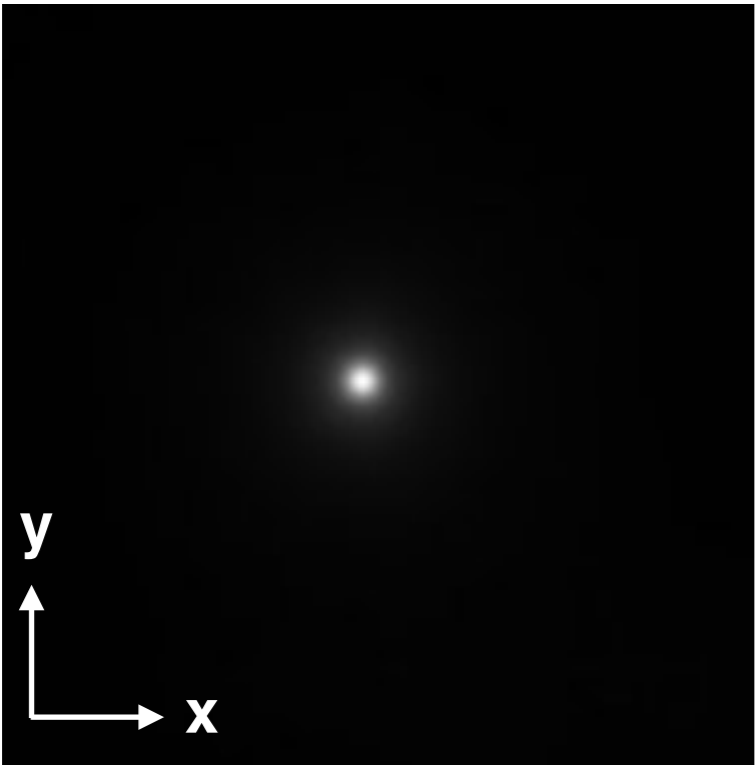
*OTF represents how spatial frequencies are handled by the optical system*

*How often it happens in space?*



# OTF (Optical transfer function) is the Fourier transform of PSF

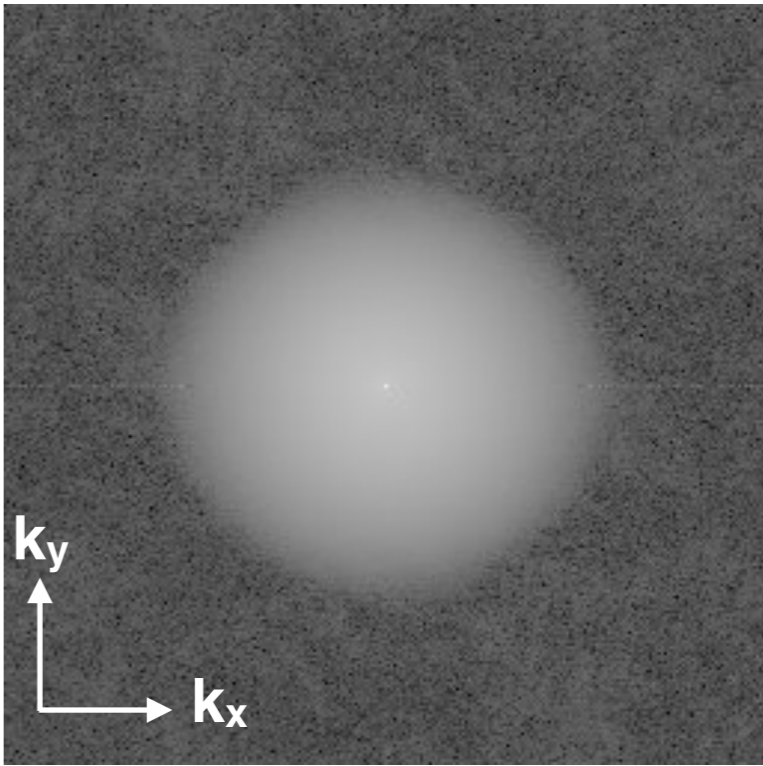
PSF



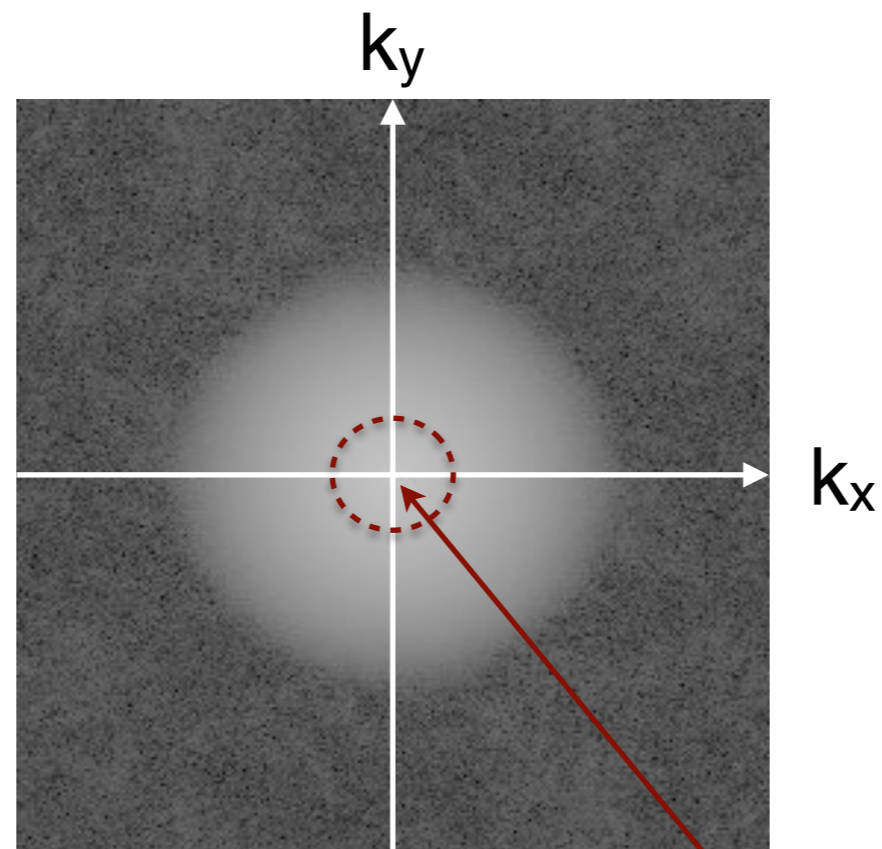
Fourier  
transform

Inverse Fourier  
transform

OTF



# OTF (Optical transfer function) is the Fourier transform of PSF



Lower frequencies  
towards the centre



What are spatial frequencies ... in an image?





Lower frequencies - blurred





Higher frequencies - sharp

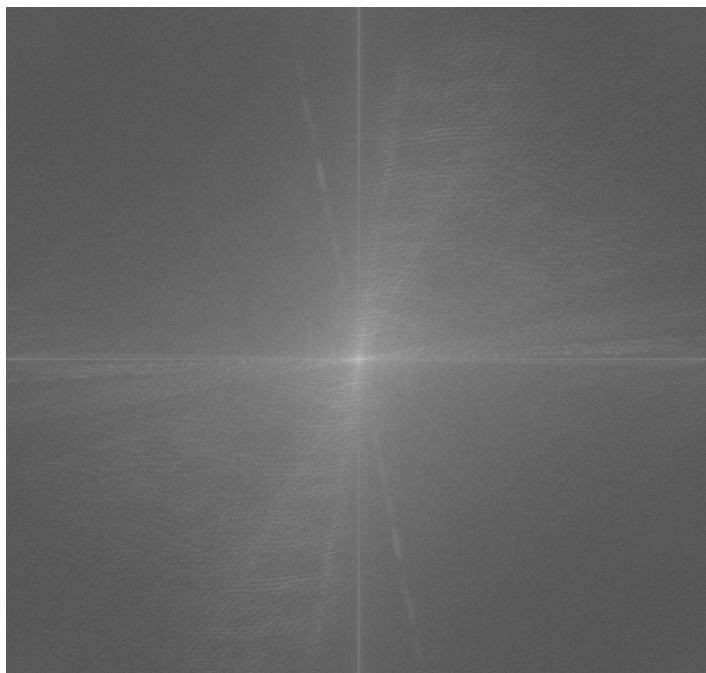




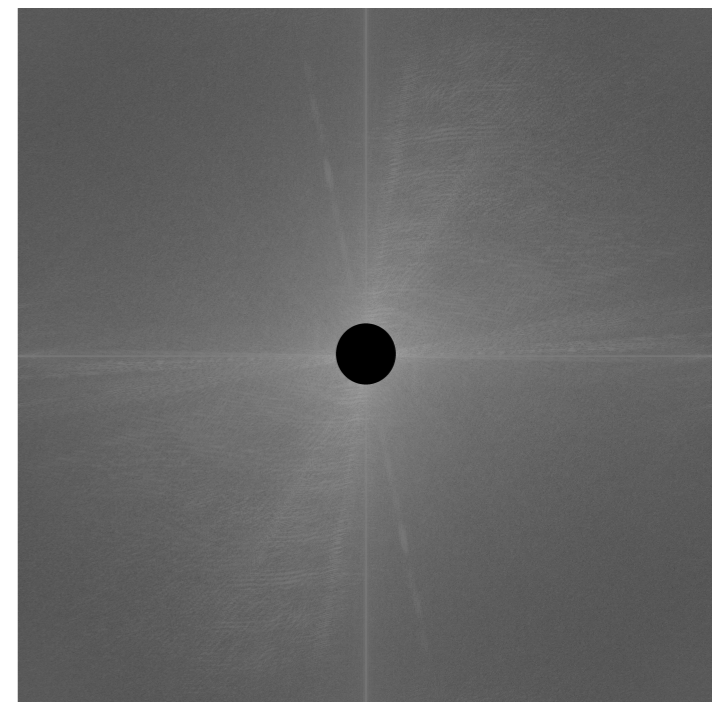
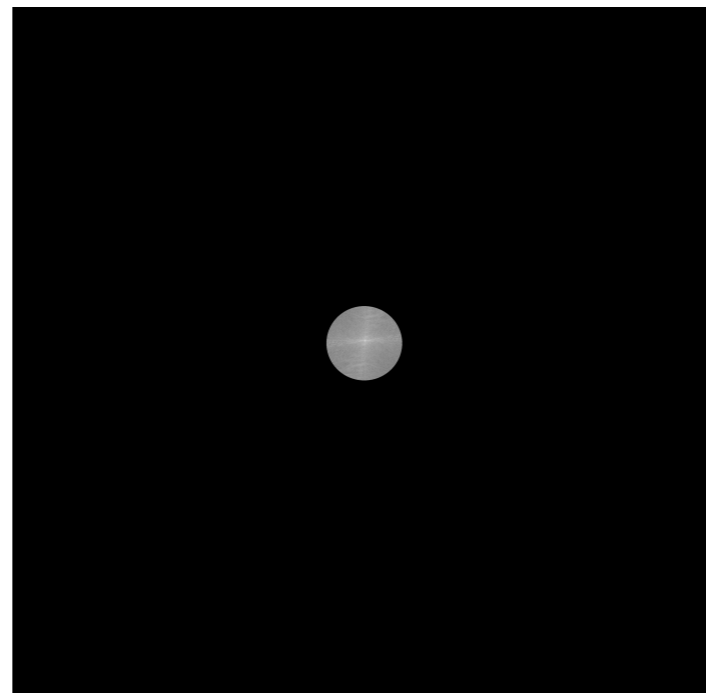
# FIJI / Process / FFT



Fourier  
transform



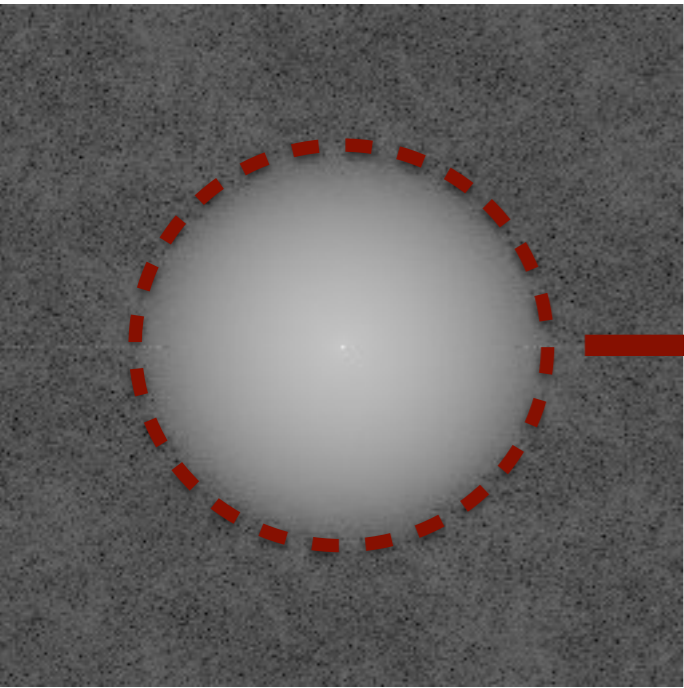
Inverse Fourier  
transform





# It's very easy to detect certain features in the frequency domain

All frequencies



*What does it represent?*

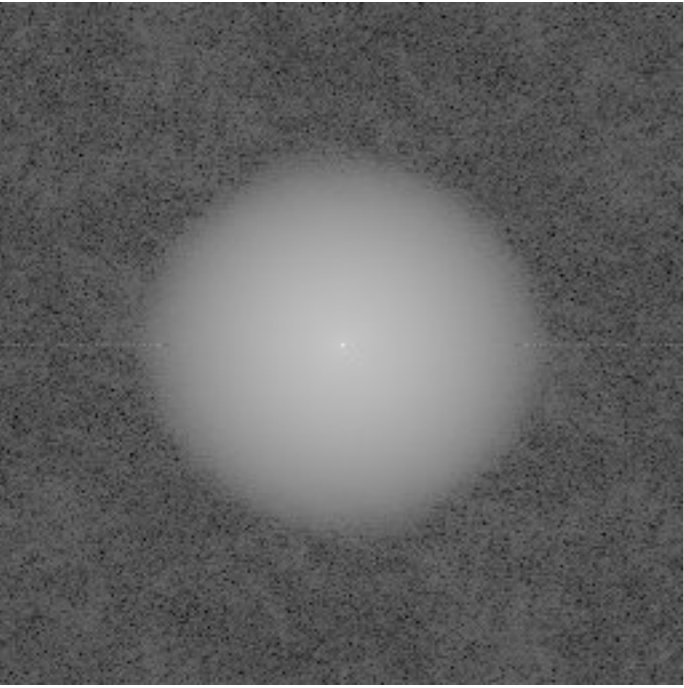
Back Aperture Objective



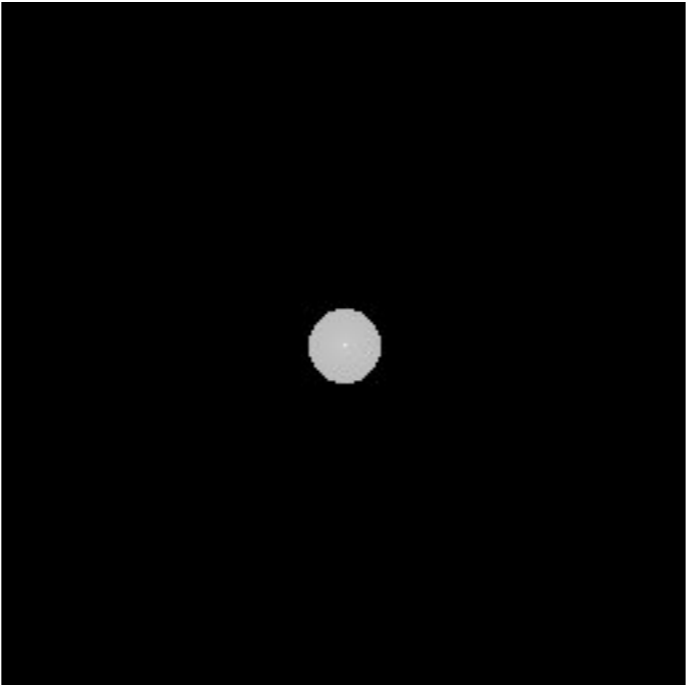
The microscope passes low frequencies (large and smooth) and excludes high frequencies

# It's very easy to detect certain features in the frequency domain

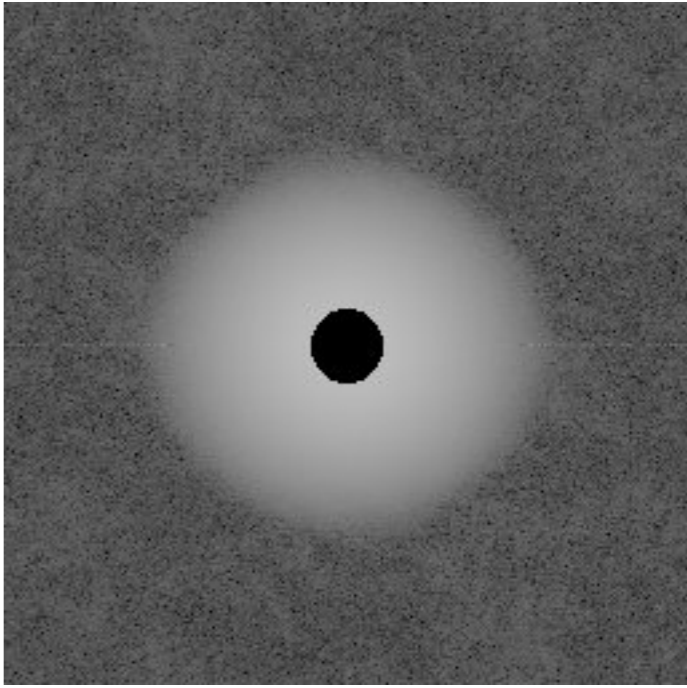
All frequencies



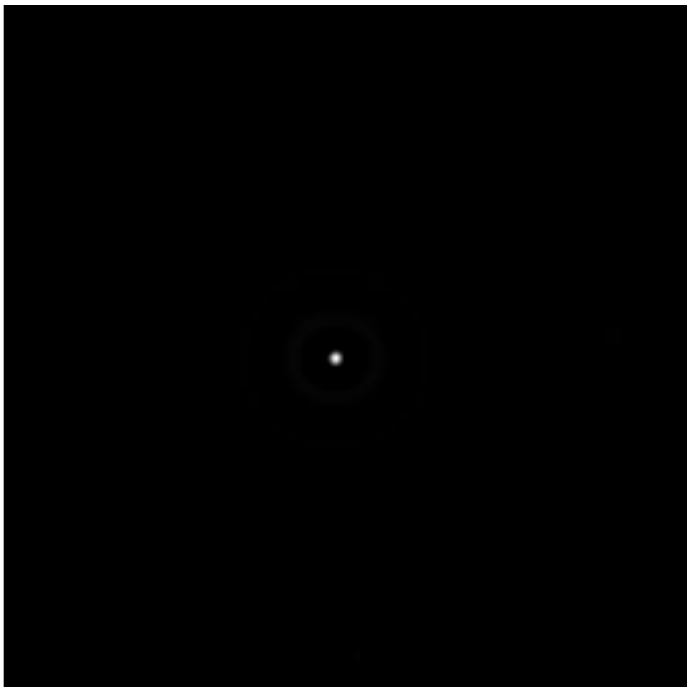
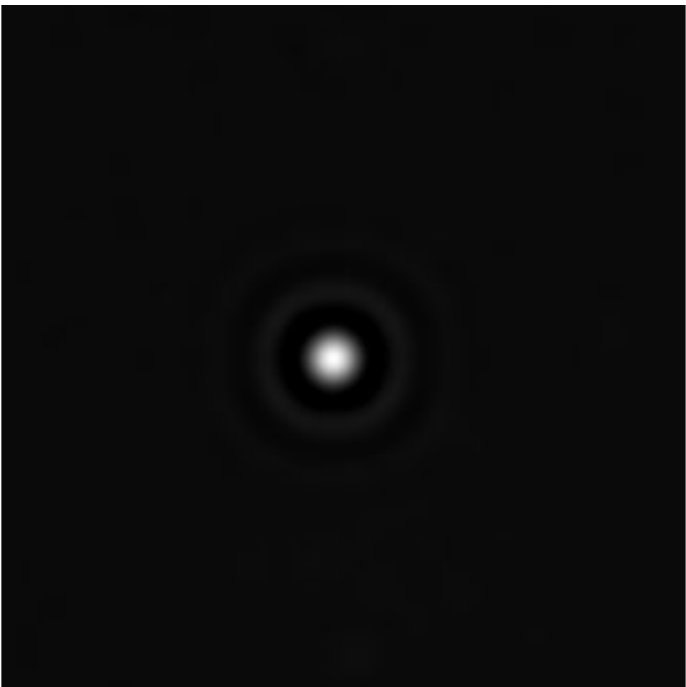
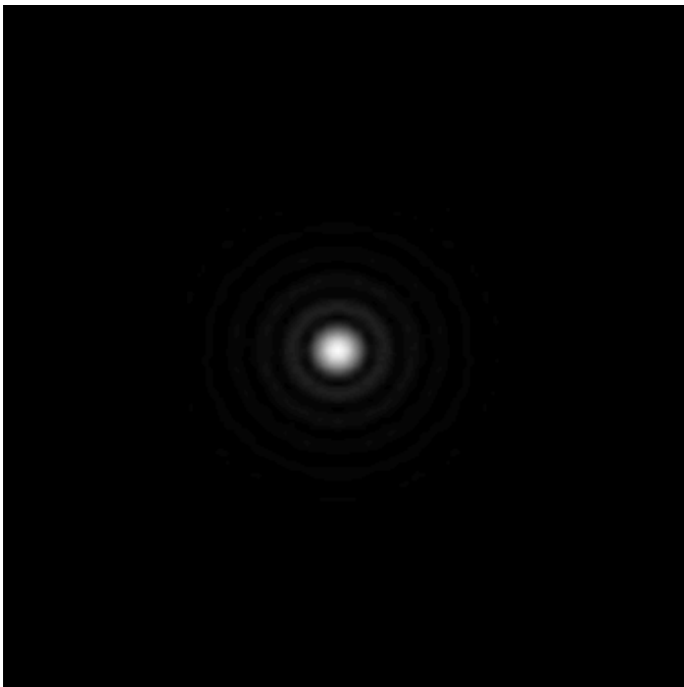
Just lower frequencies



Just higher frequencies

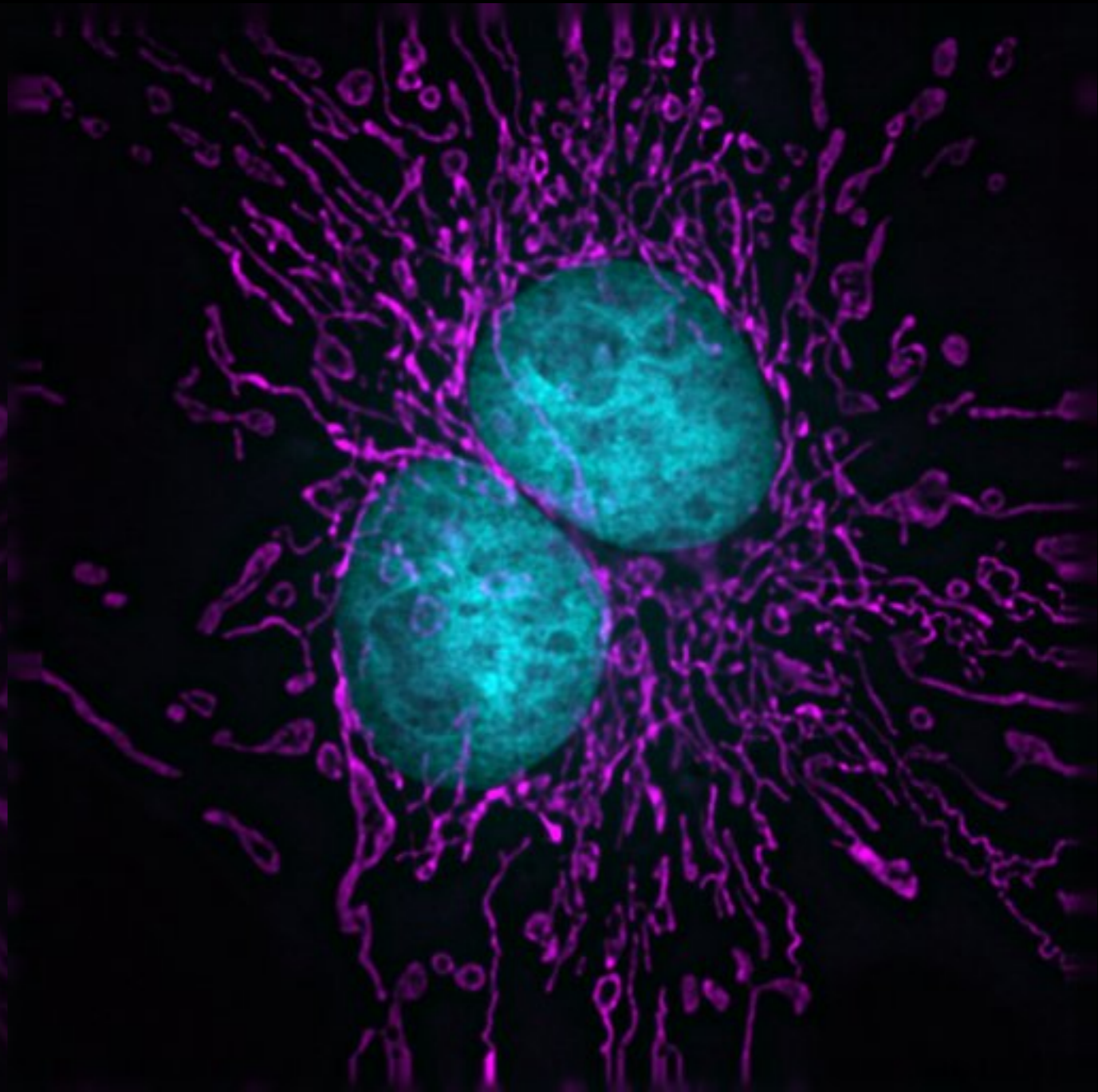
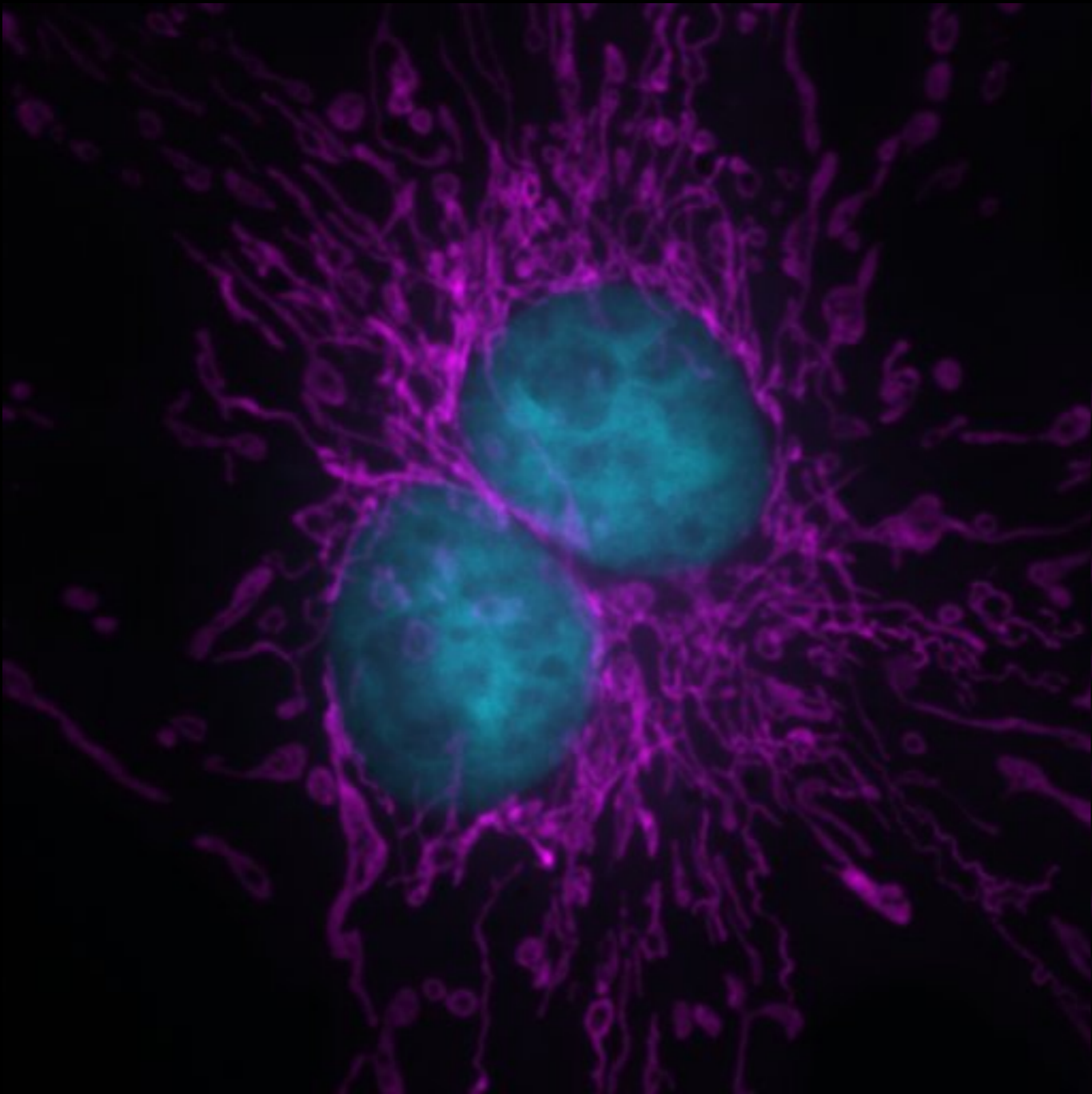


Inverse  Fourier transform 



Widefield

Deconvolution

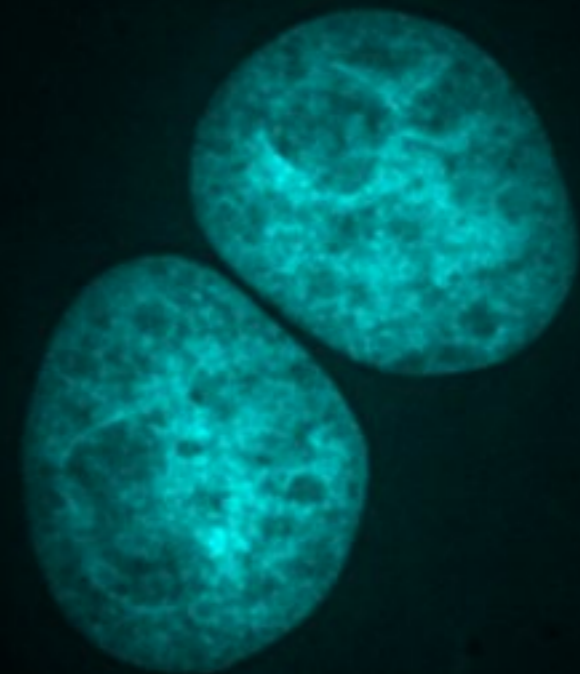
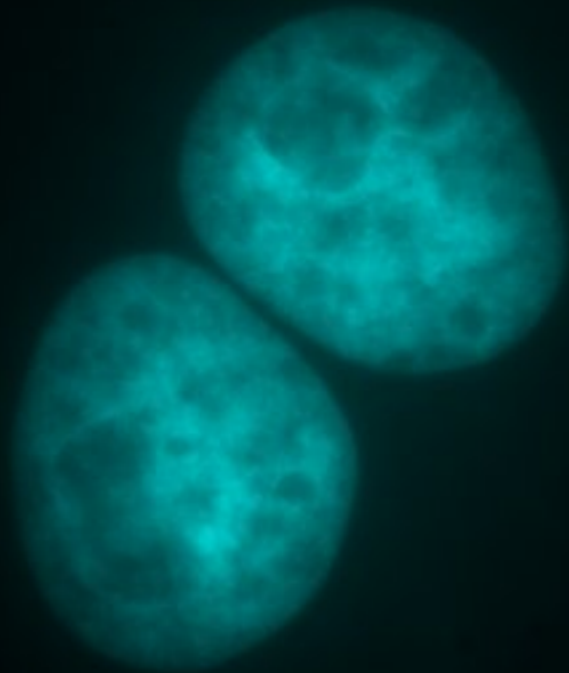


BPAE cells with Mitotracker Red (magenta) and DAPI (cyan)



Widefield

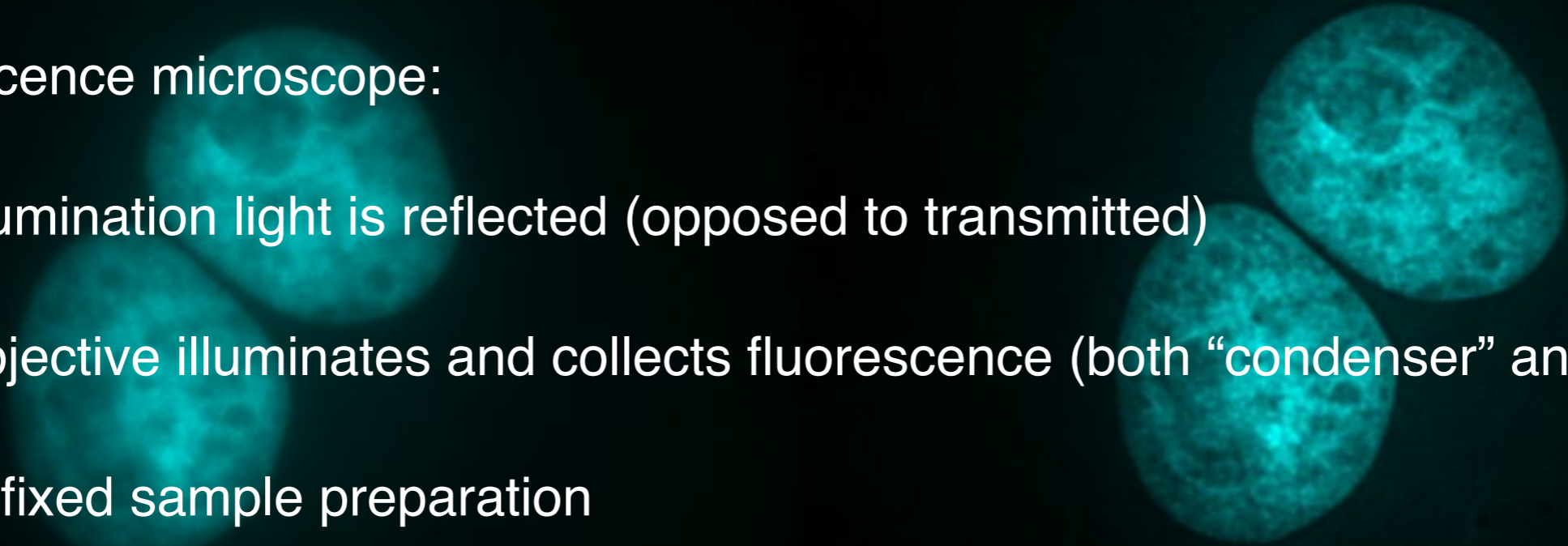
Deconvolution



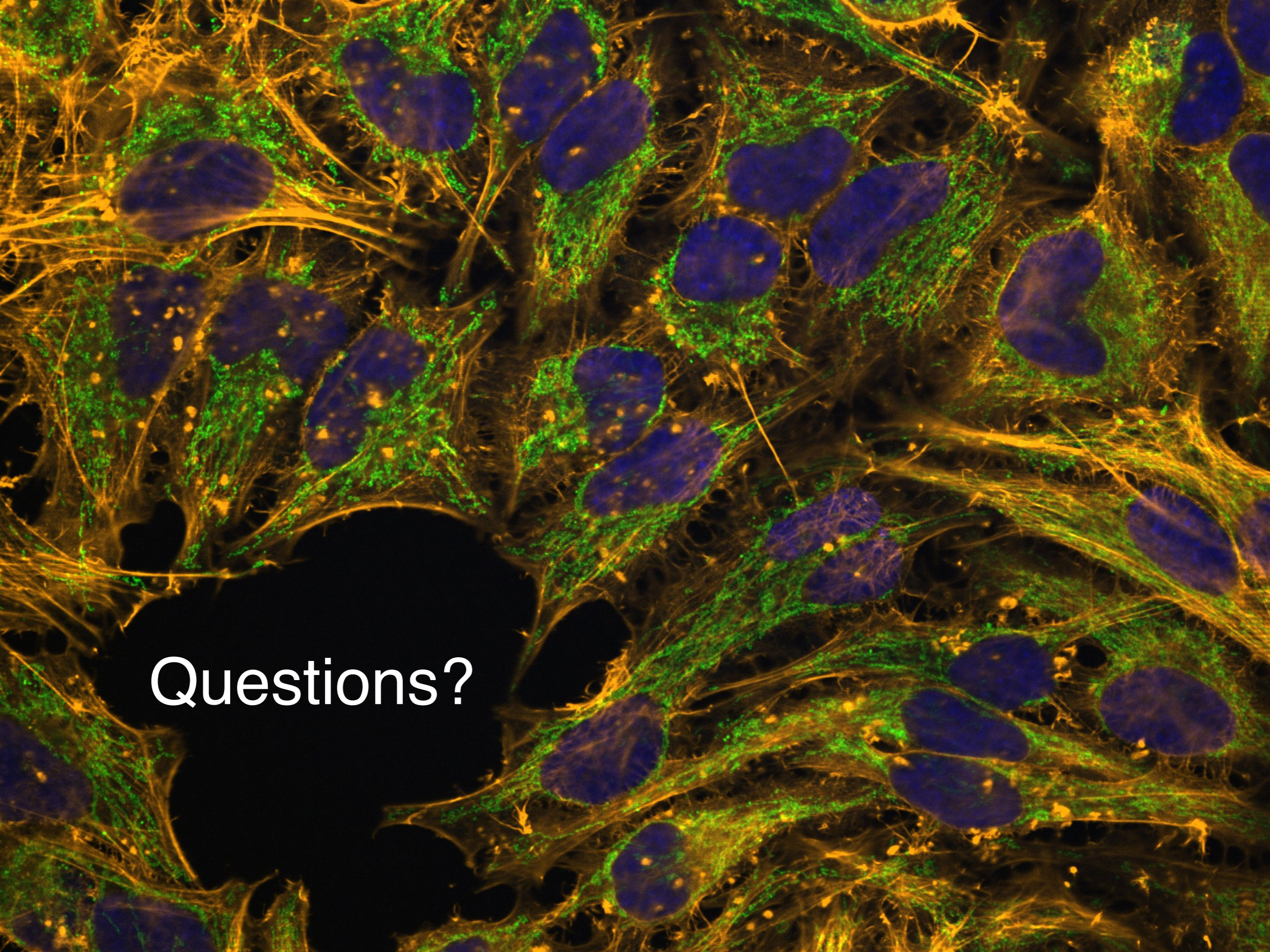
Nuclei from BPAE cells stained with DAPI (cyan)

# Conclusions

- \* Why is fluorescence? **CONTRAST**
- \* **Dichroic mirror** - separates illumination (excitation) from fluorescence (emission)
- \* Fluorescence microscope:
  - \* illumination light is reflected (opposed to transmitted)
  - \* objective illuminates and collects fluorescence (both “condenser” and objective)
- \* Tips on fixed sample preparation
- \* Point Spread Function and Optical Transfer Function
- \* Widefield fluorescence microscopy collects the **whole field** of view at once; it's **fast** and very **sensitive** and you can have **deconvolution** for free







Questions?