

Lecture 4 Understanding and applying fluorescence microscopy



Andrew Jefferson Micron Advanced Bioimaging Unit



Outline

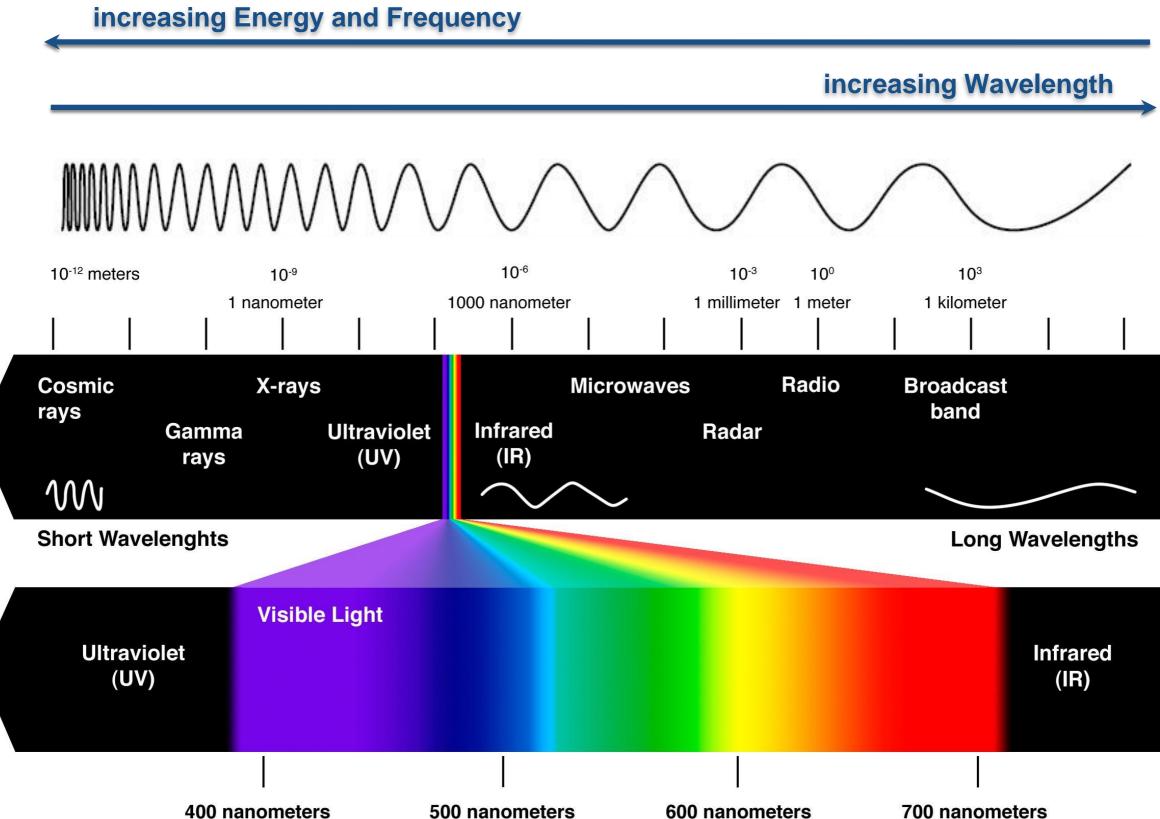
- 1. What is fluorescence?
- 2. Why fluorescence?
- 3. Principle and components of the fluorescence microscope
- 4. Fluorescent light sources
- 5. Point Spread Functions
- 6. Fixation for light microscopy



The electromagnetic spectrum







380 – 700 nm visible to the human eye



"Fluorescence is the emission of light by a substance that has absorbed light"

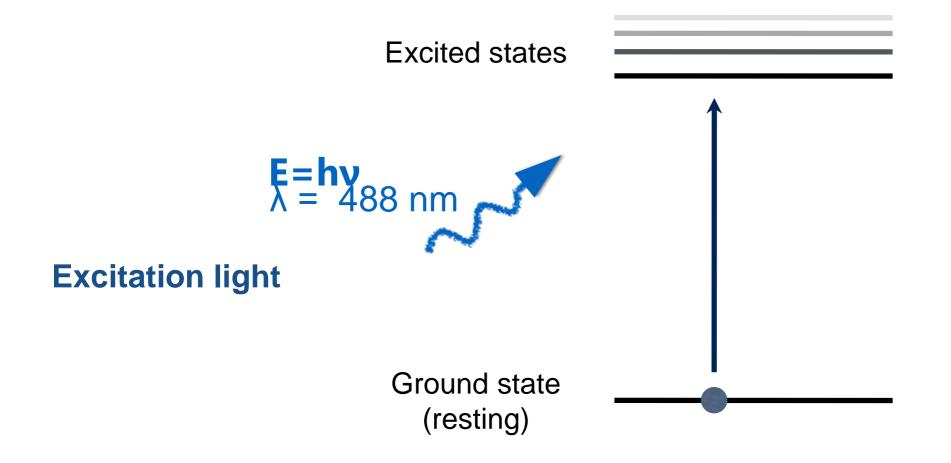
https://en.wikipedia.org/wiki/Fluorescence

Excited states	



Molecules have discrete levels of energy





A photon is the energy unit for light to interact with matter





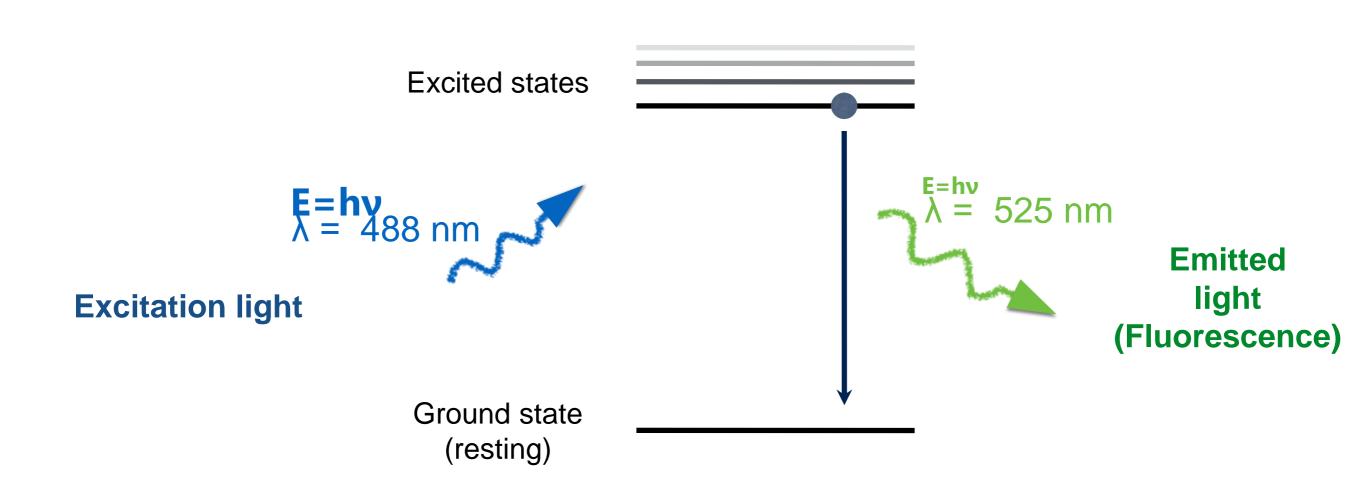




Excitation light

Ground state (resting)

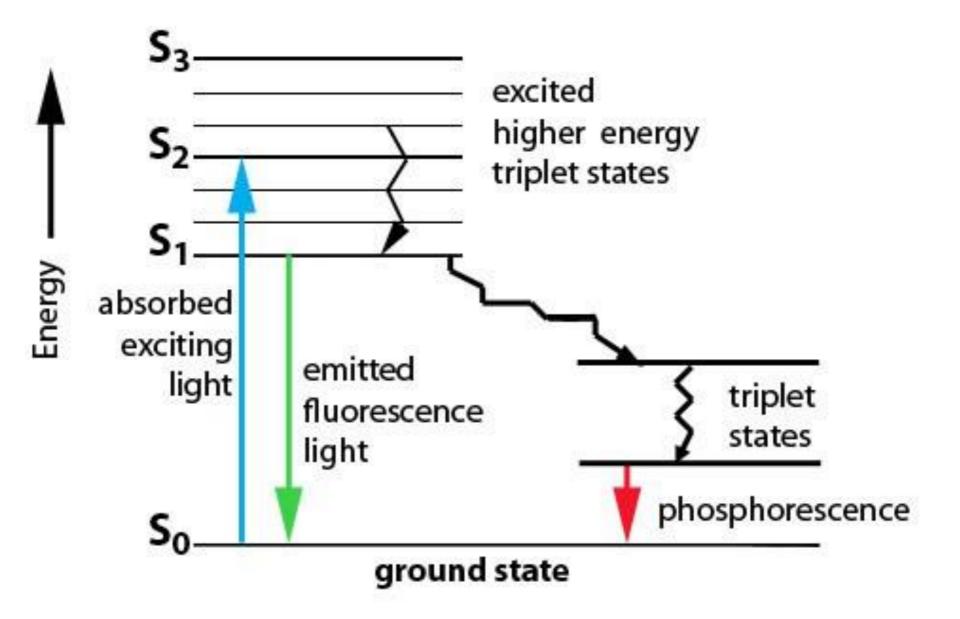




The full picture is represented by the Jablonski diagram...







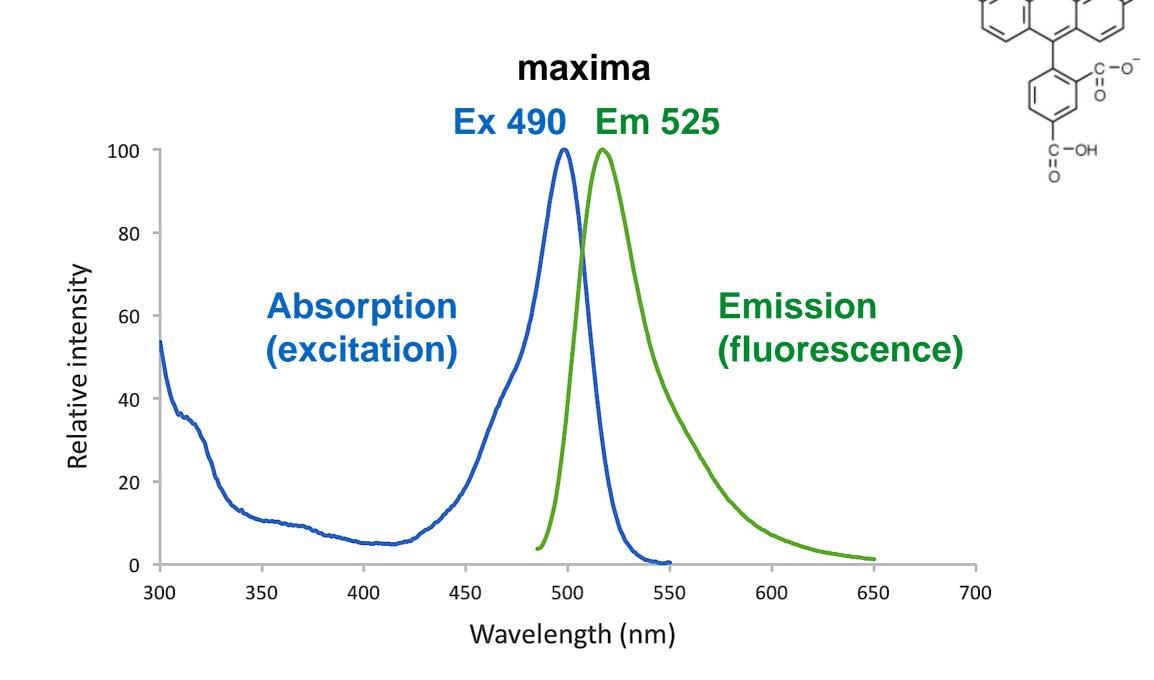
Fluorescence Spectra

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HO











Genetically encoded fluorescent proteins

• GFP, YFP, mCherry

Organic dyes

Alexa, ATTO, Fluorescein, DAPI

Fluorescent labelled antibodies (immunofluorescence)

Inorganic dyes

Quantum Dots

Endogenous species

Elastin, collagen, metabolic coenzymes (NADH, FAD)



2. Why fluorescence?

Why Fluorescence?



Chris Teren: https://www.youtube.com/watch?v=PhcITQ3g0s8

CONTRAST

Why Fluorescence?



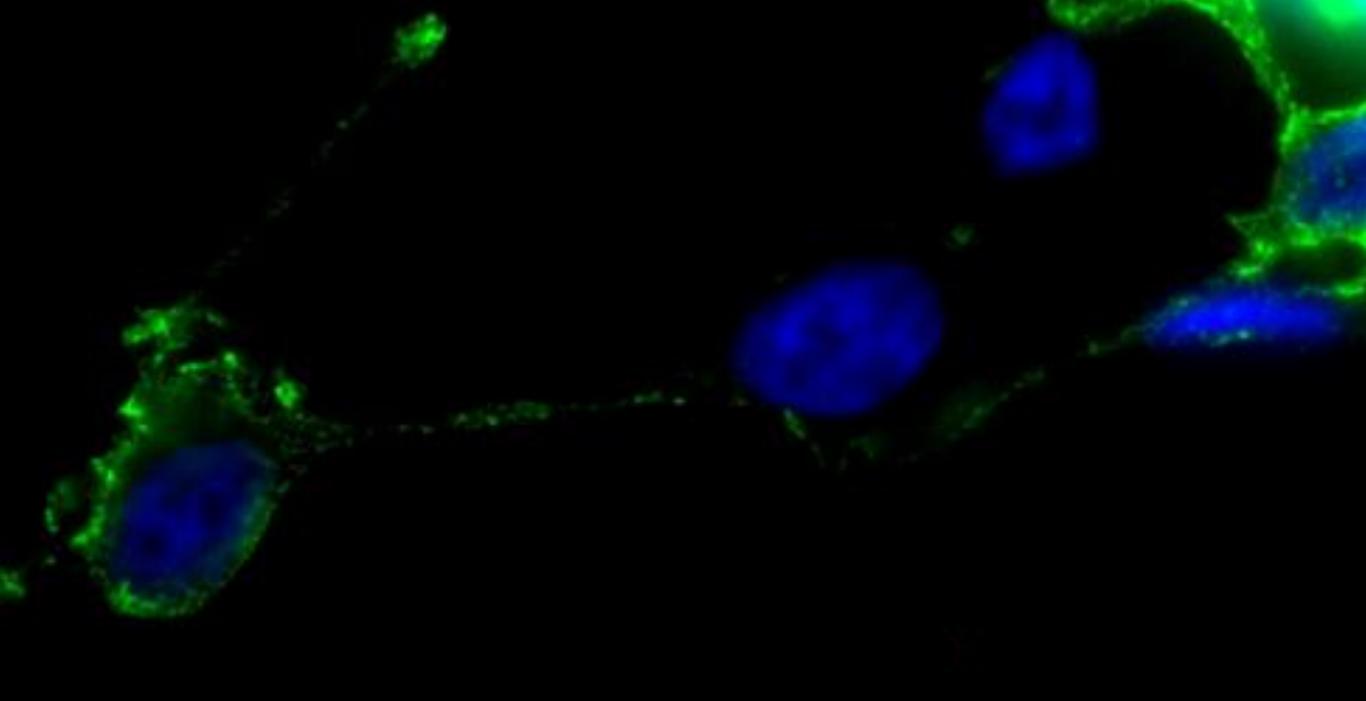
- Weak signal against dark background is easier to measure
- High signal to background contrast

Why Fluorescence?

- Selective labeling
- Ease of multiplexing
- Quantitative

Microtubules Microtubule Plus ends Nucleus

Dr Alexis J. Lomakin <u>www.olympusbioscapes.com/gallery/2011/hm29.html</u>



How do we get a black background....?

Fundamental problem in fluorescence microscopy



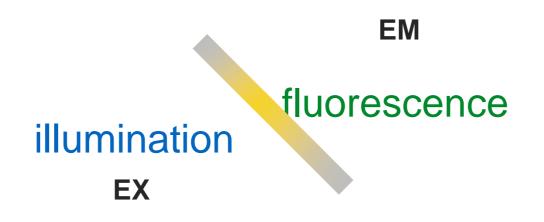


STRONG illumination

WEAK fluorescence signal



produce high-efficiency illumination of the specimen while simultaneously capturing weak fluorescence emission

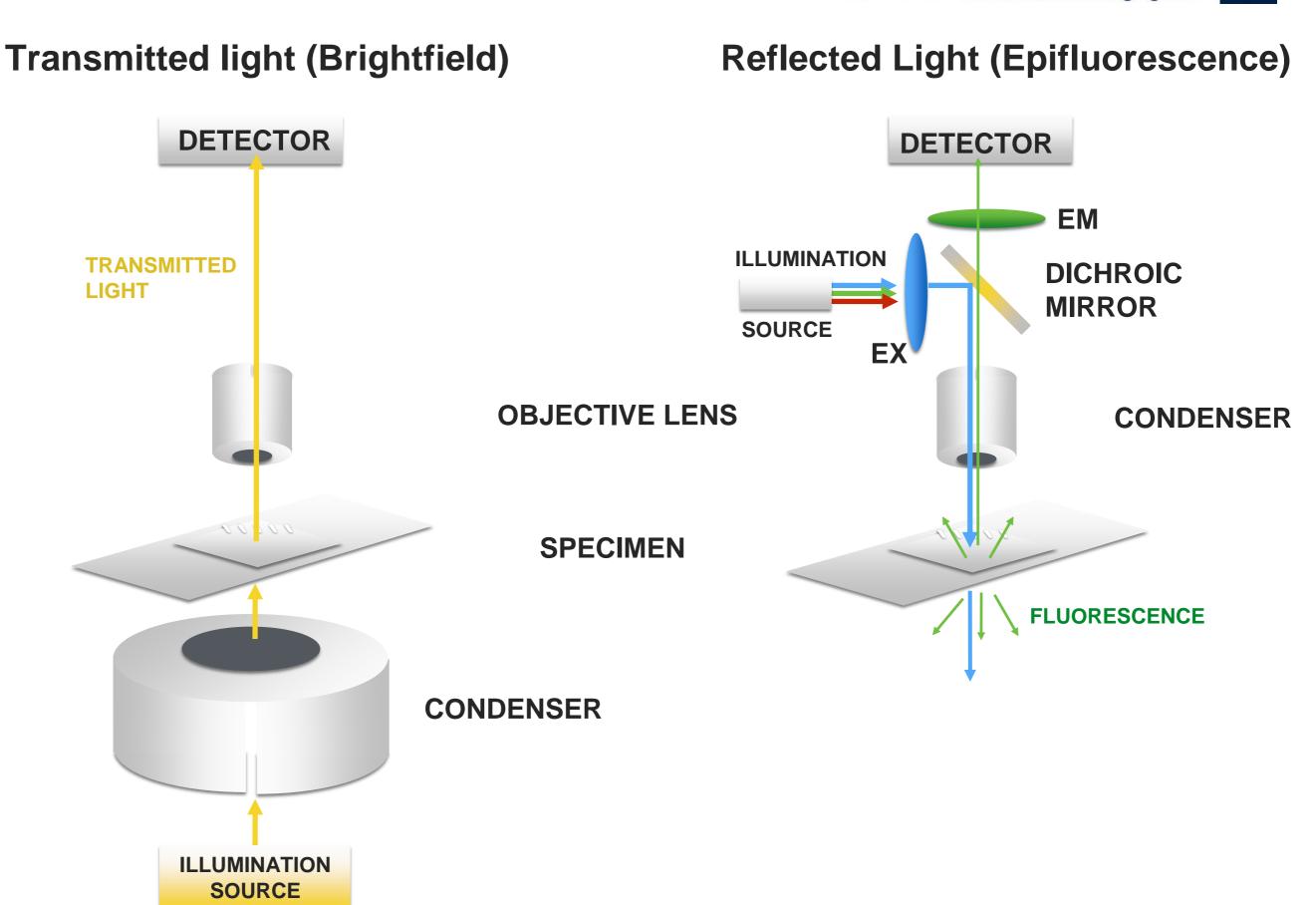




3. Principles and components of the fluorescence microscope

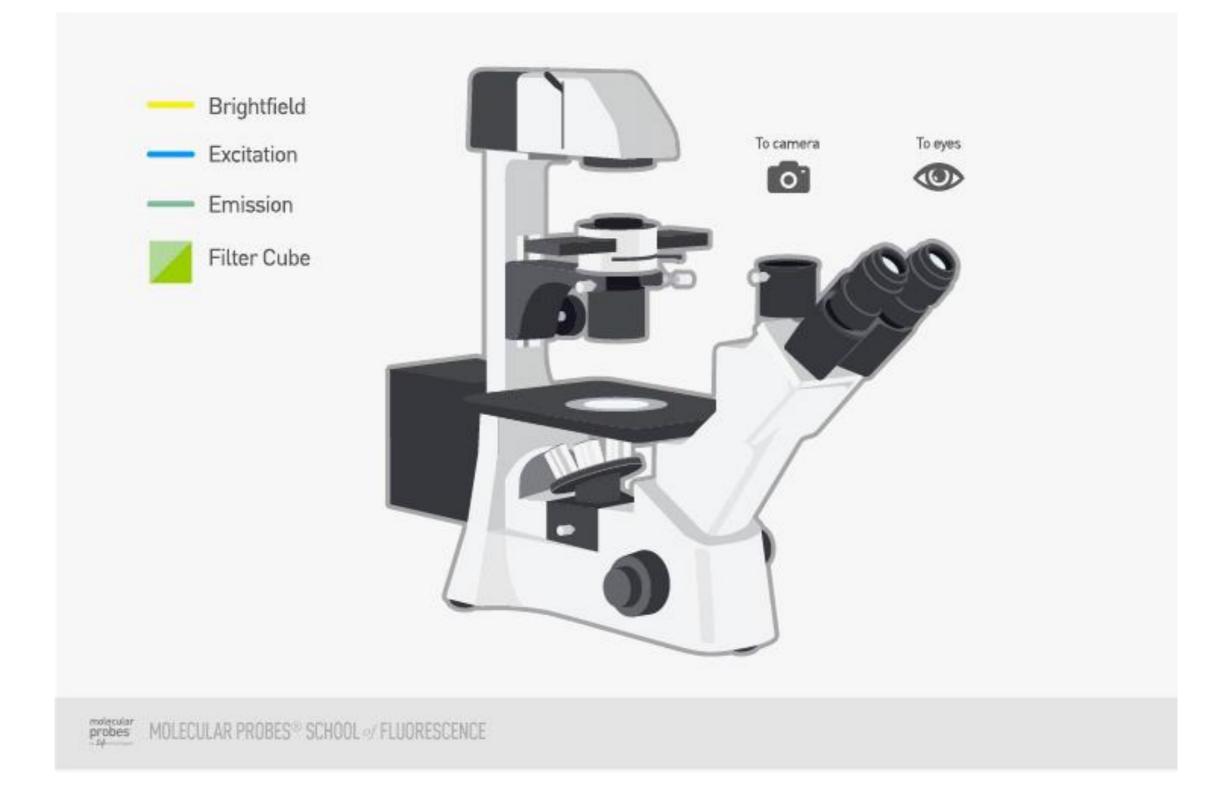


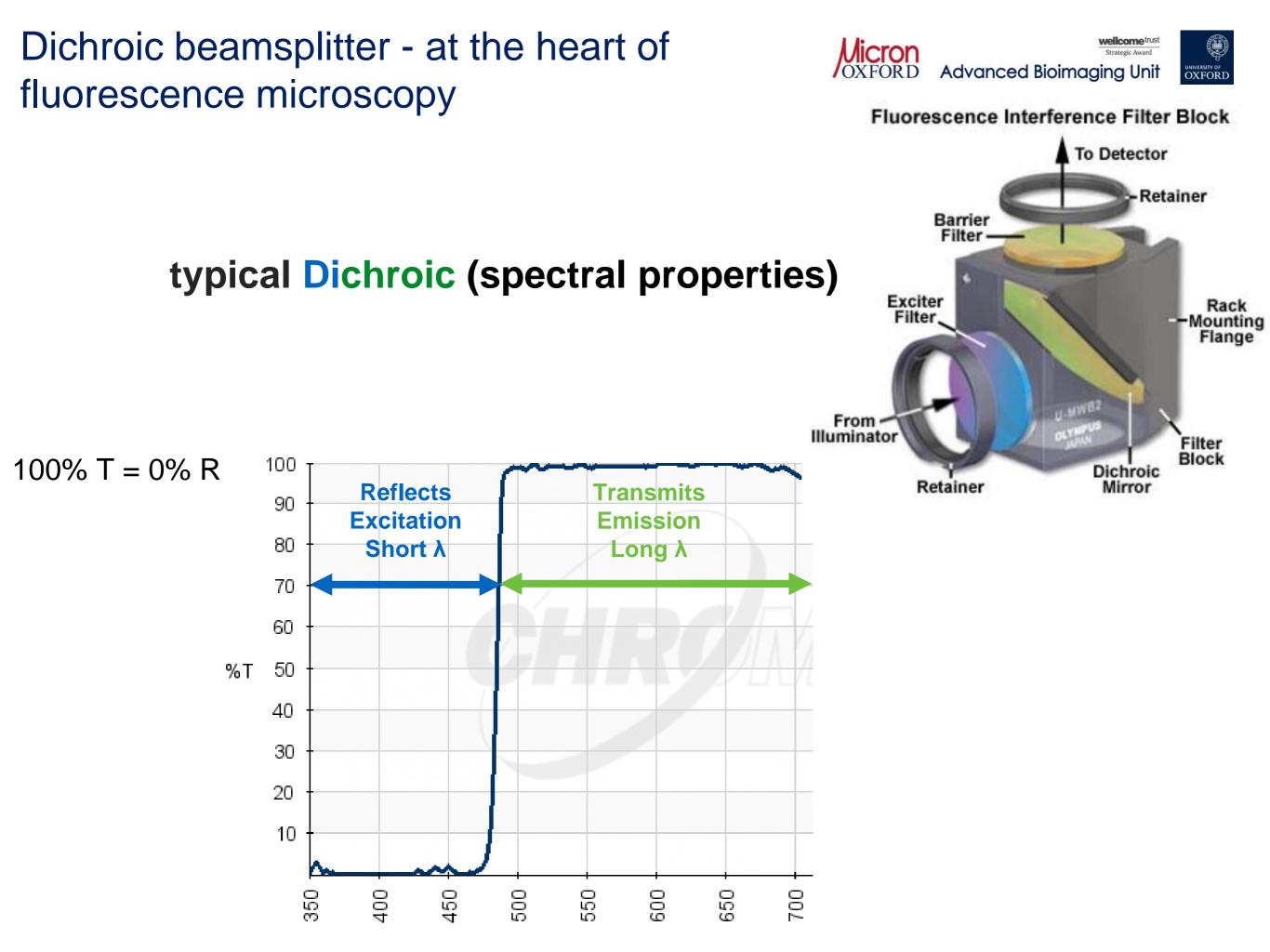
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Epifluorescence vs Transillumination light paths (inverted)

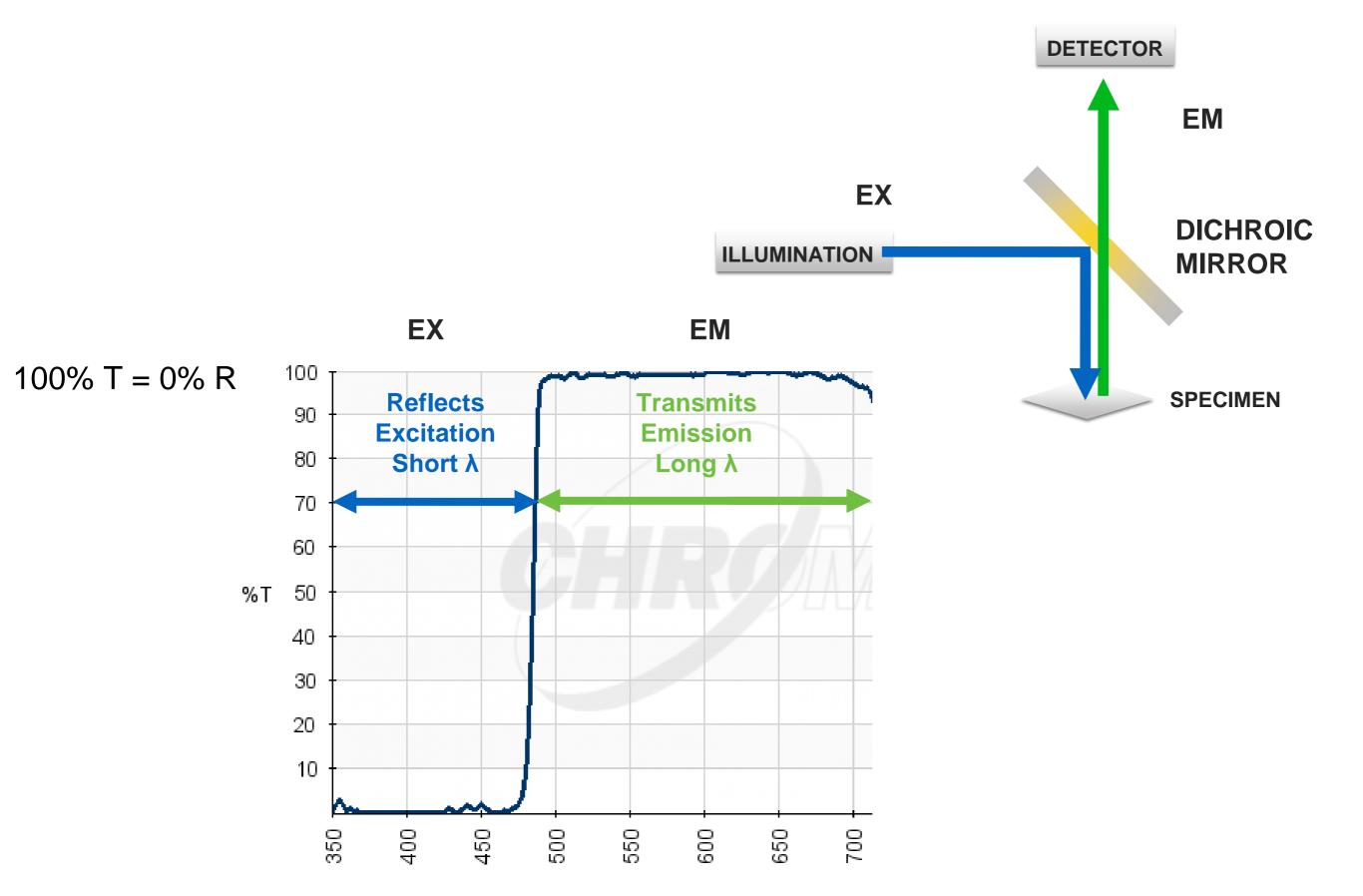


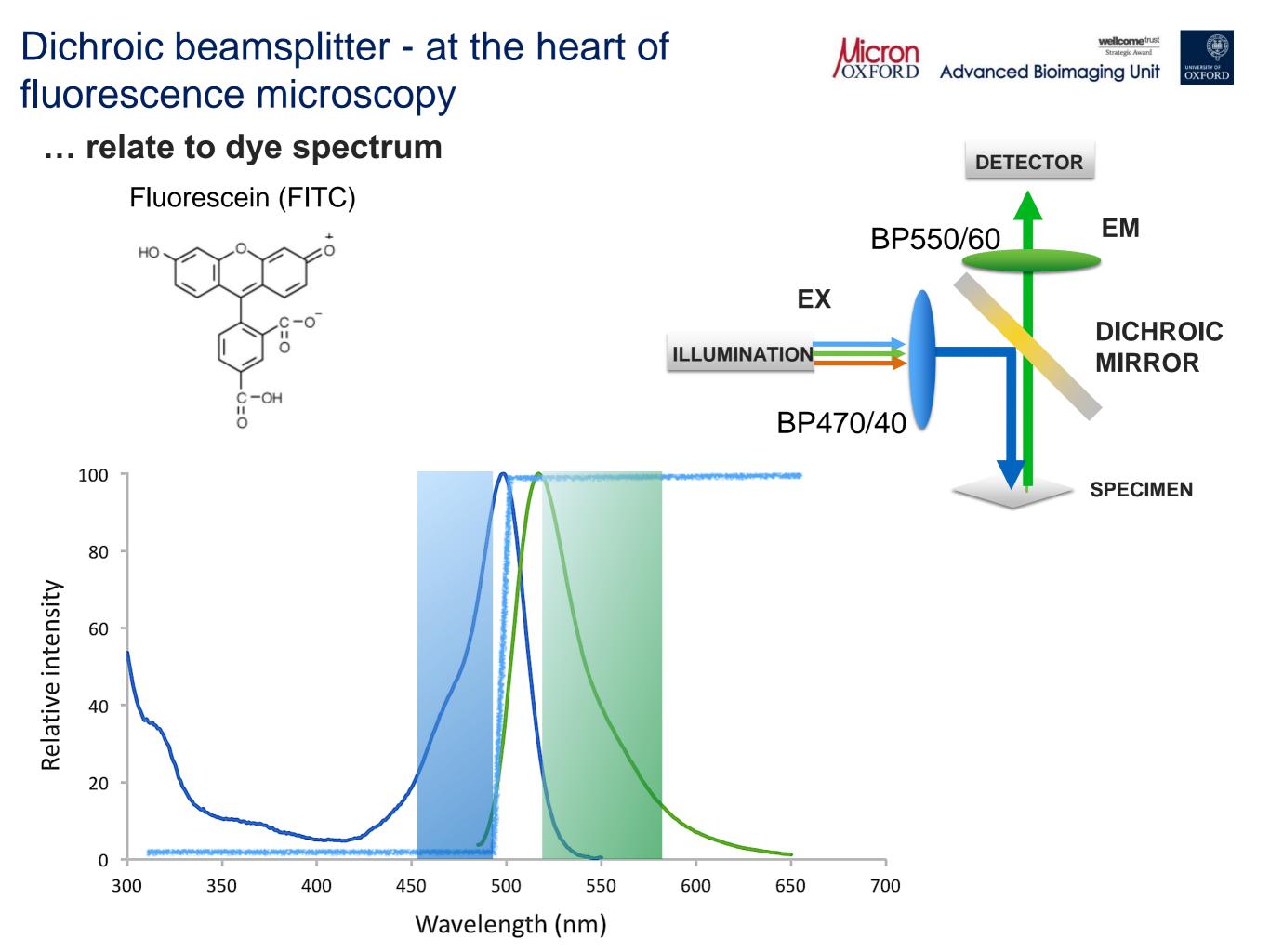




Dichroic beamsplitter - at the heart of fluorescence microscopy







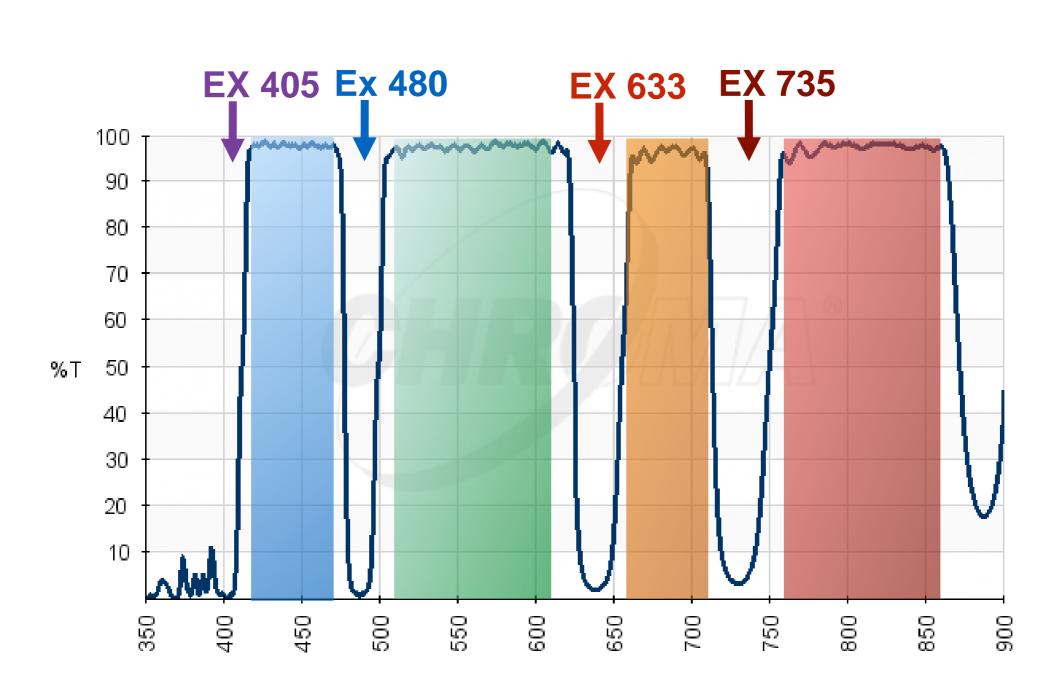
Dichroic beamsplitter - at the heart of *l*icron OXFORD Advanced Bioimaging Unit UNIVERSITY OF fluorescence microscopy DETECTOR EM ... what about multiplexing...? EX DICHROIC ILLUMINATION **MIRROR** Filter Block Turret (a) Emission **SPECIMEN** Filter Dichromatic Mirror Excitation Interference Filters

Dichroic beamsplitter - at the heart of *l*icron OXFORD Advanced Bioimaging Unit UNIVERSITY OF fluorescence microscopy DETECTOR EM ... what about multiplexing...? EX DICHROIC ILLUMINATION **MIRROR** Filter Block Turret (a) Emission **SPECIMEN** Filter Dichromatic Mirror Excitation Interference Filters

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Dichroic beamsplitter - at the heart of fluorescence microscopy





Polychroic

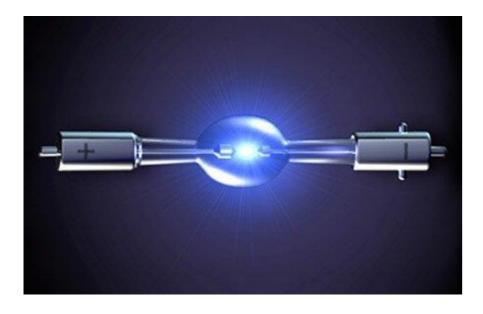


4. Fluorescent light sources

Illumination sources for widefield fluorescence microscopy

Metal arc lamps

- 200-300h lifespan
- Hazardous 0
- Generate heat 0
- Generally out of use 0

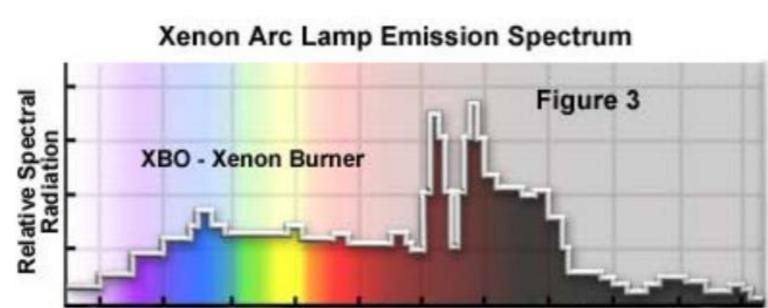


Mercury Arc Lamp UV and Visible Emission Spectrum Relative Spectral Radiation Mercury Arc Lamp (HBO)

500

400

300



600

700

800

900 1000 1100 1200 1300 300 400 500 600 800 700 Wavelength (Nanometers)

simultaneous excitation of multiple fluorophores over a wide wavelength range



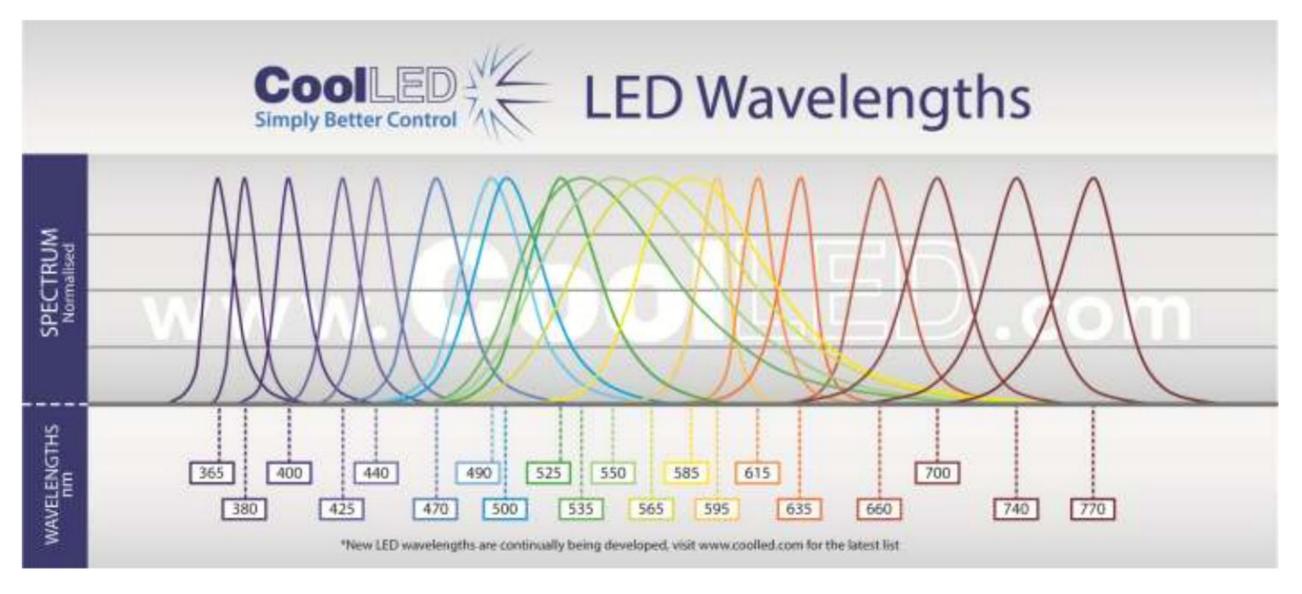


Illumination sources for widefield fluorescence microscopy



State of the art for widefield fluorescence

LEDs Light Emitting Diodes



http://www.coolled.com/product-detail/led-wavelengths/

Wide range of lines available

• 25,000 h

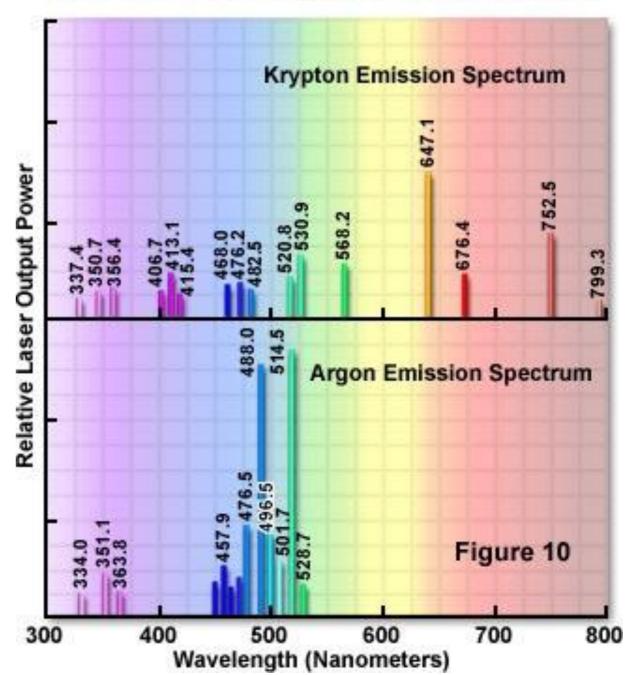
Illumination sources for widefield fluorescence microscopy



* Diode lasers * Solid State lasers

* Gas lasers

Laser Illumination Source Emission Spectra



Only discrete lines!

lines	Alex	a dye
405	405	440
440	430	540
488	488	515
514	514	540
561	568	605
633	633	645

Narrow beams of highly monochromatic,

coherent and collimated light



5. Point Spread Functions





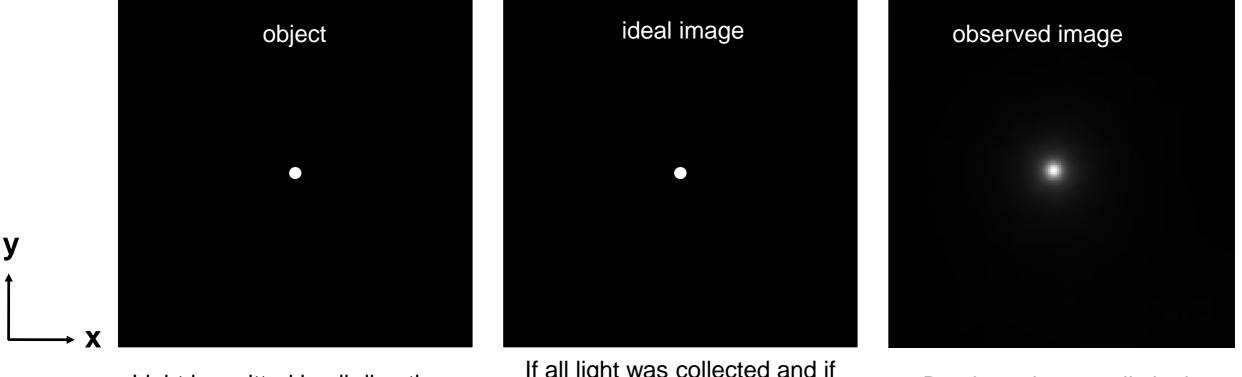
Microscope performance

Why is this important?

- x, y, z information
- Image quality
- Alignment
- Optical resolution



How does light spread out from a single point?



Light is emitted in all directions

If all light was collected and if light would travel in straight lines

But the point actually looks blurred / distorted

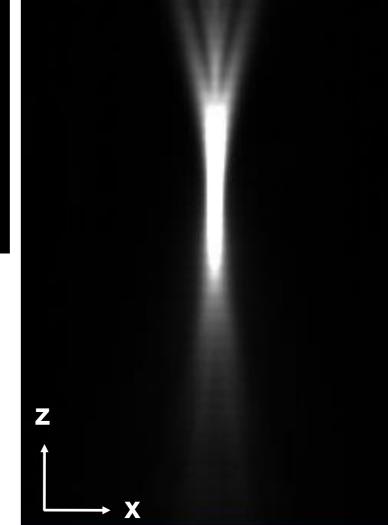
Fluorescent bead, single dye, or a fluorescent protein as a point source of light

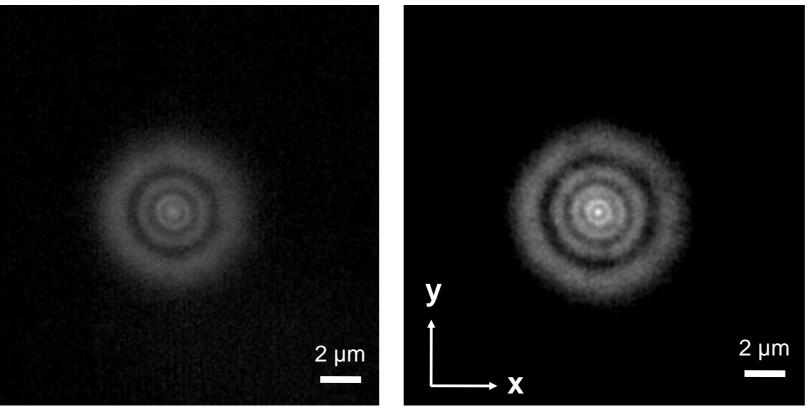
red fluorescent 170 nm bead

Airy disk diffraction pattern

(concentric rings) Light waves emitted from a point source are not focused into an infinitely small point by the objective

They converge together and interfere in the image plane







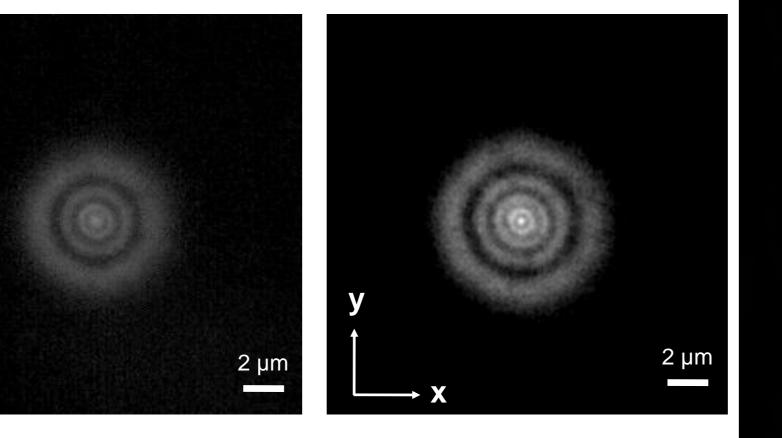
Orthogonal view

red fluorescent 100 nm bead

What can we observe?

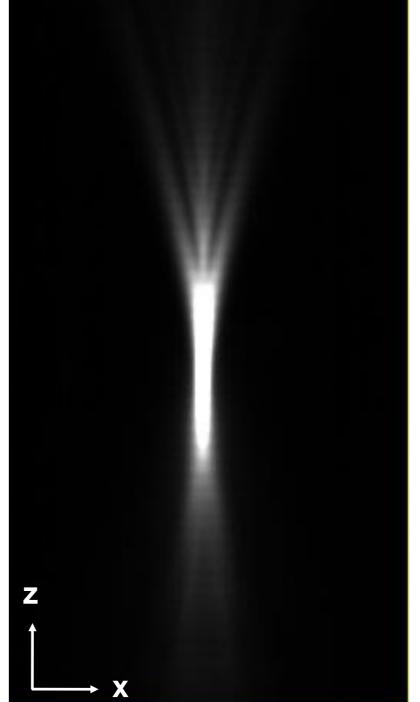
- Blur is broader in z than xy RESOLUTION
- How symmetric is the distribution

ALIGNMENT, SPHERICAL ABERRATIONS, MISMATCH REFRACTIVE INDEX





Orthogonal view

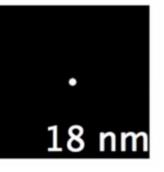


PSF of a small object









1.4NA objective $\lambda = 0.48 \ \mu m$

Image from Jeff Lichtman



6. Fixation for fluorescence microscopy



- Fixation is the preservation of the state or structure of a biological sample
- It can be achieved a number of ways, including: Chemical cross-linking UV cross-linking Dehydration

Why fix biological samples?



- 1. Convenience
- 2. Throughput
- Widely applicable molecular labeling: Immunofluorescence (cells or tissue) Fluorescence *in situ* hybridization (FISH)
- 4. Ease of multiplexing bright, relatively stable fluorescent labels

Typical Immunocytochemistry Protocol

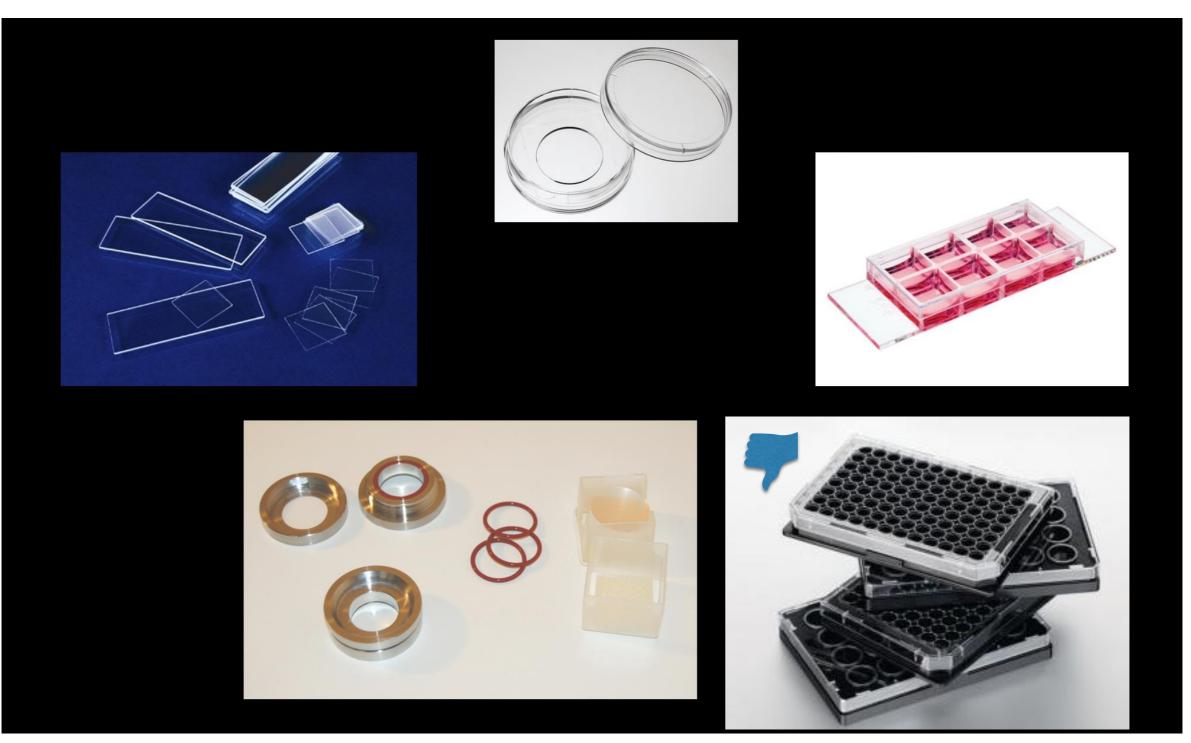
Fixation Permeabilisation Washes Blocking 1° antibody Washes 2° antibody Washes Mounting



Immobilising the specimen



Sample holder must be suitable for imaging



by Esther Garcia

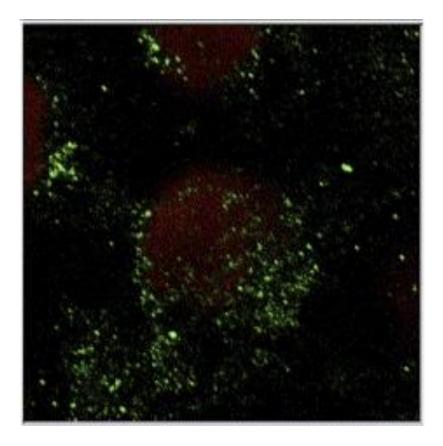
Types of Fixation



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Denaturing fixation:

Cold methanol

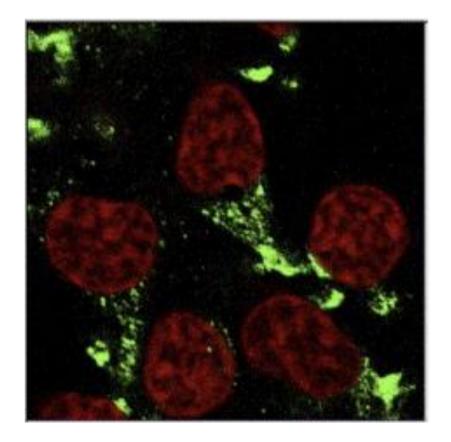


destroys 3D protein structure

dissolves lipids into micelles

Cross-linking fixation:

Formaldehyde (PFA)

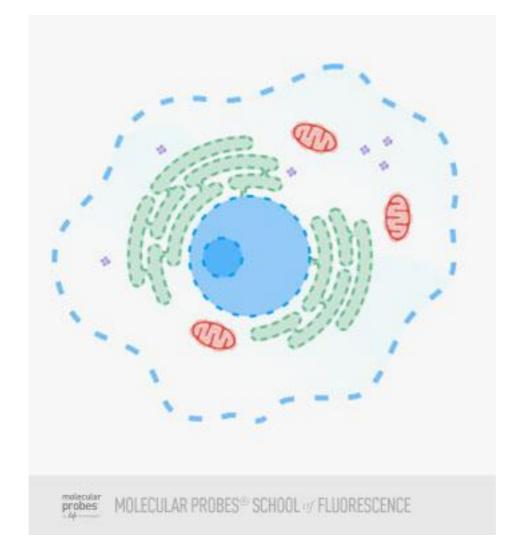


binds to proteins and some lipids, but not RNA, DNA or most sugars

Sometimes a combination of both is necessary ...



done by removing some lipids with detergents





Triton X-100

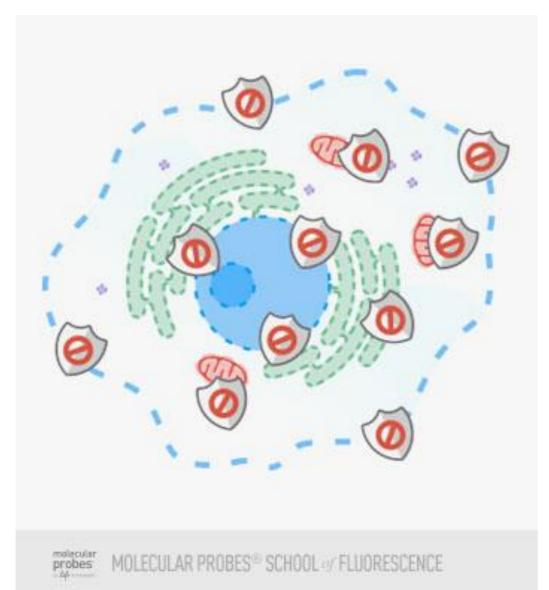
To allow large labels (antibodies) to penetrate fixed cells/tissue





Reduction of nonspecific "background" staining

done with a solution containing excess of protein



Bovine Serum Albumin (BSA)

Casein (or non-fat dry milk)

blocking proteins prevent low-affinity antibody interactions elsewhere in the sample

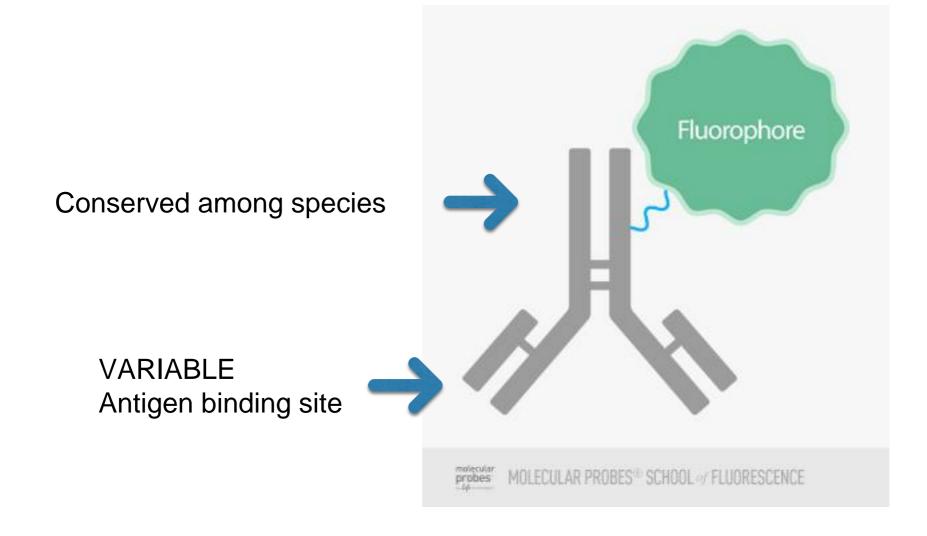
Immunolabeling (antibodies)



Antibody (large Y-shaped protein called immunoglobulin)

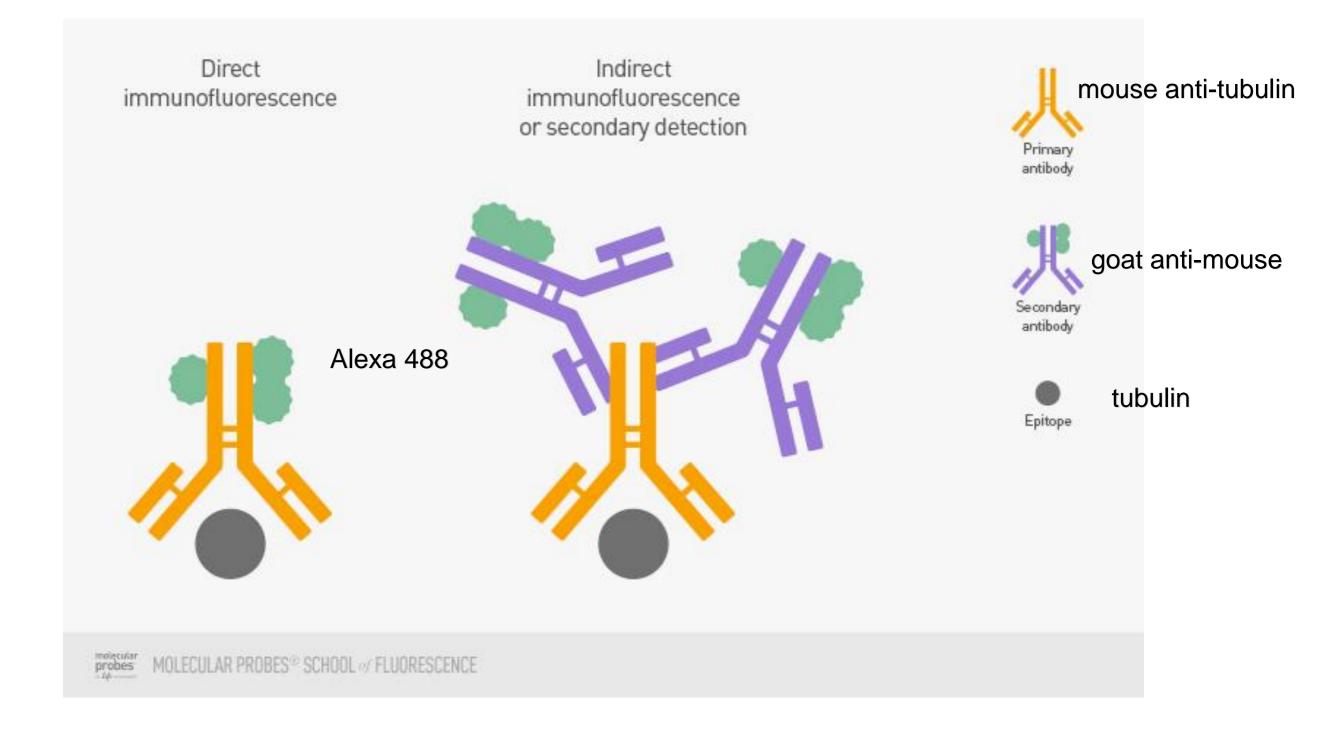
produced by the immune system, found in the blood or other body fluids of vertebrates.

The antibody recognises unique parts of the foreign target called an antigen.



Immunolabeling (antibodies)

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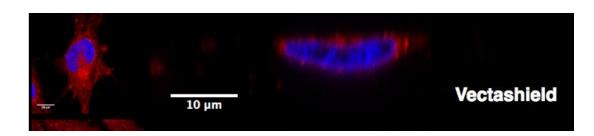


Mounting

0

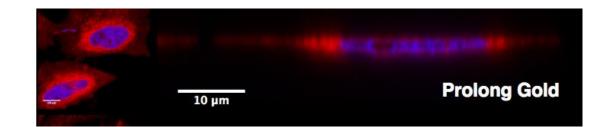
Non-hardening

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- Short-term storage (days to few weeks)
- Dabco, Glycerol, Vectashield (antifading agent, but does not work with FarRed dyes)

- Hardening
 - Long term storage (months)



- It can flatten the cell if polymerises too fast
- Prolong Gold, Vectashield hardset

Thank you for listening

Any questions?