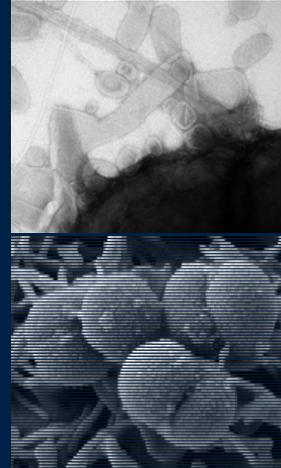


# Basic Principles of Electron Microscopy (EM)

Dr Errin Johnson  
Head of the Dunn School EM Facility



## Basic Principles *Brief history of EM*

1873 – Hermann von Helmholtz & Ernst Abbe show that the wavelength of light affects optical resolution

1924 – Louis de Broglie theorised the wave/particle duality of electrons

1926 – Hans Busch demonstrated that magnetic lenses can manipulate the path of electrons in the same way as optical lenses do with light

1932 – Ernst Ruska & Max Knoll invent the TEM

1934 – Ladislaus Marton publishes the first biological EM micrograph

1937 – Manfred von Ardenne builds the first SEM

1951 – Erwin Muller develops the field emission microscope for atomic resolution

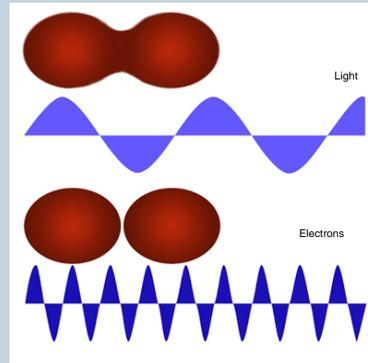
1951 – Albert Claude *et al* publish the first TEM image of an intact cell

For a detailed history of EM, see:  
1. Haguenu et al. (2003) Key events in the history of electron microscopy. *Microscopy & Microanalysis*, 9(2): 96-138.  
2. Masters, B (2009) History of the electron microscope in cell biology. eLS, Wiley.



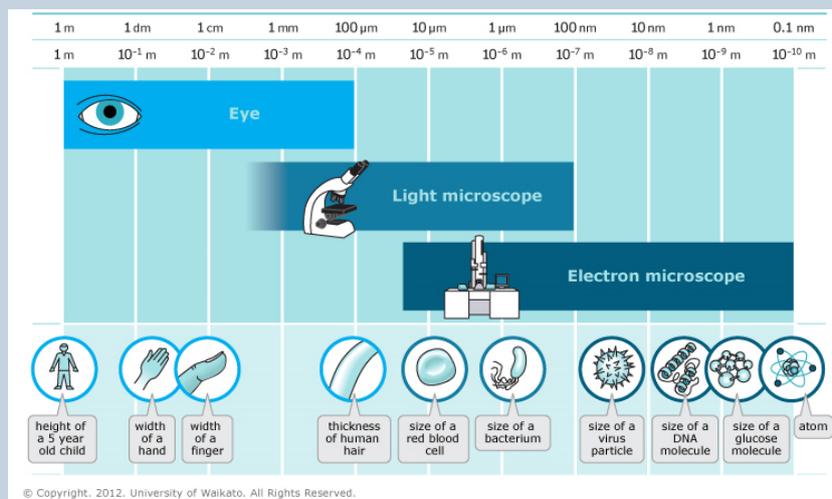
## Basic Principles *Resolution*

- Resolution is the smallest distance at which two neighbouring points can be distinguished and is dependent on wavelength
- The wavelength of accelerated electrons (6 pm) is several orders of magnitude shorter than that of light (600 nm)

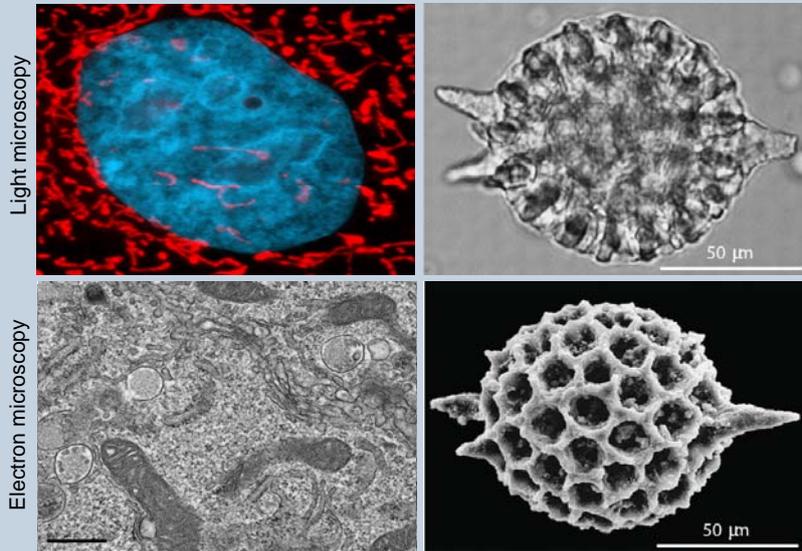


www.ammr.org

## Basic Principles *Resolution*



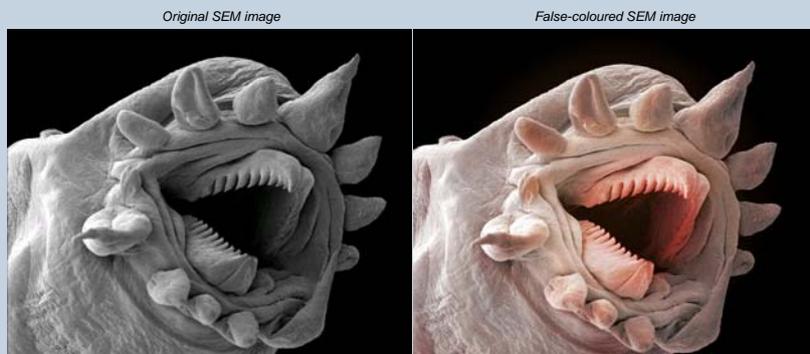
## Basic Principles *Resolution and Contrast*



TEM image of fibroblast cell (E Johnson). Bottom: Confocal image of a kidney cell stained with DAPI (blue) and MitoTracker (red) (Hamamatsu.magnet.fsu.edu) Radiolarian imaged with both SEM (top) and light microscopy (bottom). From: General Chemistry: Principles, Patterns, and Applications, B. Averill & P. Elderege

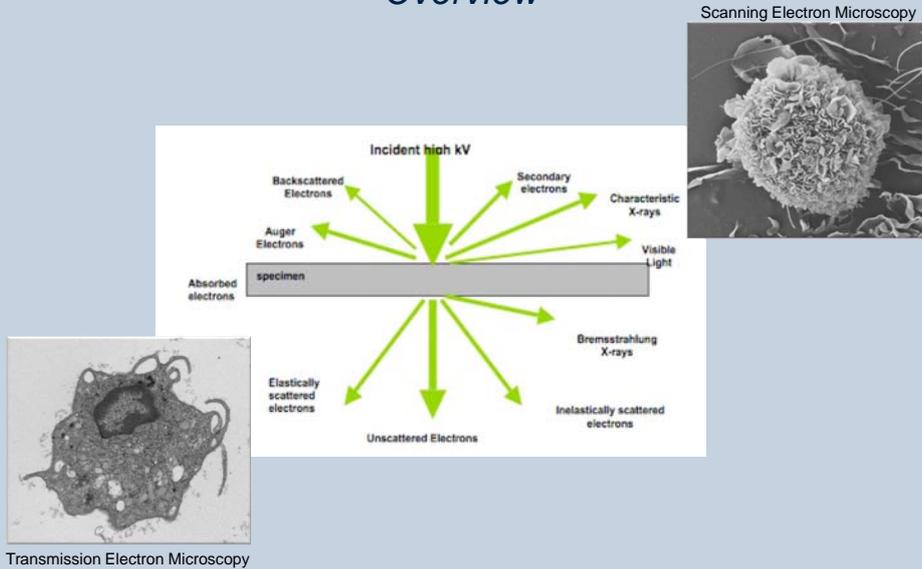
## Basic Principles *Signal detection*

- EM images are monochromatic and are essentially intensity maps of the number of electrons that are detected from a given point. False colour may be added during post-processing of the image, if desired.

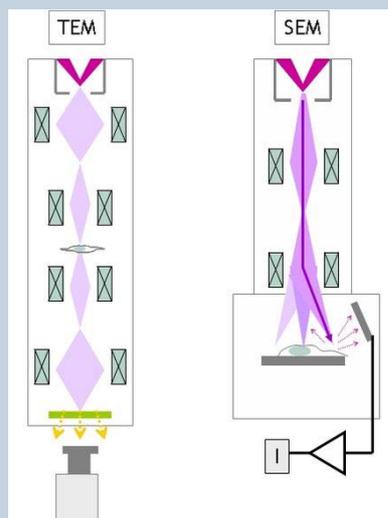


Hydrothermal worm (x525) by Philippe Crassous, FEI.com

## Features of Electron Microscopes Overview



## Features of Electron Microscopes Overview



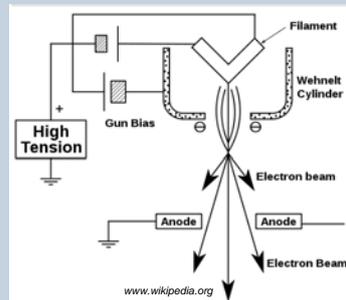
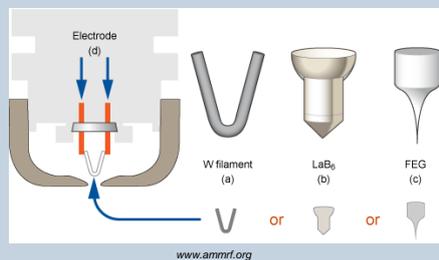
The main components of an electron microscope are:

- An electron gun
- Electromagnetic lens system
- Vacuum system
- Camera/detector
- Computer

## Features of Electron Microscopes

### *The Electron gun*

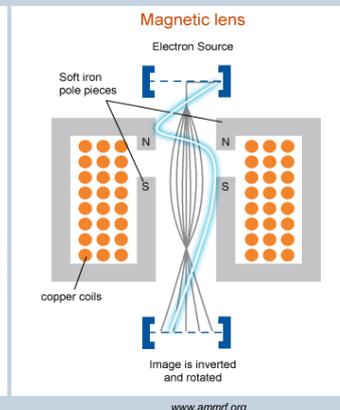
- The gun consists of an electron source, electrode, Wehnelt assembly and anode
- Electron sources are typically Tungsten or LaB6 and can be thermionic or field emission (FEG). A current is run through the filament/crystal to heat it or, a field is applied to the tip, resulting in the emission of electrons from the tip. The high voltage difference between the cap and the anode causes the electrons to accelerate and form a beam



## Features of Electron Microscopes

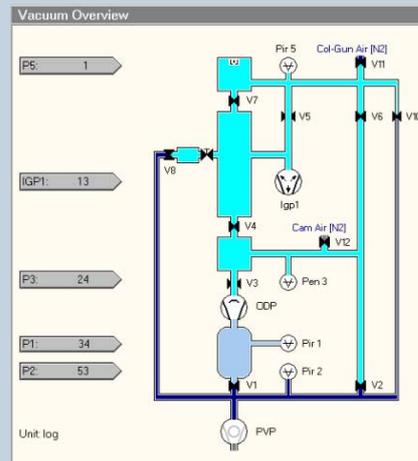
### *Electromagnetic Lenses*

- EM lenses are electromagnetic, creating precise, circular magnetic fields that manipulate the electron beam, much the same way that optical lenses focus and direct light



## Features of Electron Microscopes *Vacuum systems*

- EMs have elaborate pumping systems to ensure that the microscope is operated under a high vacuum ( $10^{-4}$  Pa)
- Maintains the integrity of the electron beam, as any interaction with gas atoms will cause the beam to scatter
- Avoids arcing between the cathode and ground (and damage to the filament)

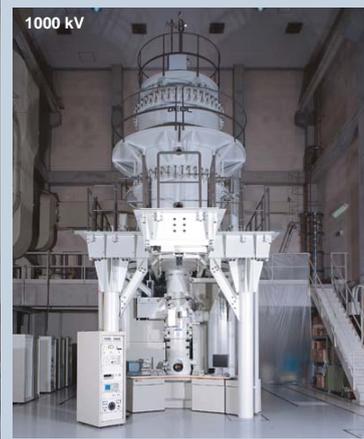


## Transmission Electron Microscopy (TEM)

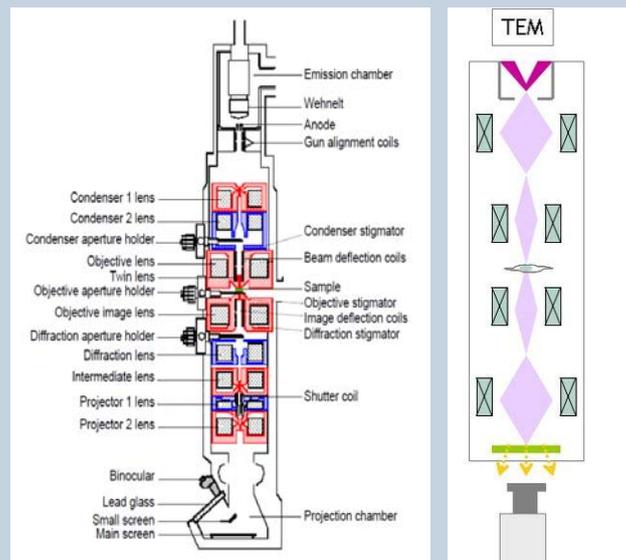


## The TEM

- Resolution depends on a number of factors, including the accelerating voltage, the type of electron source used and how you setup the microscope
- Accelerating voltage (kV) is typically 80-300 kV for biological specimens

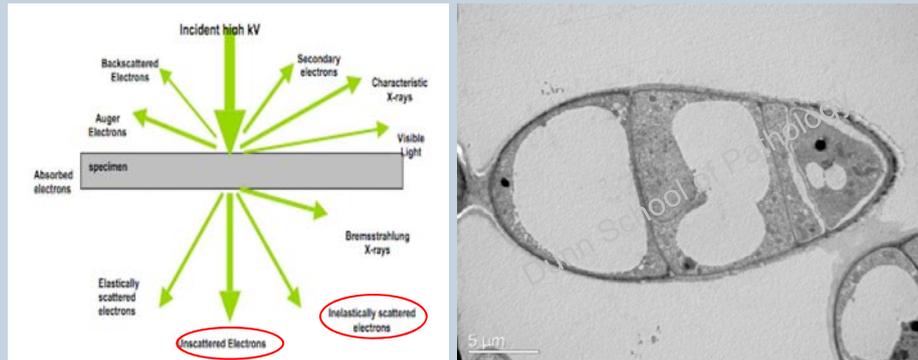


## The TEM



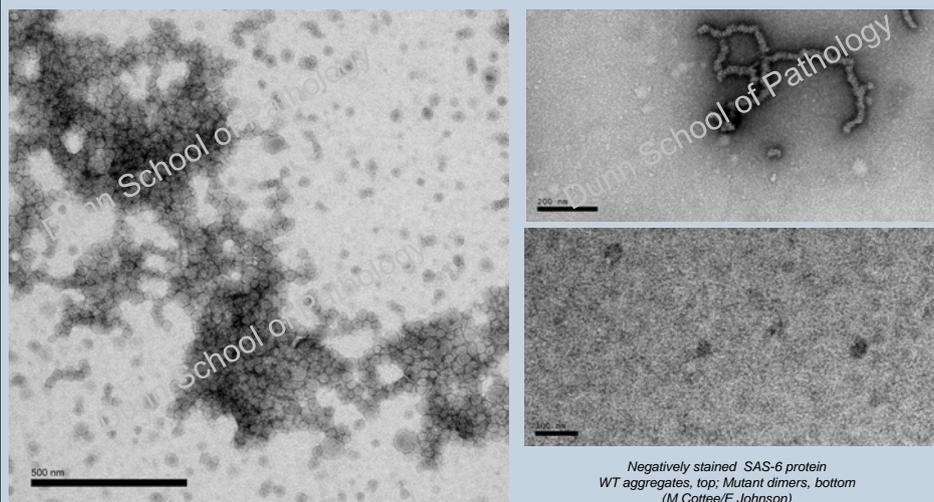
## The TEM Contrast

- Contrast is generated by density differences within the sample.
- Darker areas in the image are where few electrons have been transmitted through the sample, due to thickness or high atomic number.



Lavender trichome, E Johnson

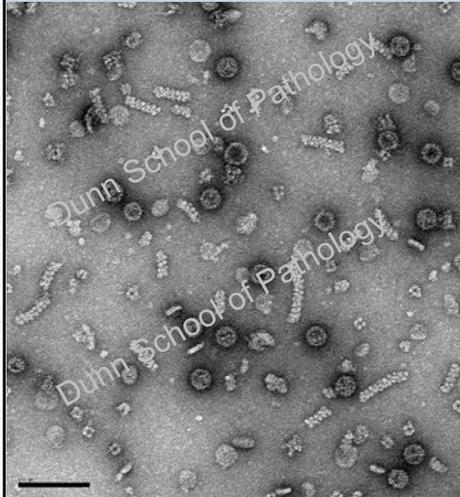
## TEM applications Particulate samples



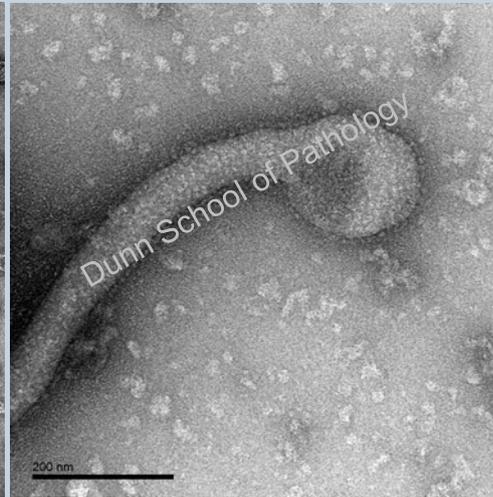
Negatively stained  $\alpha$ -synuclein  
E Johnson

Negatively stained SAS-6 protein  
WT aggregates, top; Mutant dimers, bottom  
(M Cottee/E Johnson)

## TEM imaging *Particulate samples - Viruses*

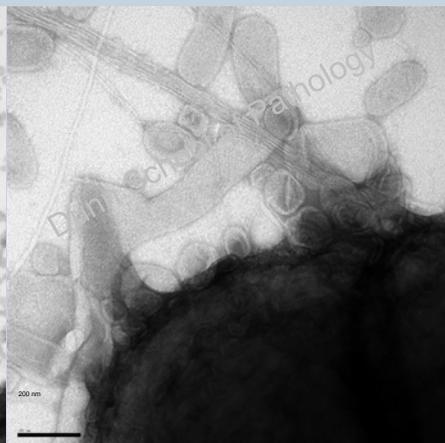
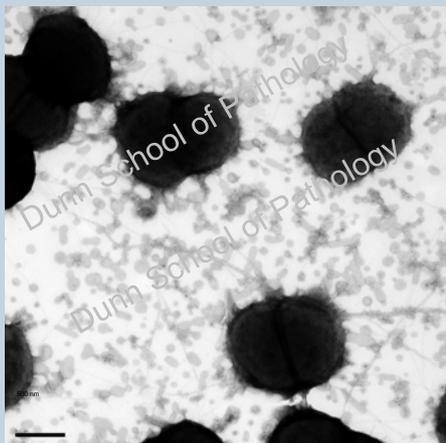


Negatively stained virus-like particles  
(D Leneghan/E Johnson)



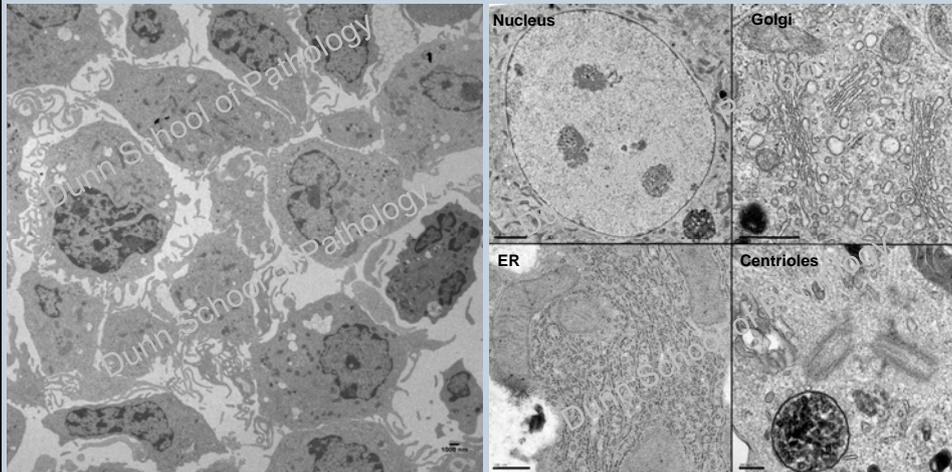
Negatively stained virus-like particles (Ebola)  
(E Johnson)

## TEM imaging *Particulate samples - Bacteria*



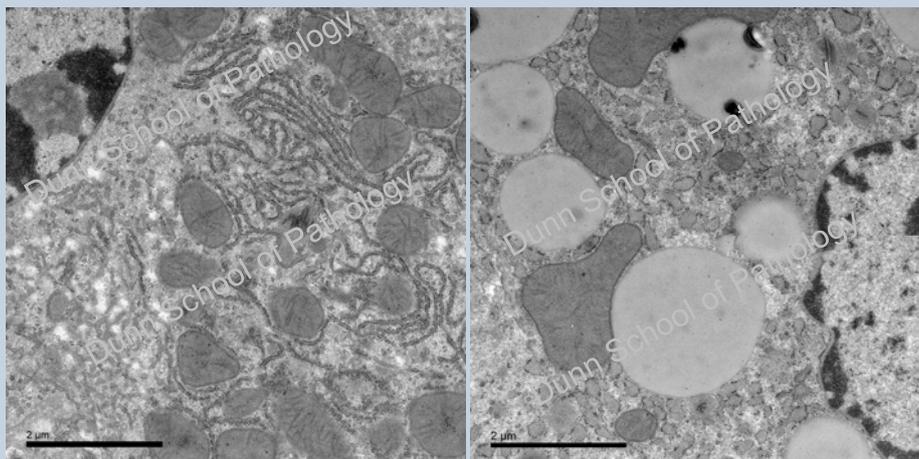
Negatively stained *Neisseria* sp. (R Exley/EJohnson)

## TEM Applications *Ultrastructure - Cells*



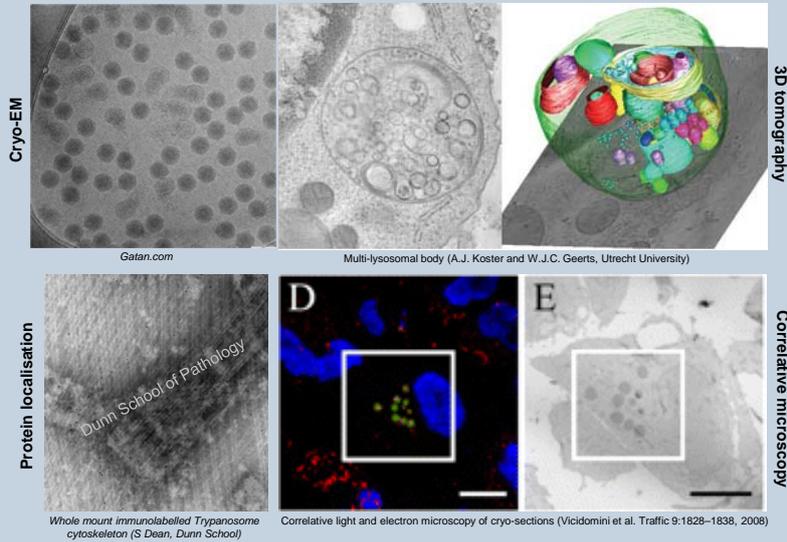
Mammalian culture cells (E Johnson)

## TEM Applications *Ultrastructure - Tissue*



Mouse liver tissue, untreated (left) and under ER stress (right) (V Liebe & E Johnson)

## TEM Applications *Advanced techniques*

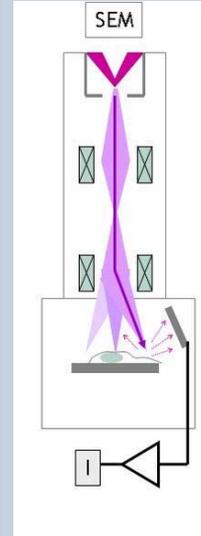
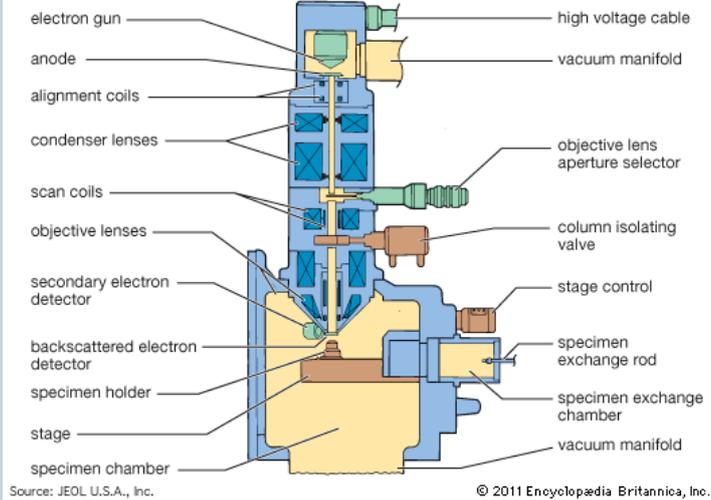


## Scanning Electron Microscopy (SEM)



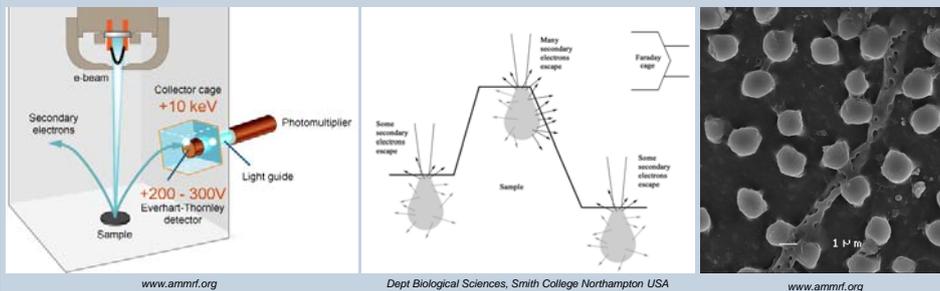
## The SEM

### Parts of a scanning electron microscope



## The SEM Signal detection

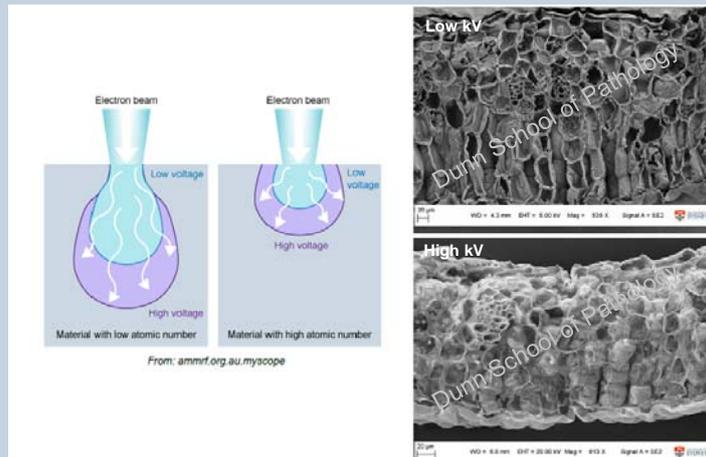
- Secondary electrons (SEs) provides surface morphology and topology information.
- SEs are captured by the Everhart-Thornley detector



## The SEM

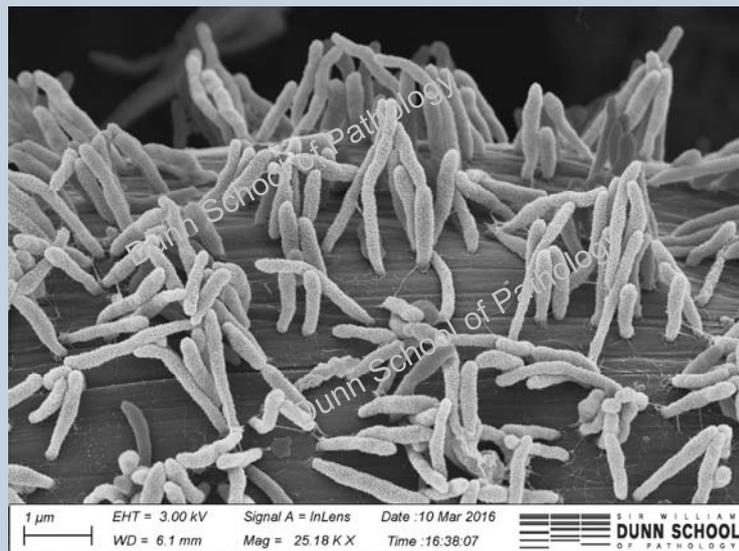
### *Accelerating voltage*

- Resolution depends on a number of factors, including the accelerating voltage, the type of electron source used and how you setup the microscope and the scan
- Accelerating voltage (kV) is typically 1-10 kV for biological specimens



## SEM Applications

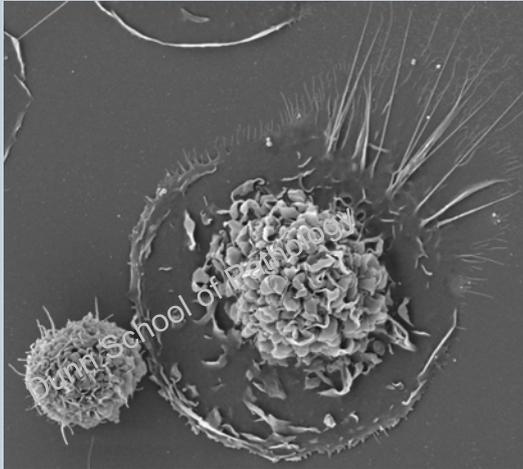
### *Topography - Cells*



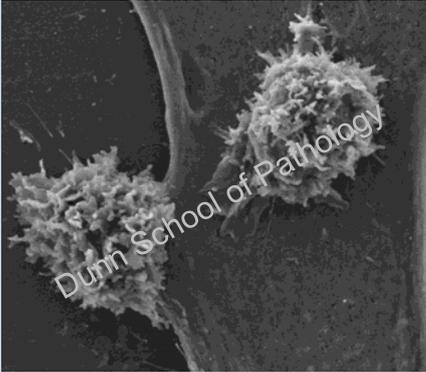
Bacteria on carbon electrodes  
(s Putra/E. Johnson)

# SEM Applications

## Topography - Cells



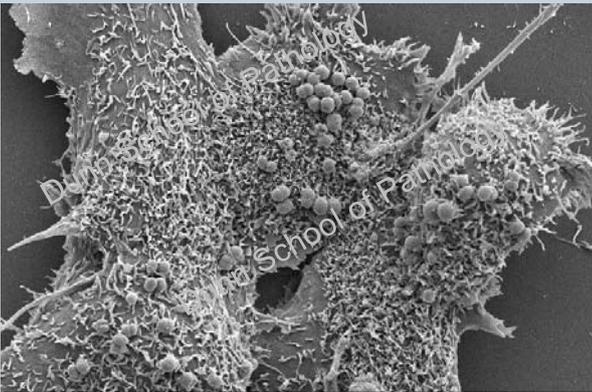
Monocyte and macrophage  
(B van Wilgenburg/E Johnson)



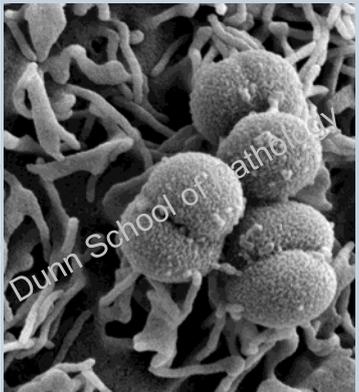
HEK 293 cell interacting with streptavidin coated  
dynabeads plus biotinylated anti-EPCAM (M Brenner/E Johnson)

# SEM Applications

## Topography - Cells



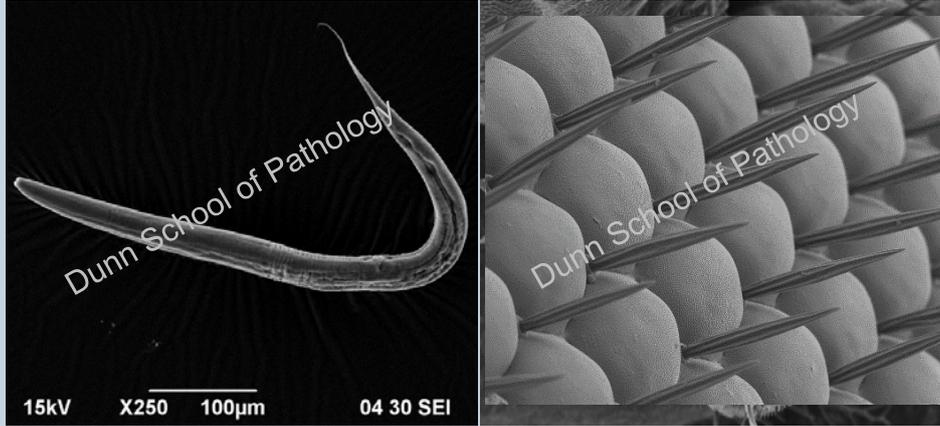
2 µm EHT = 3.50 kV Signal A = SE2 Date: 14 May 2016  
WD = 7.1 mm Mag = 7.40 K X Time: 12:11:49 ZEISS



200 nm EHT = 3.50 kV Signal A = SE2  
WD = 7.1 mm Mag = 61.74 K X

*Neisseria sp. on epithelial cells* (R Exley/E Johnson)

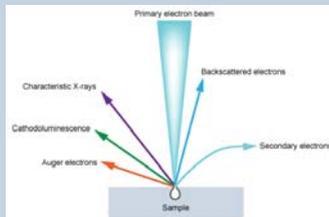
## SEM Applications *Topography - Organisms*



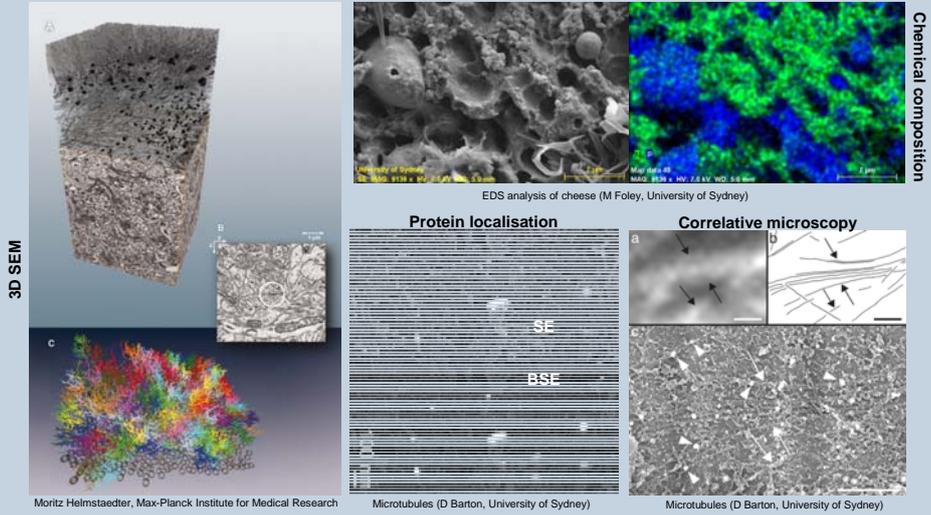
*C. elegans*  
(E Johnson/A Moloney, Dunn School)

*Drosophila* rough eye phenotype  
(M Elschami, NDCN)

## The SEM *Diverse imaging capabilities*



## Biological Applications SEM - Advanced



## Questions?



A happy Leishmania nucleus  
(J Valli)