

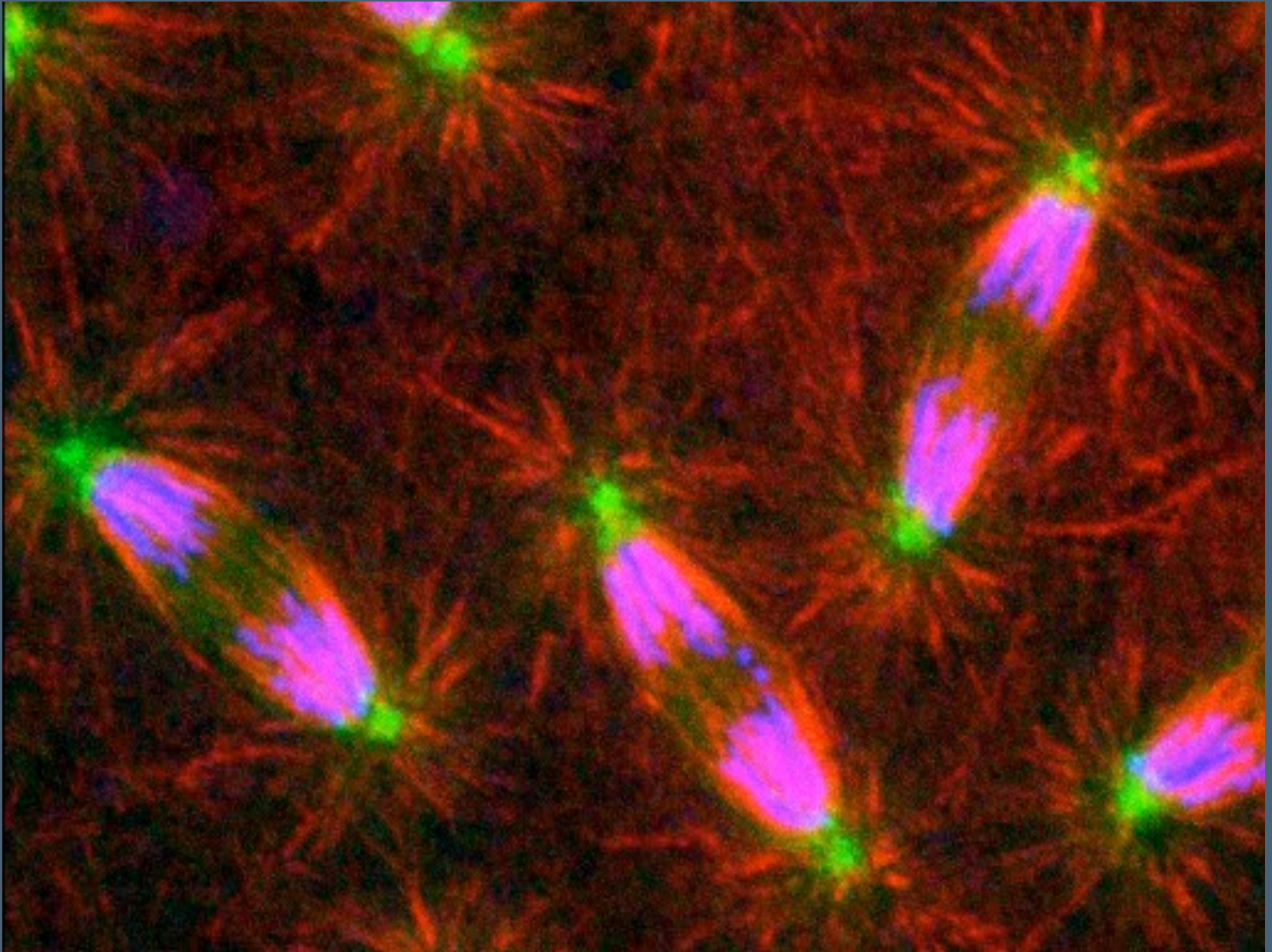
Confocal Microscopy

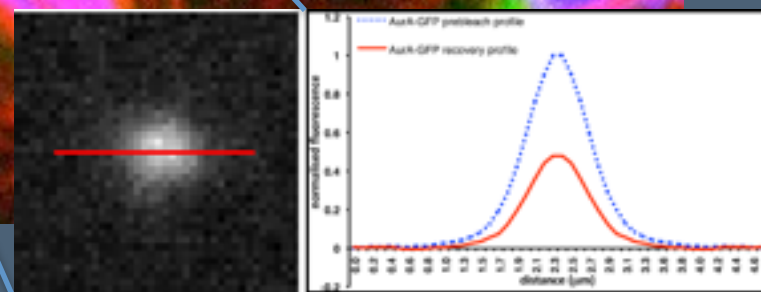
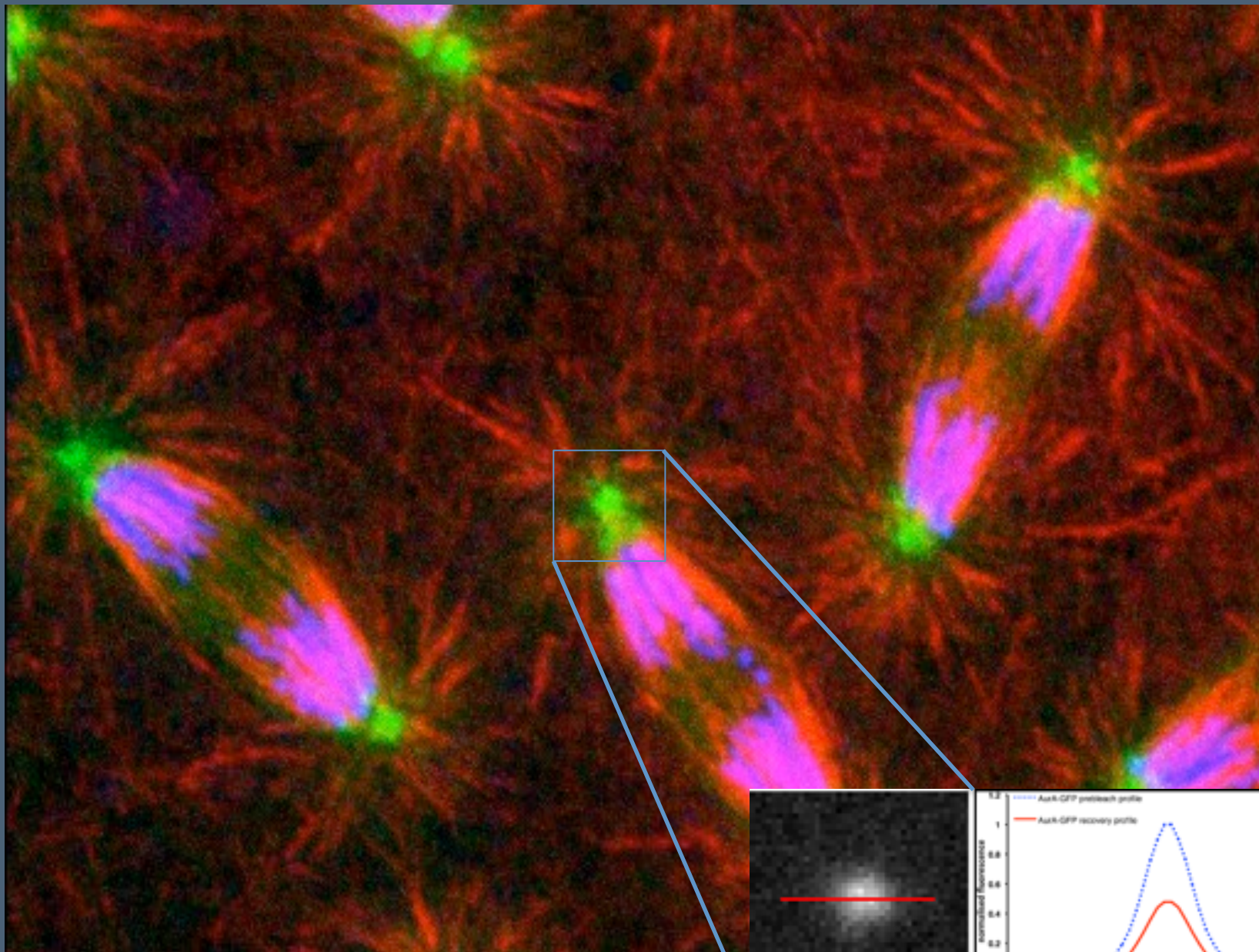
(Increasing contrast and resolution using optical sectioning)

Lecture 7

November 2016



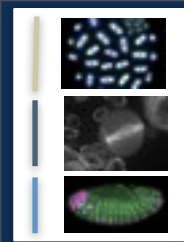




3 Flavours of Microscope

Problem:
Out of Focus
Light

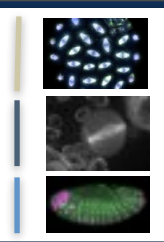
3 Flavours of Microscope



Problem:
Out of Focus
Light



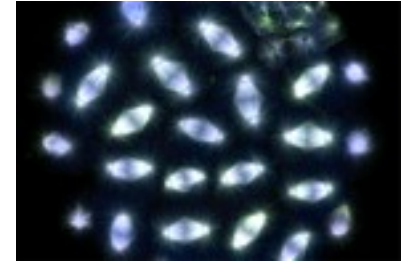
3 Flavours of Microscope



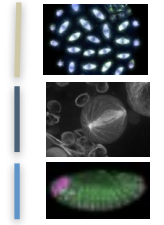
Problem:
Out of Focus
Light



Laser
Scanning



3 Flavours of Microscope

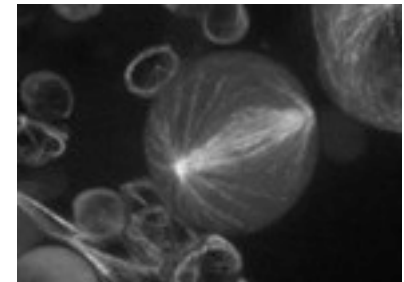
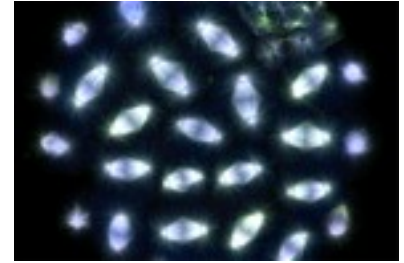


Problem:
Out of Focus
Light

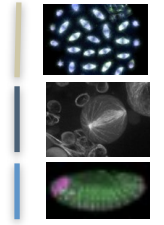


Laser
Scanning

Spinning disc



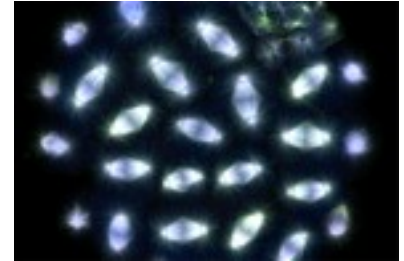
3 Flavours of Microscope



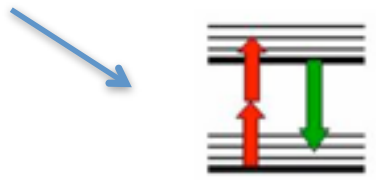
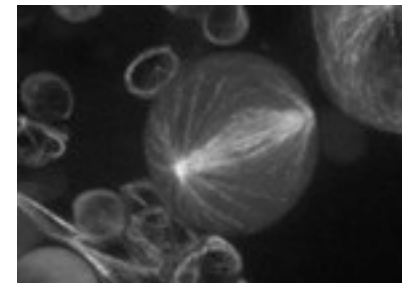
Problem:
Out of Focus
Light



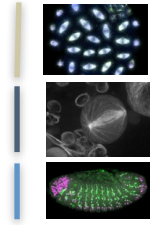
Laser
Scanning



Spinning disc



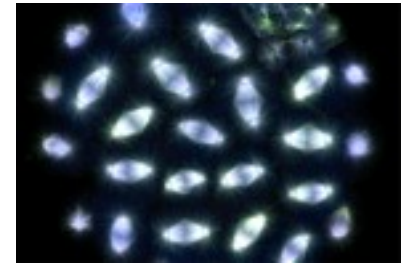
3 Flavours of Microscope



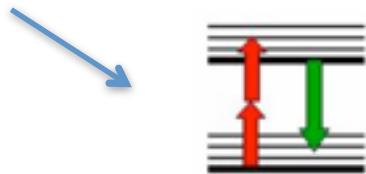
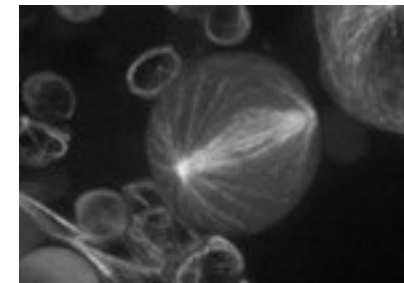
Problem:
Out of Focus
Light



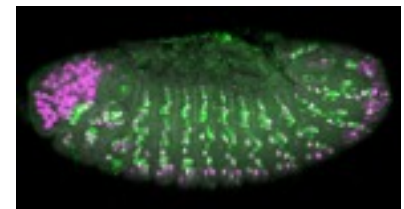
Laser
Scanning



Spinning disc

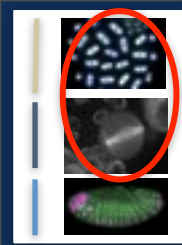


2-Photon





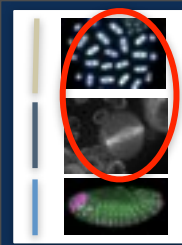
short History of Confocal Microscope



short History of Confocal Microscope

Confocal “concept’ patented by Marvin Minsky in 1957



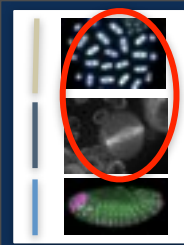


short History of Confocal Microscope

Confocal “concept’ patented by Marvin Minsky in 1957



Eggar and Petran developed “spinning disc” confocal in late 1960s



short History of Confocal Microscope

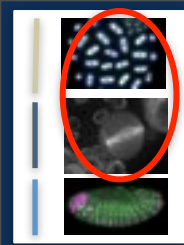
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Brakenhoff, Stelzer developed “stage” scanning confocal in late 1970





short History of Confocal Microscope

Confocal “concept’ patented by Marvin Minsky in 1957



Eggar and Petran developed “spinning disc” confocal in late 1960s

Brakenhoff, Stelzer developed “stage” scanning confocal in late 1970



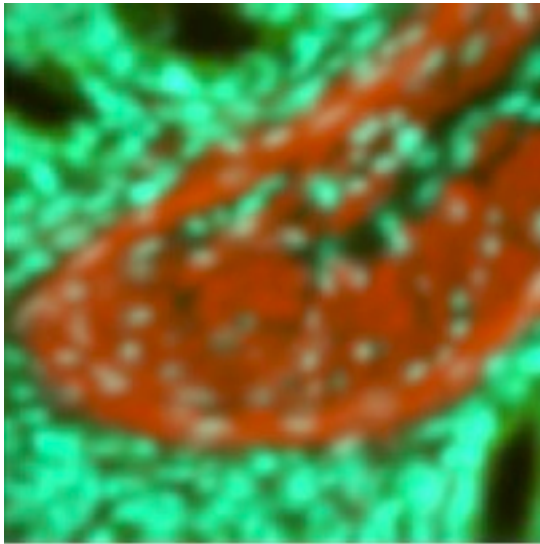
White, Amos and Wilson developed the MRC500 point scanning confocal
-Marketed commercially in 1987





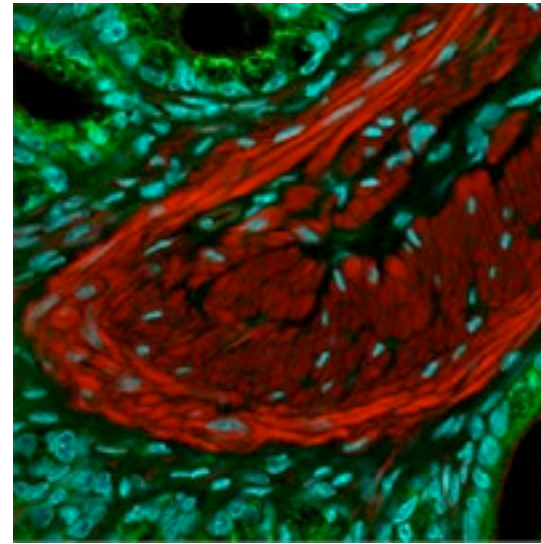
Comparison Widefield Vs Confocal

Widefield

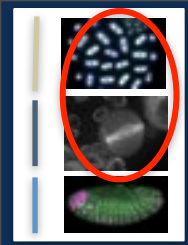


Out of focus light 'blurs' image

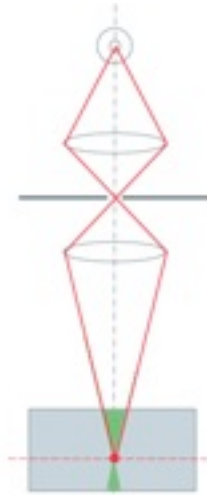
Confocal



Out of focus light is blocked



Principle of Confocal Microscopes Pinhole

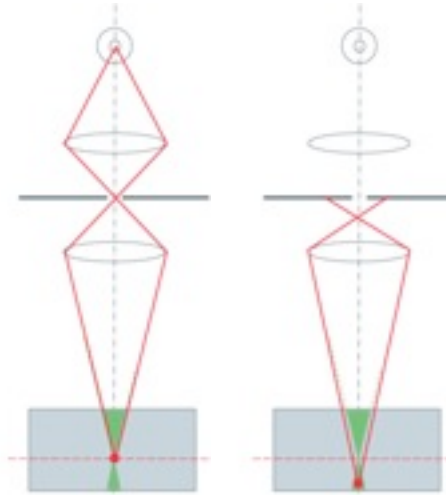


Pinhole diaphragm in the
Conjugated focal plane =
CONFOCAL

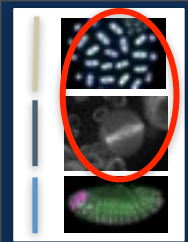
in focus light (from the optical section) passes
through the pinhole and into the detector



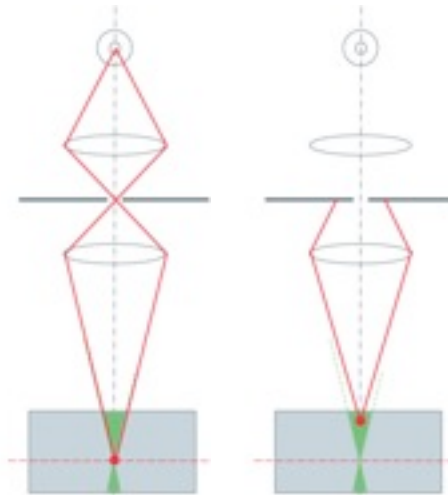
Pinhole – blocks out-of-focus light



light from below the optical section crosses in front of the pinhole and doesn't pass through the pinhole aperture

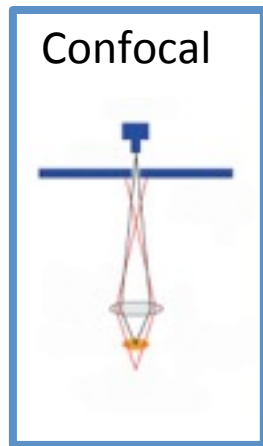


Pinhole – blocks out-of-focus light

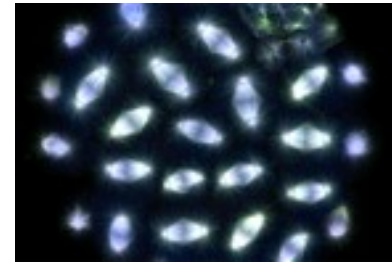


light from above the optical section also doesn't
pass through the pinhole aperture

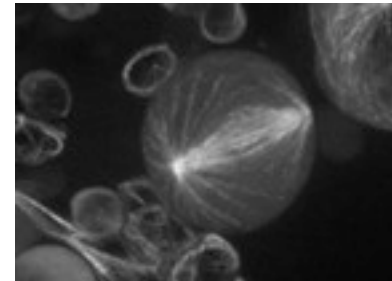
Confocal Microscopes



Laser
Scanning



Spinning disc



Laser Scanning Confocal

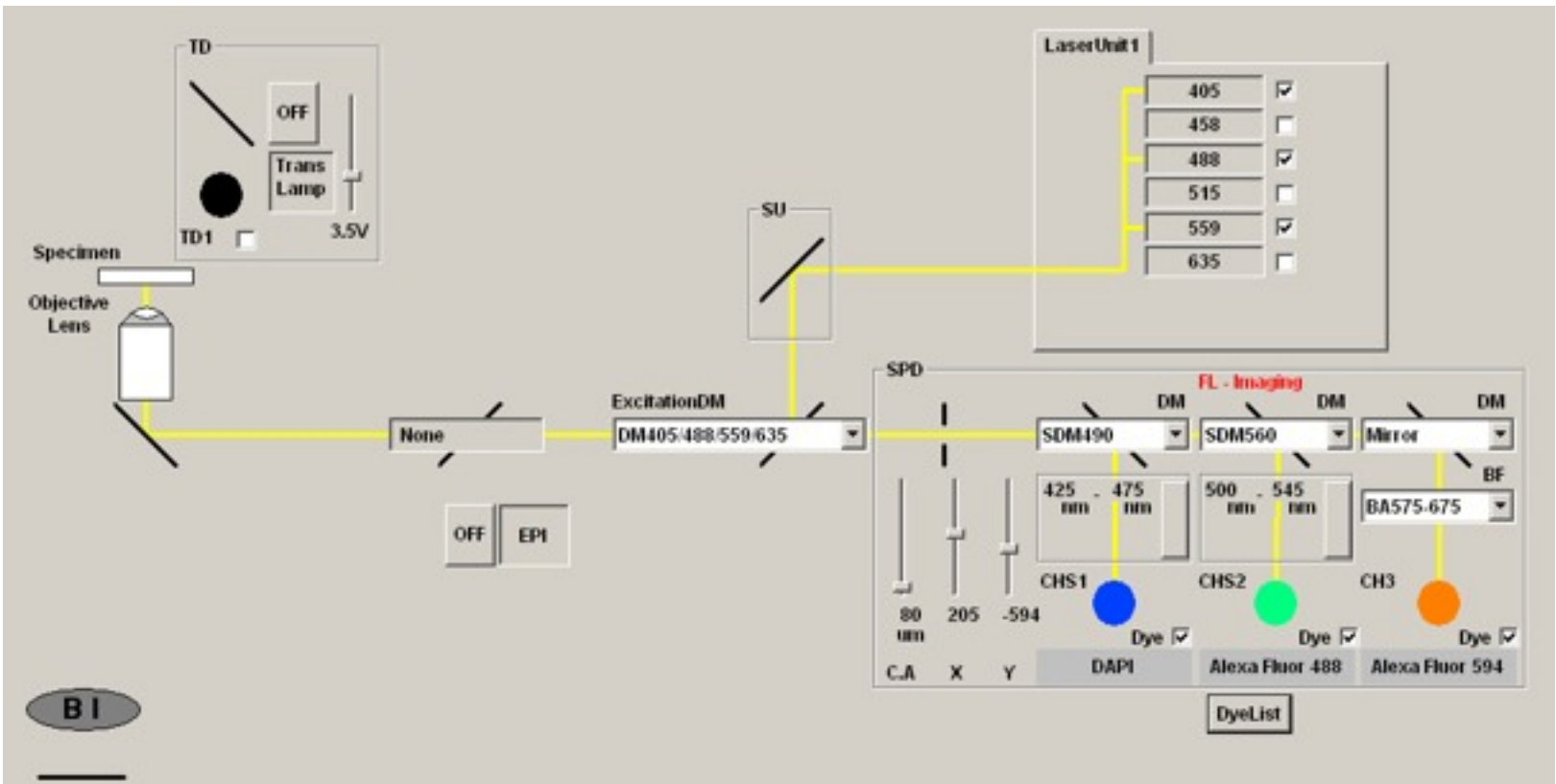


Laser Scanning Confocal

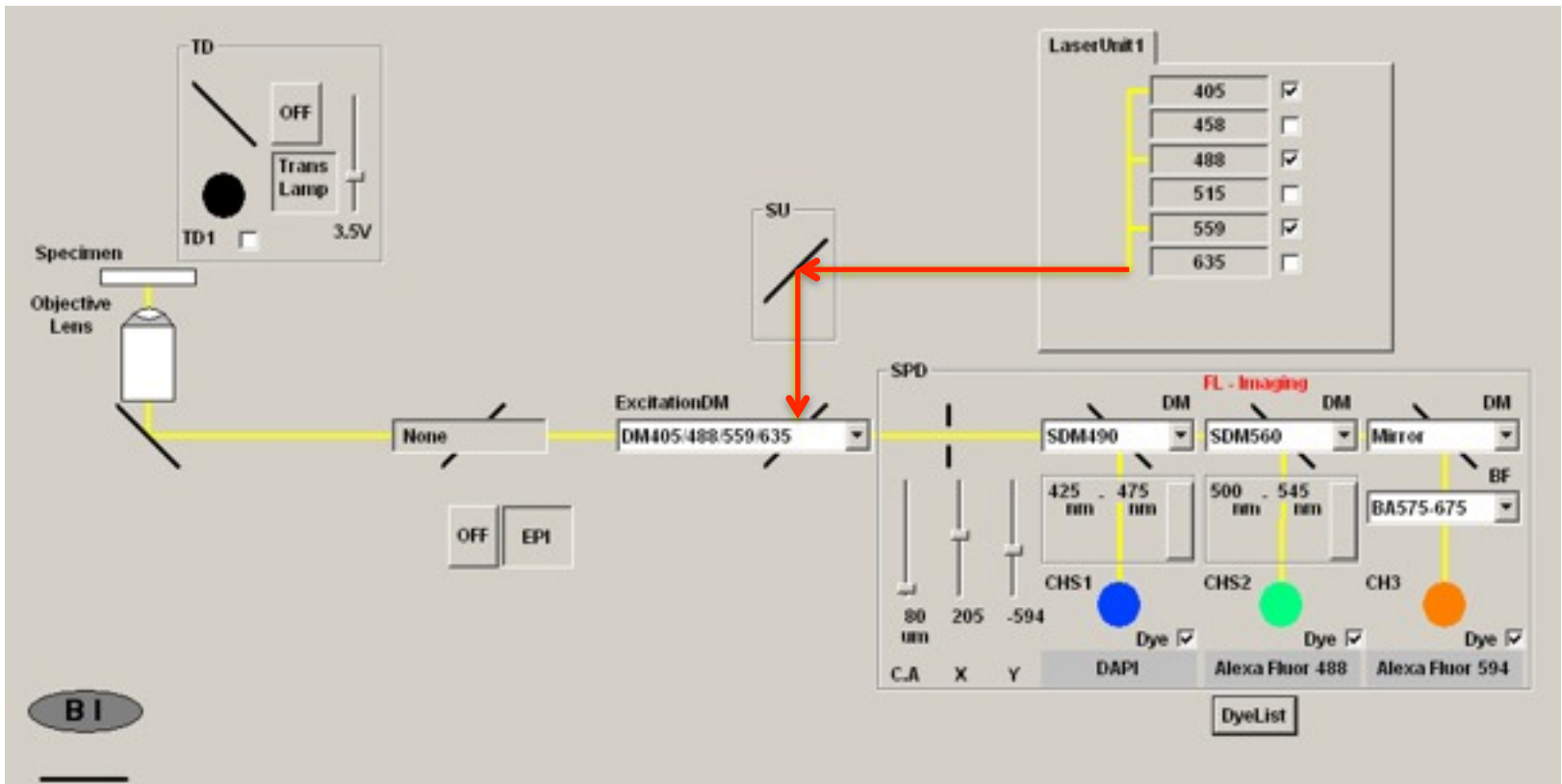


Laser Scanning Confocals are great to get 'pretty' images

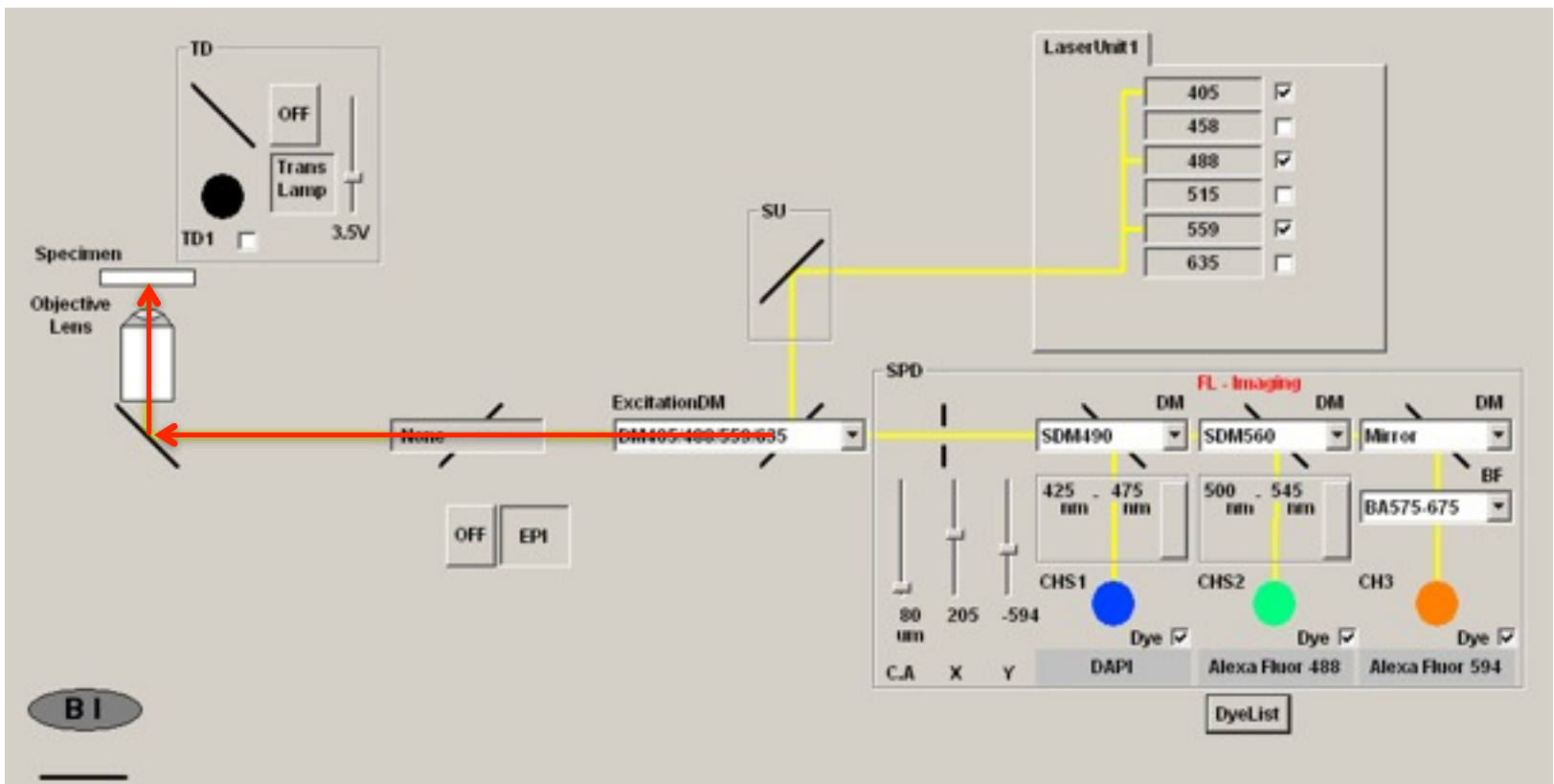
Laser Scanning Confocal



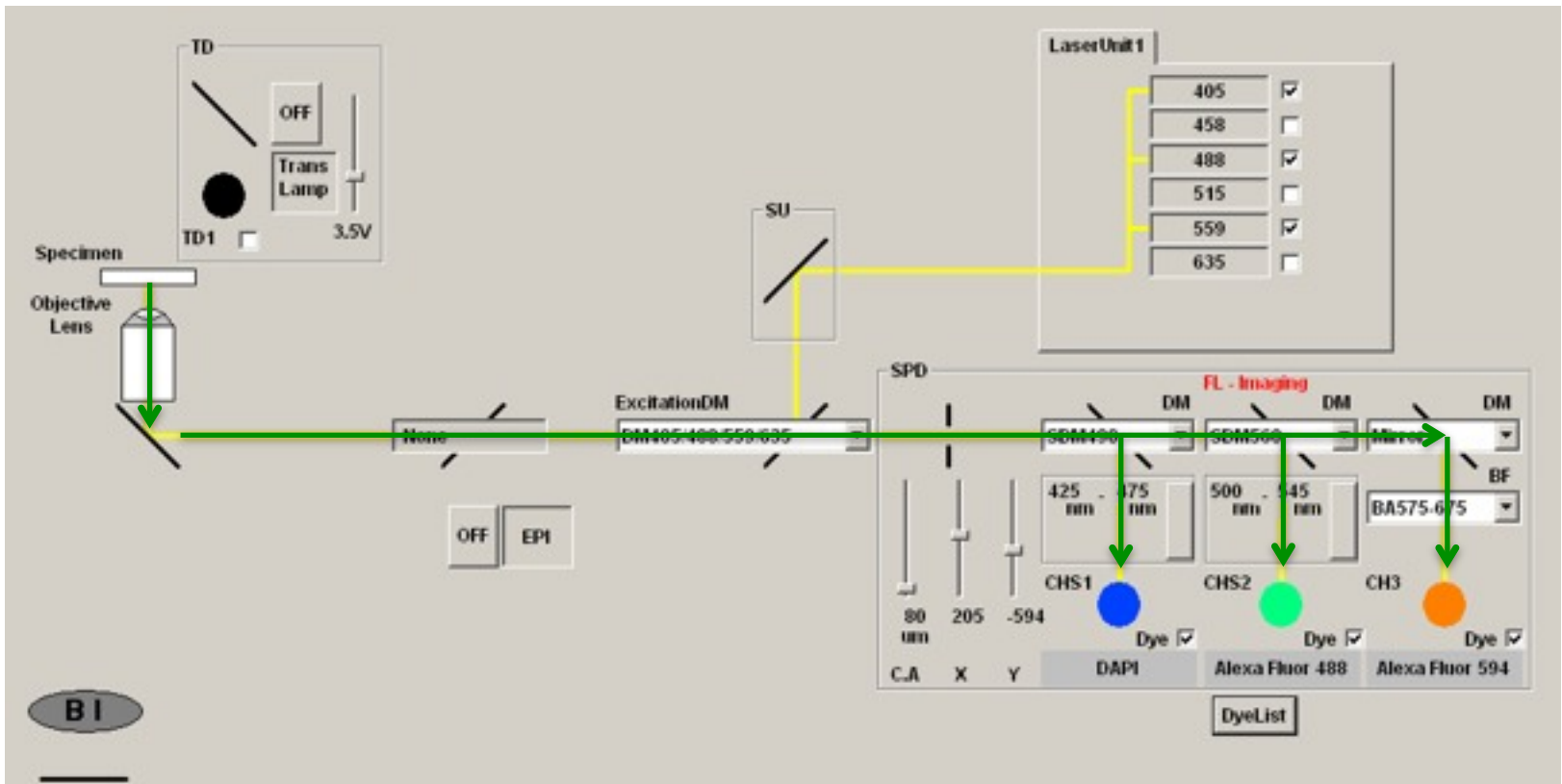
Laser Scanning Confocal



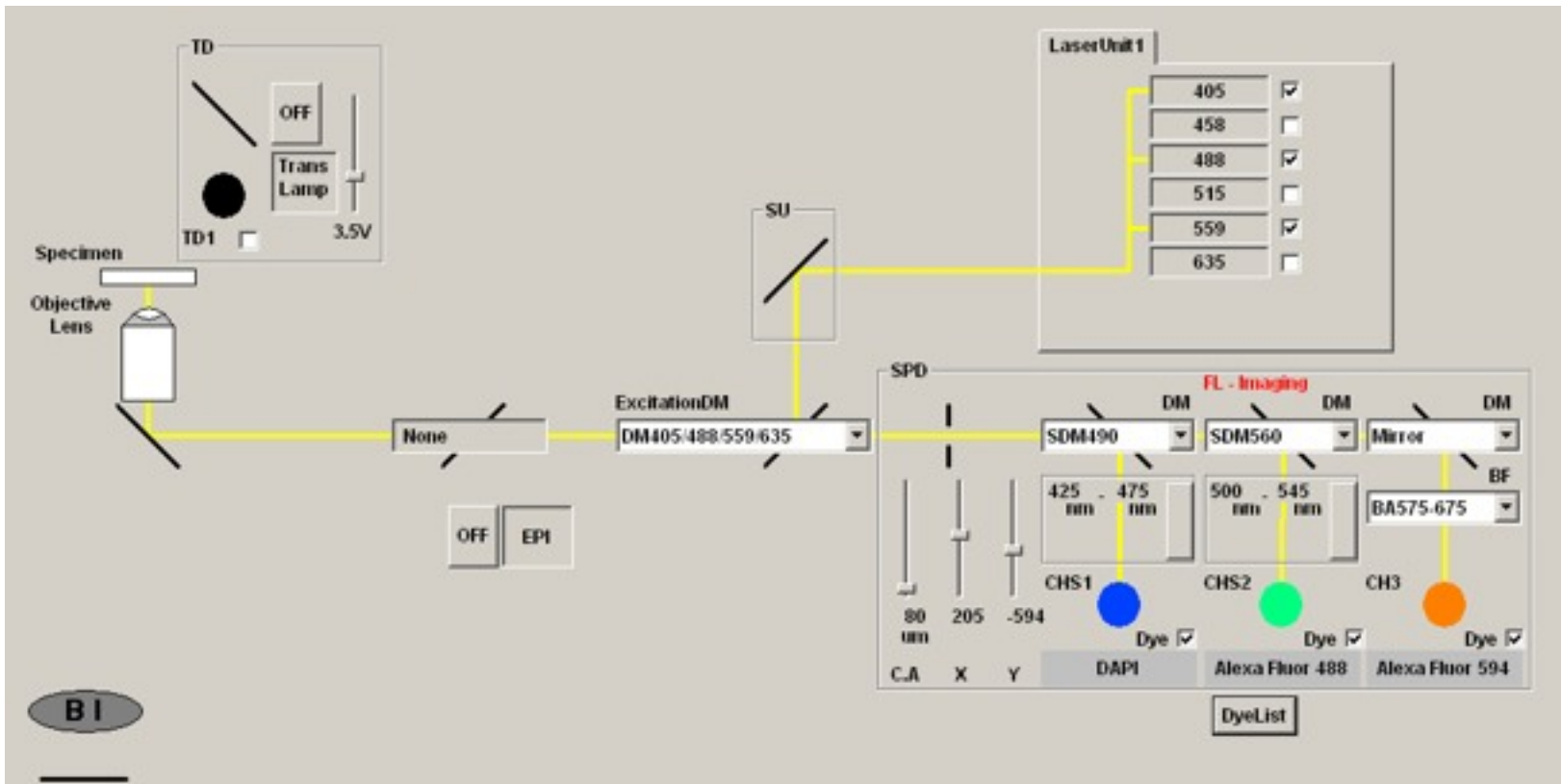
Laser Scanning Confocal



Laser Scanning Confocal

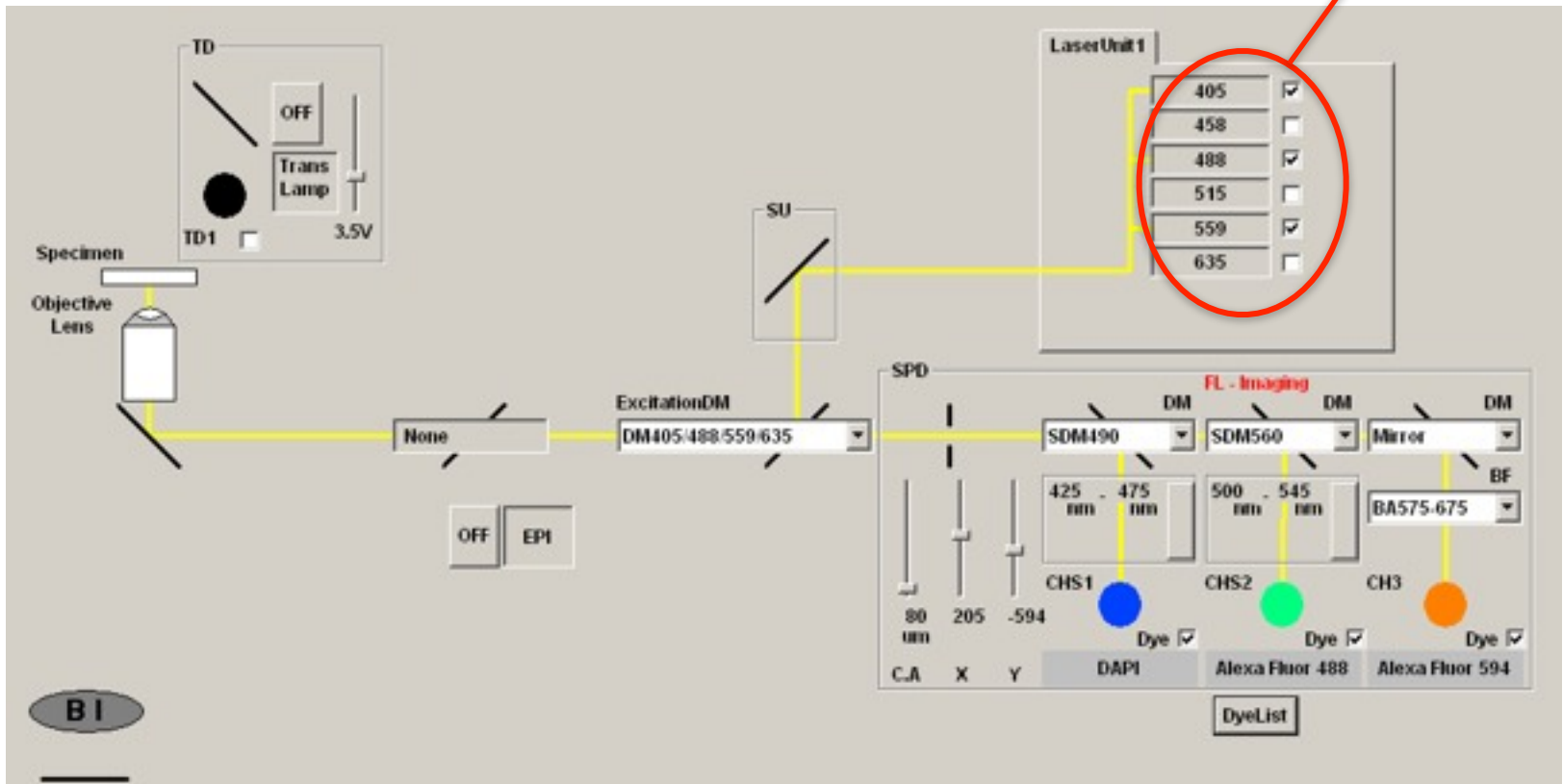


Laser Scanning Confocal



Laser Light Source

laser light source



Laser Light Source



Laser Emission Spectra

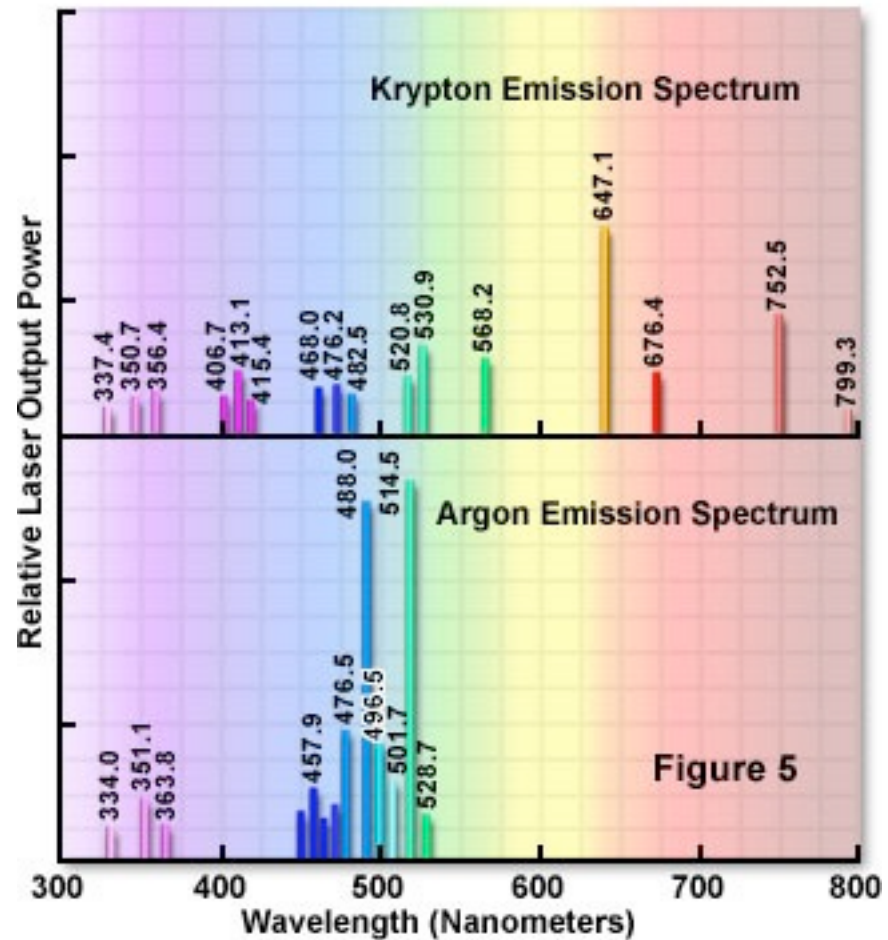


Figure 5

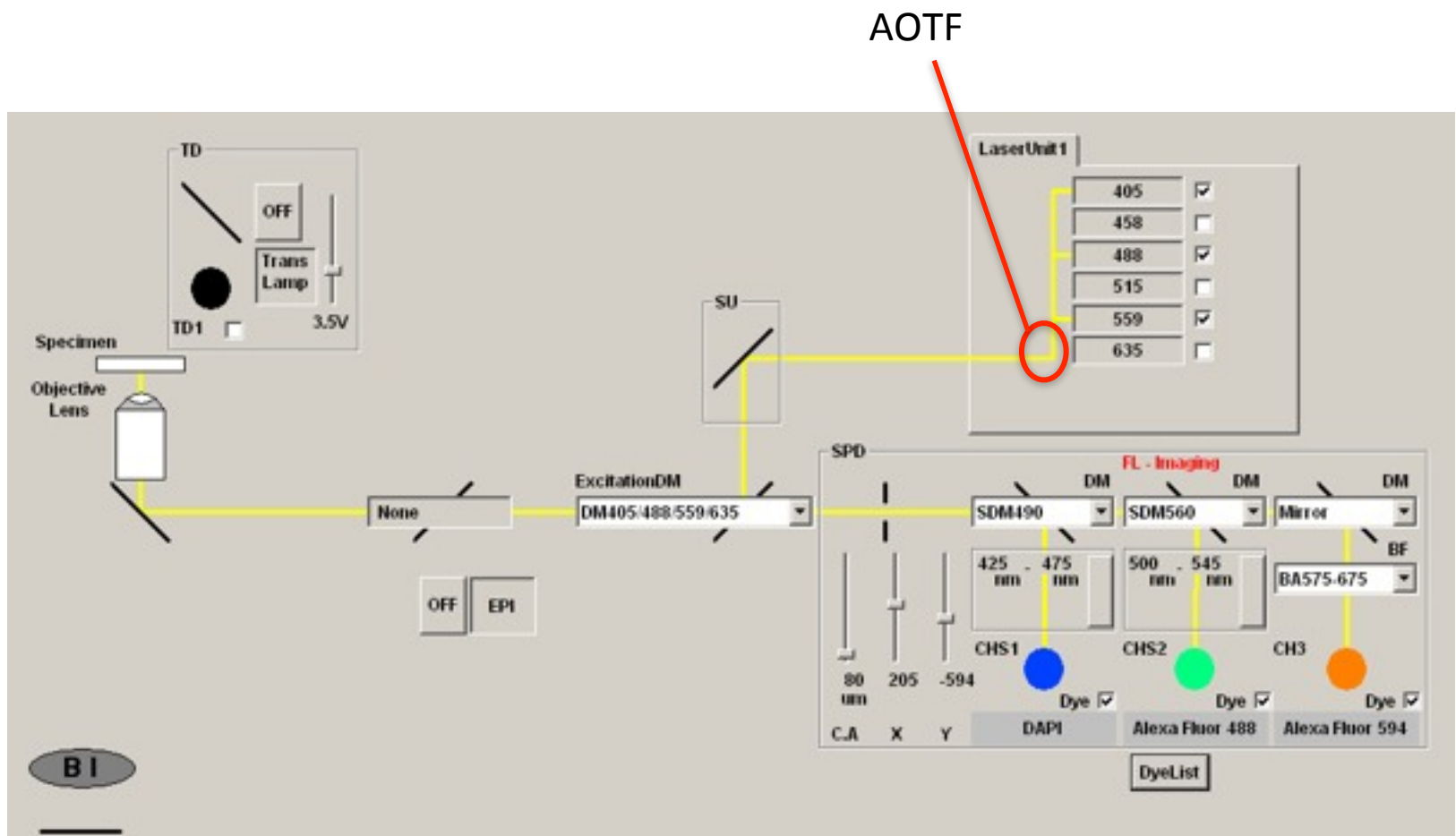


enables tighter control of fluorophores excited



AOTF

Acousto-Optic Tunable Filter

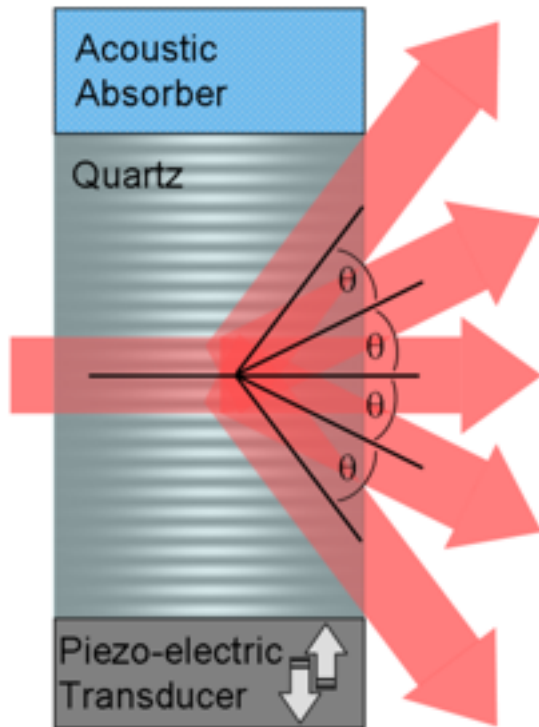




THEORY

AOTF

Acousto-Optic Tunable Filter



acousto-optic effect:

Acoustic wave excited within the quartz gives rise to variations in the refractive index

The wavelength of the diffracted light is dependent on the acoustic frequency in the quartz. By tuning the frequency of the acoustic wave, the desired wavelength of the optical wave can be diffracted acousto-optically.



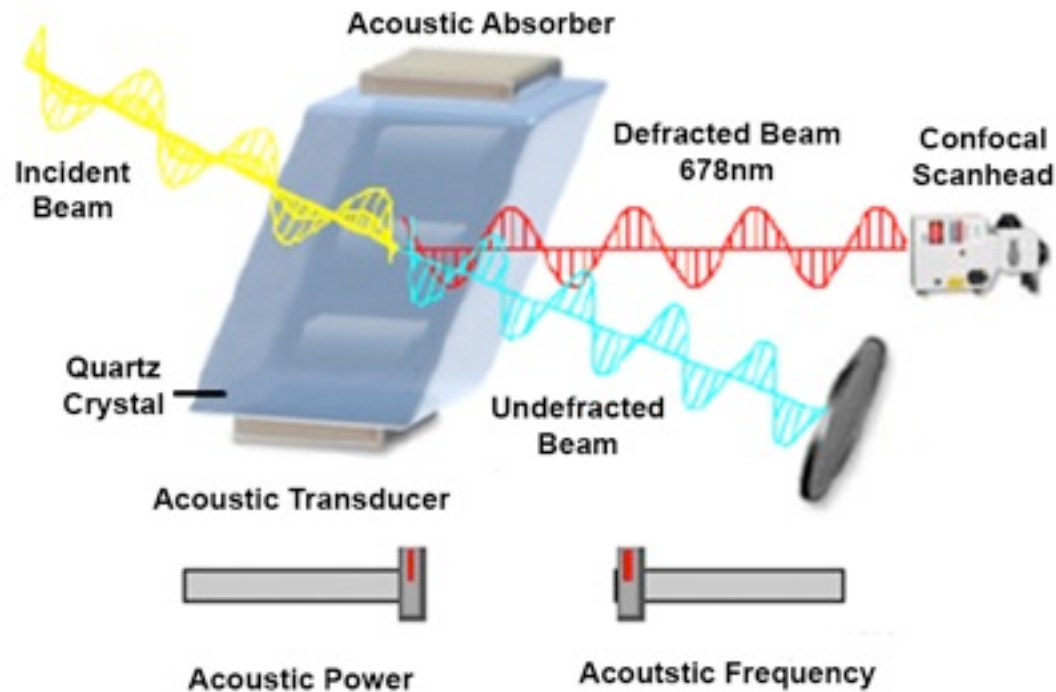
AOTF

Acousto-Optic Tunable Filter



AOTF

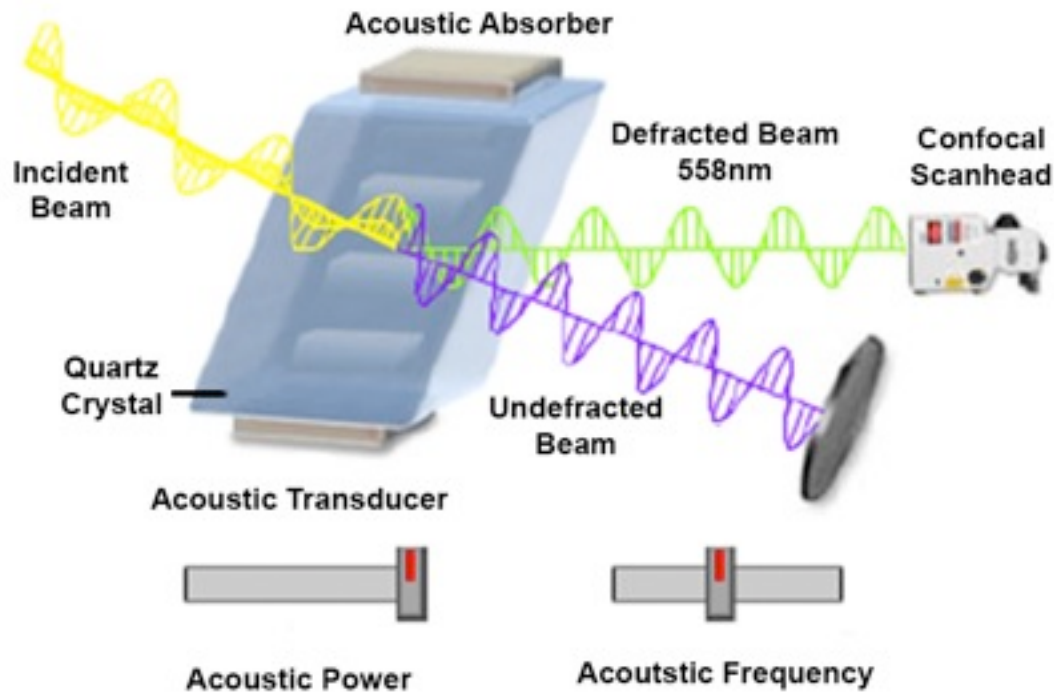
Acousto-Optic Tunable Filter





AOTF

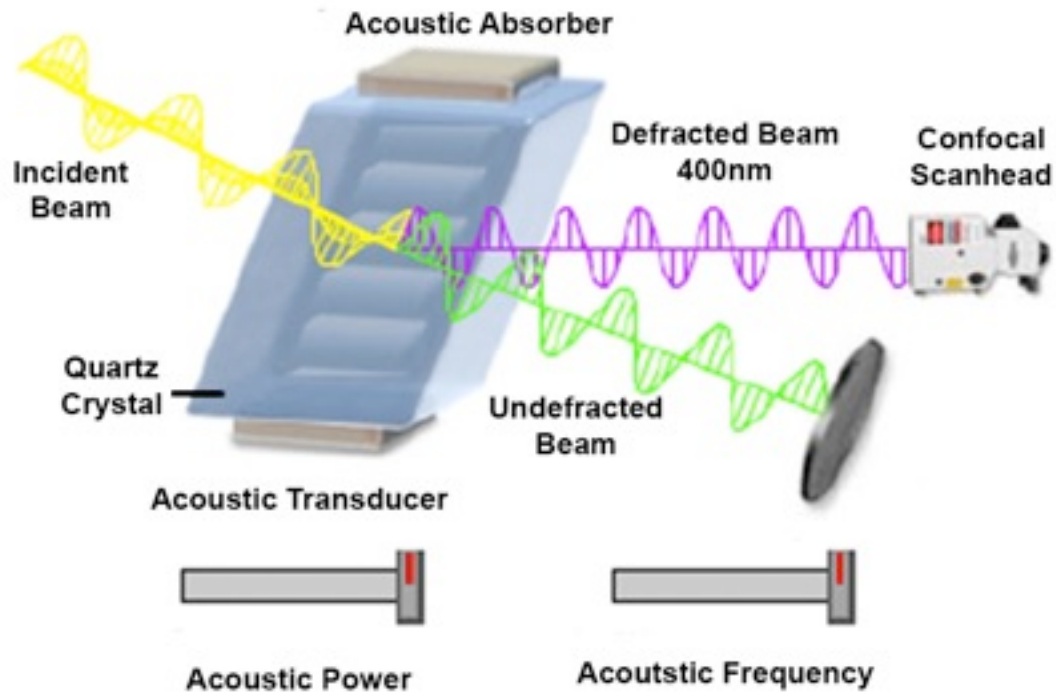
Acousto-Optic Tunable Filter





AOTF

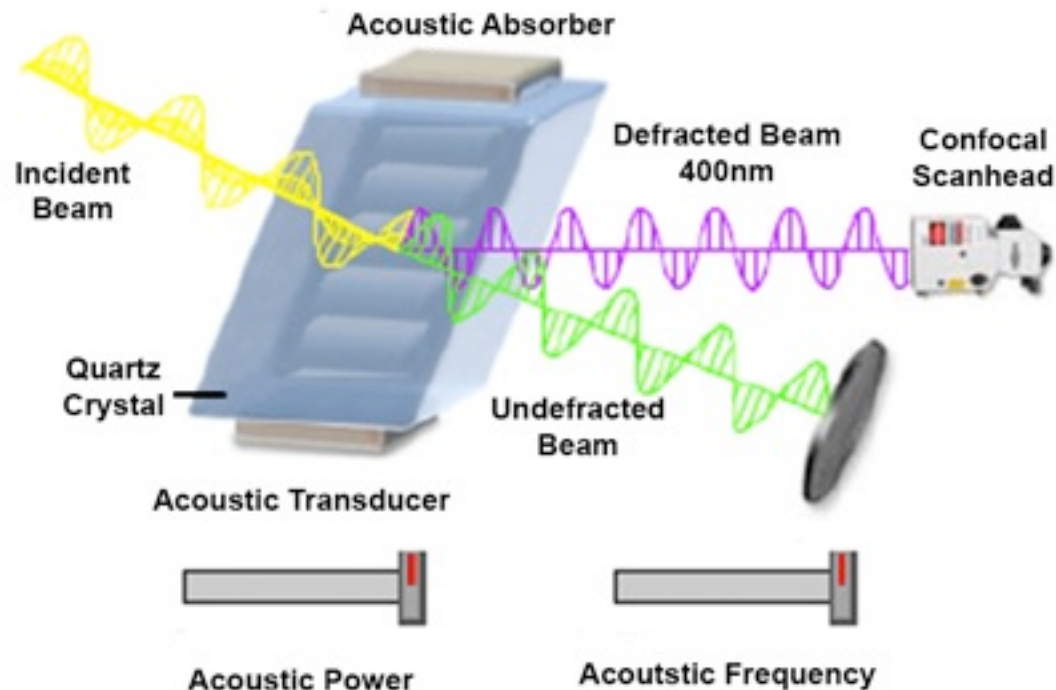
Acousto-Optic Tunable Filter





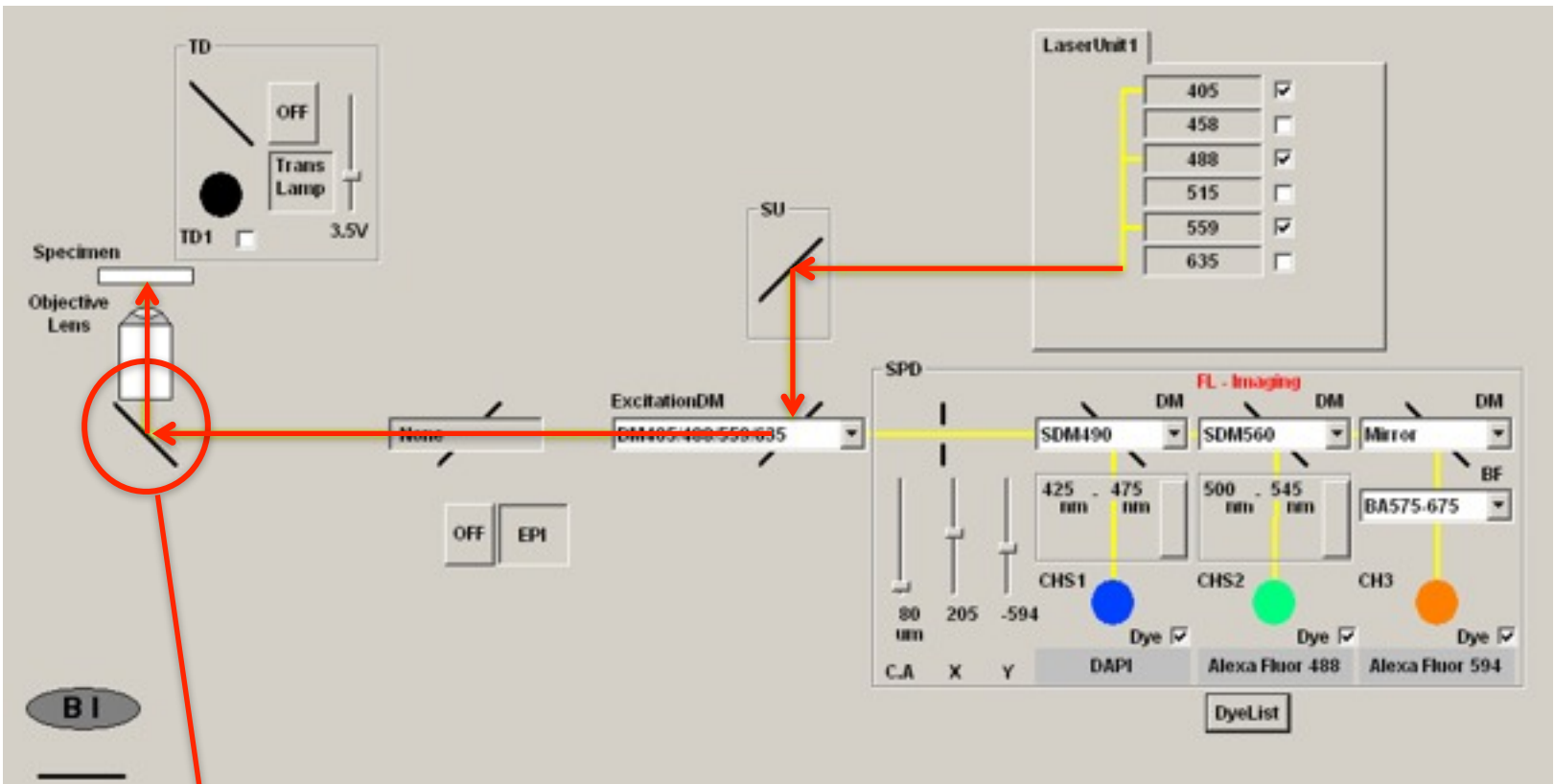
AOTF

Acousto-Optic Tunable Filter



Quick On/Off of lasers
Very fast changes between excitation wavelengths

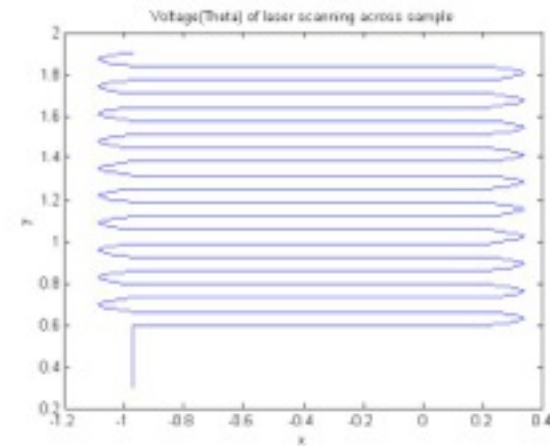
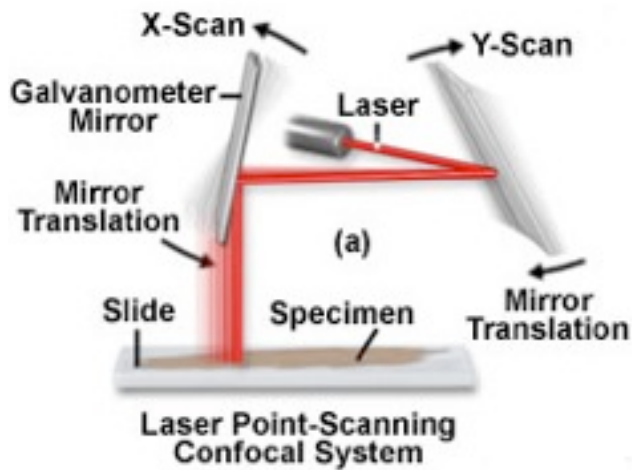
Galvo Scanning Mirrors



Galvo Scanning Mirrors



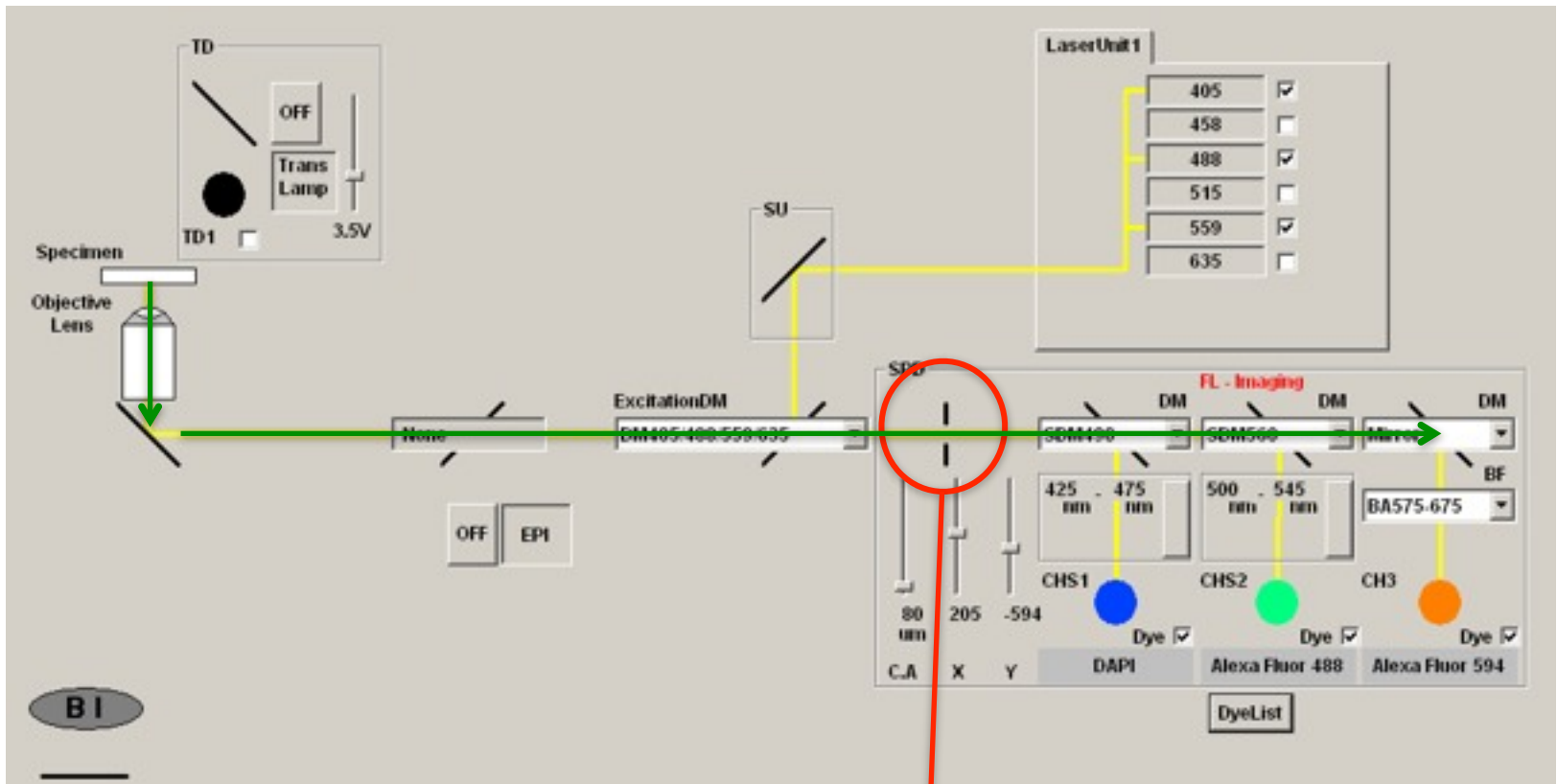
Galvo Scanning Mirrors



Sample excited at one point at a time
Relatively slow

Adjustable Pinhole

AOTF

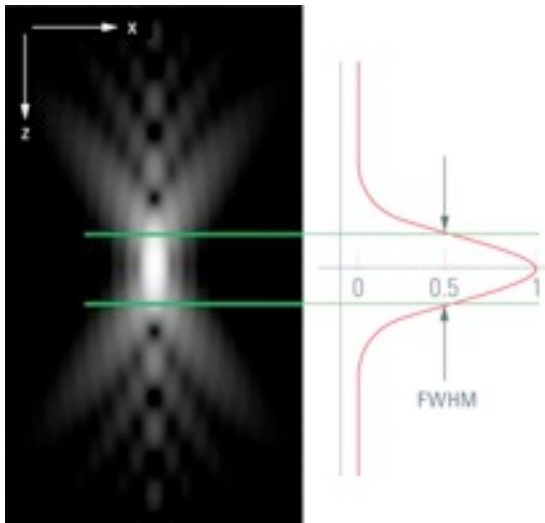


pinhole



THEORY

Pinhole – Optical Sectioning



$$\text{FWHM}_{\text{axial}} = \sqrt{\left(\frac{\lambda_{\text{exc}} \cdot n}{NA^2}\right)^2 + \left(\frac{n \cdot \sqrt{2} \cdot PH}{NA}\right)^2}$$

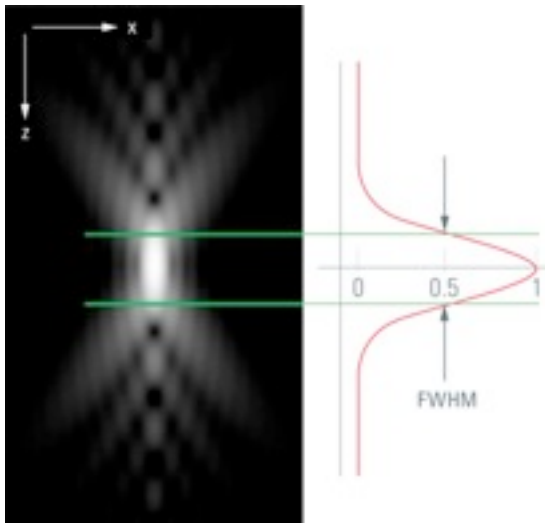
FWHM=Full Width Half-Maximum



THEORY

Pinhole – Optical Sectioning

Shorter the wavelength the thinner the optical section



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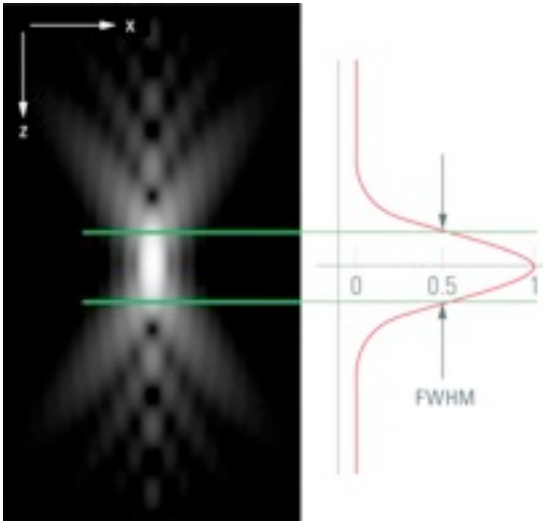


THEORY

Pinhole – Optical Sectioning

Shorter the wavelength the thinner the optical section

Diameter of the pinhole: Smaller pinhole thinner optical section



$$\text{FWHM}_{\text{axial}} = \sqrt{\left(\frac{\lambda_{\text{exc}} \cdot n}{NA^2}\right)^2 + \left(\frac{n \cdot \sqrt{2} \cdot \text{PH}}{NA}\right)^2}$$

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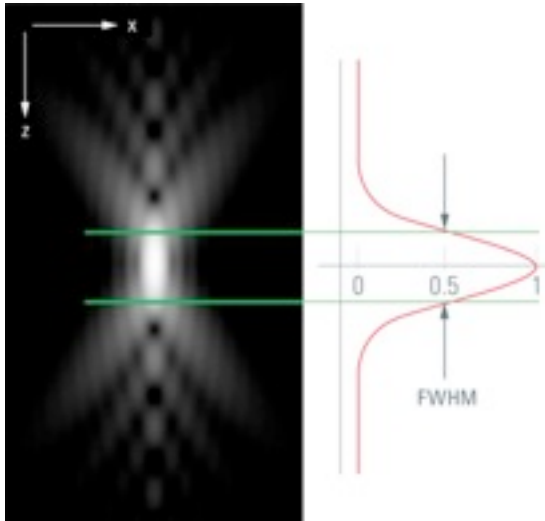


THEORY

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The higher the NA.
the thinner the section

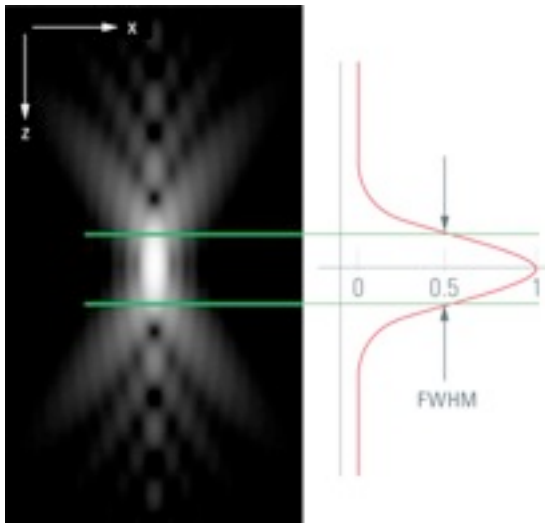


THEORY

Pinhole – Optical Sectioning

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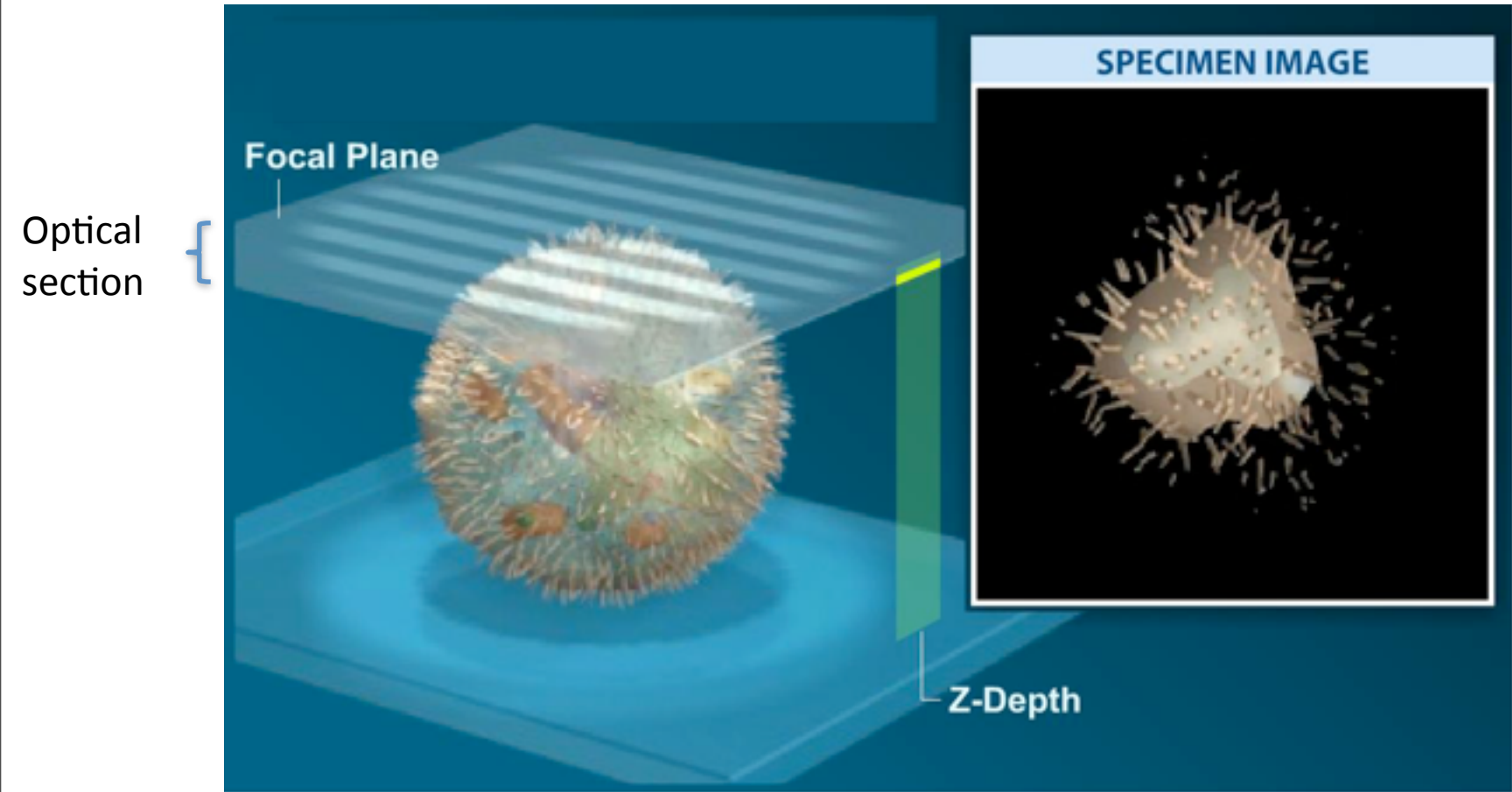
$$FWHM_{axial} = \sqrt{\left(\frac{\lambda_{exc} \cdot n}{NA^2}\right)^2 + \left(\frac{n \cdot \sqrt{2} \cdot PH}{NA}\right)^2}$$

The higher the NA. the thinner the section

Weak signal > open pinhole > more light but thicker section

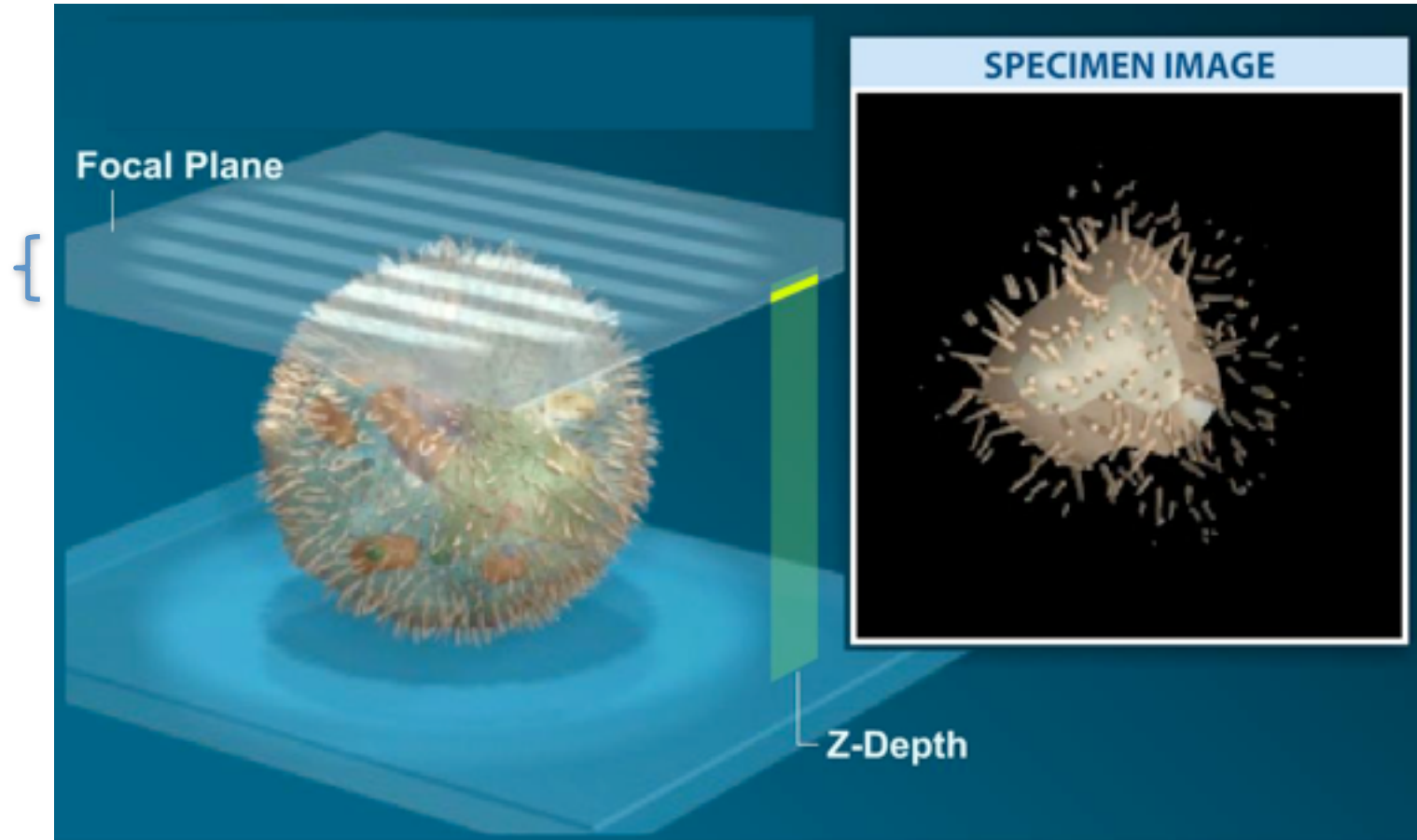


Confocal enables 3D reconstruction





Confocal enables 3D reconstruction

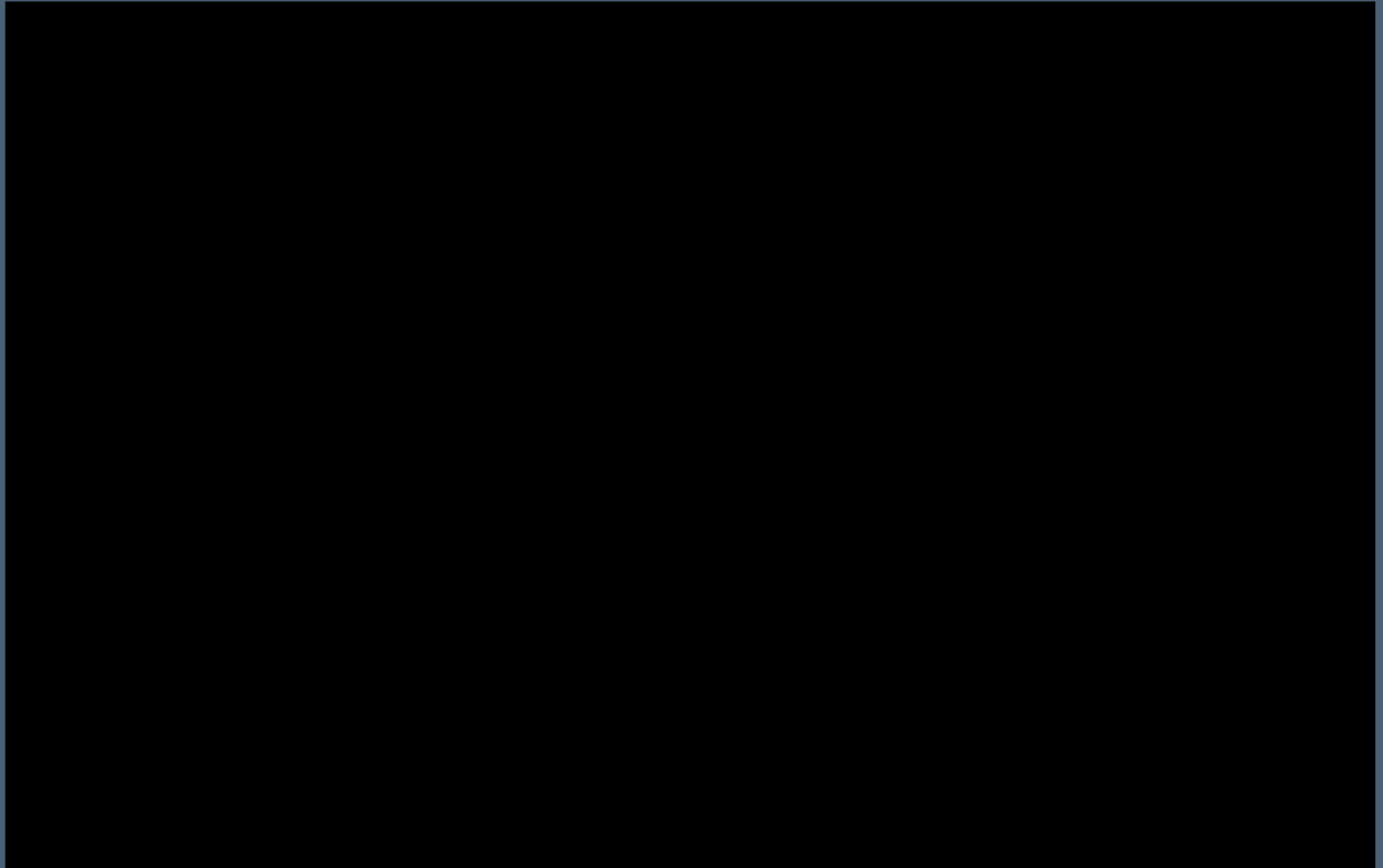




Confocal enables 3D reconstruction



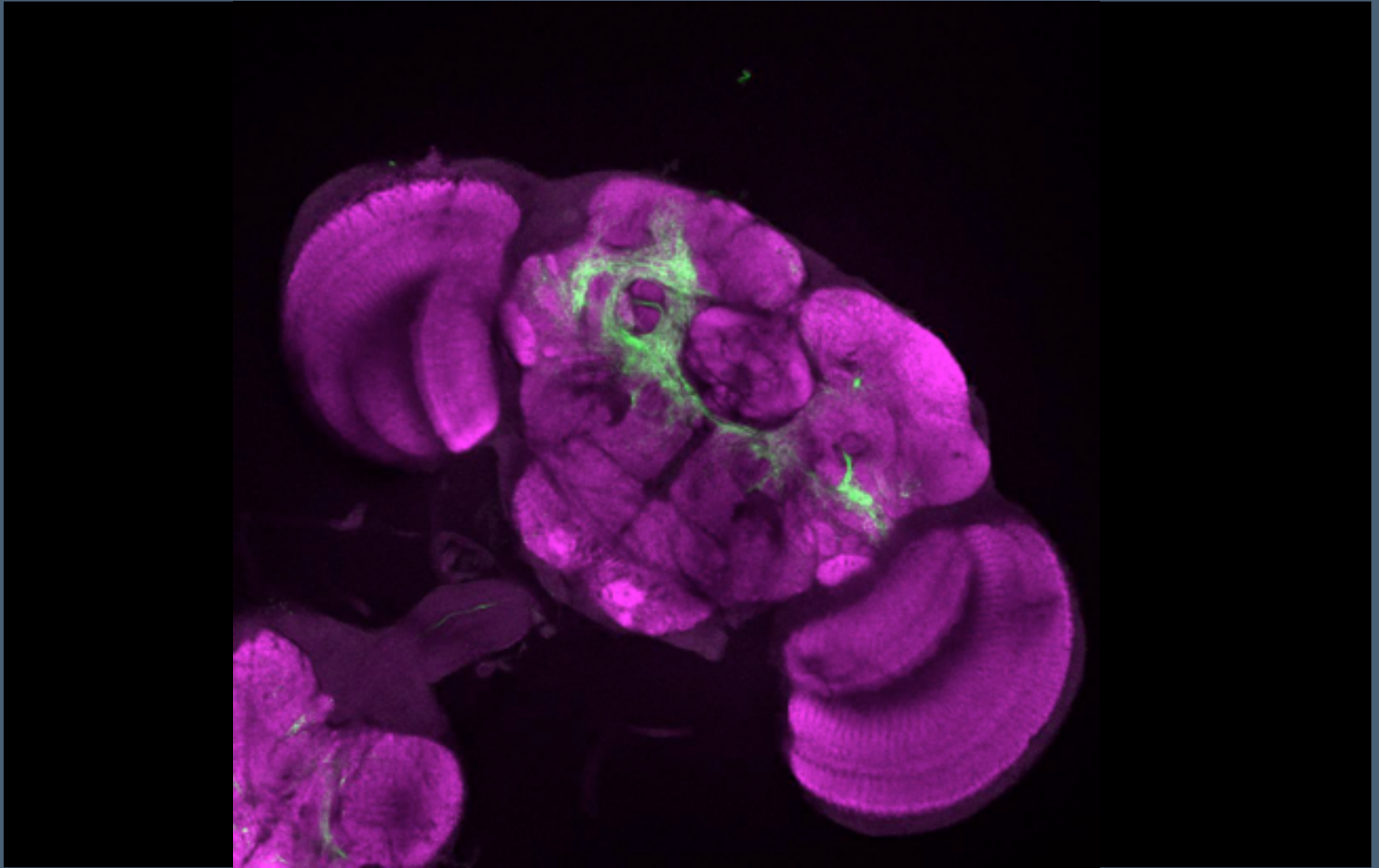
Confocal enables 3D reconstruction



Adult Drosophila head (C. Rezeval Goodwin Lab)

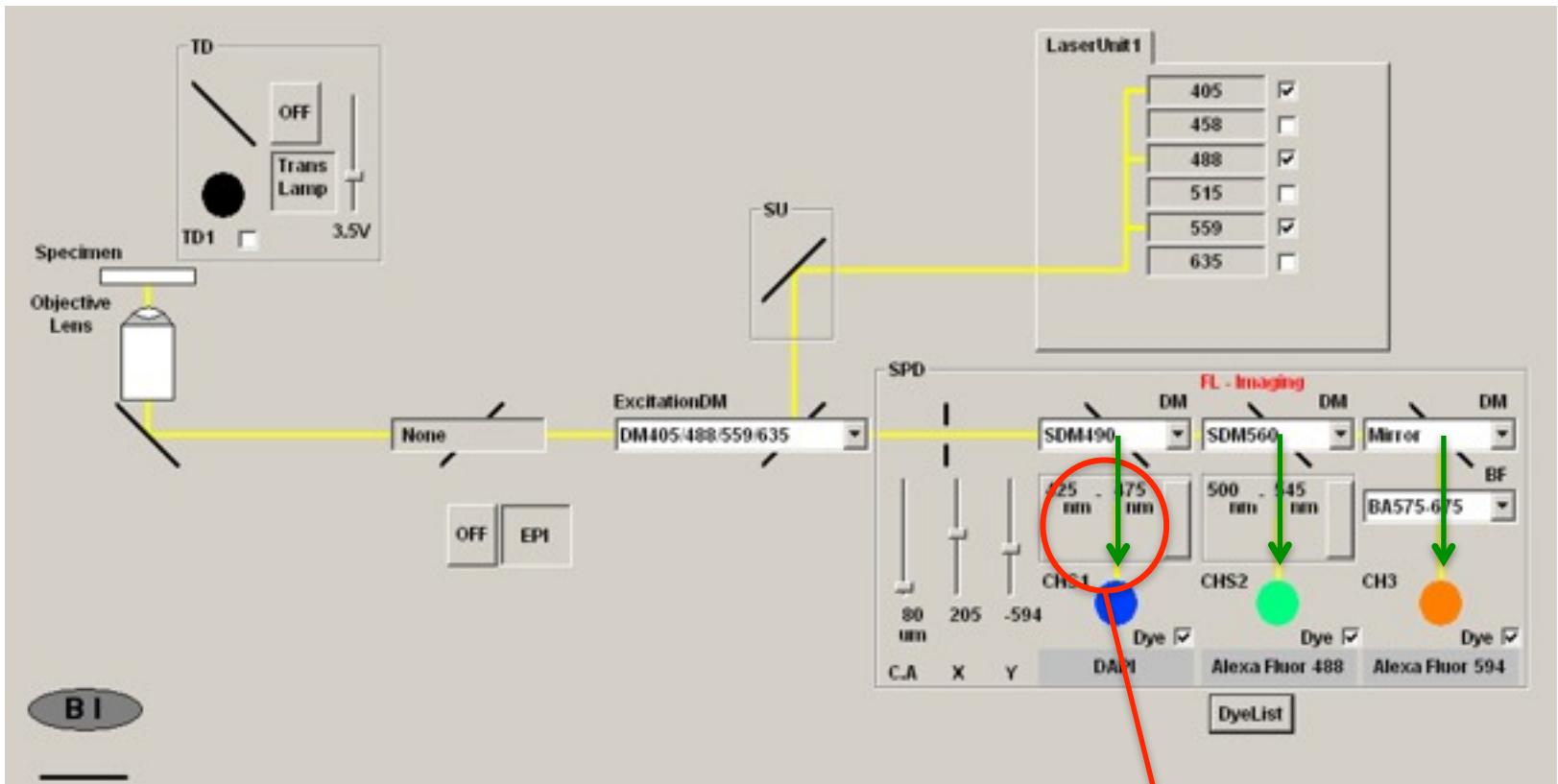


Confocal enables 3D reconstruction



Adult Drosophila head (C. Rezeval Goodwin Lab)

Variable Detector Slit

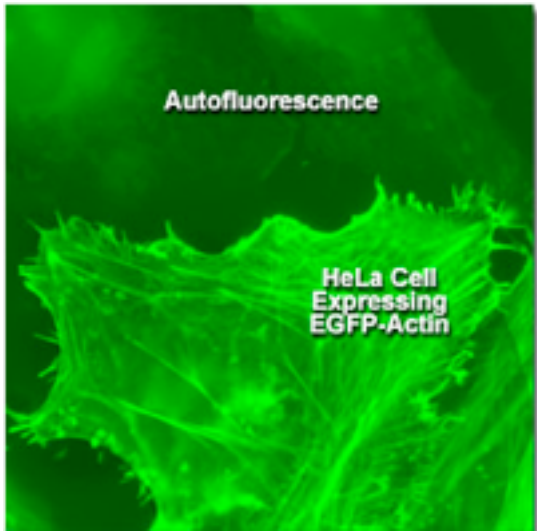


variable
detector slit

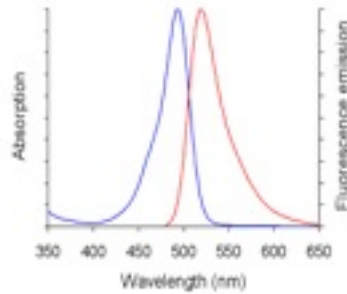
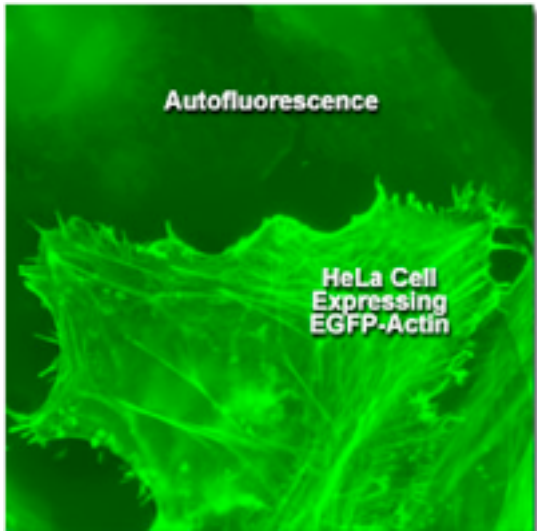
Spectral Unmixing



Spectral Unmixing



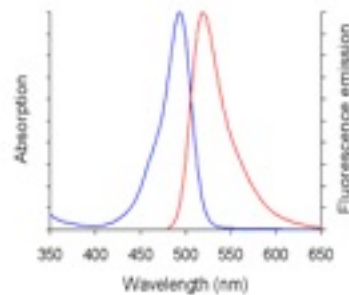
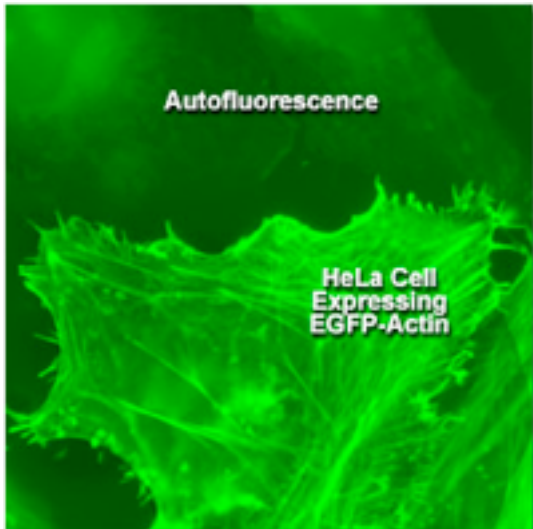
Spectral Unmixing



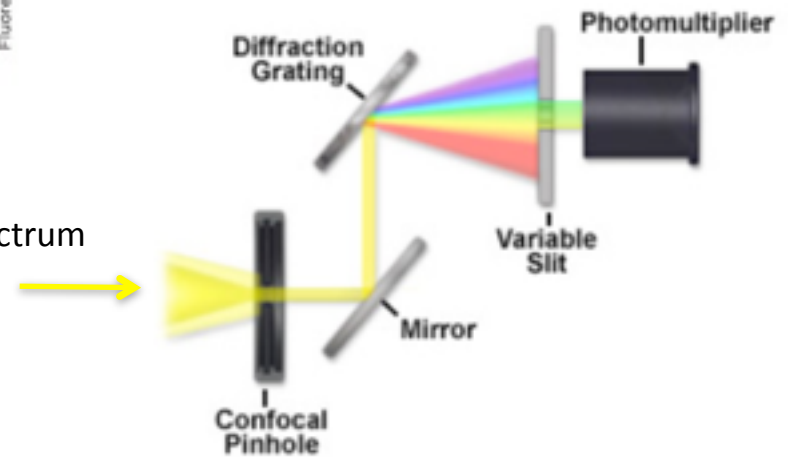
Light emitted from
fluorophore as a spectrum



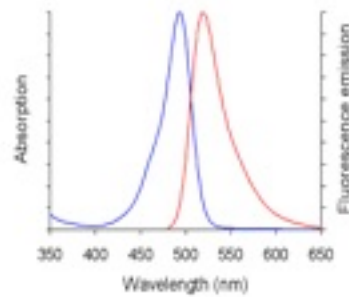
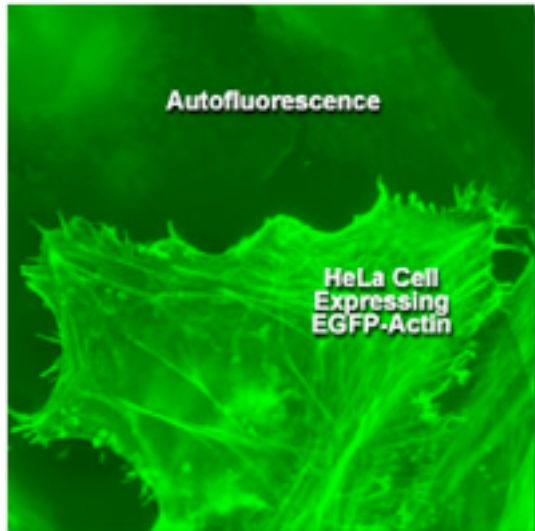
Spectral Unmixing



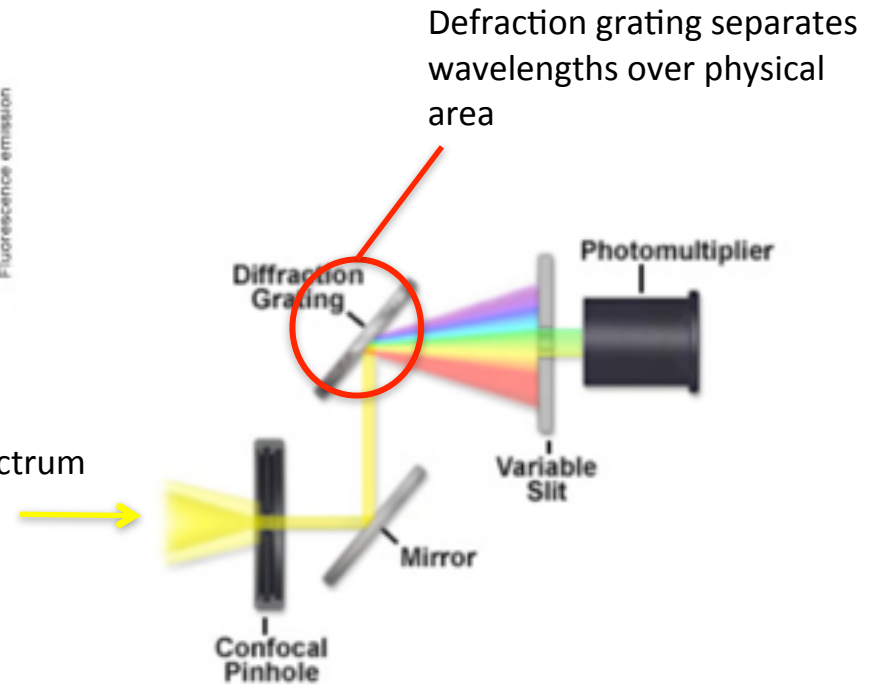
Light emitted from fluorophore as a spectrum



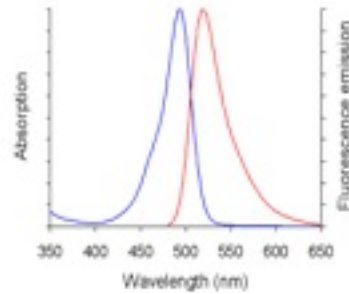
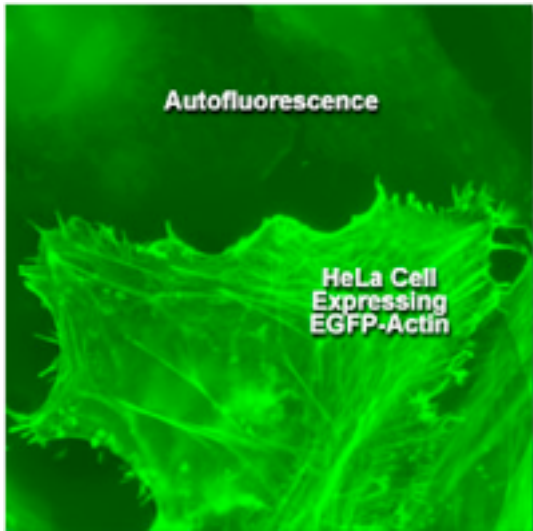
Spectral Unmixing



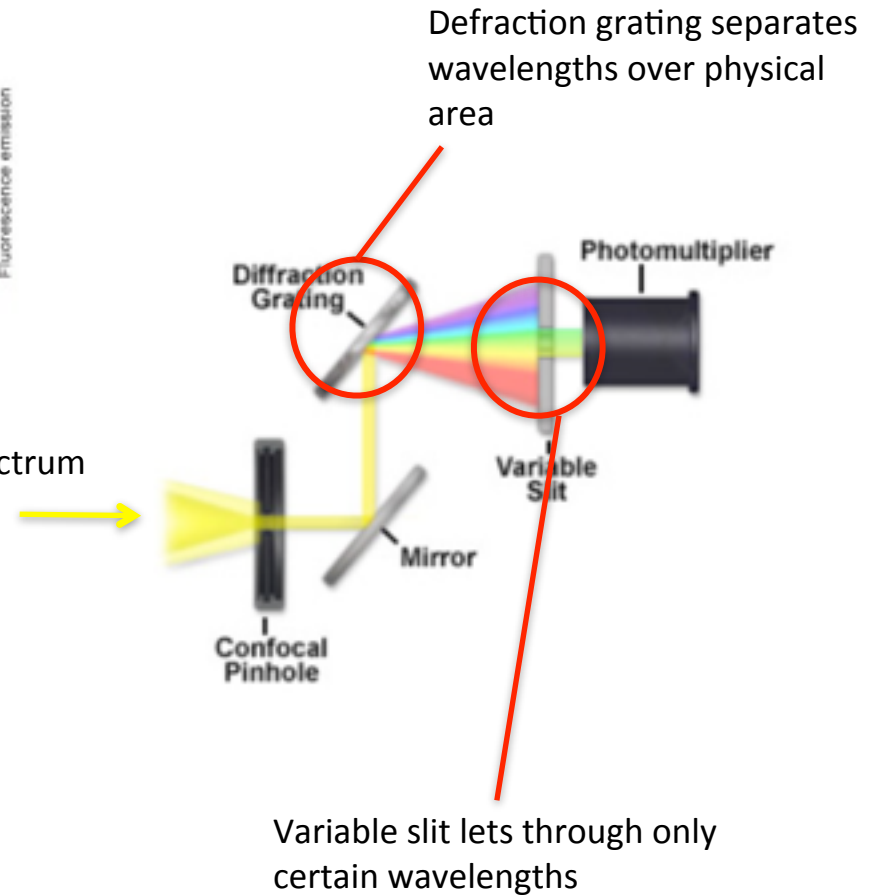
Light emitted from fluorophore as a spectrum



Spectral Unmixing

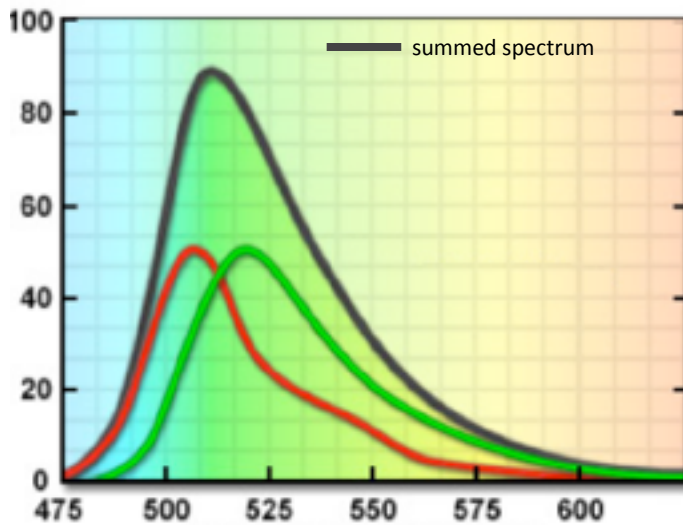
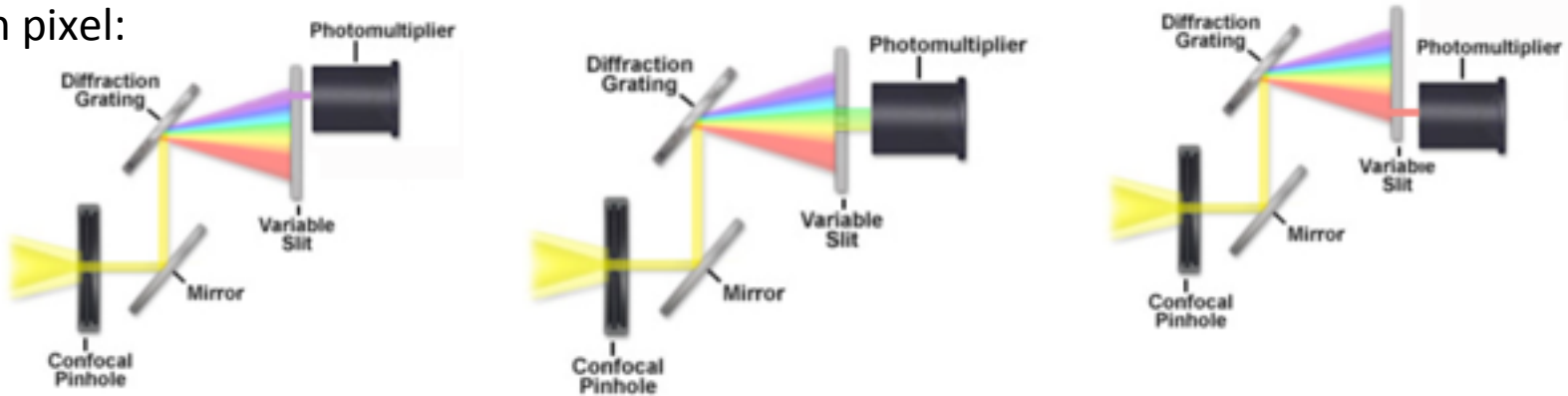


Light emitted from fluorophore as a spectrum



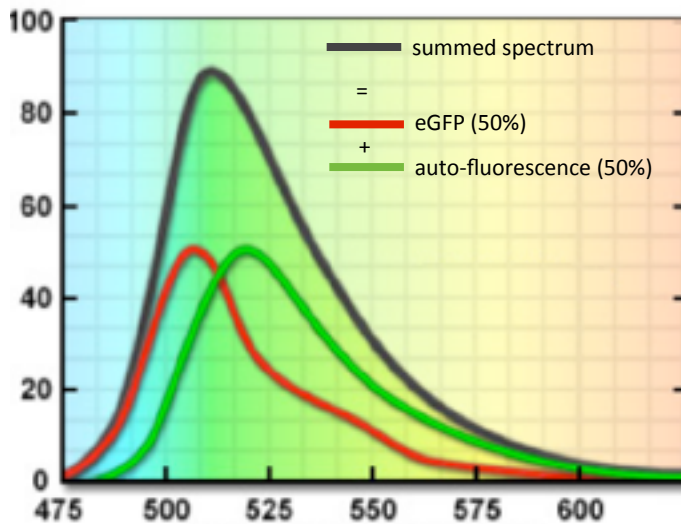
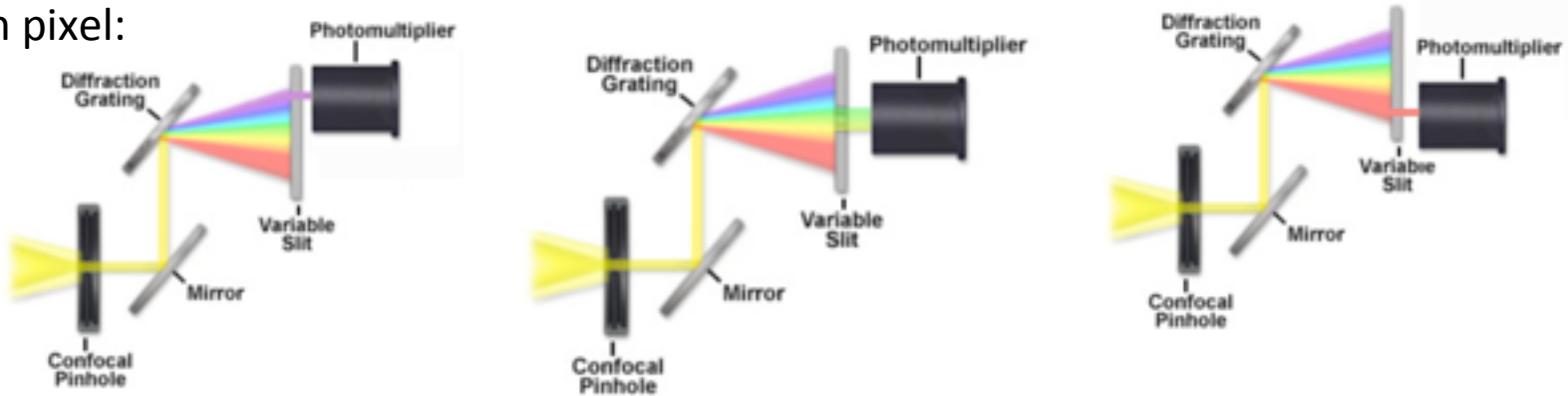
Spectral Unmixing

At each pixel:



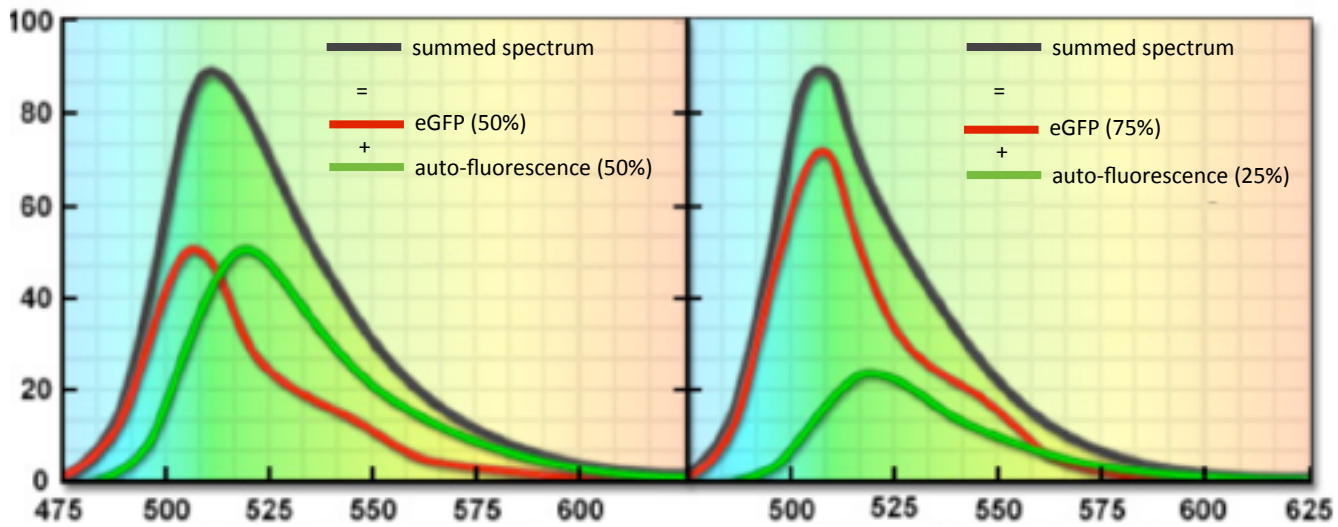
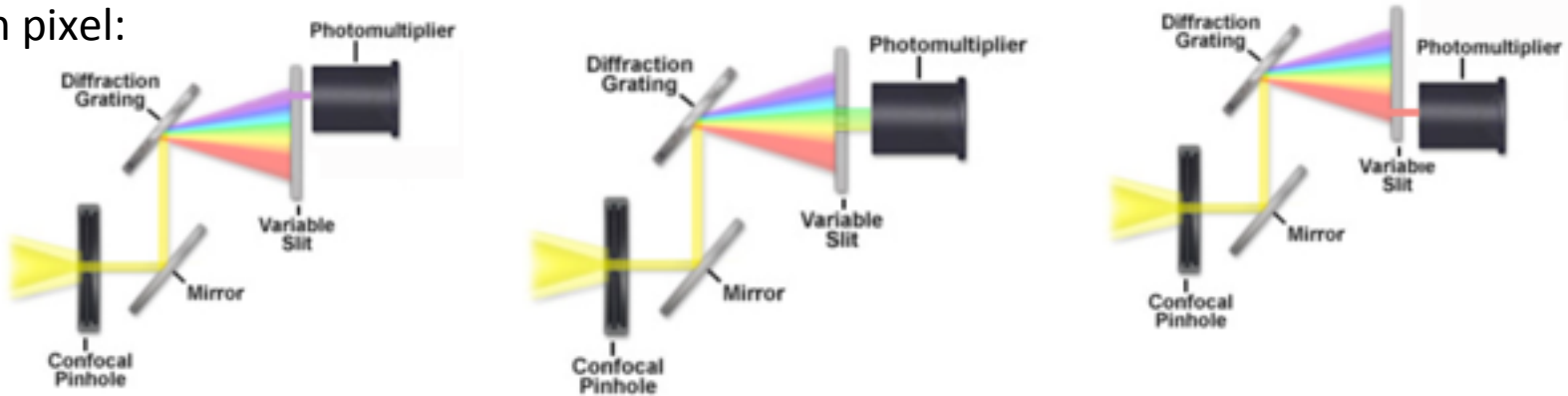
Spectral Unmixing

At each pixel:



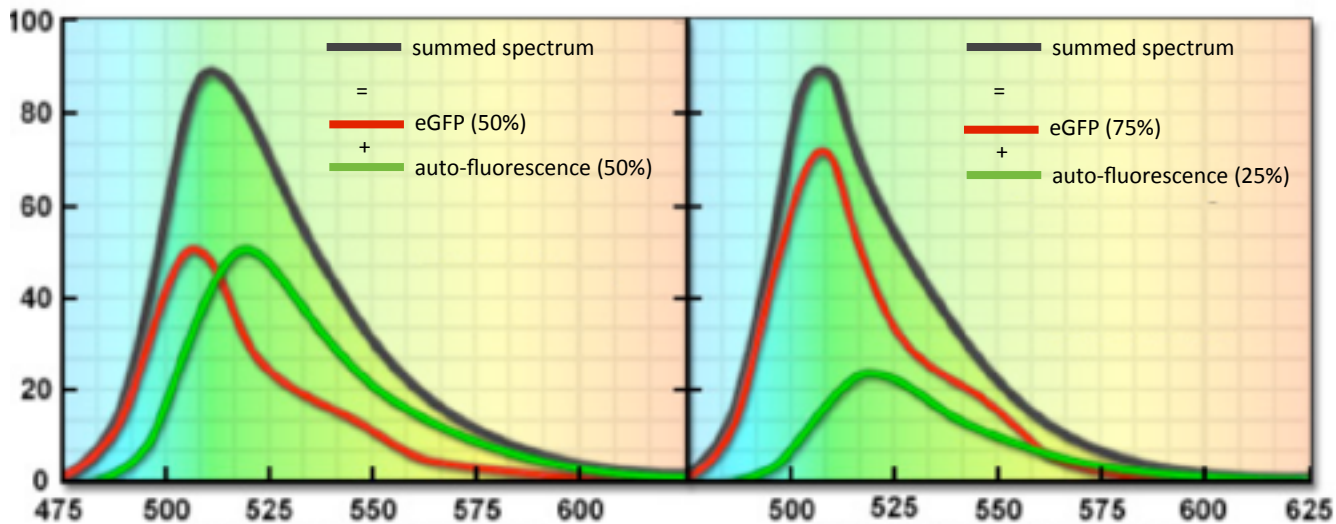
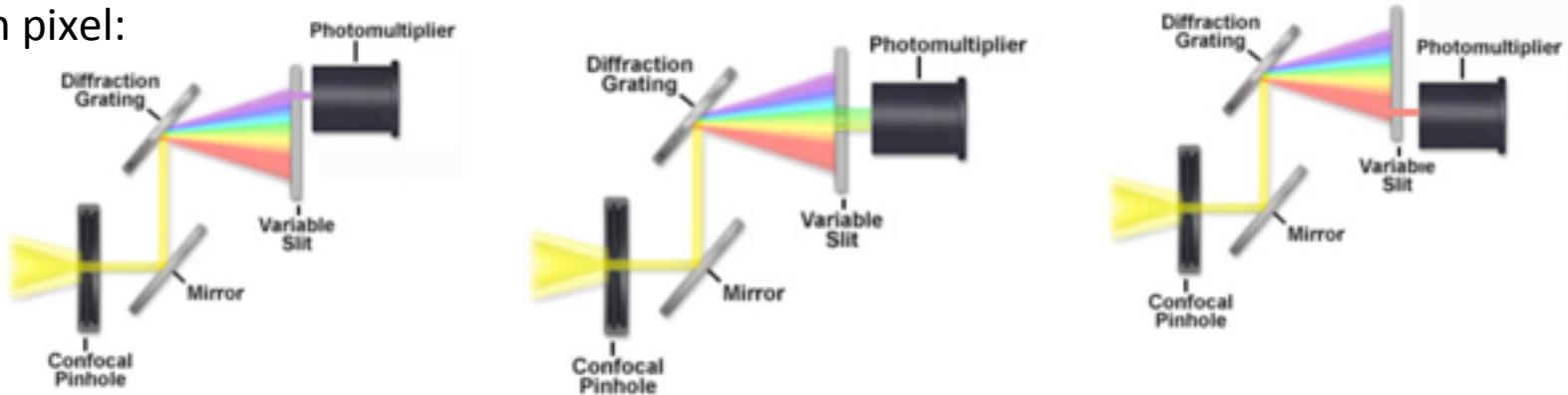
Spectral Unmixing

At each pixel:



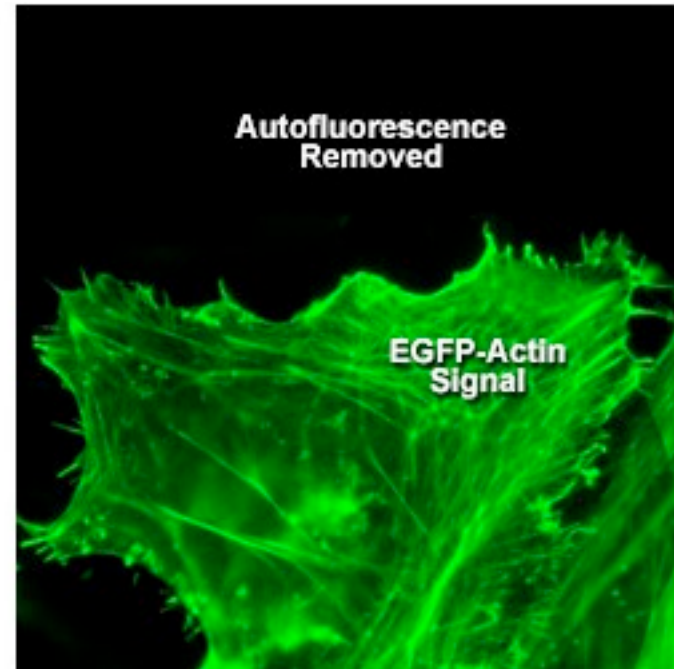
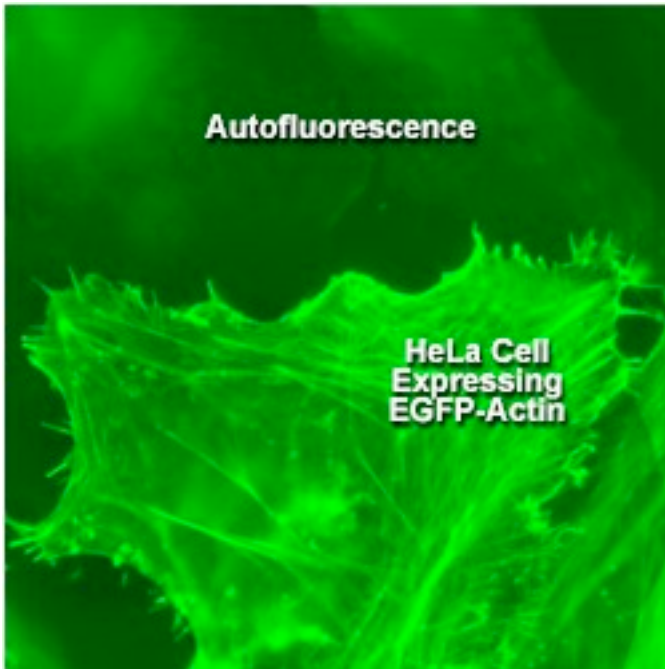
Spectral Unmixing

At each pixel:



Match the summed spectrum with all possible summed combinations from a library
At each pixel you therefore know the proportion of each fluorophore present

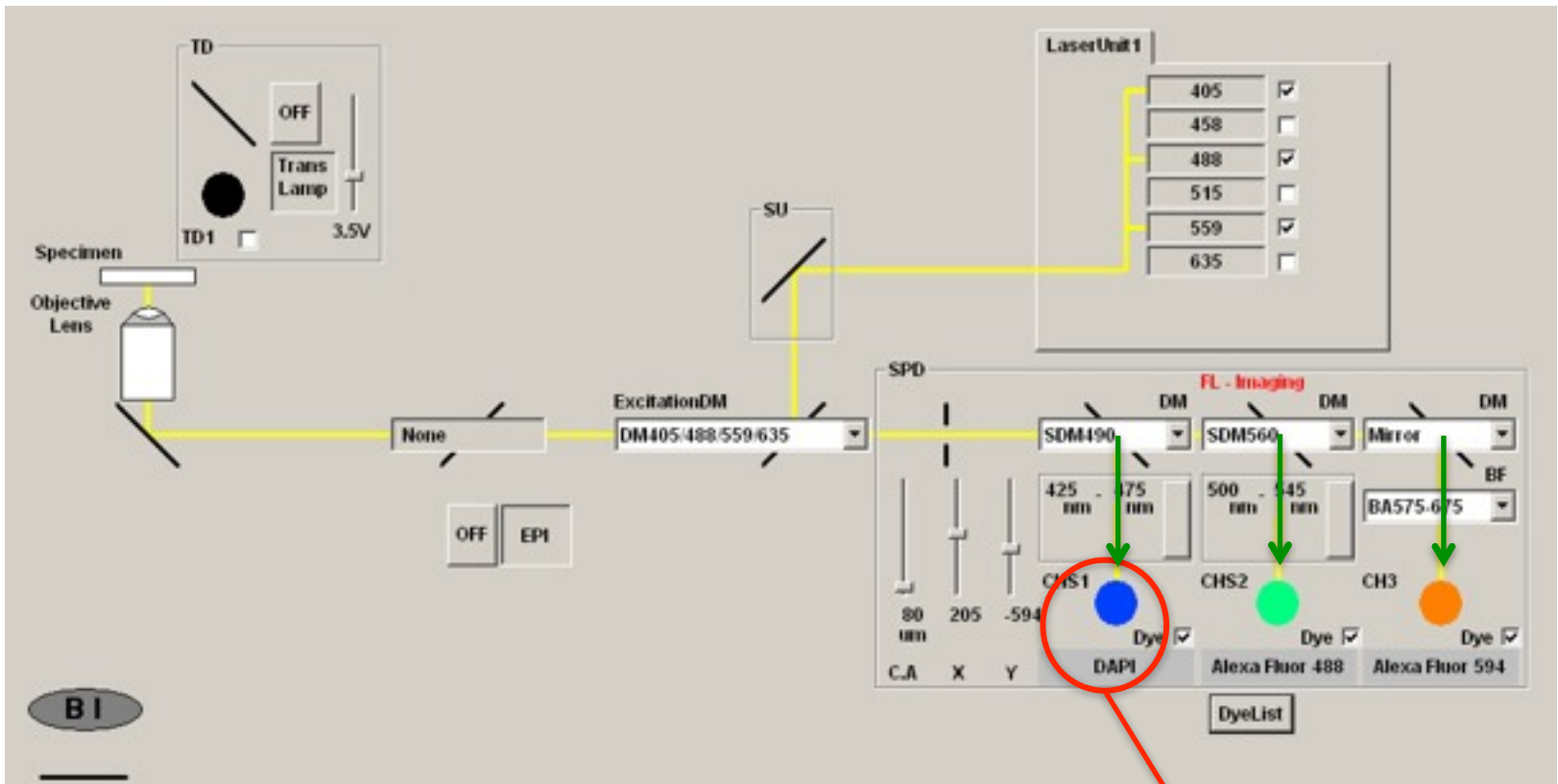
Spectral Unmixing removal of autofluorescence



At each pixel:

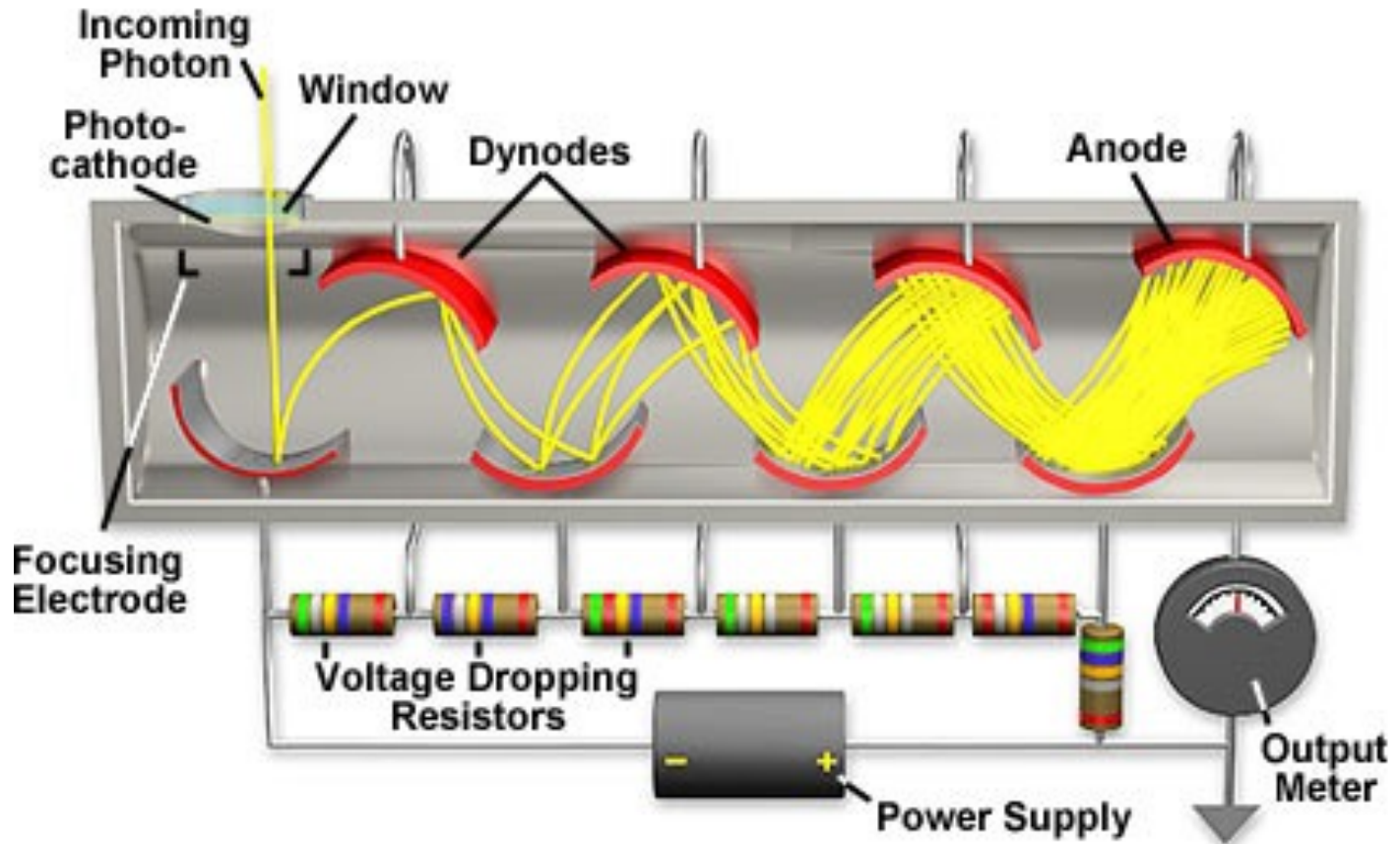
Calculate the proportion of the pixel is due to autofluorescence.
Subtract the autofluorescence from the 'true' GFP value.

PMT – Photon Multiplier Tube

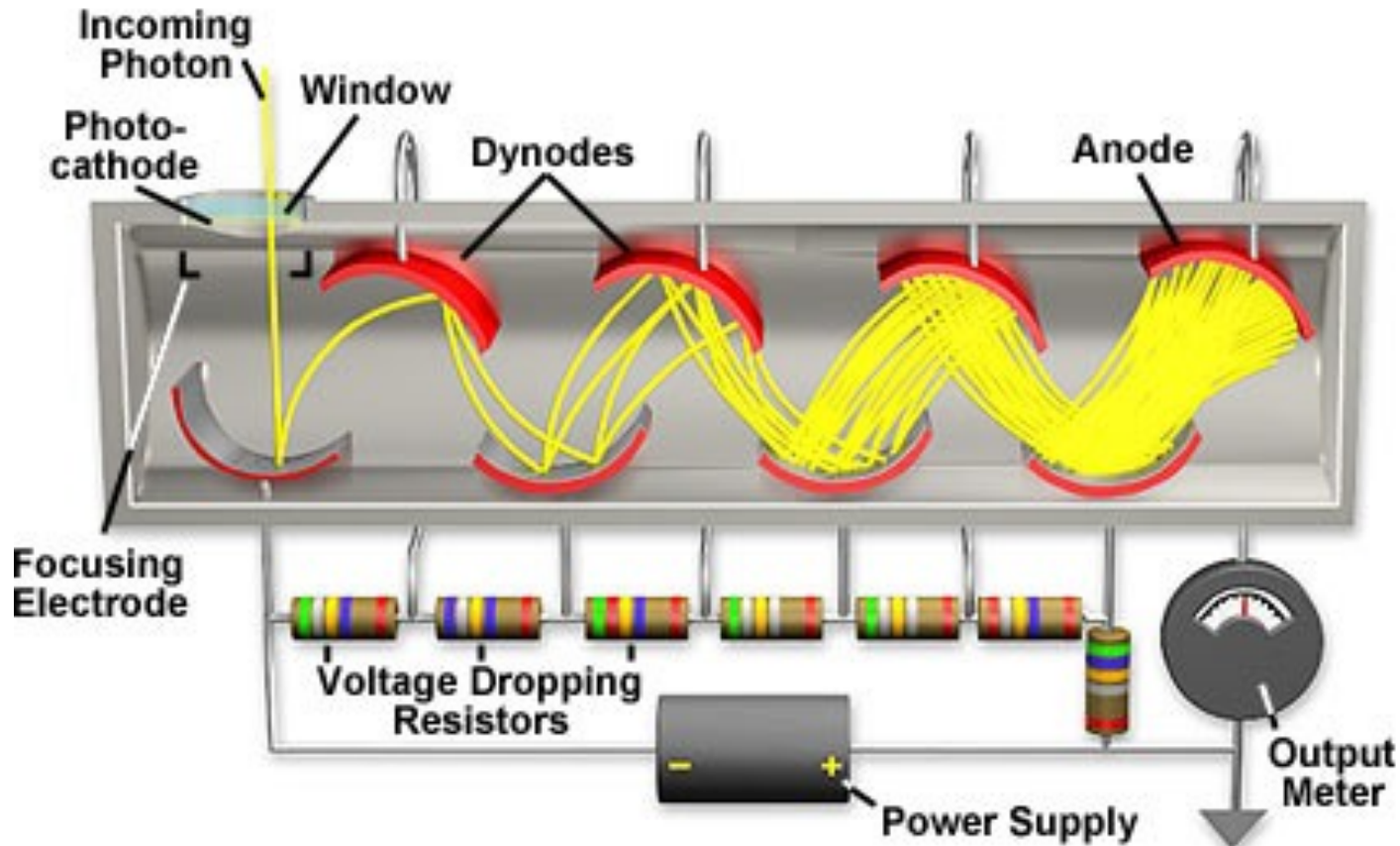


PMT detectors

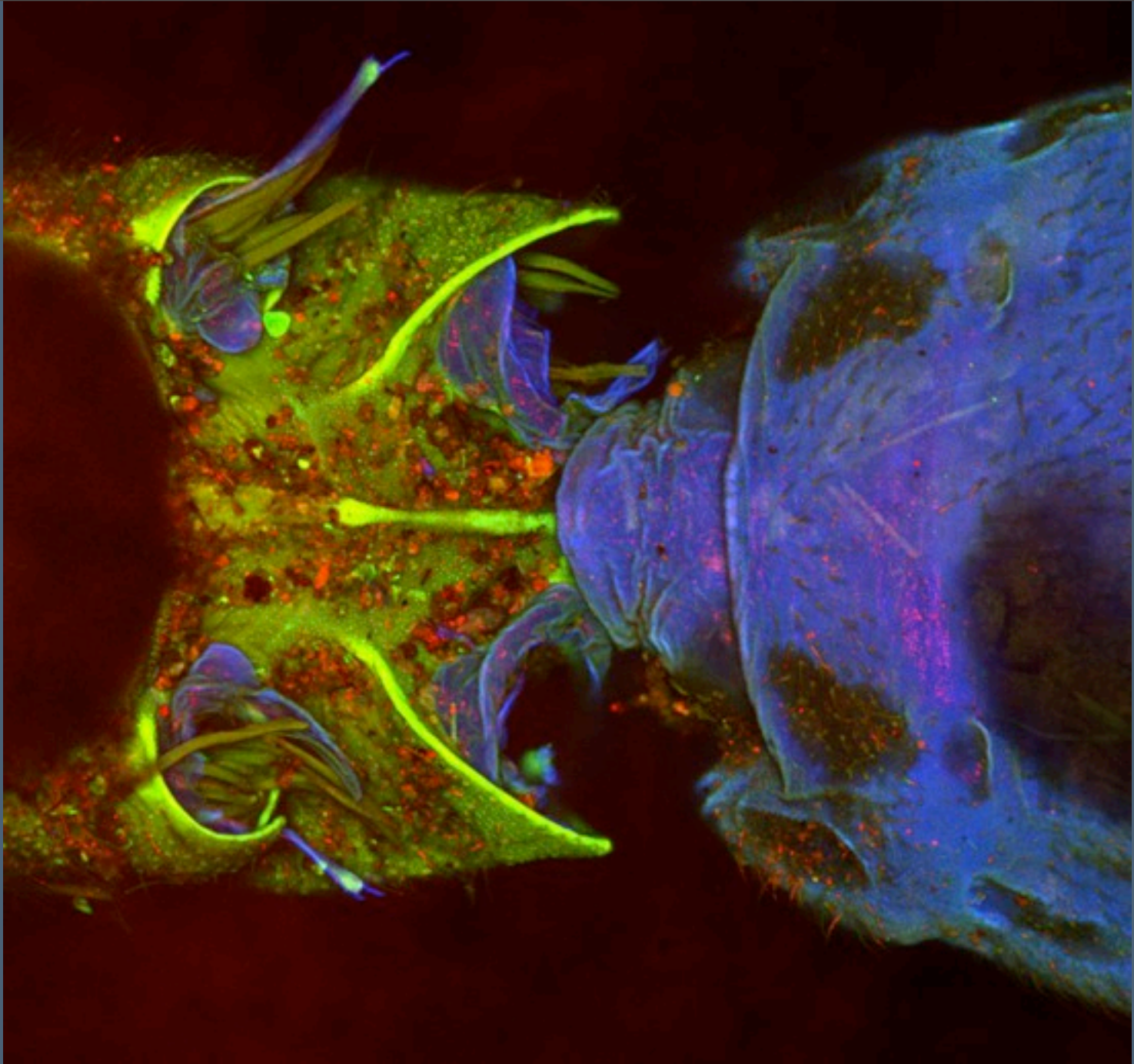
PMT – Photon Multiplier Tube



PMT – Photon Multiplier Tube



Very Low Noise
Huge Signal Amplification ($\sim 1 \times 10^8$)



insect autofluorescence



'Airy-Scan' technology





THEORY

'Airy-Scan' technology

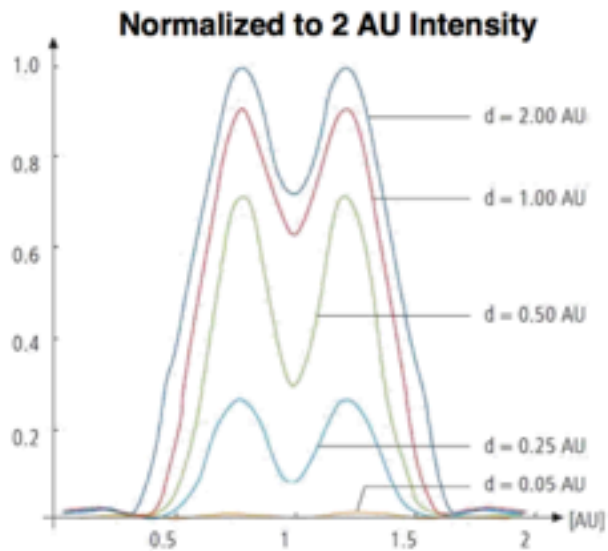
Small Pinhole, signal loss but resolution gain..



THEORY

'Airy-Scan' technology

Small Pinhole, signal loss but resolution gain..



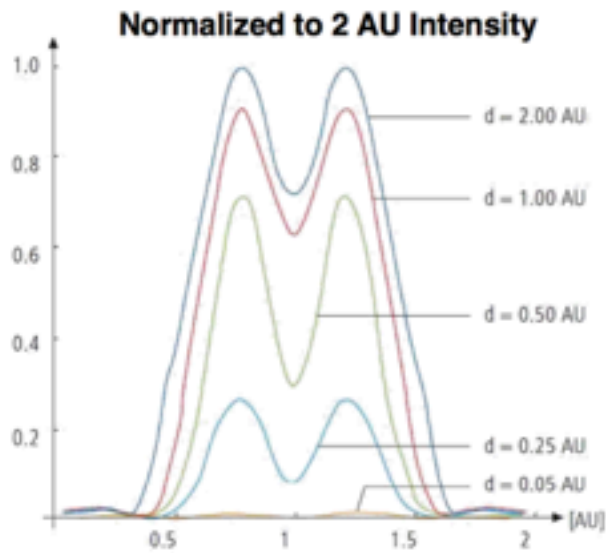
However, constricting the pinhole actually yields a drastic reduction in signal below 1 AU



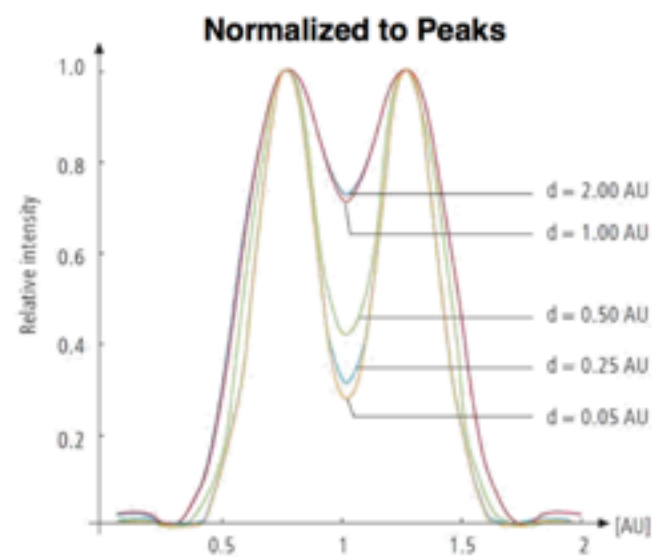
THEORY

'Airy-Scan' technology

Small Pinhole, signal loss but resolution gain..



However, constricting the pinhole actually yields a drastic reduction in signal below 1 AU



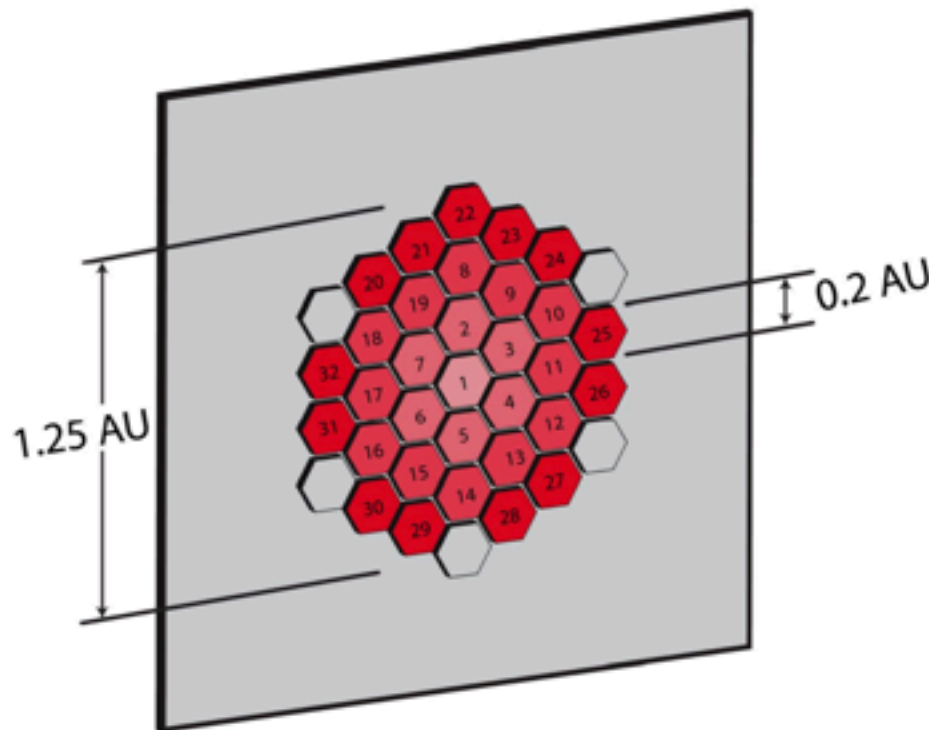
Small pinhole diameters lead to improved resolution steadily until about 0.2 AU, results in deeper dips between two objects



THEORY

'Airy-Scan' technology

let through all the emitted light
capture 0.2AU on each detector



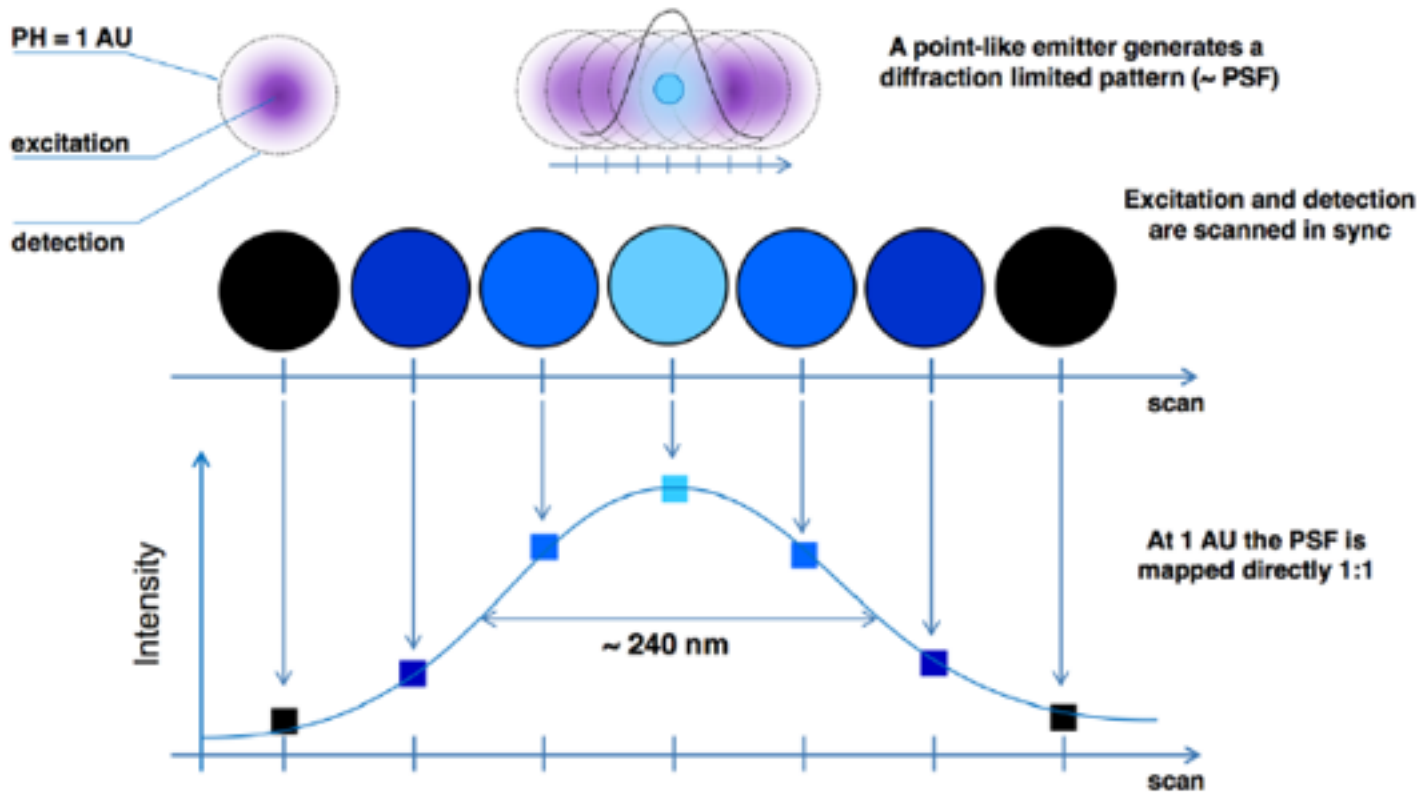
- 32 GaAsP detectors in hexagonal lattice
- Each detector approximately 0.2 AU in diameter
- Total detection area approximately 1.25 AU in diameter
- **Simultaneous improvement in resolution and signal**



THEORY

'Airy-Scan' technology

point of light scanned with 1 AU 'standard' detector

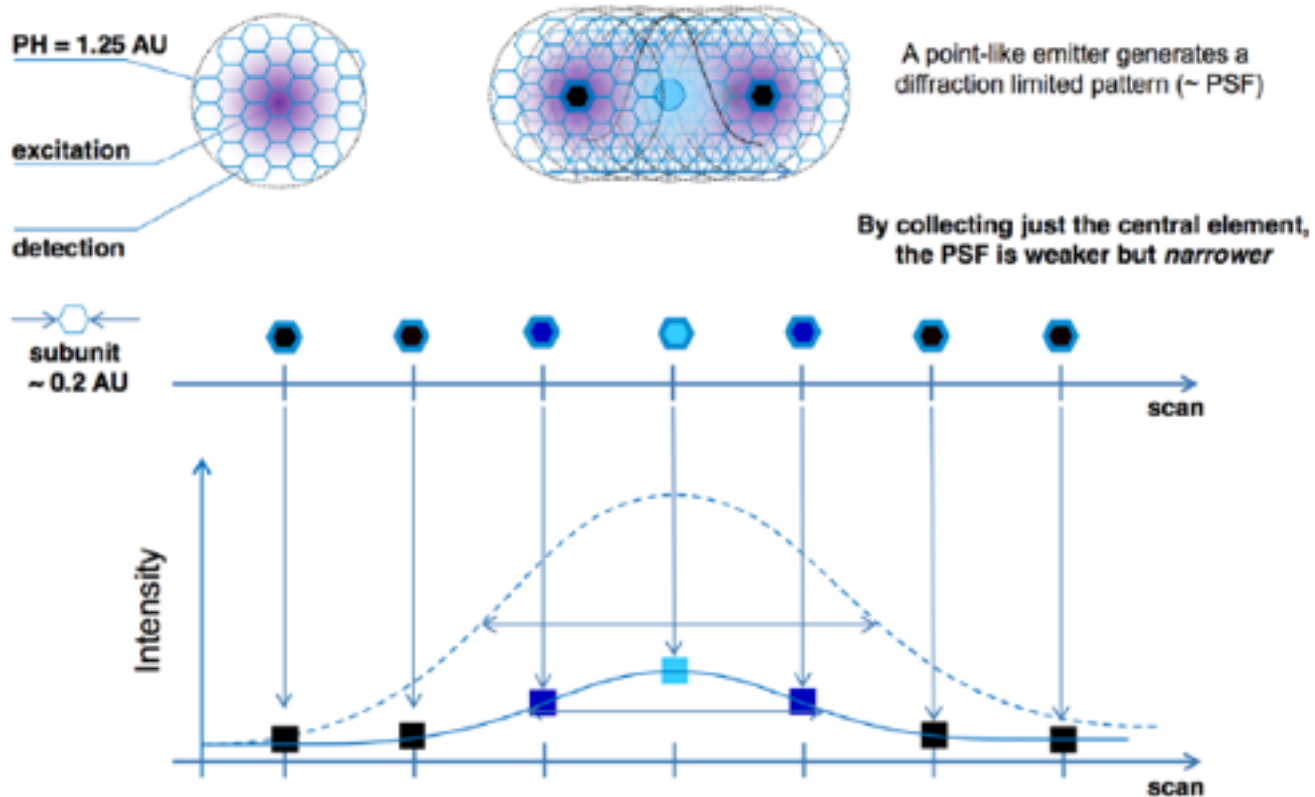




THEORY

'Airy-Scan' technology

point of light scanned with 0.2AU 'Airyscan' detector
>increased resolution

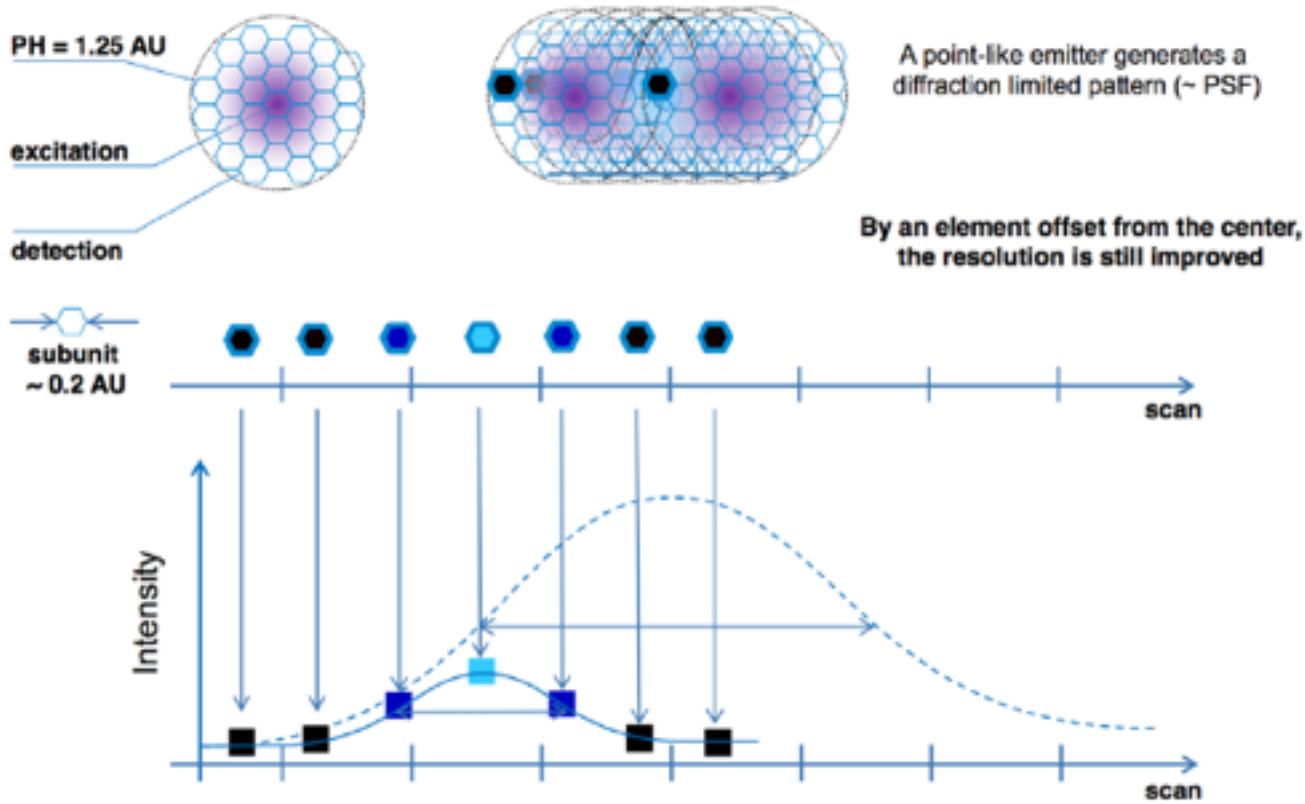




THEORY

'Airy-Scan' technology

each 0.2AU 'Airyscan' detector provides
>increased resolution

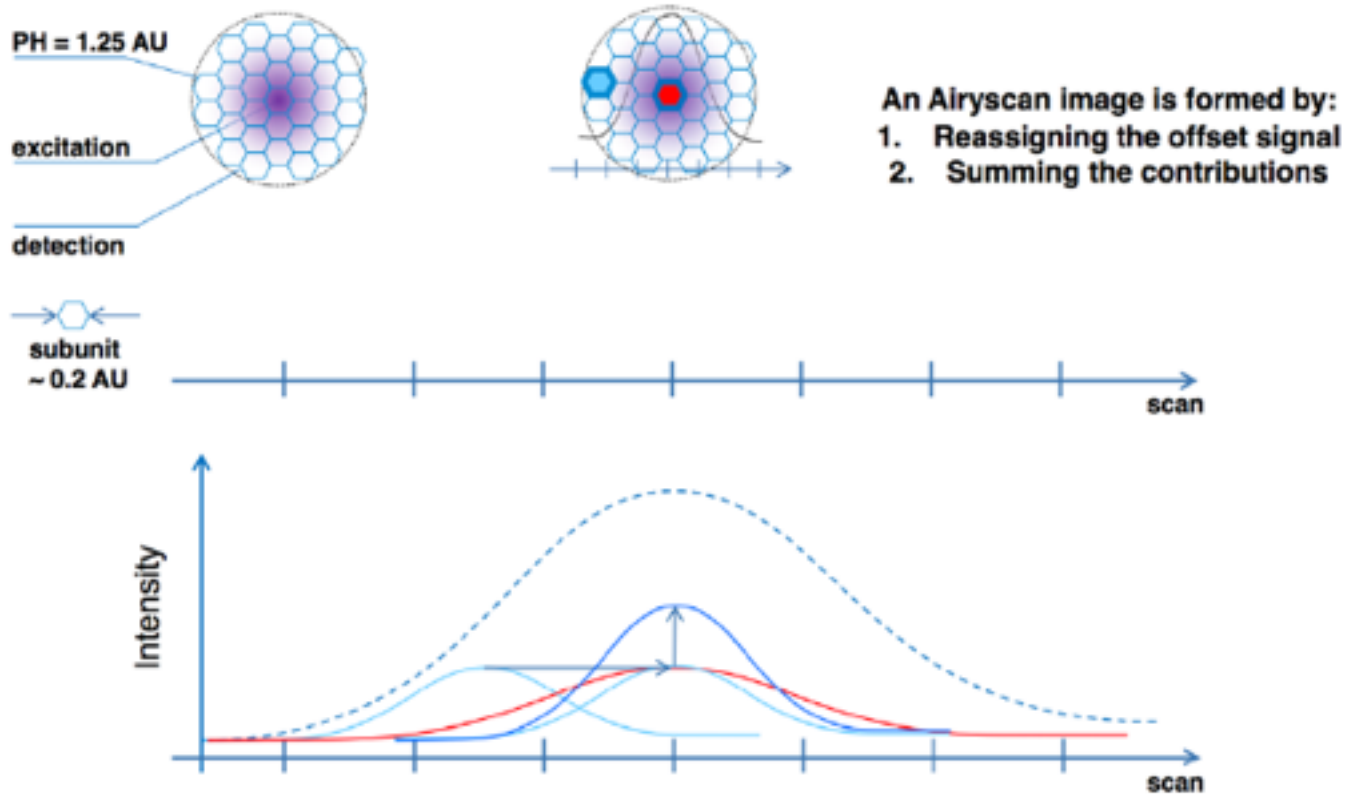




THEORY

'Airy-Scan' technology

each 0.2AU 'Airyscan' detector info
is reassigned and summed

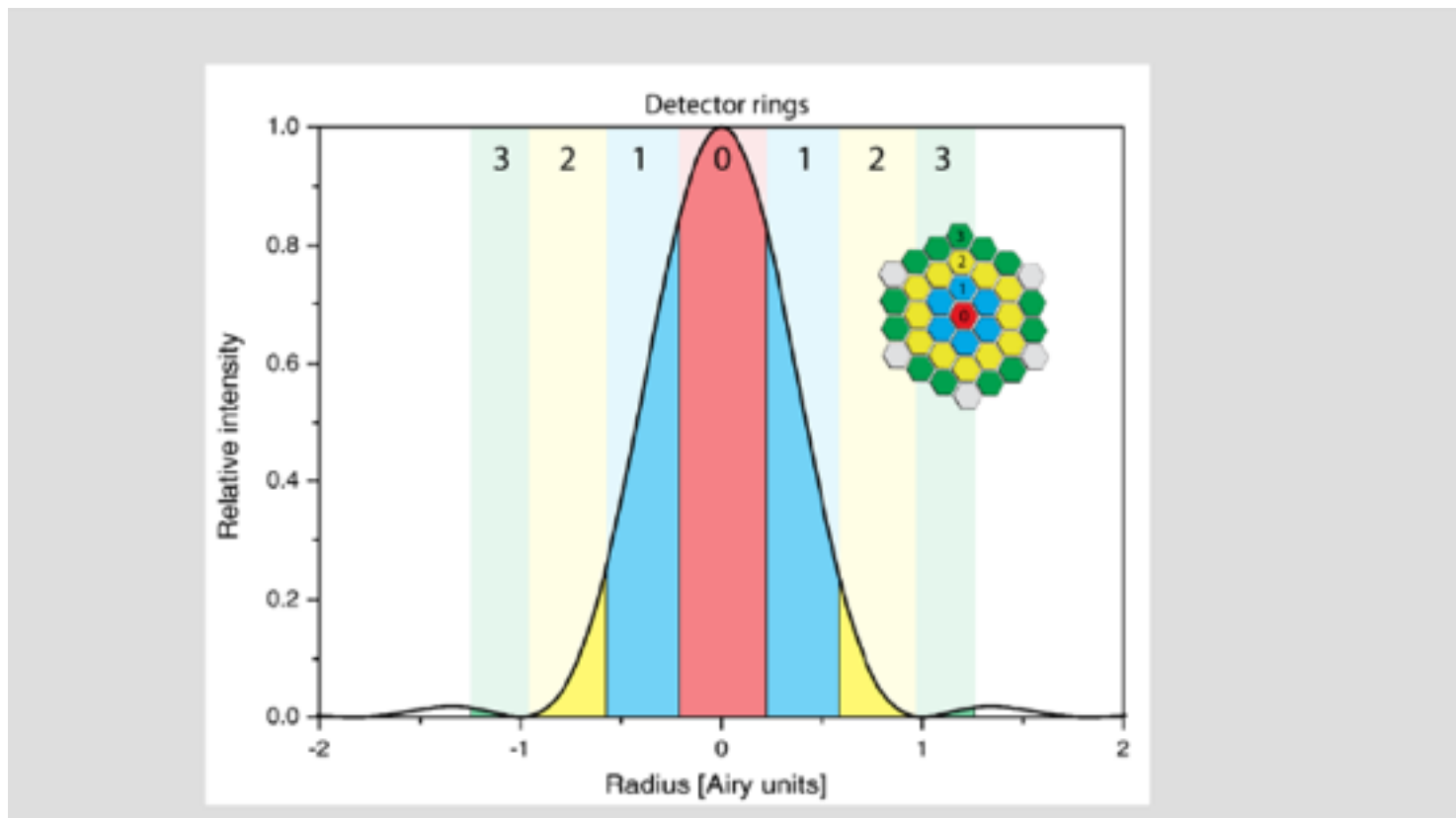


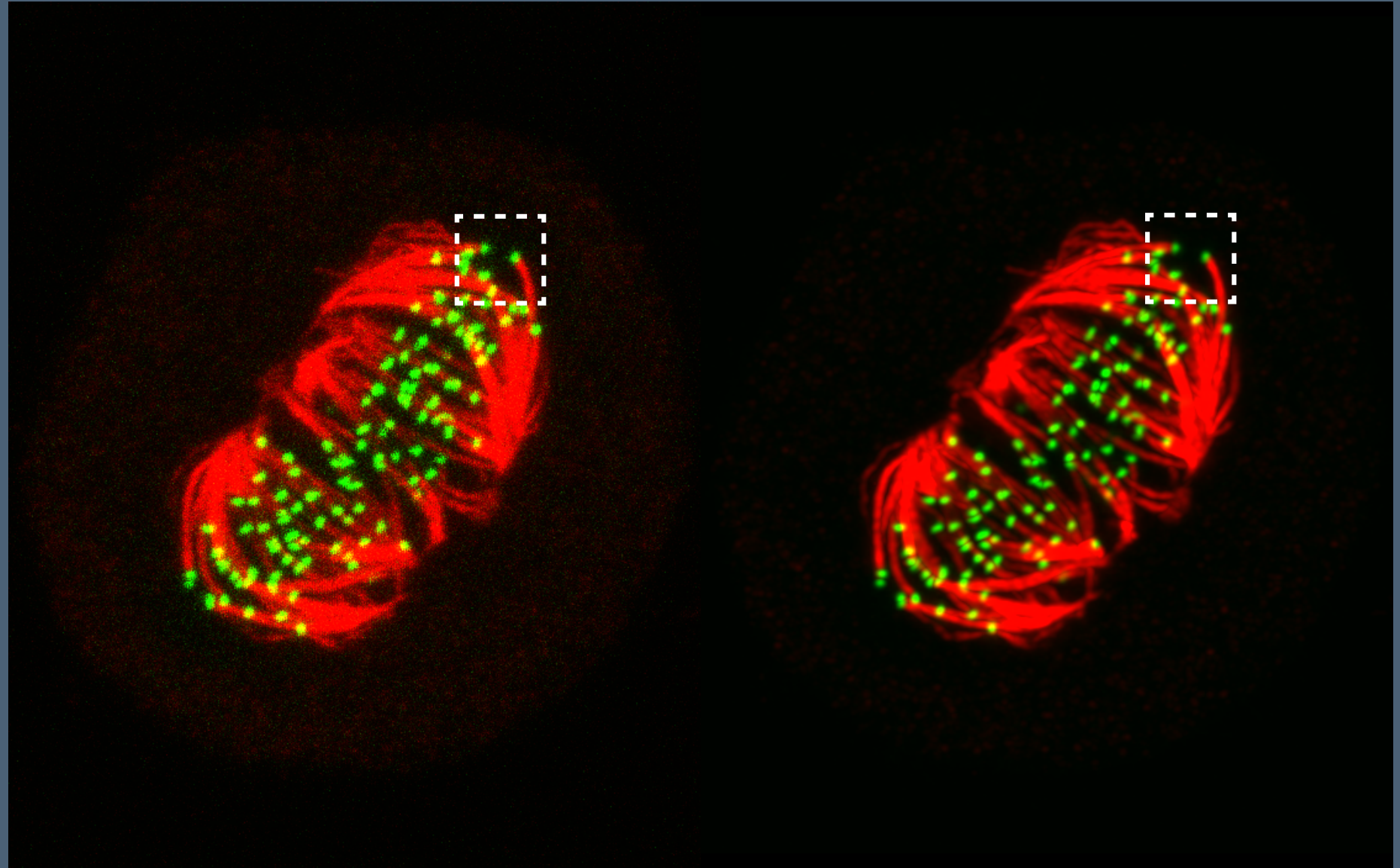


THEORY

'Airy-Scan' technology

effective PSF is now smaller.. > increased resolution (1.4x - 1.7x)

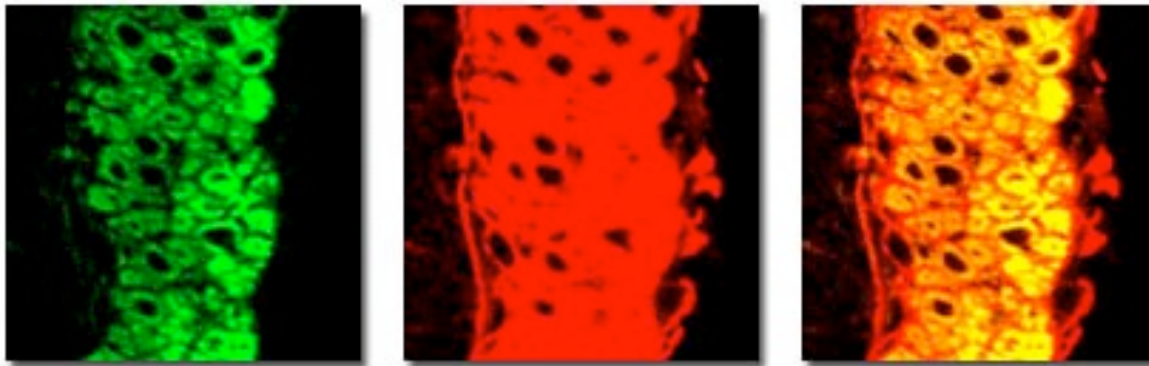




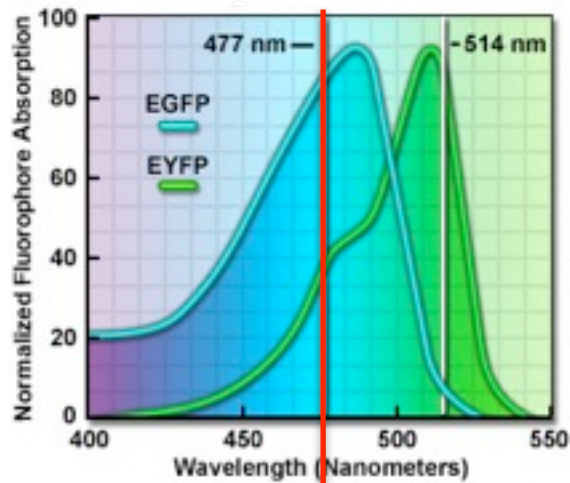
kinetochores (James Banecroft, Gruneberg Lab)



'bleed-through'

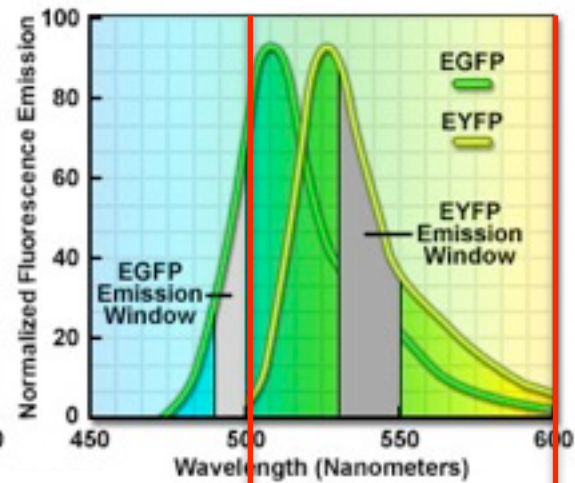


Absorption spectral profiles



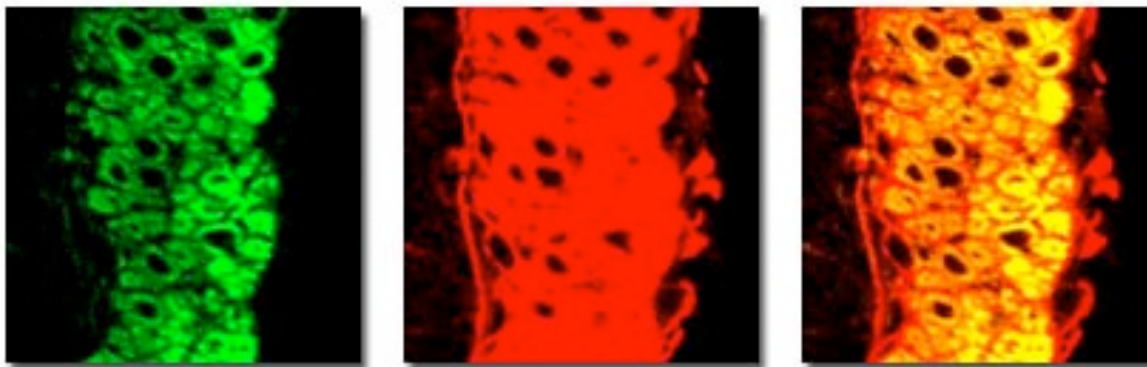
Excite at 477nm

Emission spectral profiles

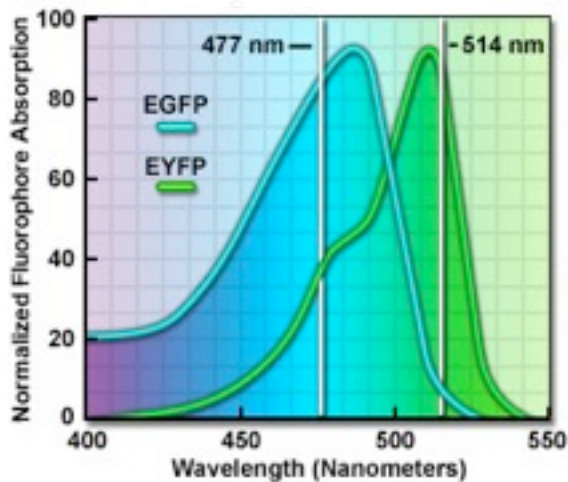


overlapping emission

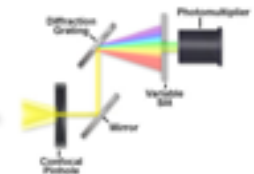
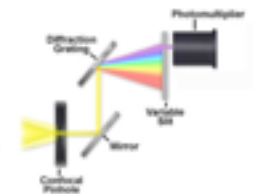
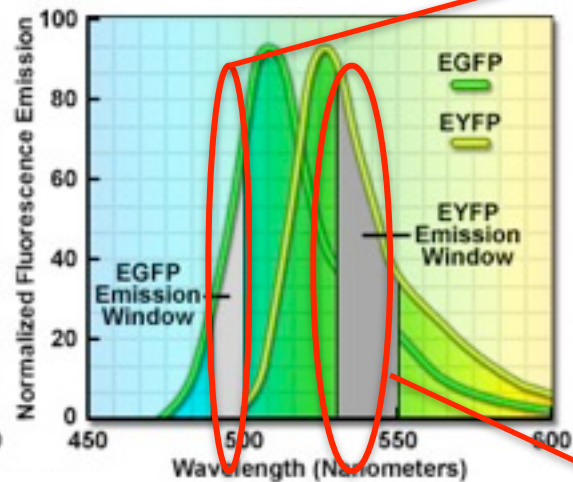
minimising 'bleed-through' Variable Slits



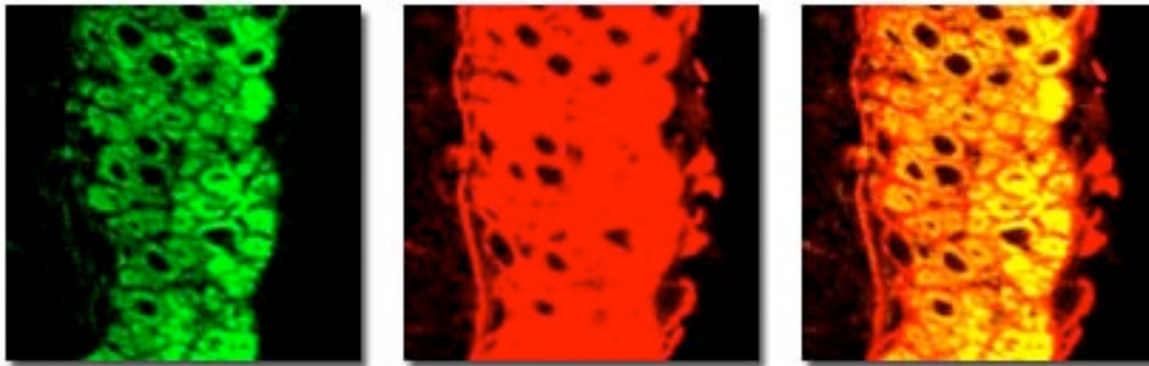
Absorption spectral profiles



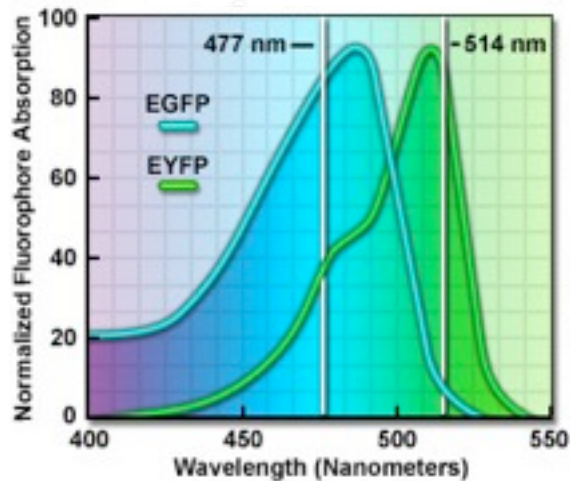
Emission spectral profiles



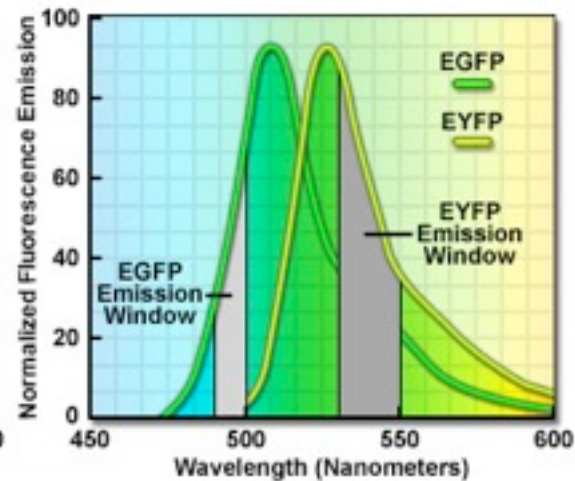
minimising 'bleed-through' Sequential Scanning



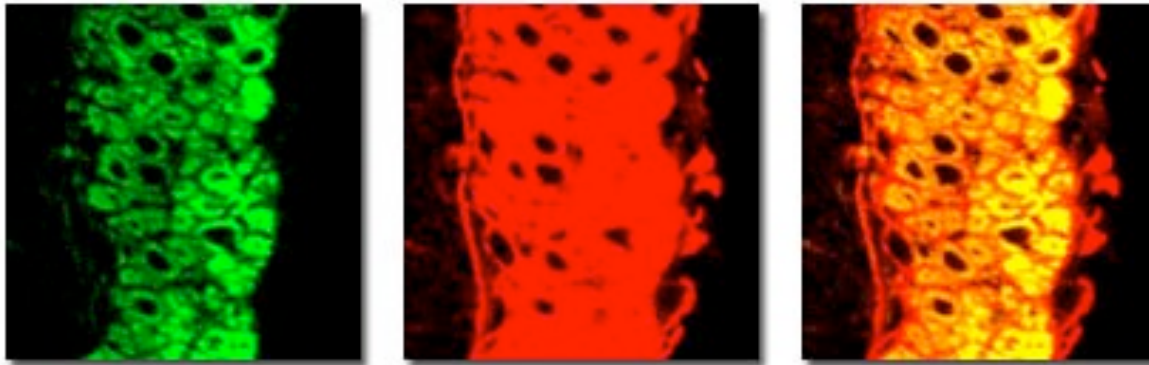
Absorption spectral profiles



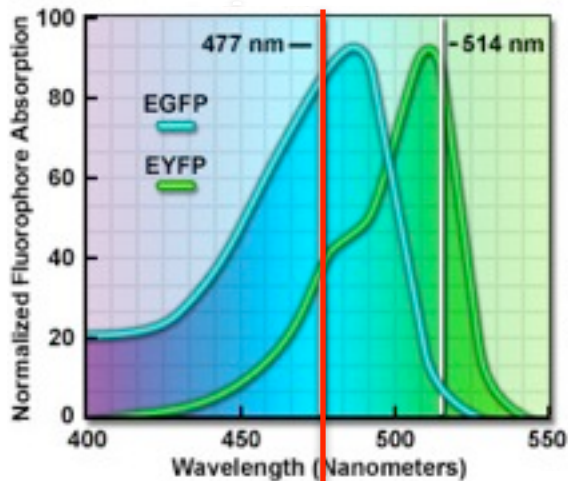
Emission spectral profiles



minimising 'bleed-through' Sequential Scanning

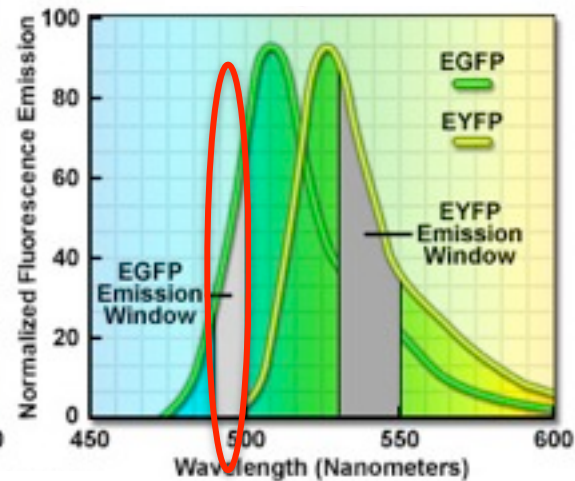


Absorption spectral profiles

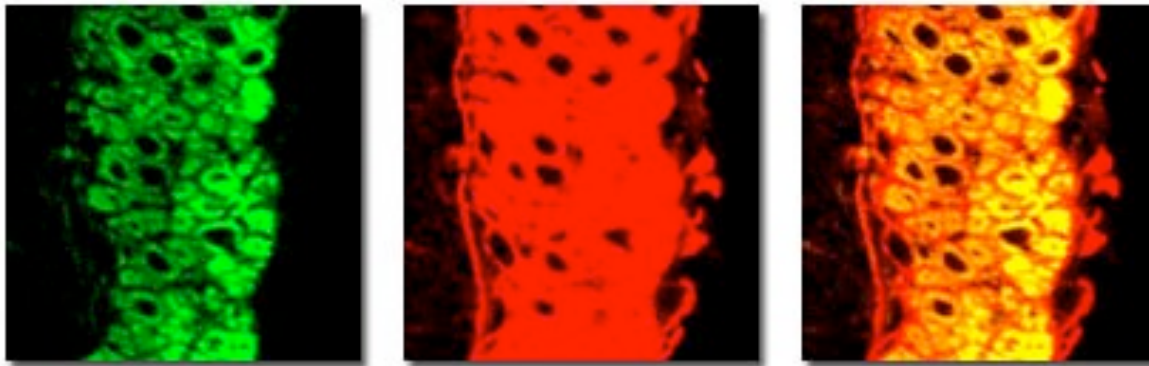


Excite at 477nm

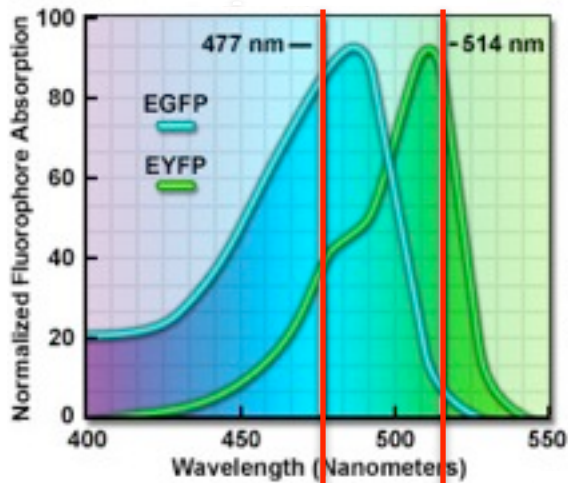
Emission spectral profiles



minimising 'bleed-through' Sequential Scanning



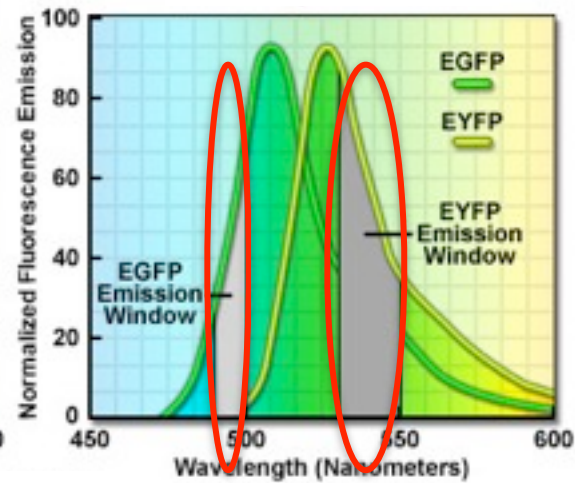
Absorption spectral profiles



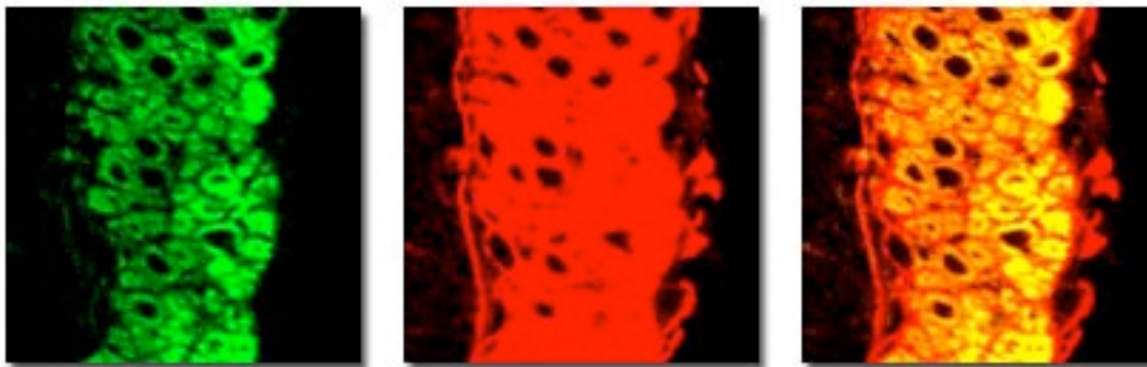
Excite at 477nm

Excite at 514nm

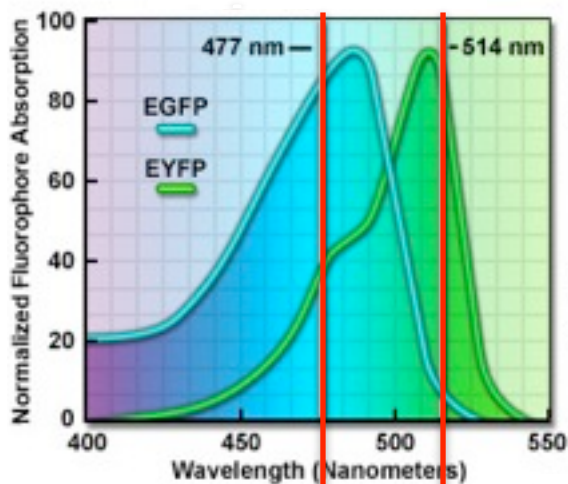
Emission spectral profiles



minimising 'bleed-through' Sequential Scanning



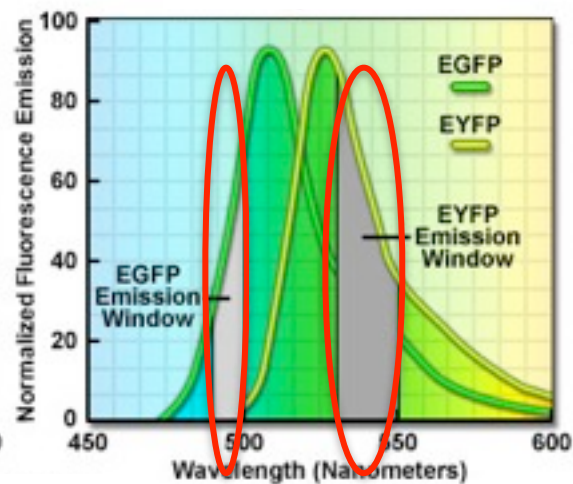
Absorption spectral profiles



Excite at 477nm

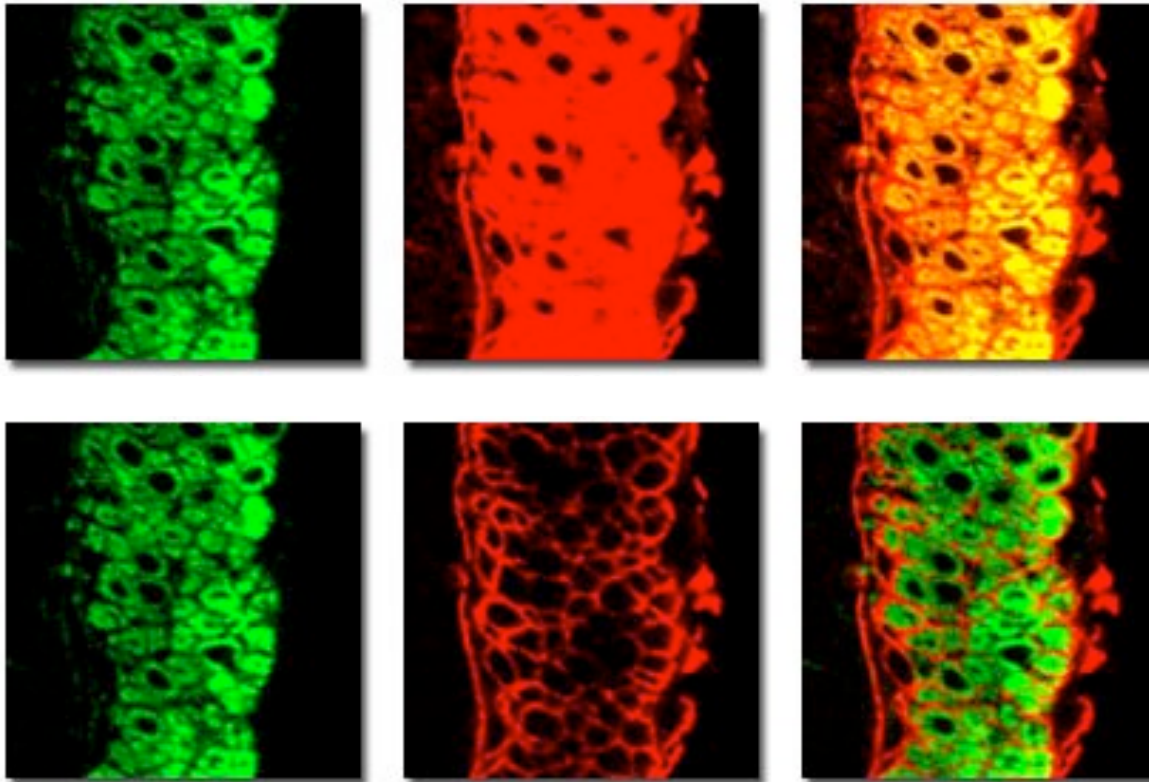
Excite at 514nm

Emission spectral profiles

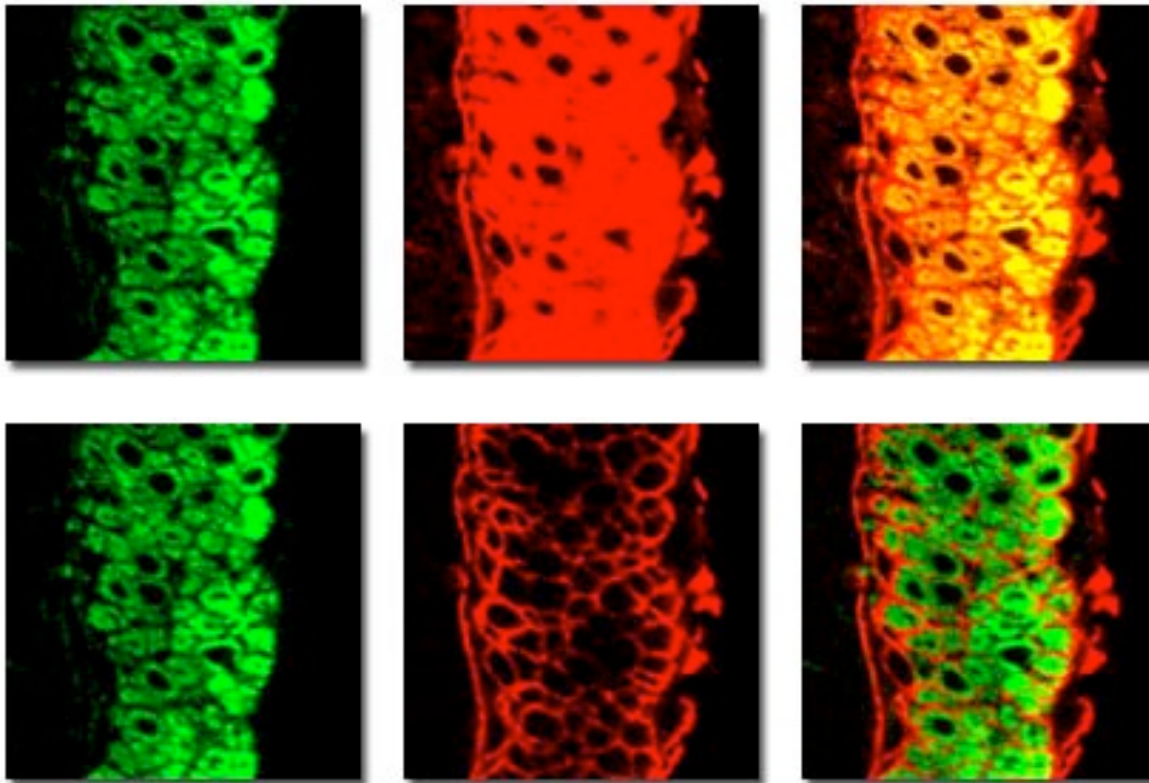


Temporal separation

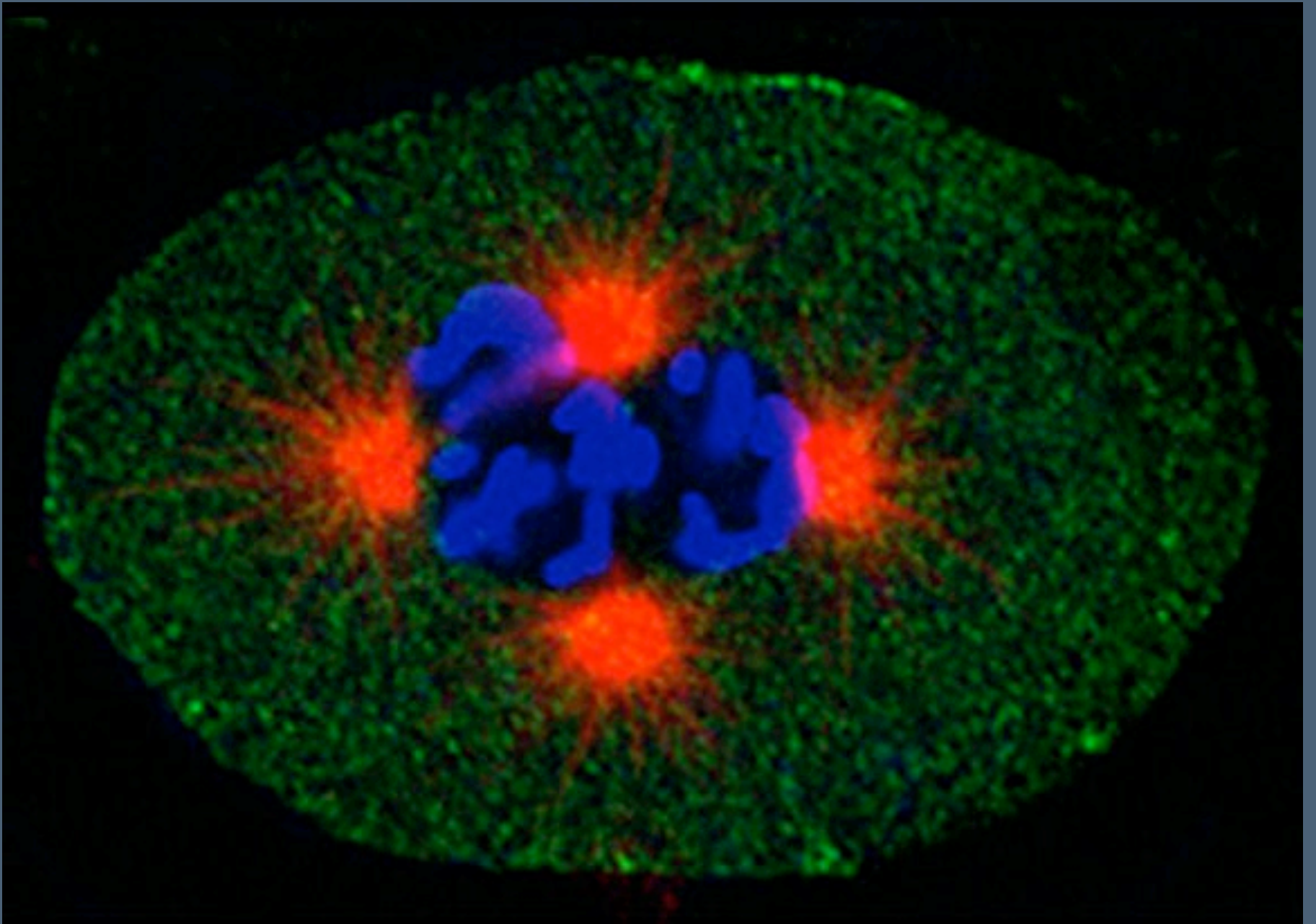
minimising 'bleed-through'

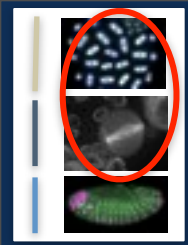


minimising 'bleed-through'



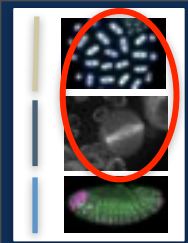
Adjust detector slit widths
Use sequential scanning





Confocal Microscopes

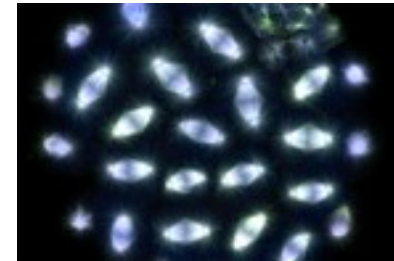
Confocal Microscopes



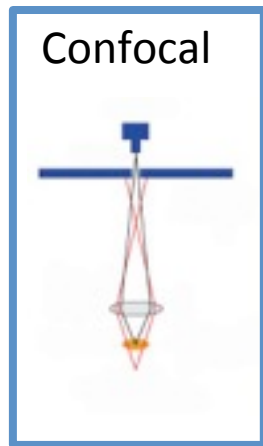
Confocal Microscopes



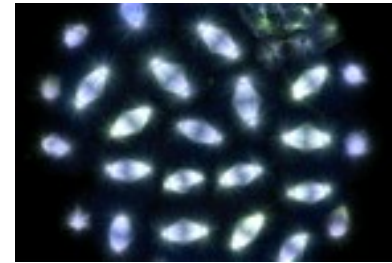
Laser
Scanning



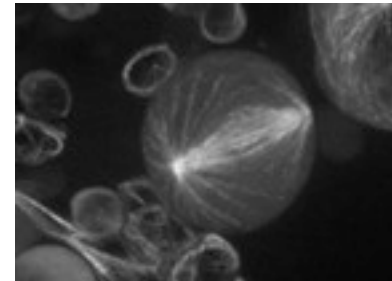
Confocal Microscopes



Laser
Scanning



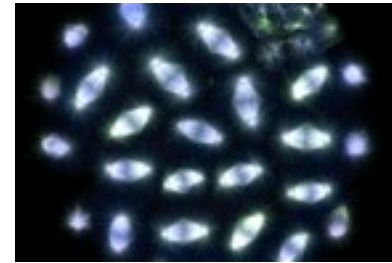
Spinning disc



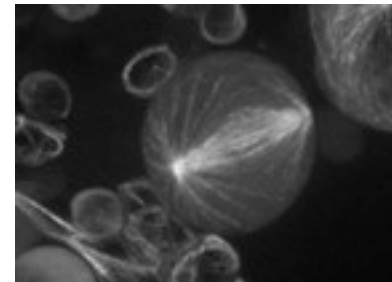
Confocal Microscopes



Laser
Scanning

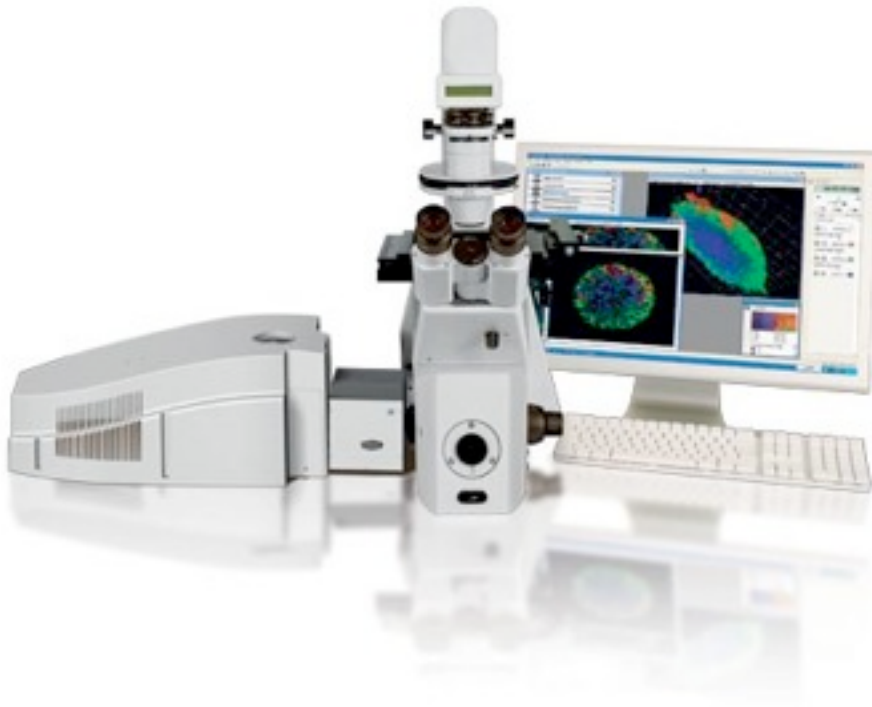


Spinning disc



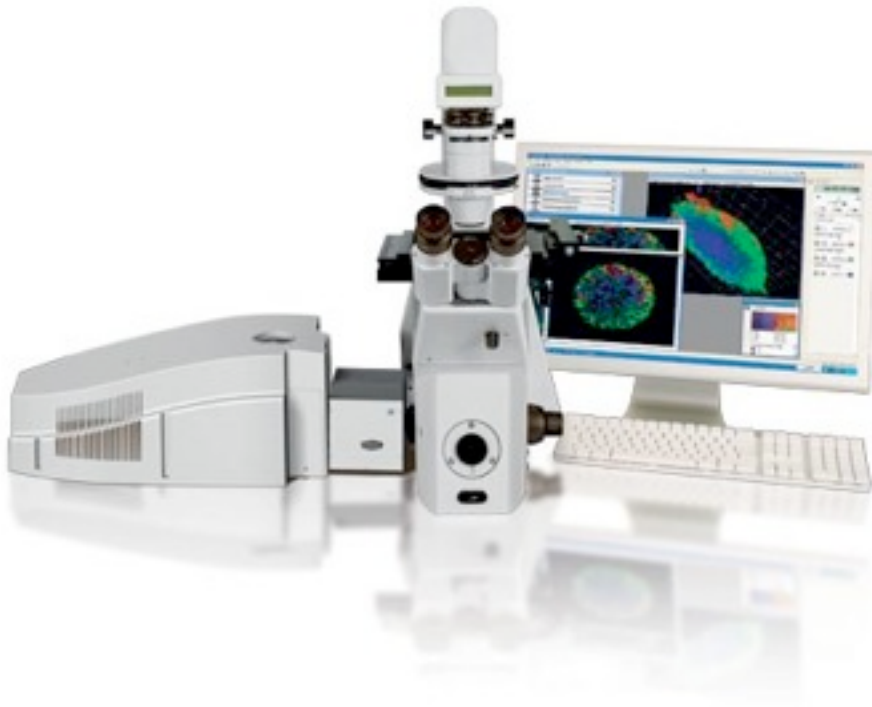
Both are confocals

Spinning Disc Confocal



Great for live cell imaging

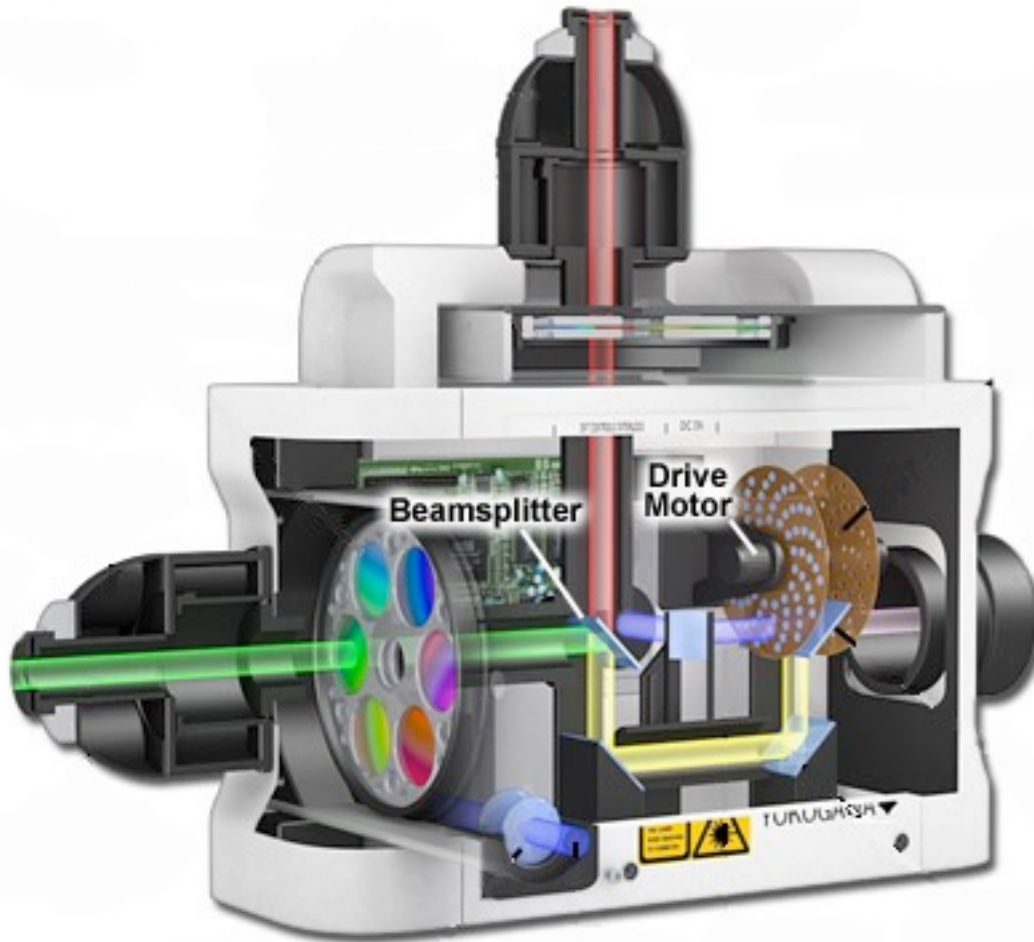
Spinning Disc Confocal



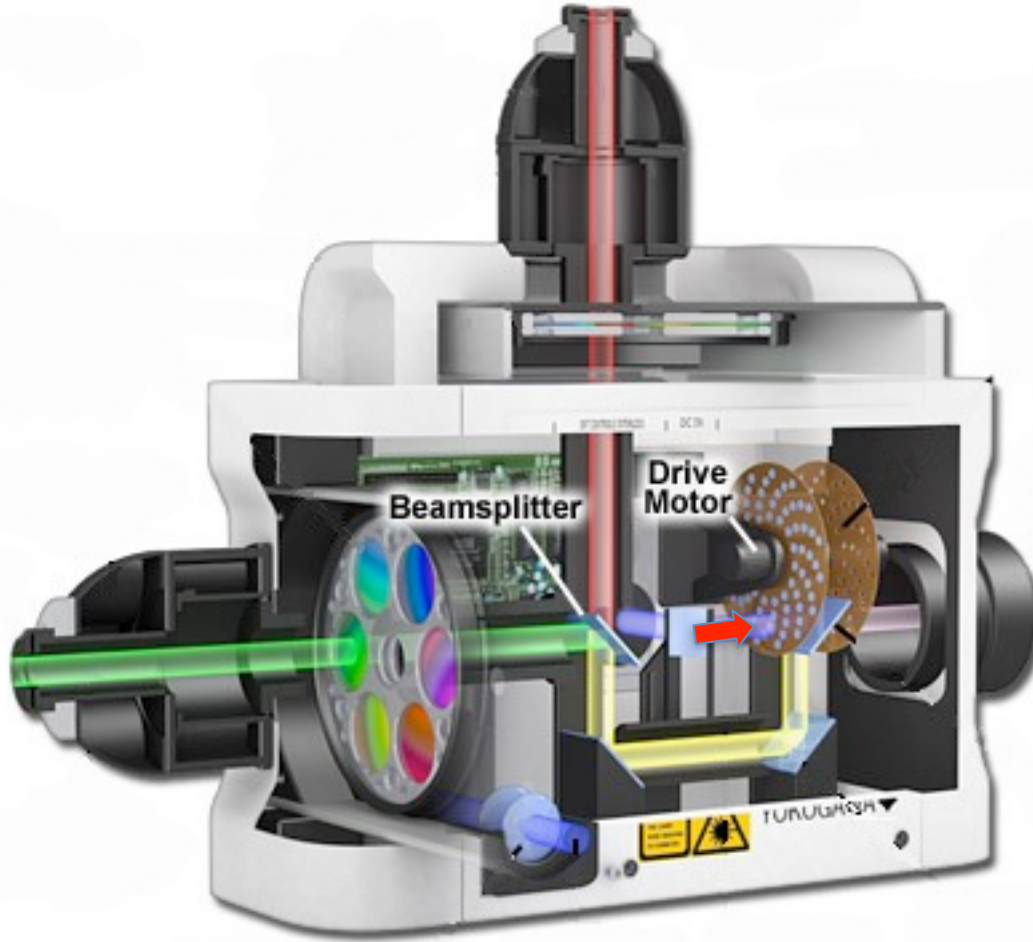
Great for live cell imaging

Can collect many images per second

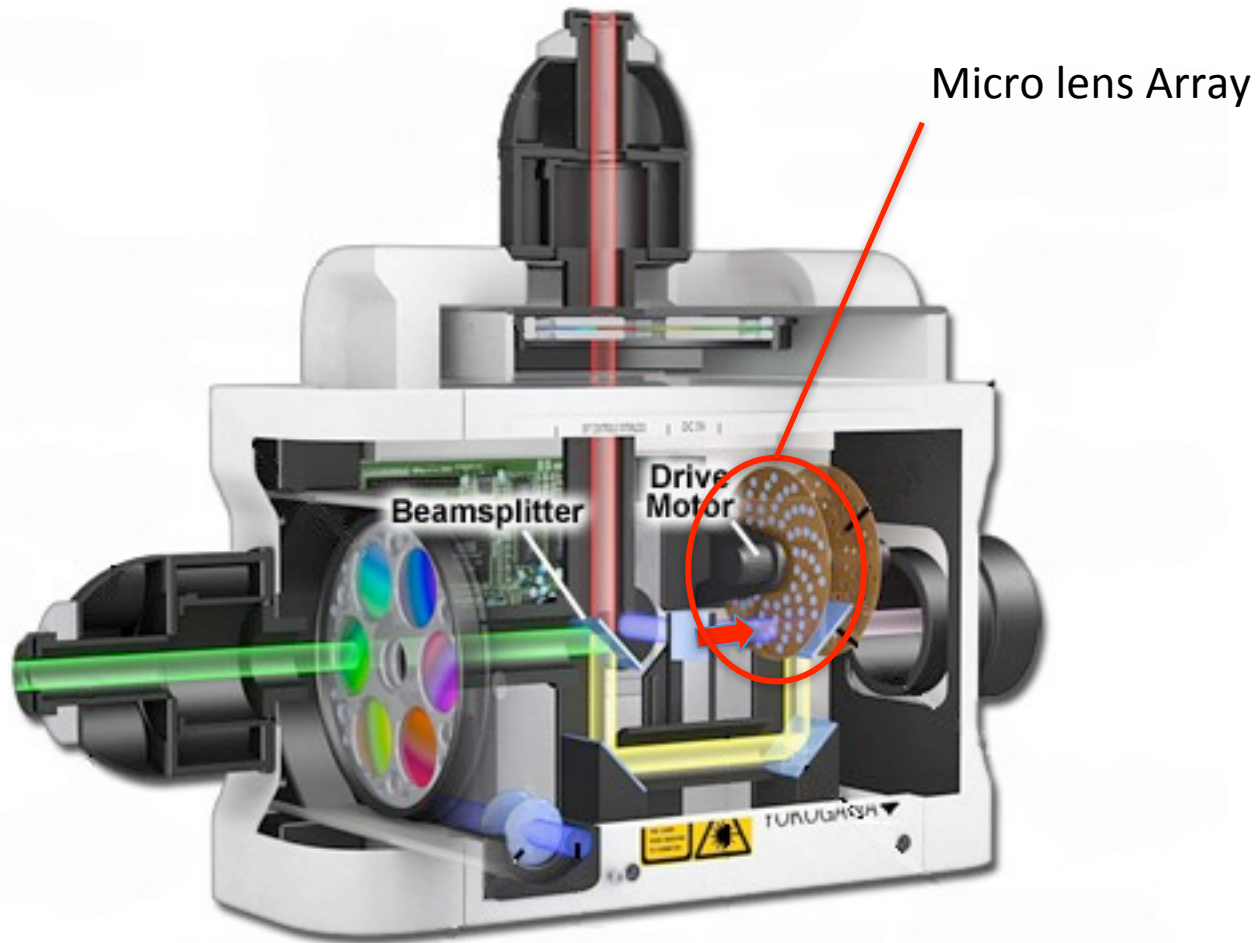
Yokogawa CSU-X1



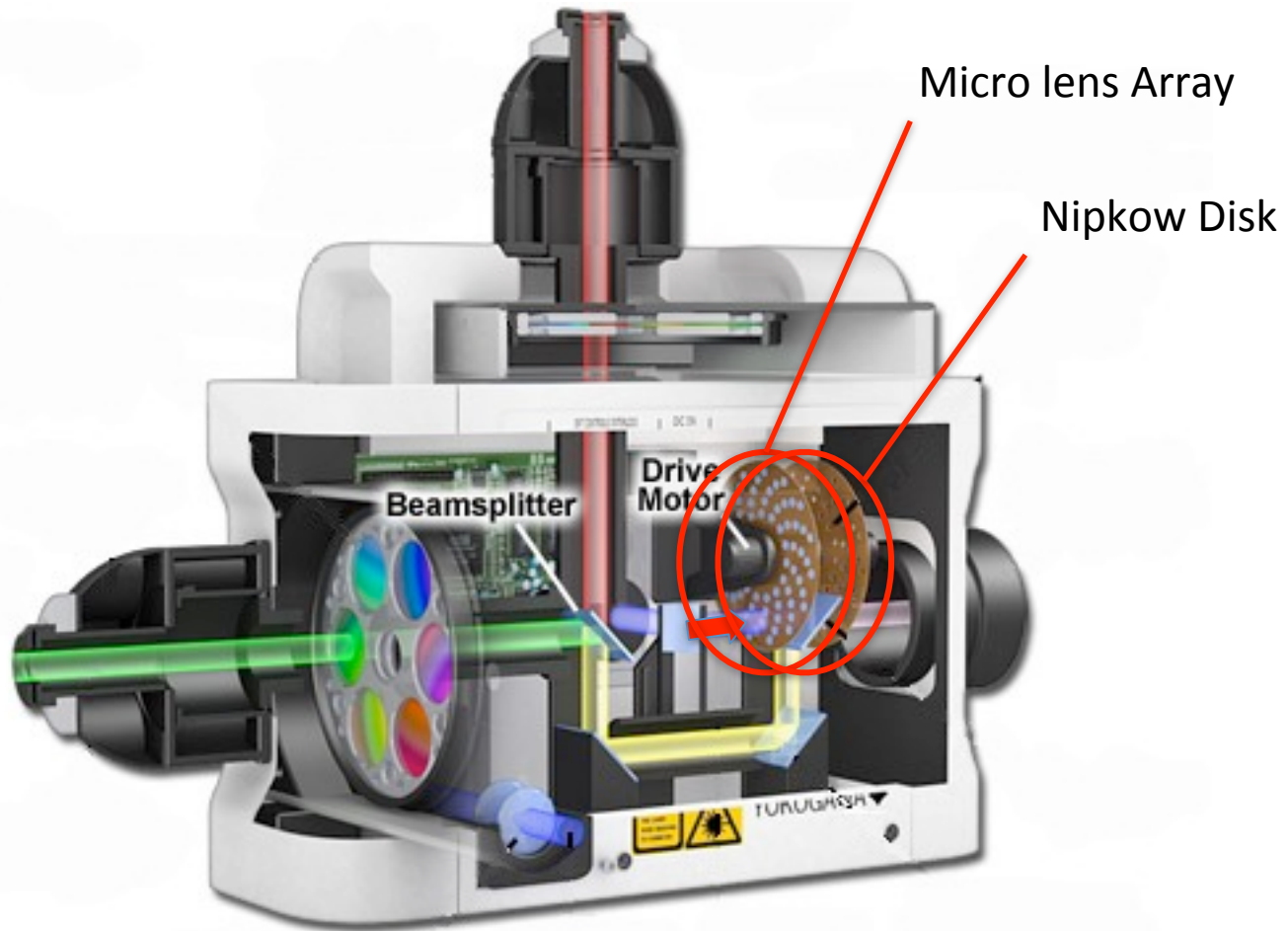
Yokogawa CSU-X1



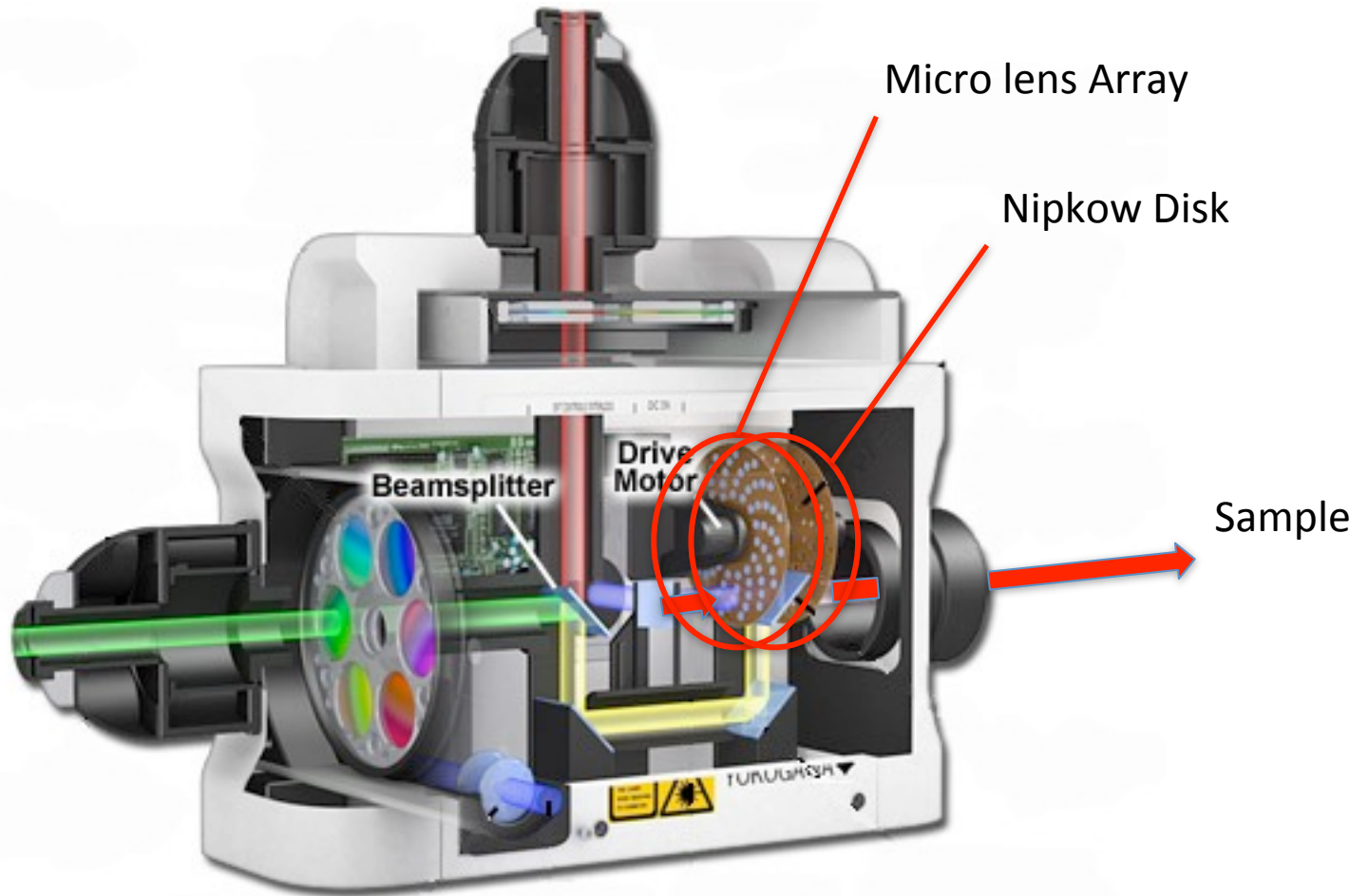
Yokogawa CSU-X1



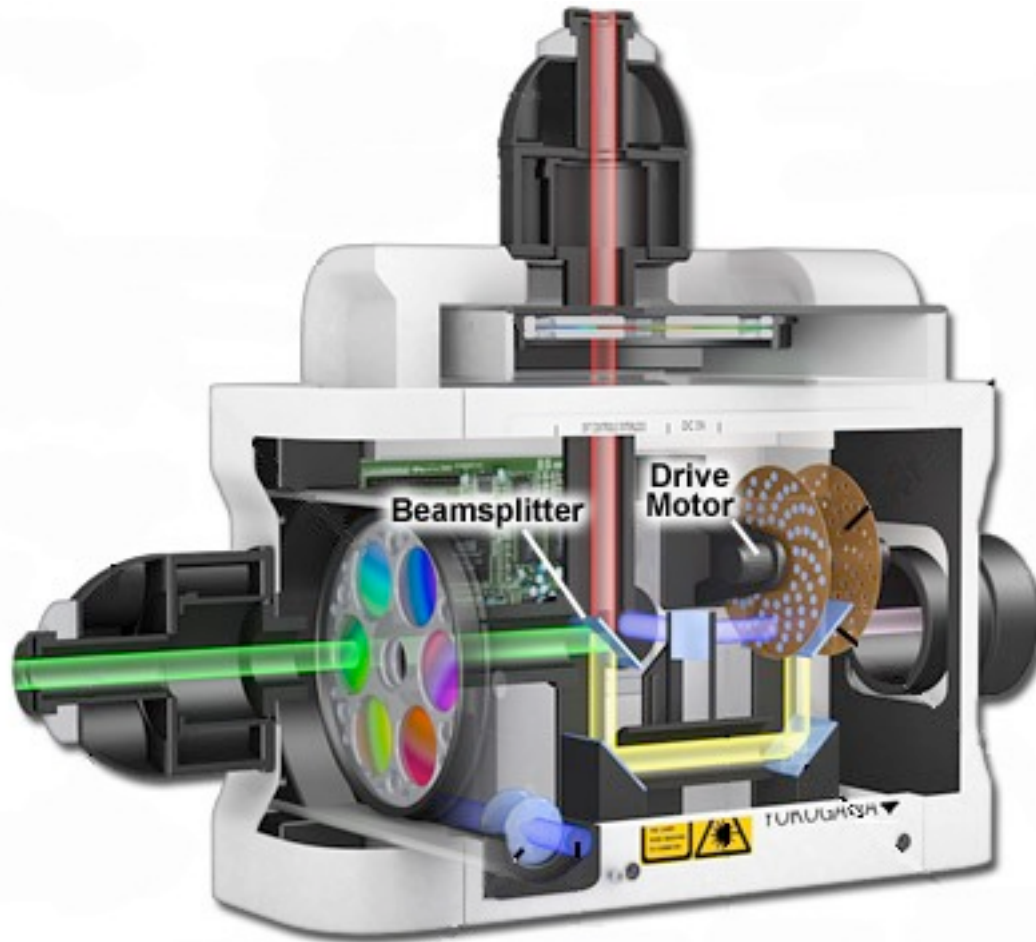
Yokogawa CSU-X1



Yokogawa CSU-X1

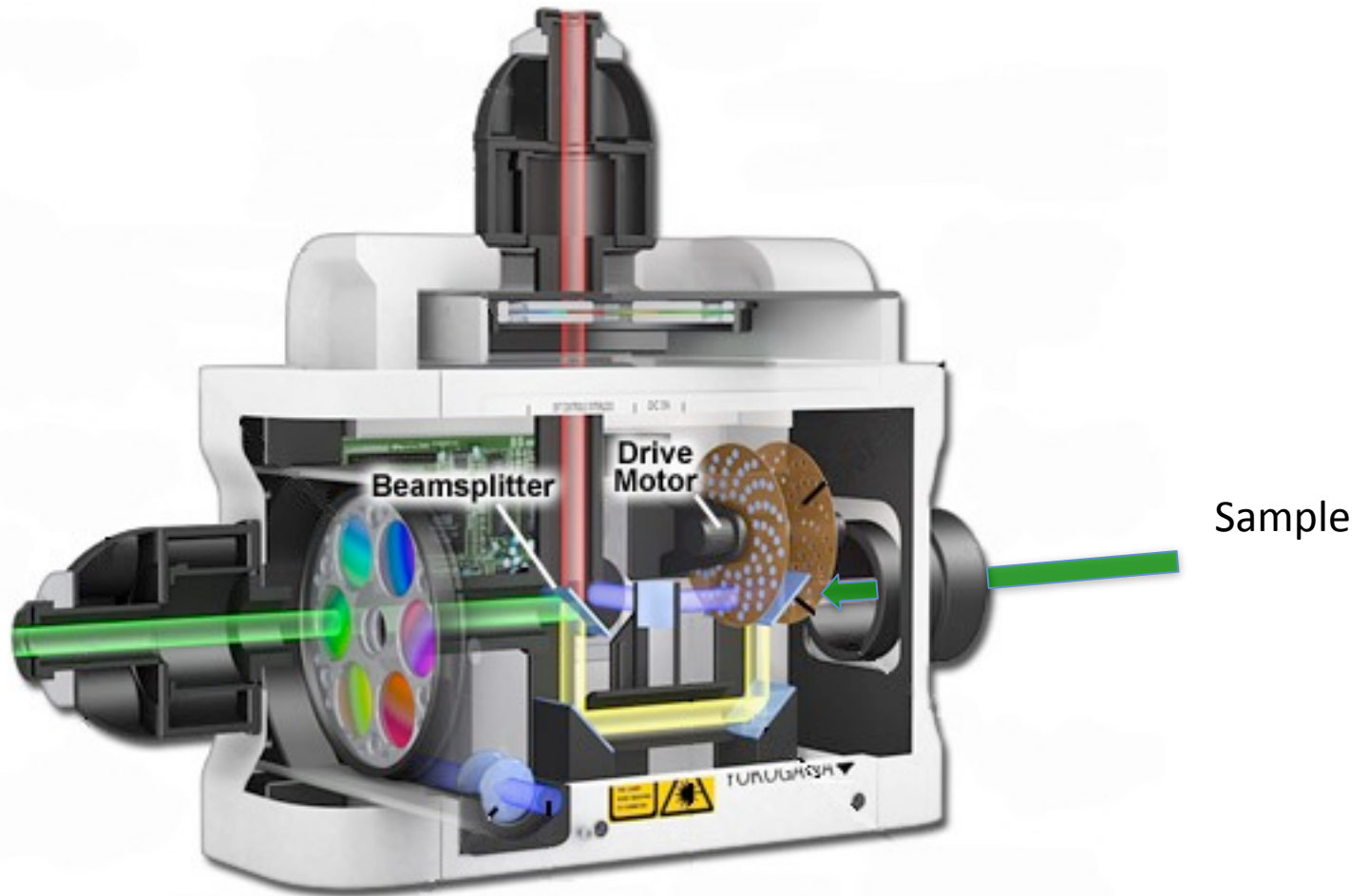


Yokogawa CSU-X1

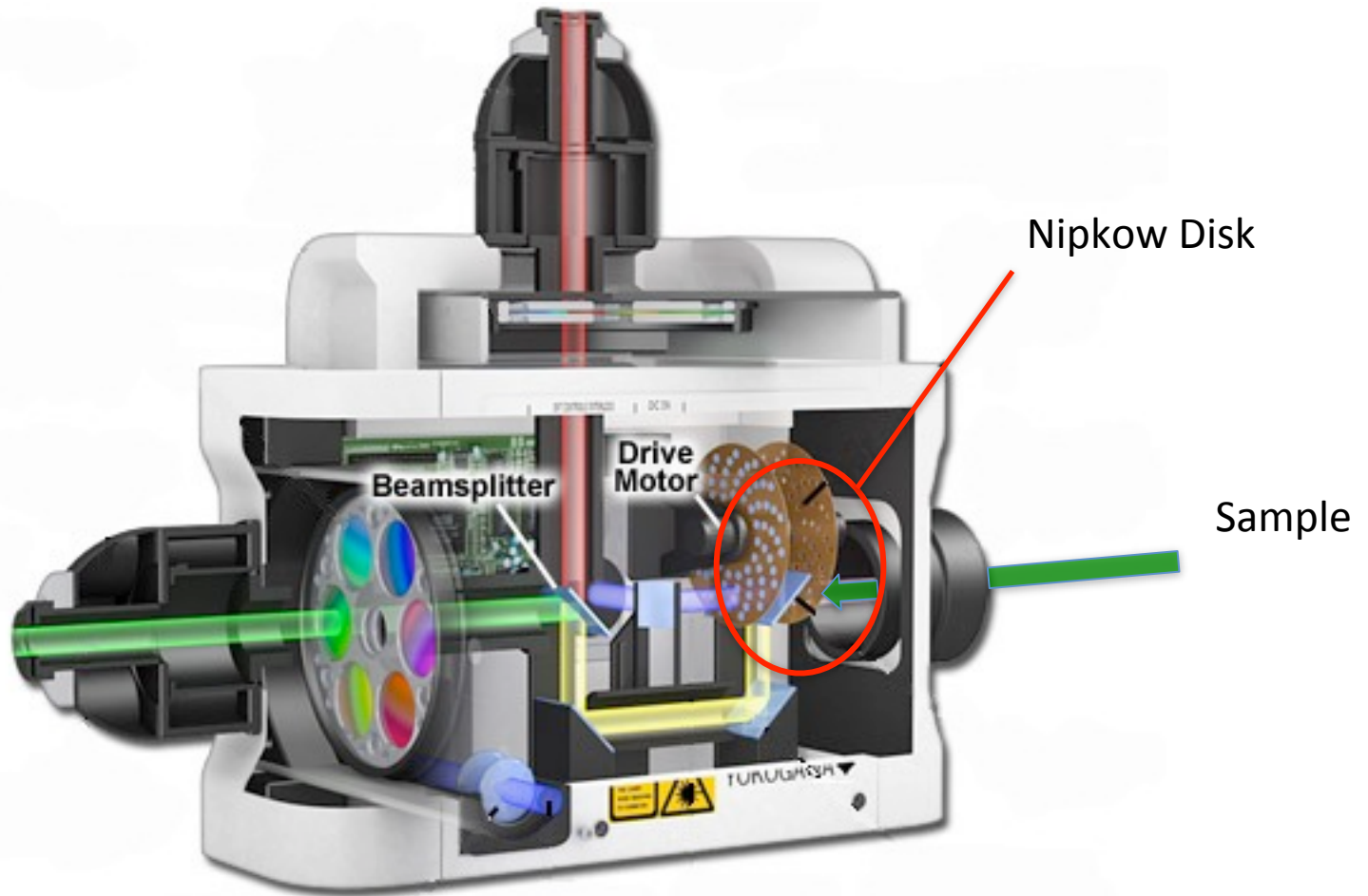


Sample

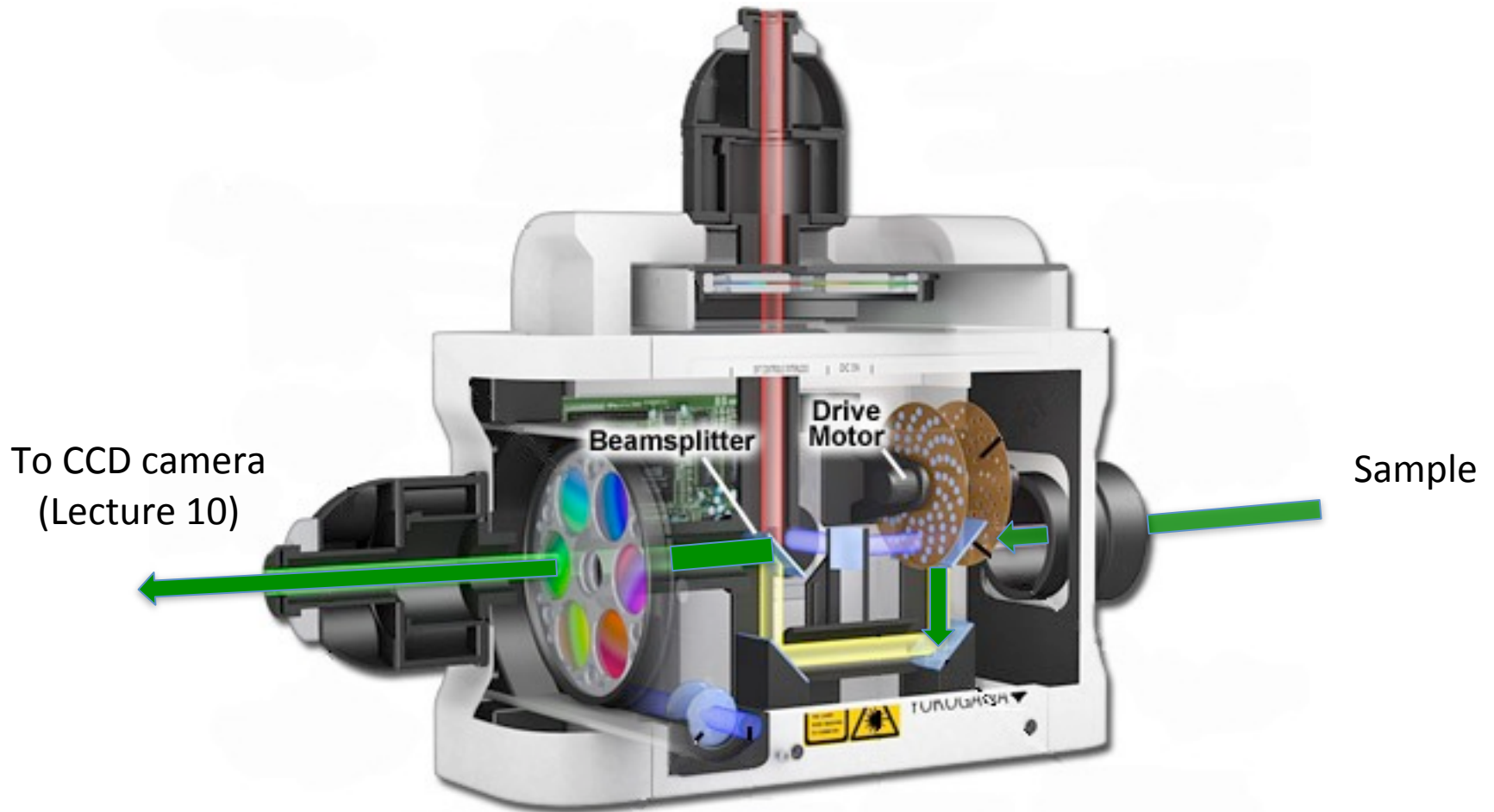
Yokogawa CSU-X1



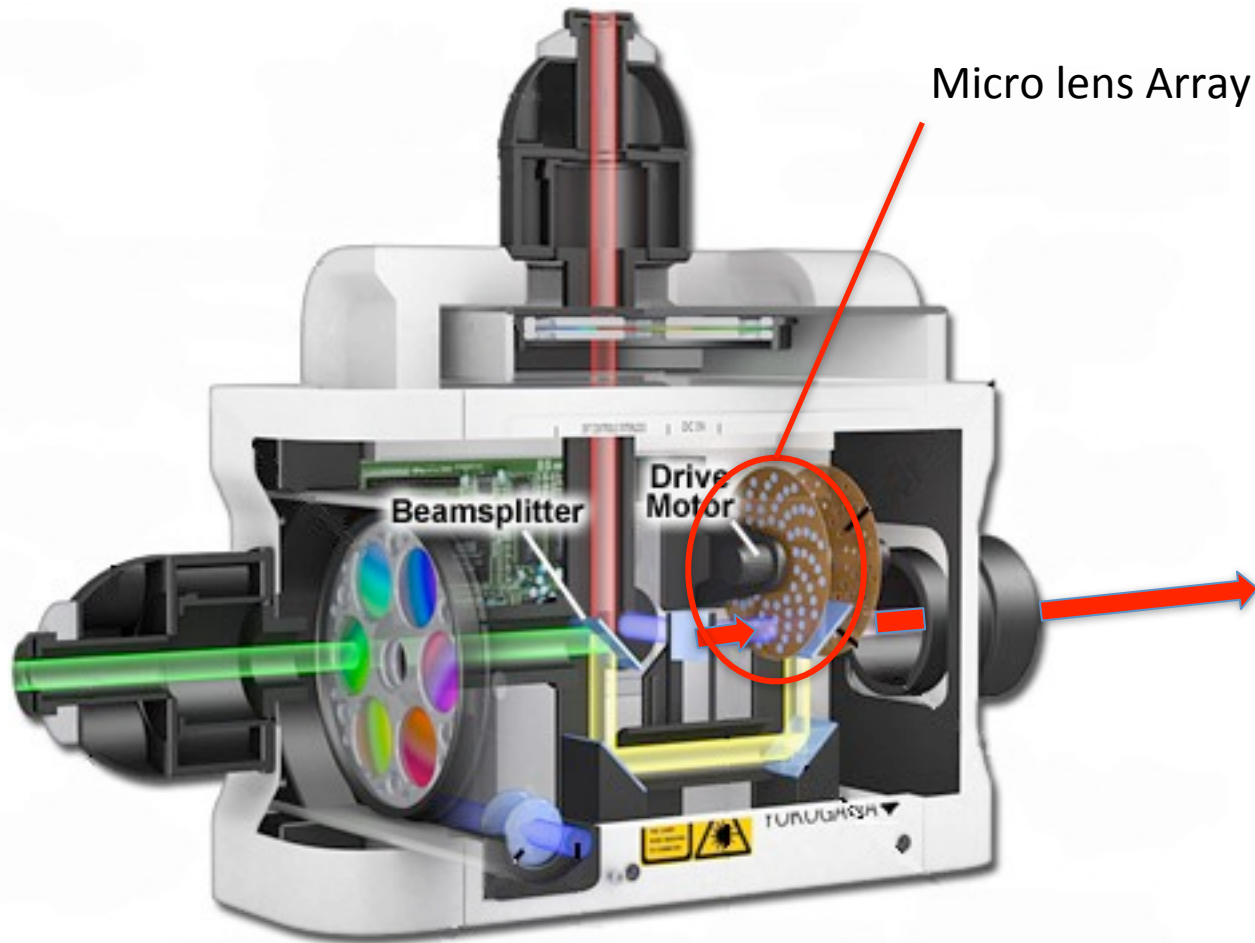
Yokogawa CSU-X1



Yokogawa CSU-X1

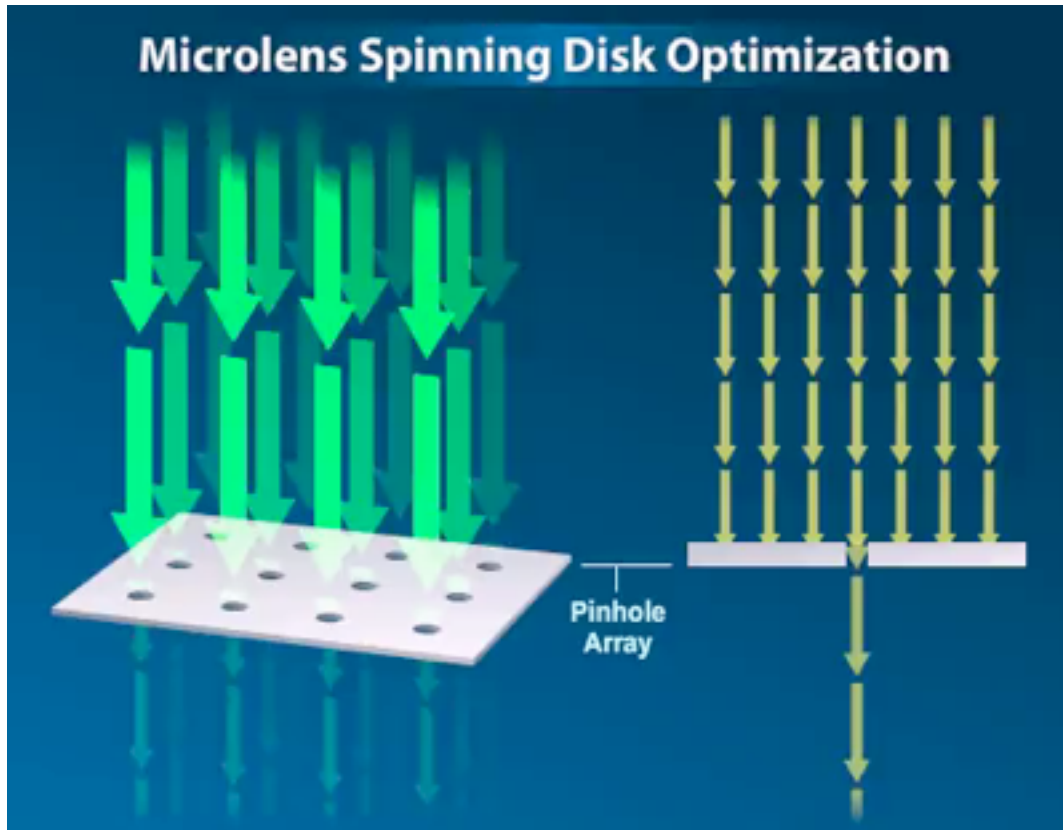


Yokogawa CSU-X1



Yokogawa Spinning Disc Confocal

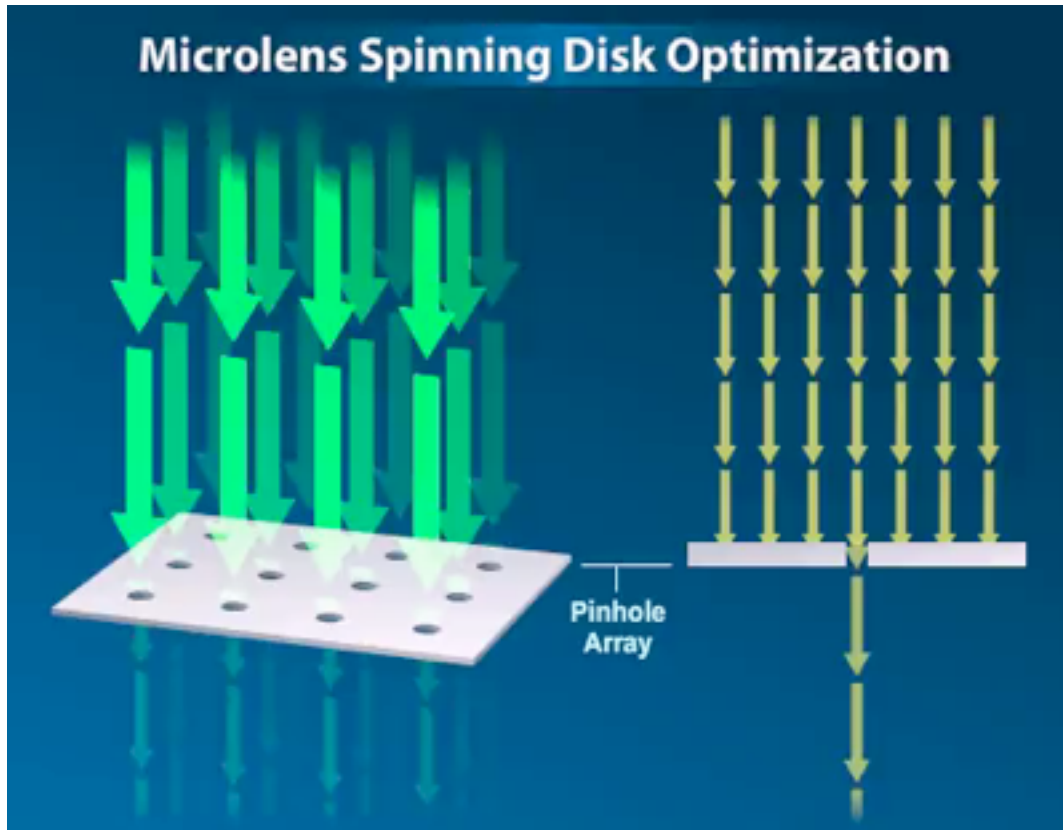
Microlens Spinning Disk Optimization



just a pinhole array –
Optimised for ‘cofocality’
and ‘crosstalk’

too much light is blocked
from reaching the specimen

Yokogawa Spinning Disc Confocal



just a pinhole array –
Optimised for 'cofocality'
and 'crosstalk'

too much light is blocked
from reaching the specimen

Only 4% light passes through disc



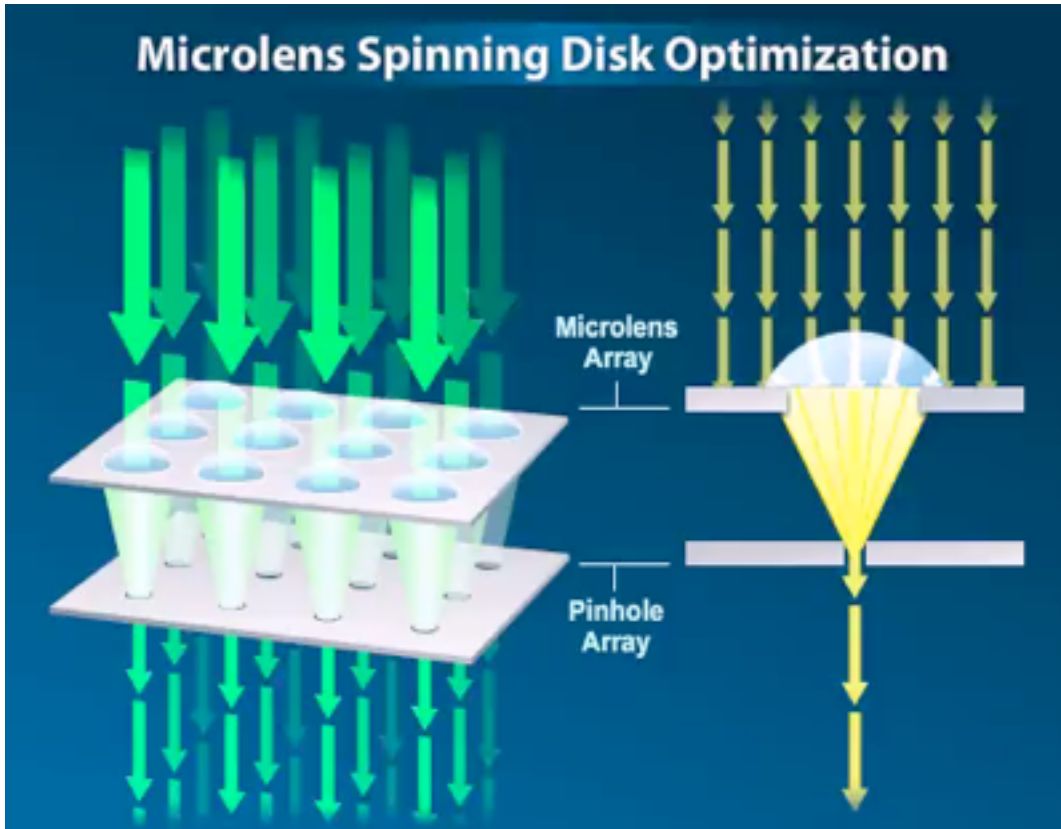
Yokogawa Spinning Disc Confocal

just a pinhole array –
Optimised for ‘cofocality’
and ‘crosstalk’

too much light is blocked
from reaching the specimen

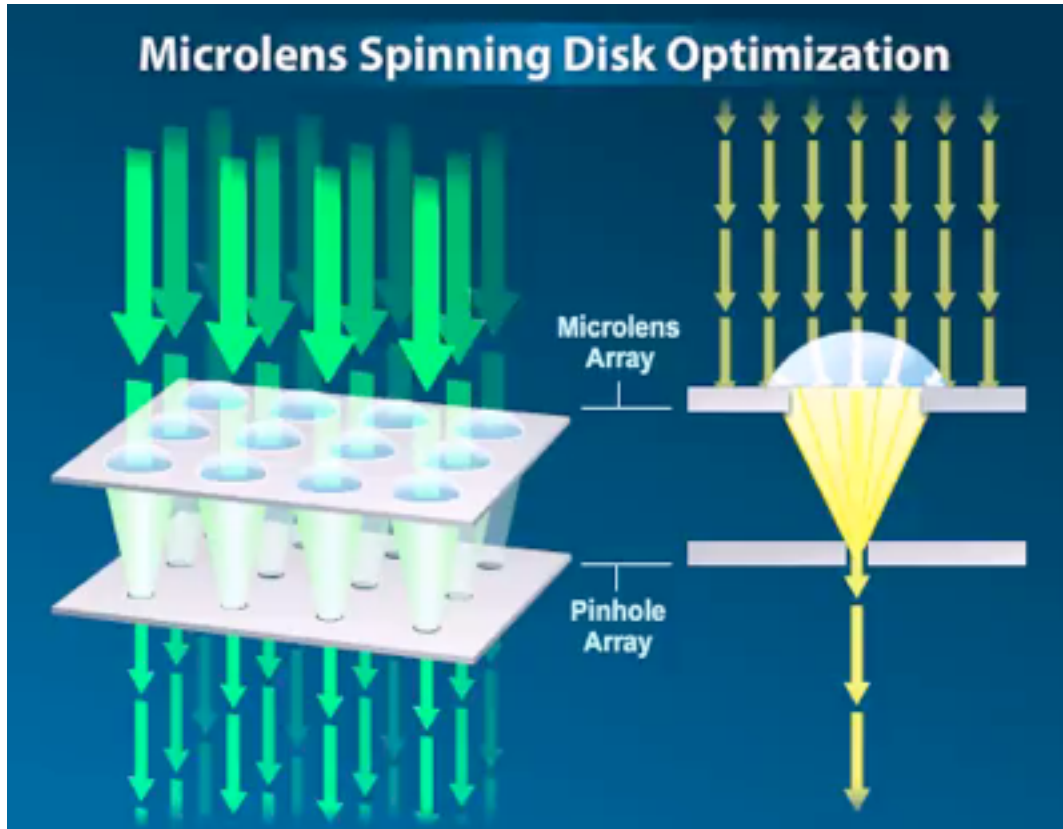
Only 4% light passes through disc

Yokogawa Spinning Disc Confocal



micro-lens array increase
the light
reaching the specimen

Yokogawa Spinning Disc Confocal



micro-lens array increase the light reaching the specimen

Typically 56% light passes through disc

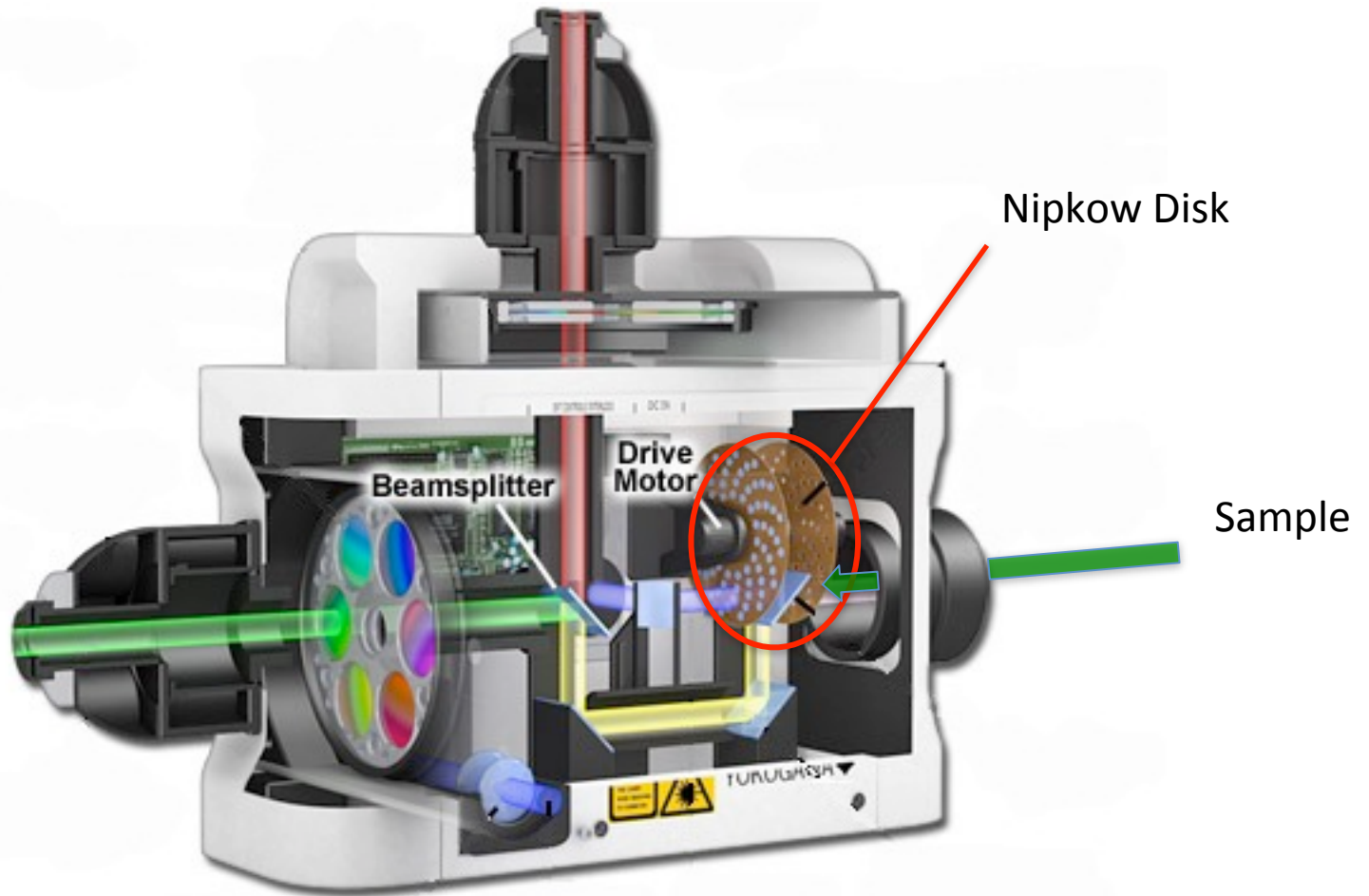


Yokogawa Spinning Disc Confocal

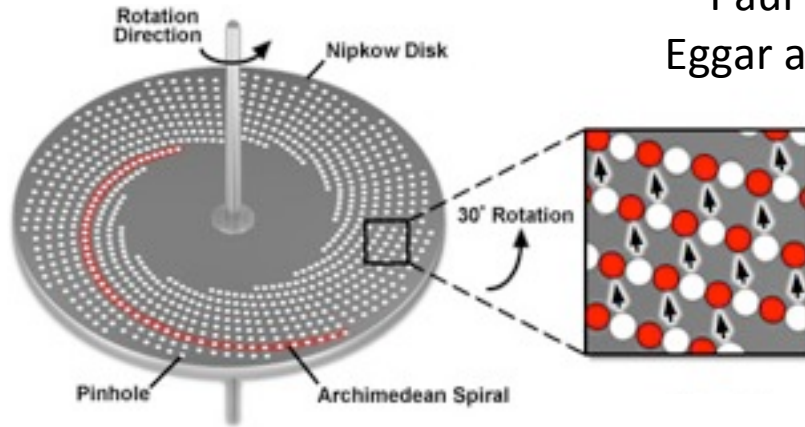
micro-lens array increase
the light
reaching the specimen

Typically 56% light passes through disc

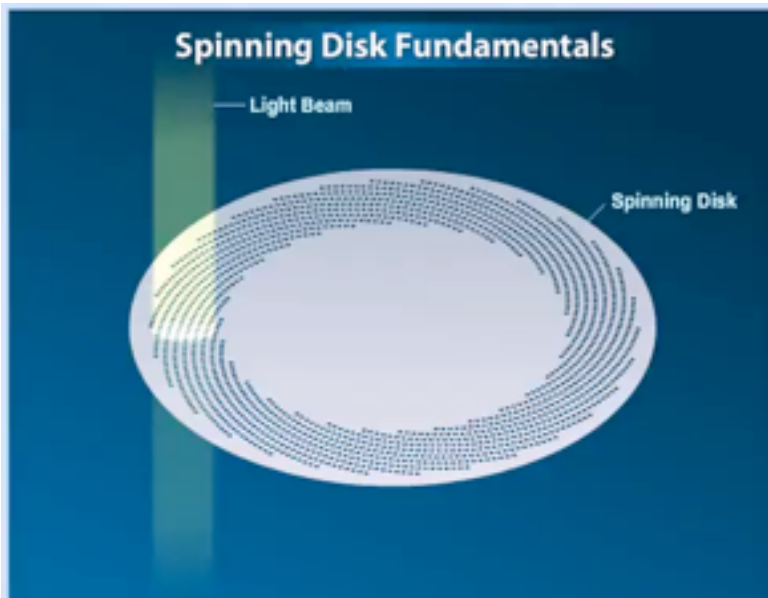
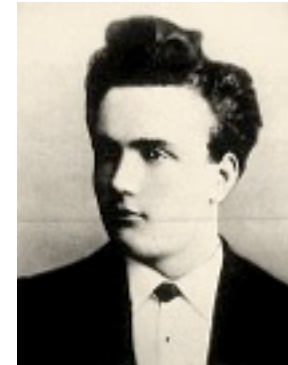
Yokogawa CSU-X1



The Nipkow Disk



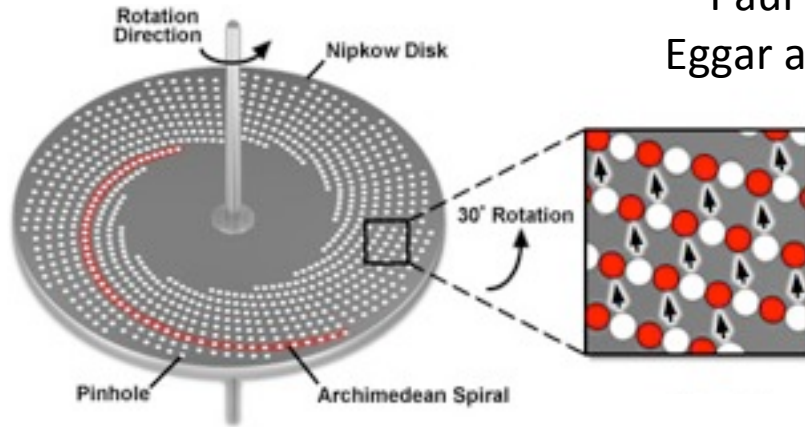
Paul Nipkow, 1884
Eggar and Petran, 1967



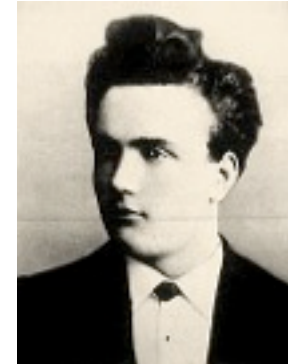
Approx. 1000 pinholes

Single frame created with each
30-degree of rotation of disc
(12 frames per rotation)

The Nipkow Disk



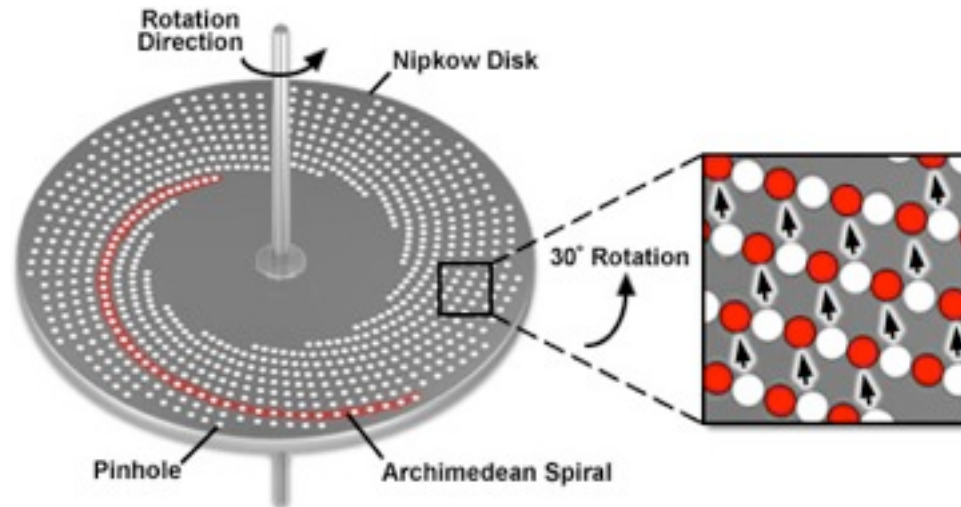
Paul Nipkow, 1884
Eggar and Petran, 1967



Approx. 1000 pinholes

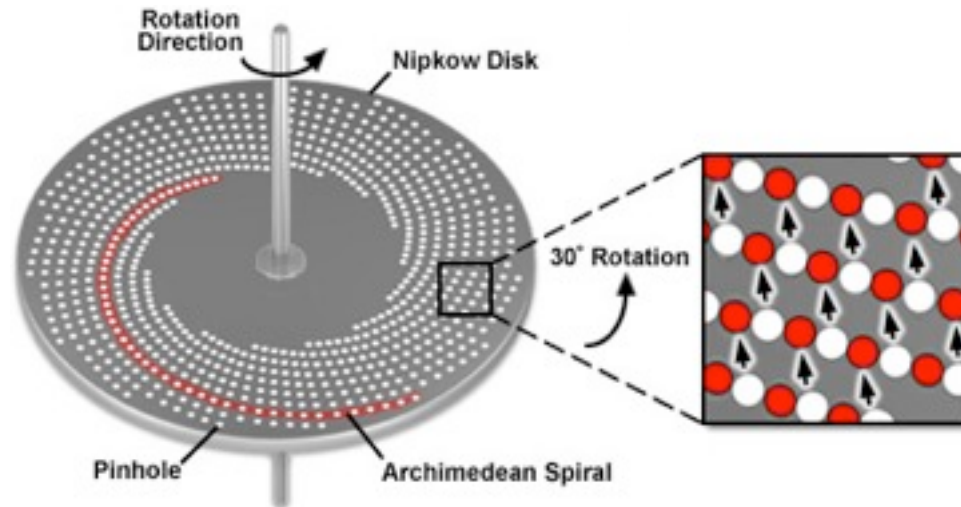
Single frame created with each
30-degree of rotation of disc
(12 frames per rotation)

The Nipkow Disk



Larger pinholes - brighter image, but less “confocal”

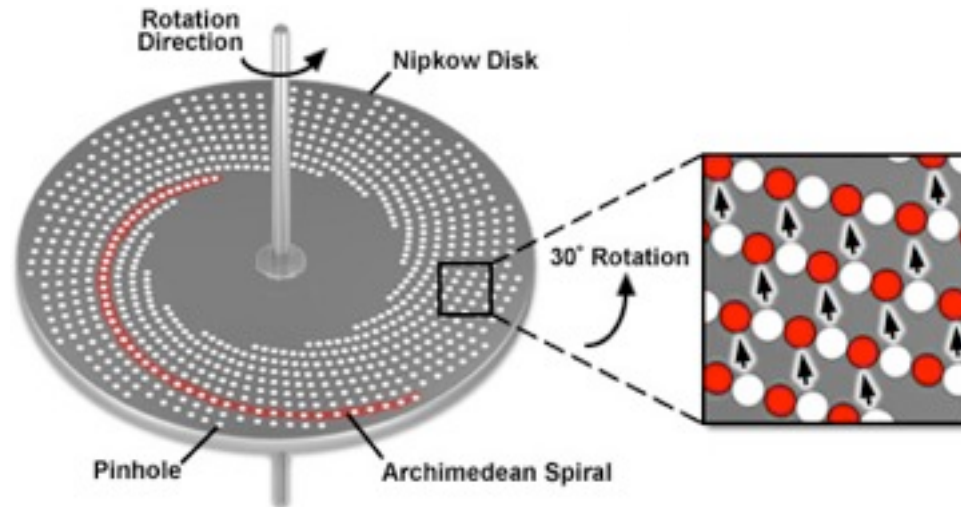
The Nipkow Disk



Larger pinholes - brighter image, but less “confocal”

Pinholes fixed size: Typically = 50um

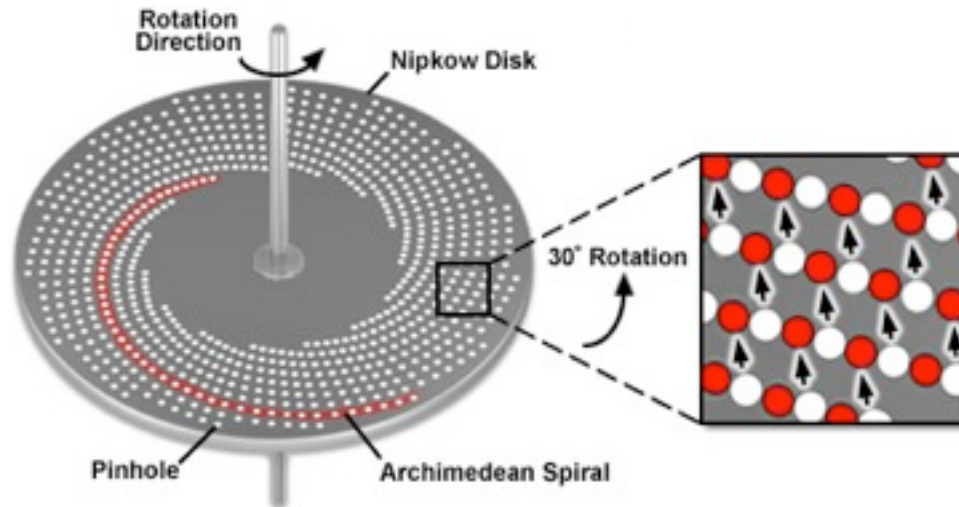
The Nipkow Disk



Larger pinholes - brighter image, but less “confocal”

Pinholes fixed size: Typically = 50 μ m
(optimised for biology)

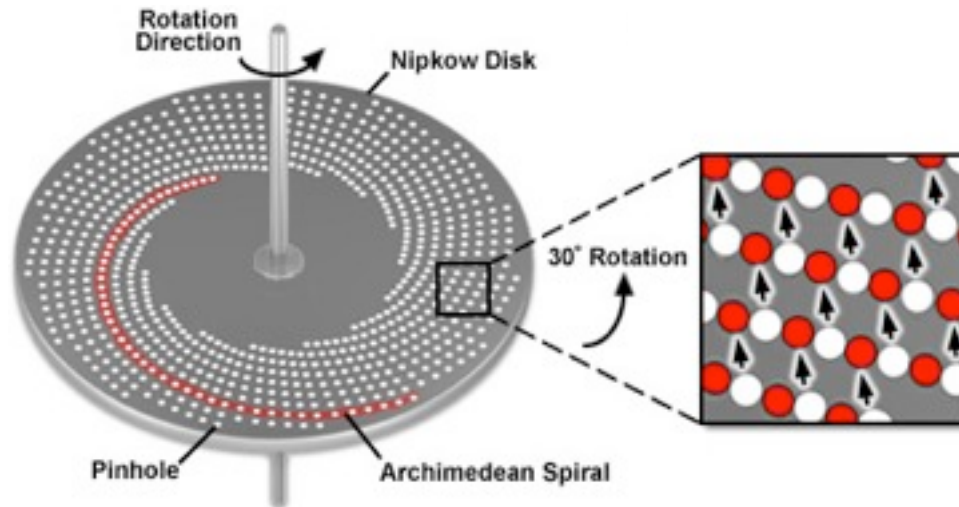
The Nipkow Disk



Constant Battle:

Smaller spacing - more light gets through, but “crosstalk”

The Nipkow Disk



Constant Battle:

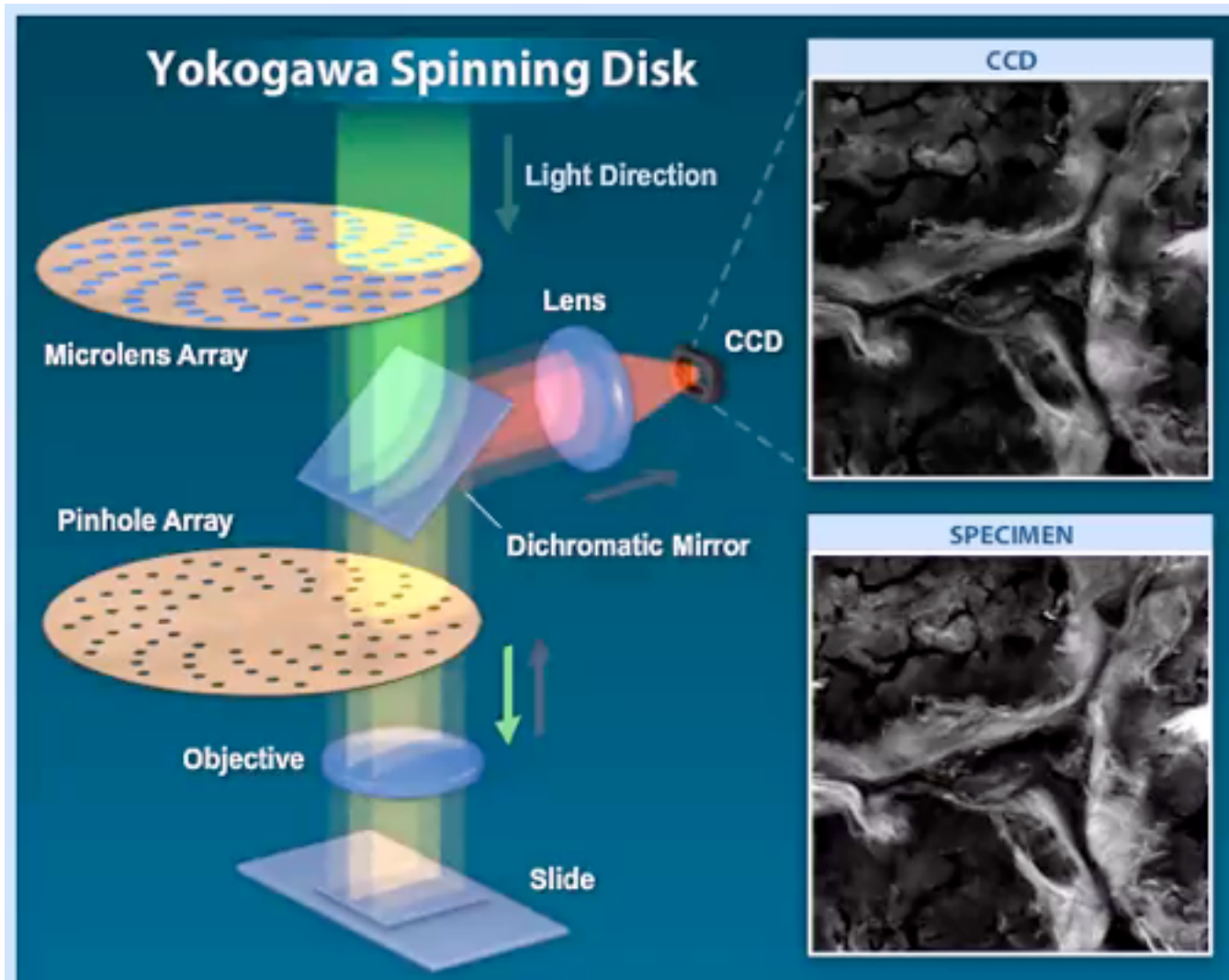
Smaller spacing - more light gets through, but “crosstalk”

Pinhole Spacing Typically = 2.5 μ m apart

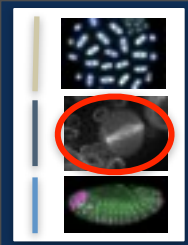
Yokogawa

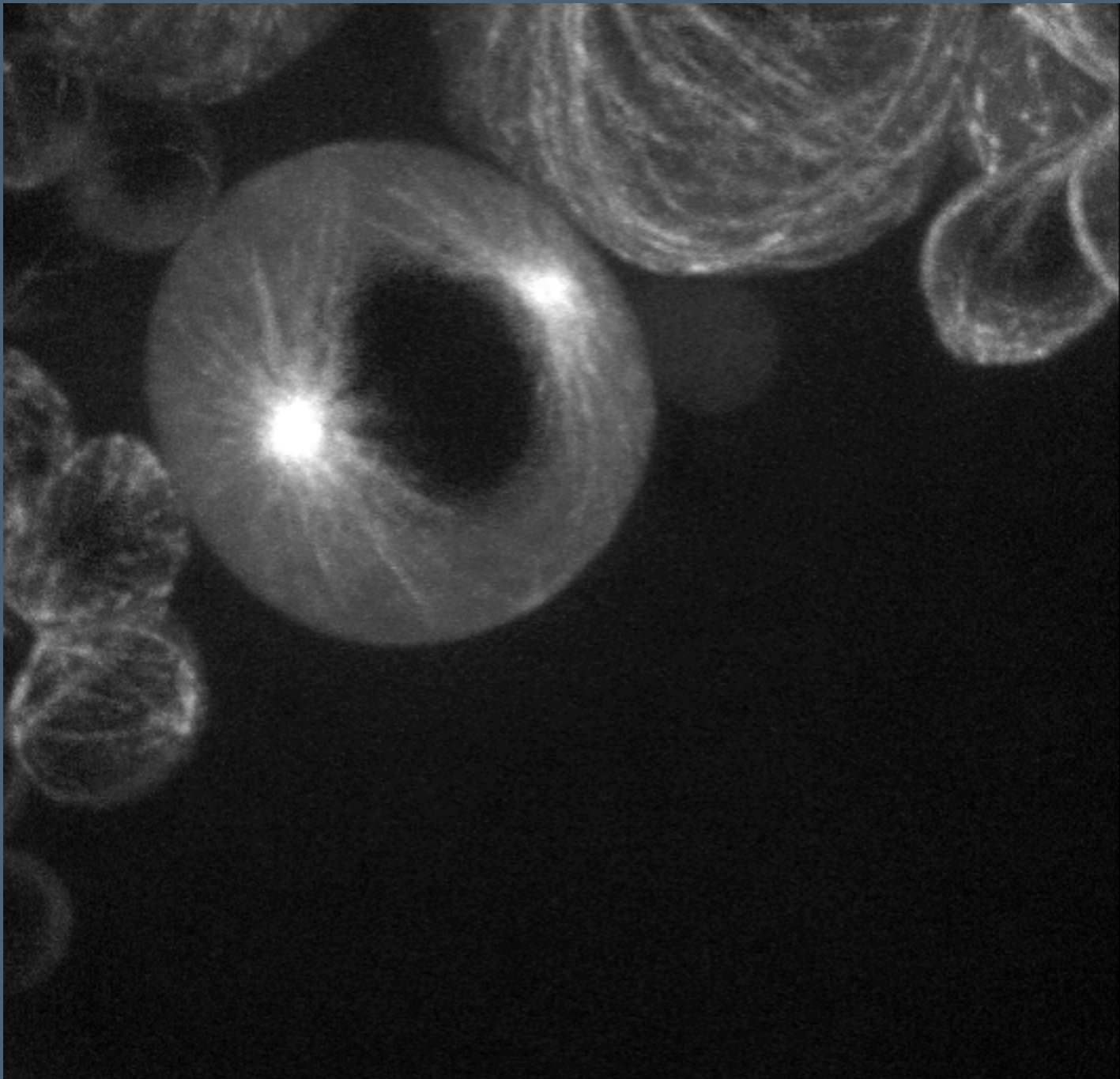


Yokogawa

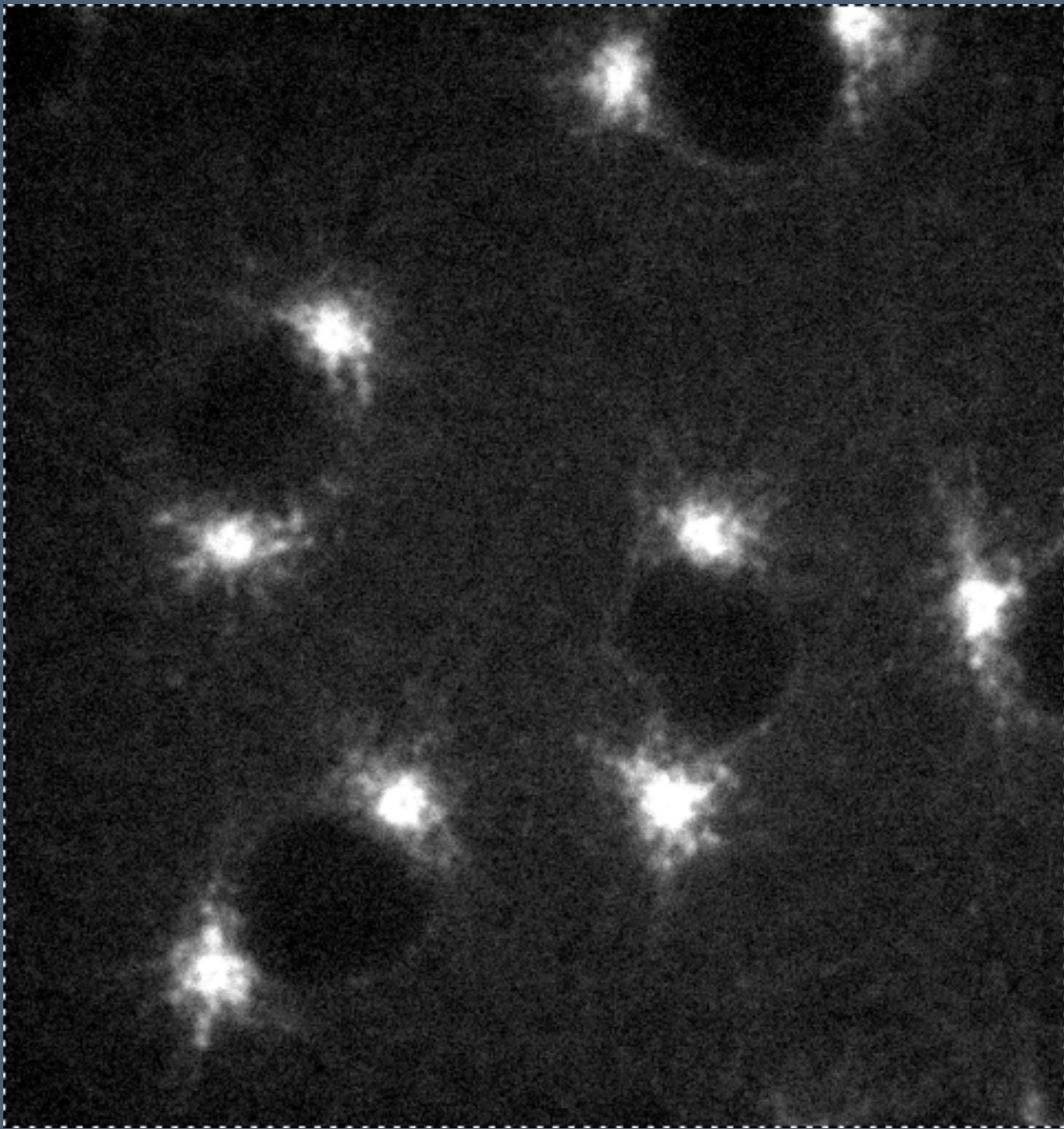


Yokogawa





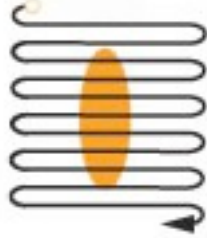
Cell division in brain stem cells (neuroblasts), Raff Lab



MT binding protein in *Drosophila* embryo, Raff Lab

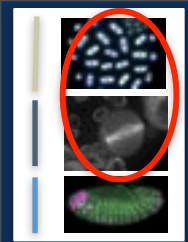
Point Scanning Vs Spinning Disc

Point Scanning



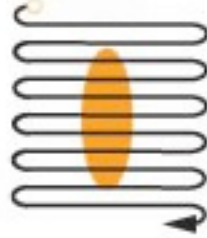
Spinning Disc





Point Scanning Vs Spinning Disc

Point Scanning



Spinning Disc



Speed

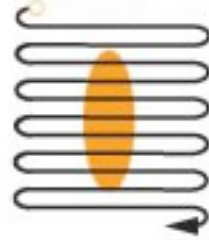
Slow (secs)

Fast (msecs)



Point Scanning Vs Spinning Disc

Point Scanning



Spinning Disc



Speed

Slow (secs)

Fast (msecs)

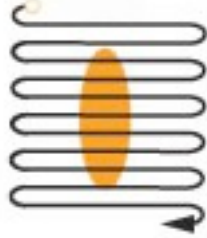
Sensitivity

OK

OK

Point Scanning Vs Spinning Disc

Point Scanning



Spinning Disc



Speed

Slow (secs)

Fast (msecs)

Sensitivity

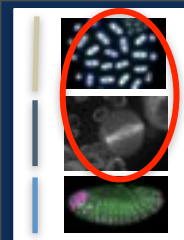
OK

OK

Flexibility

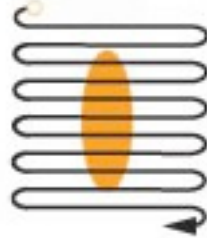
Good

Poor



Point Scanning Vs Spinning Disc

Point Scanning



Spinning Disc



Speed

Slow (secs)

Fast (msecs)

Sensitivity

OK

OK

Flexibility

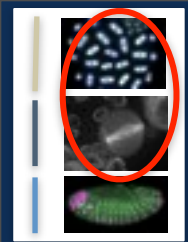
Good

Poor

Bleaching

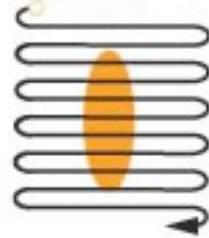
Poor

Good



Point Scanning Vs Spinning Disc

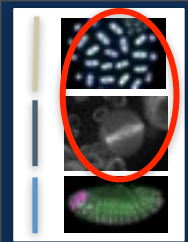
Point Scanning



Spinning Disc

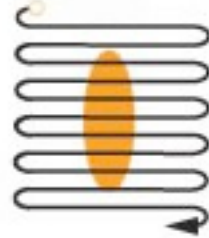


Speed	Slow (secs)	Fast (msecs)
Sensitivity	OK	OK
Flexibility	Good	Poor
Bleaching	Poor	Good
Pretty Pictures	Unbeatable!	Pretty damn good!



Point Scanning Vs Spinning Disc

Point Scanning

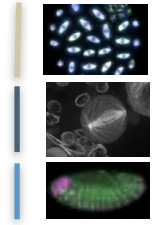


Spinning Disc



Speed	Slow (secs)	Fast (msecs)
Sensitivity	OK	OK
Flexibility	Good	Poor
Bleaching	Poor	Good
Pretty Pictures	Unbeatable!	Pretty damn good!
Pretty Movies	Good – if process slow	Unbeatable!

3 Flavours of Microscope

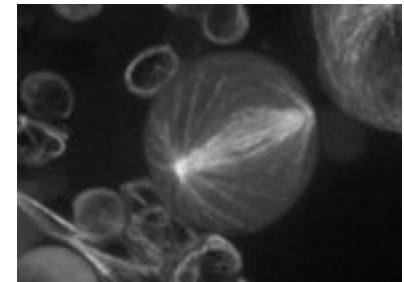
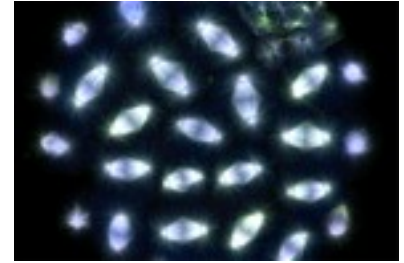


Problem:
Out of Focus
Light

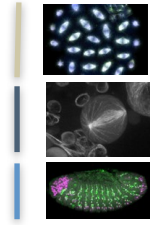


Laser
Scanning

Spinning disc



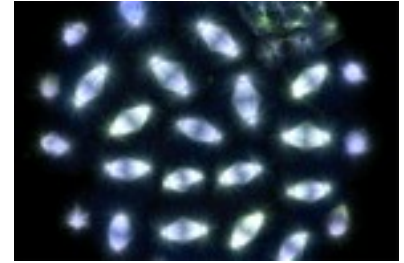
3 Flavours of Microscope



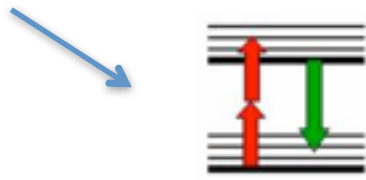
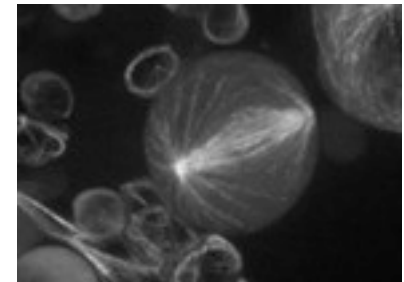
Problem:
Out of Focus
Light



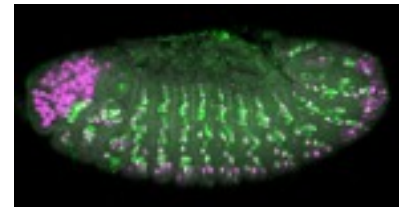
Laser
Scanning



Spinning disc



2-Photon



2-photon Microscope



Not a 'confocal'

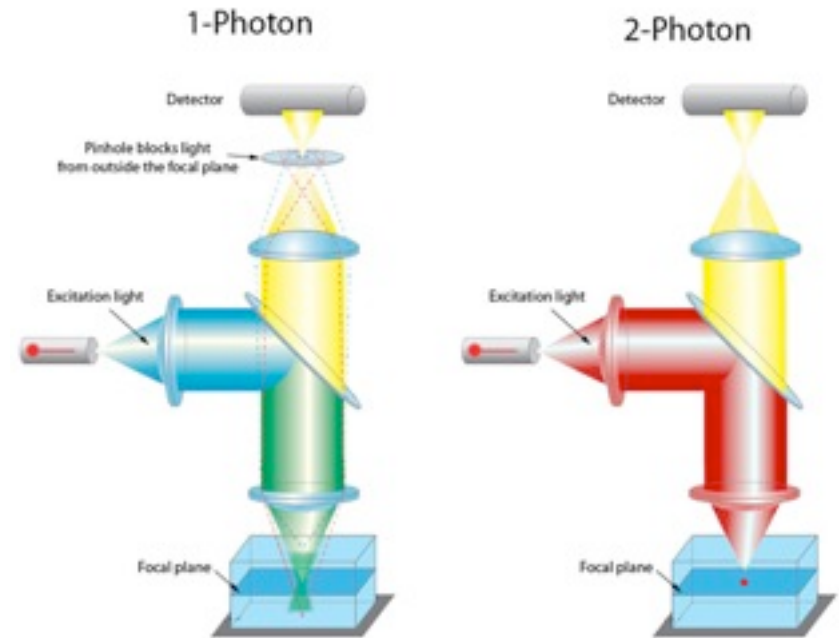
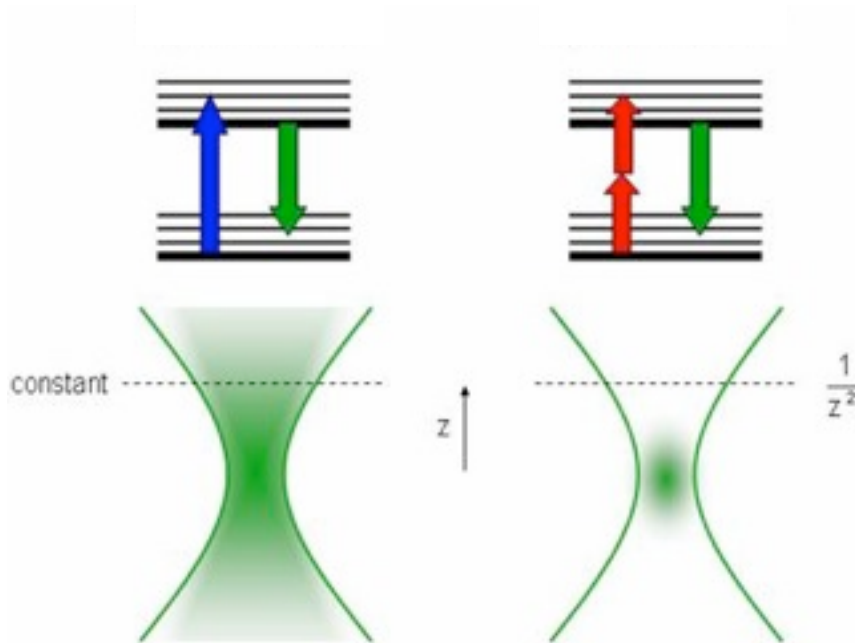
for imaging deeper into thick specimens

less damaging to biological samples

Confocal Vs 2-photon

1 Photon Excitation

2 Photon Excitation

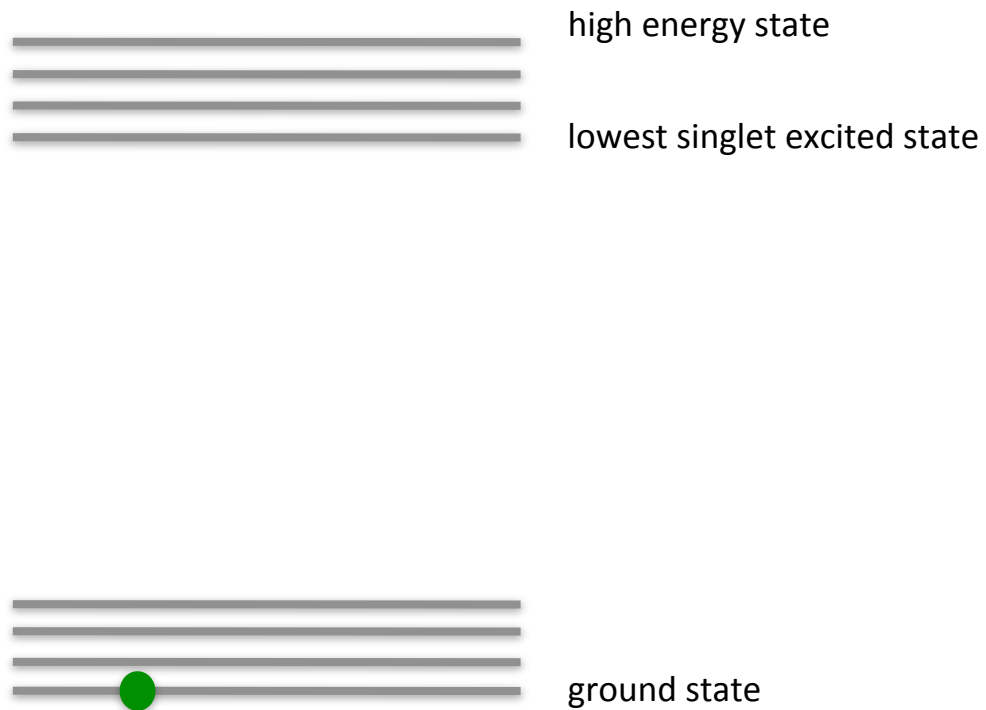


There is no out of focus light



THEORY

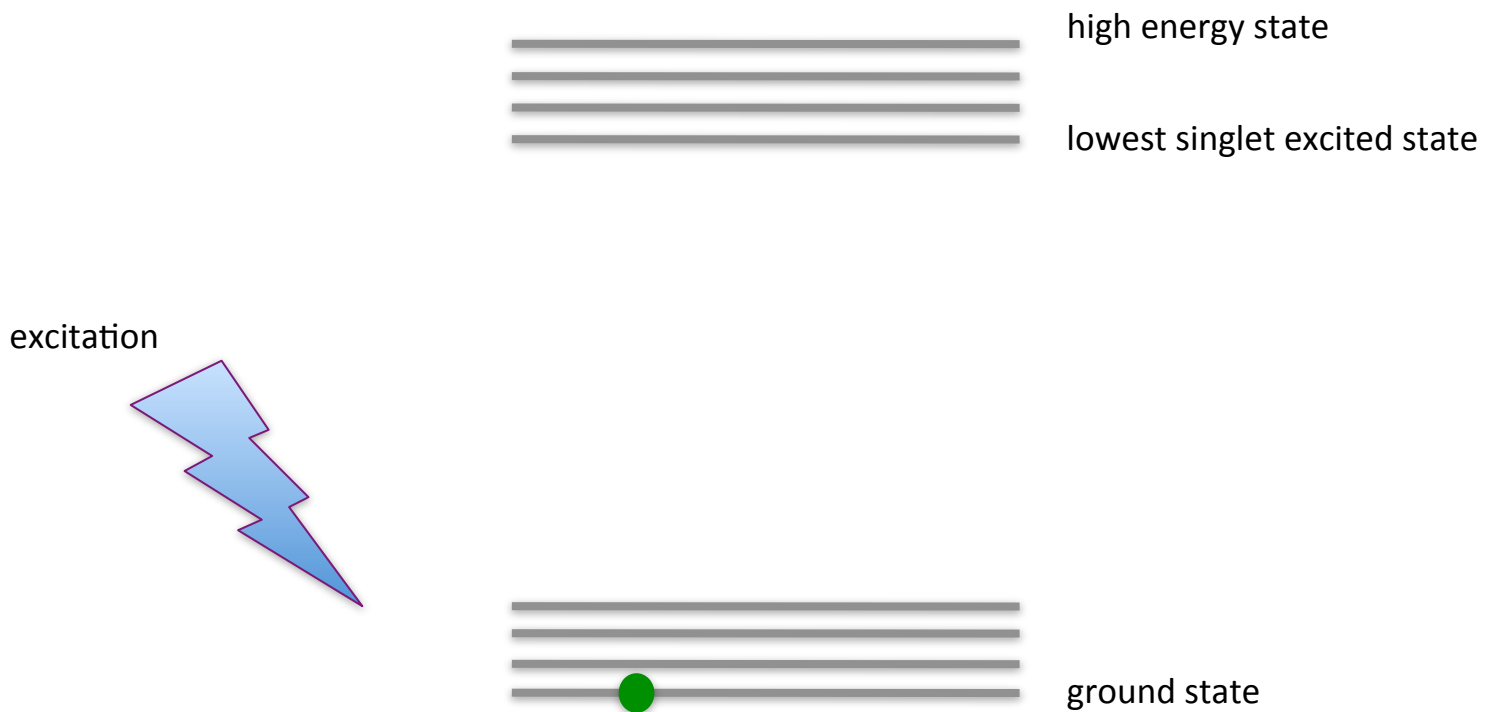
1 Photon Excitation





THEORY

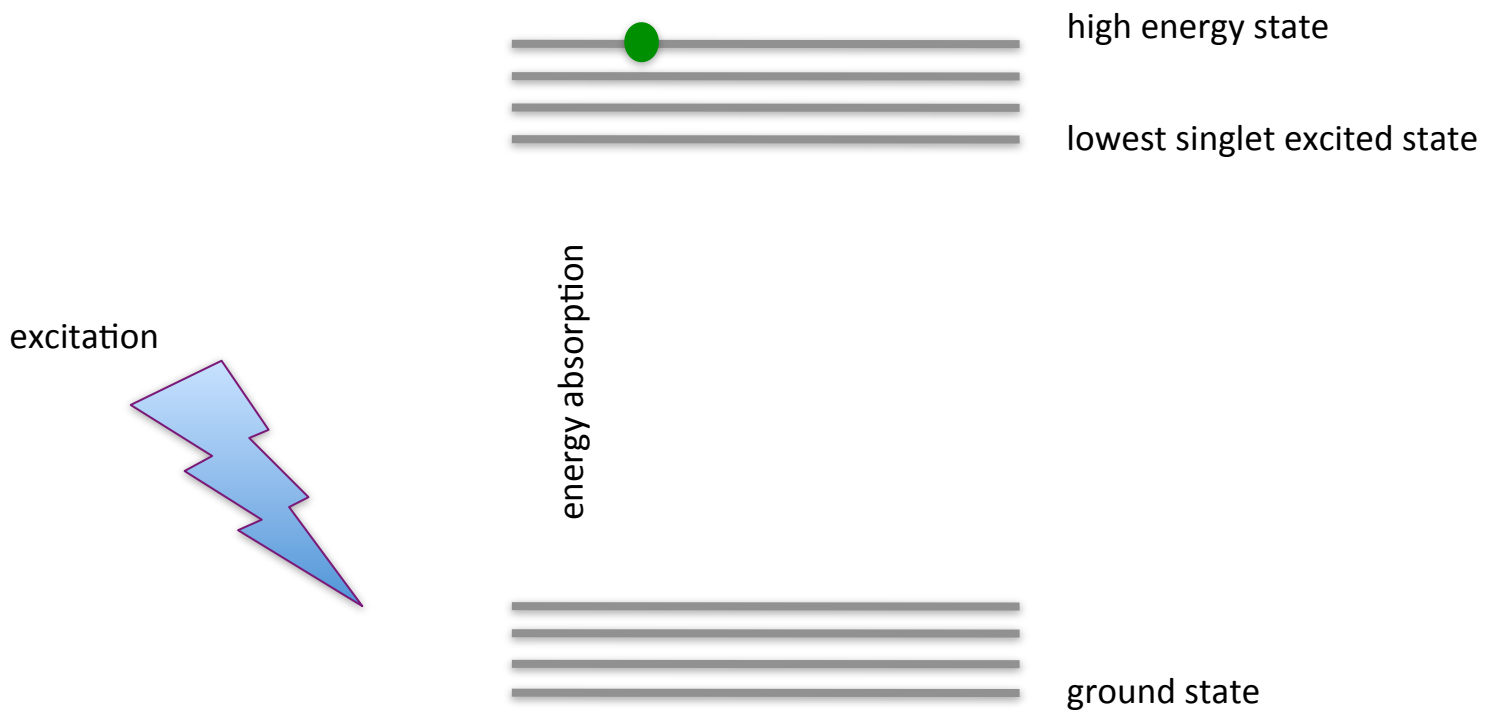
1 Photon Excitation





THEORY

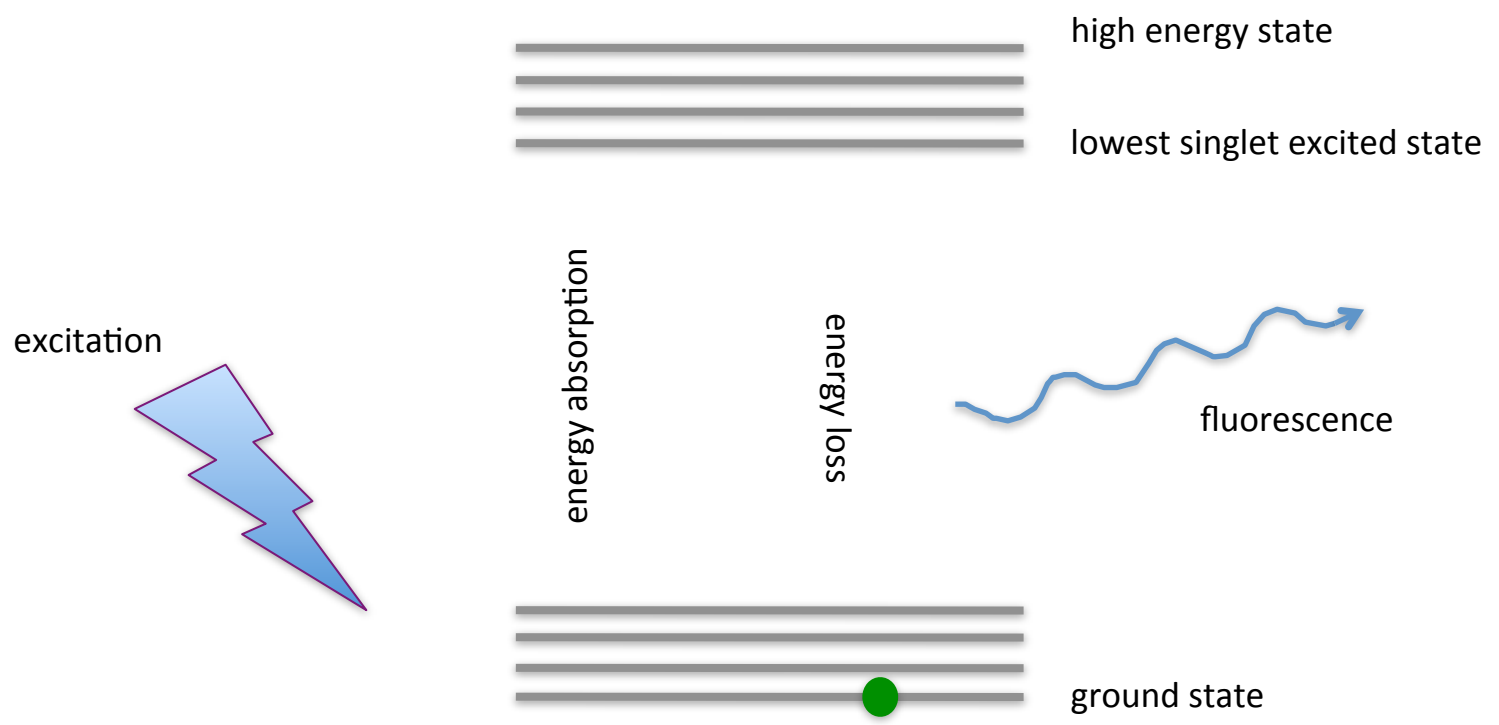
1 Photon Excitation





THEORY

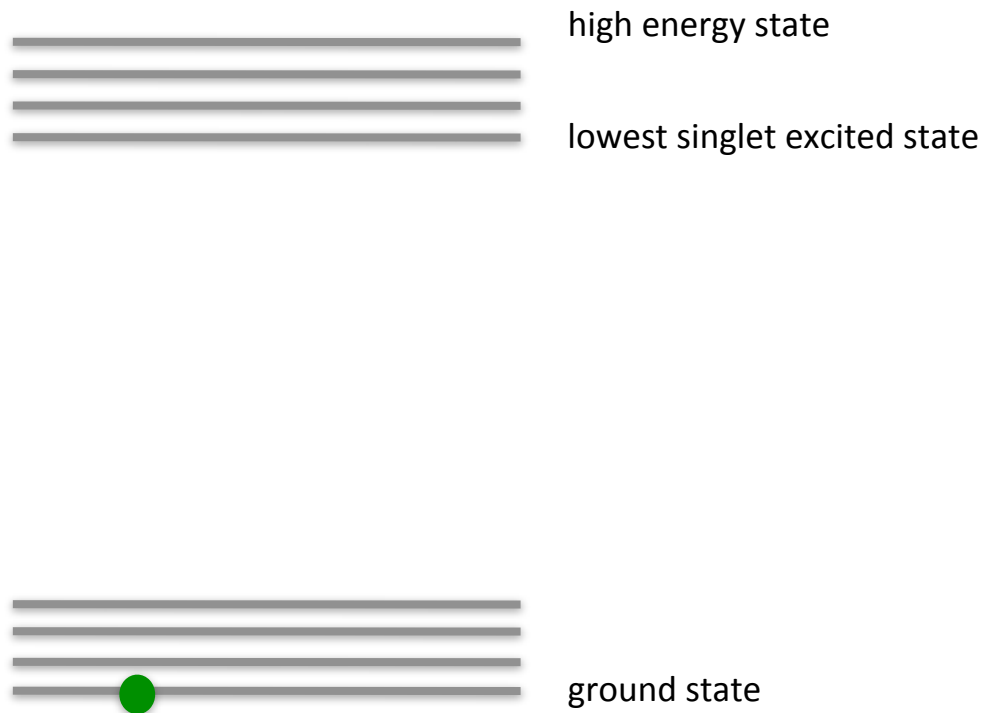
1 Photon Excitation





THEORY

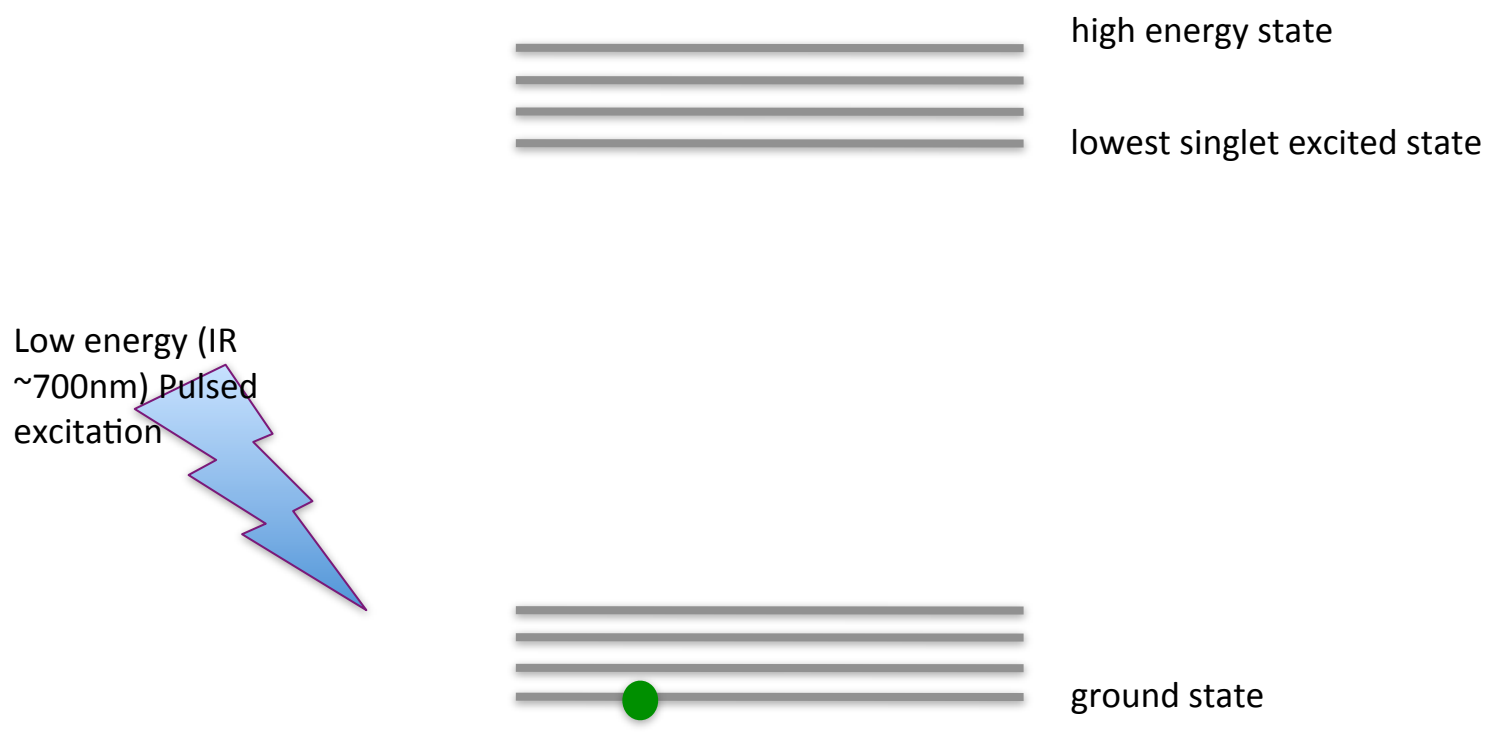
2 Photon Excitation





THEORY

2 Photon Excitation





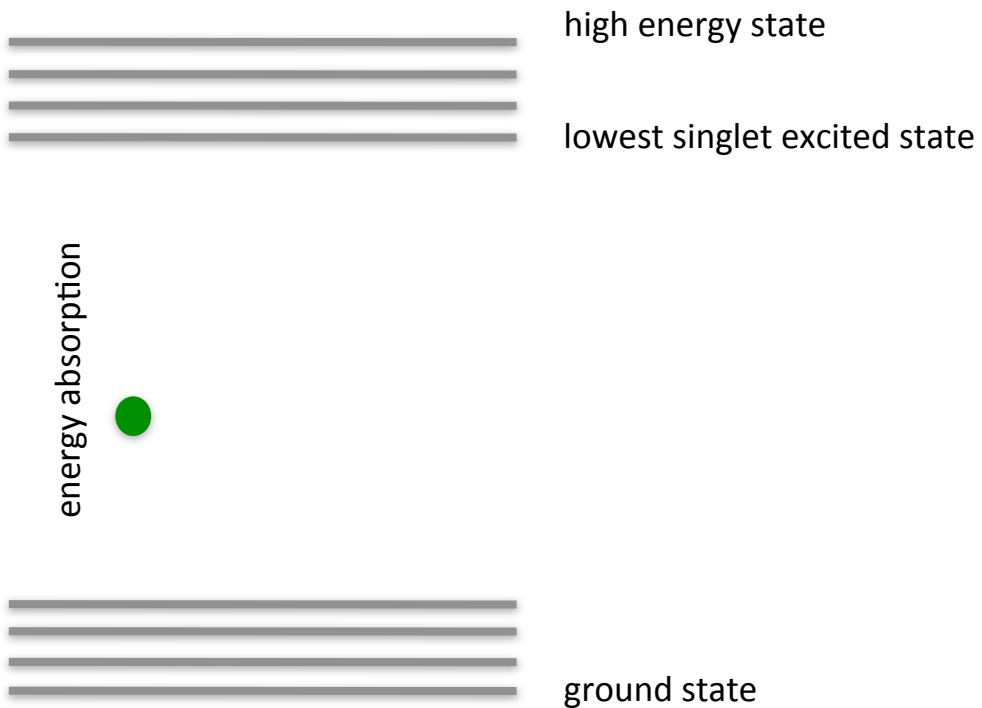
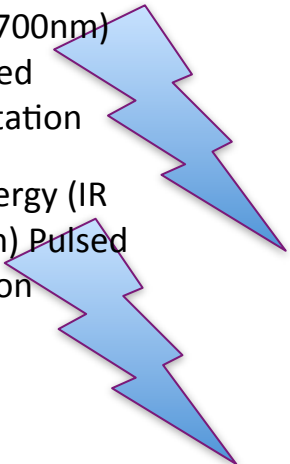
THEORY

2 Photon Excitation

Almost simultaneous

2nd low energy
(IR~700nm)
pulsed
excitation

Low energy (IR
~700nm) Pulsed
excitation





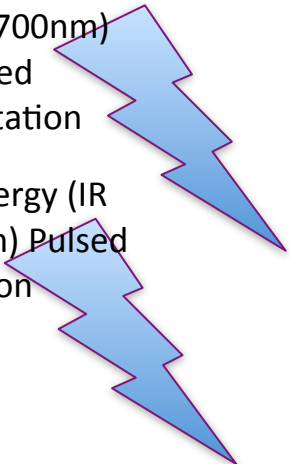
THEORY

2 Photon Excitation

Almost simultaneous

2nd low energy
(IR~700nm)
pulsed
excitation

Low energy (IR
~700nm) Pulsed
excitation



high energy state

lowest singlet excited state

energy absorption



ground state



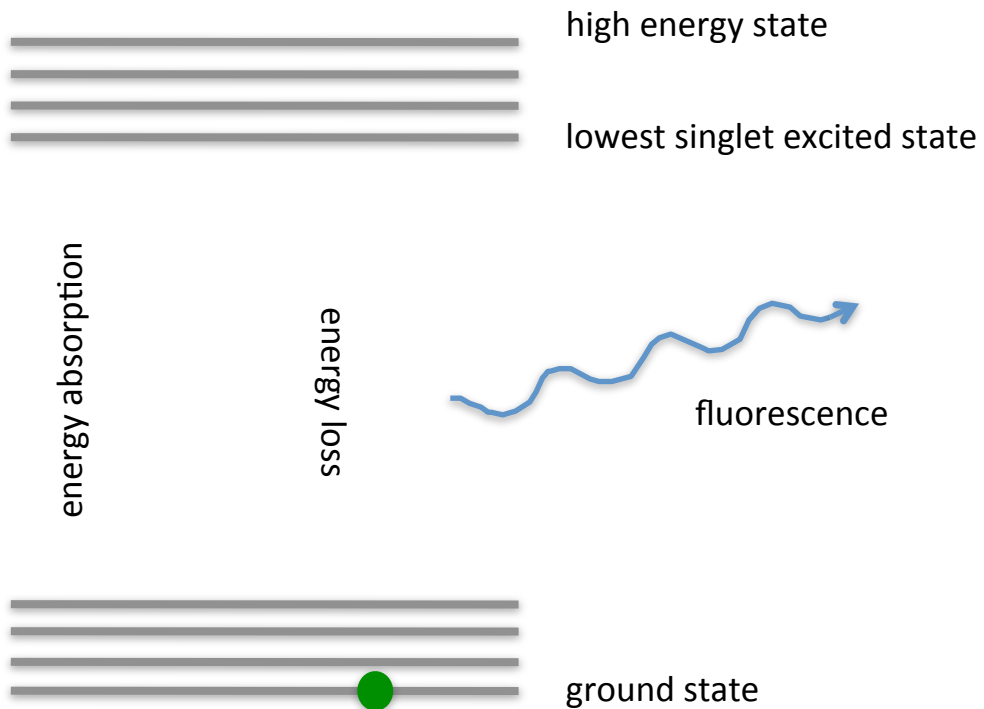
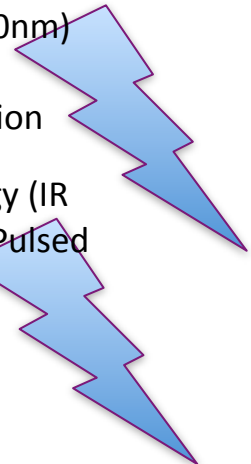
THEORY

2 Photon Excitation

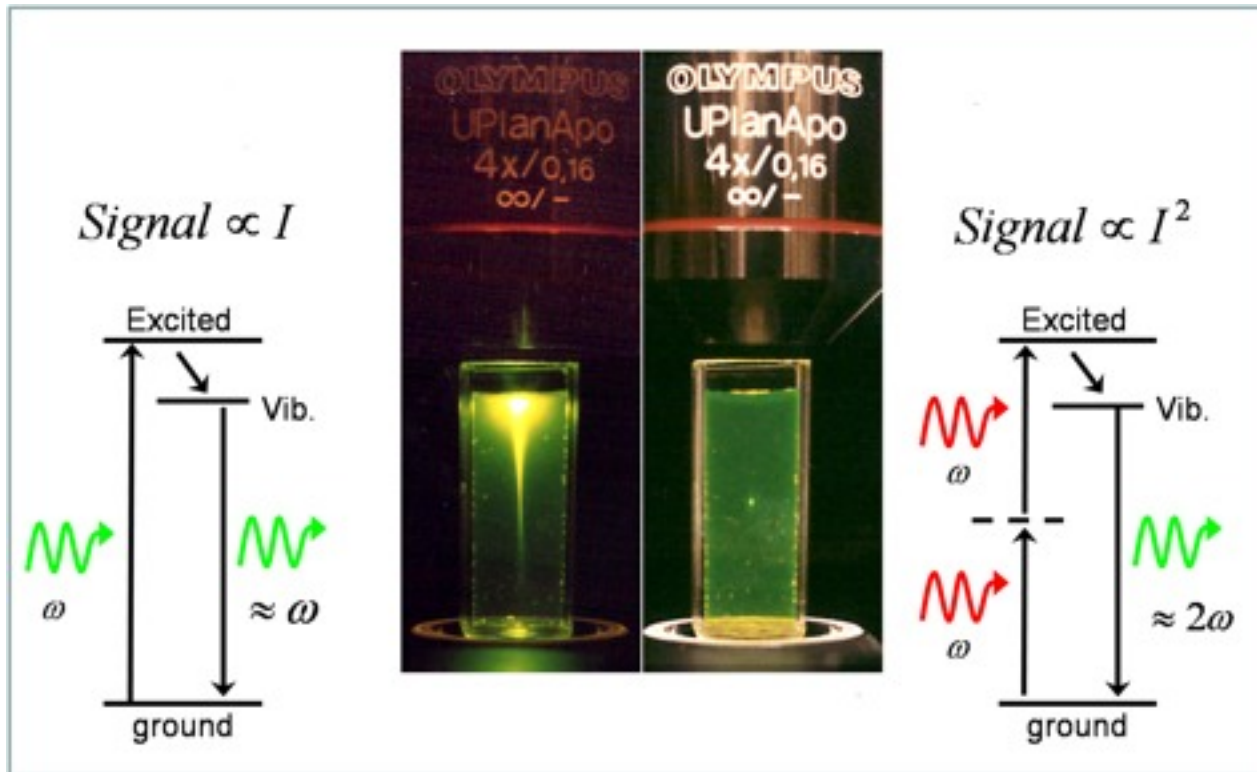
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2nd low energy
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Low energy (IR
~700nm) Pulsed
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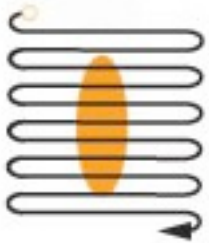
Principle of 2-photon Microscope



Near simultaneous, two photon event highly unlikely, only really possible a focal point

Tightly focused excitation

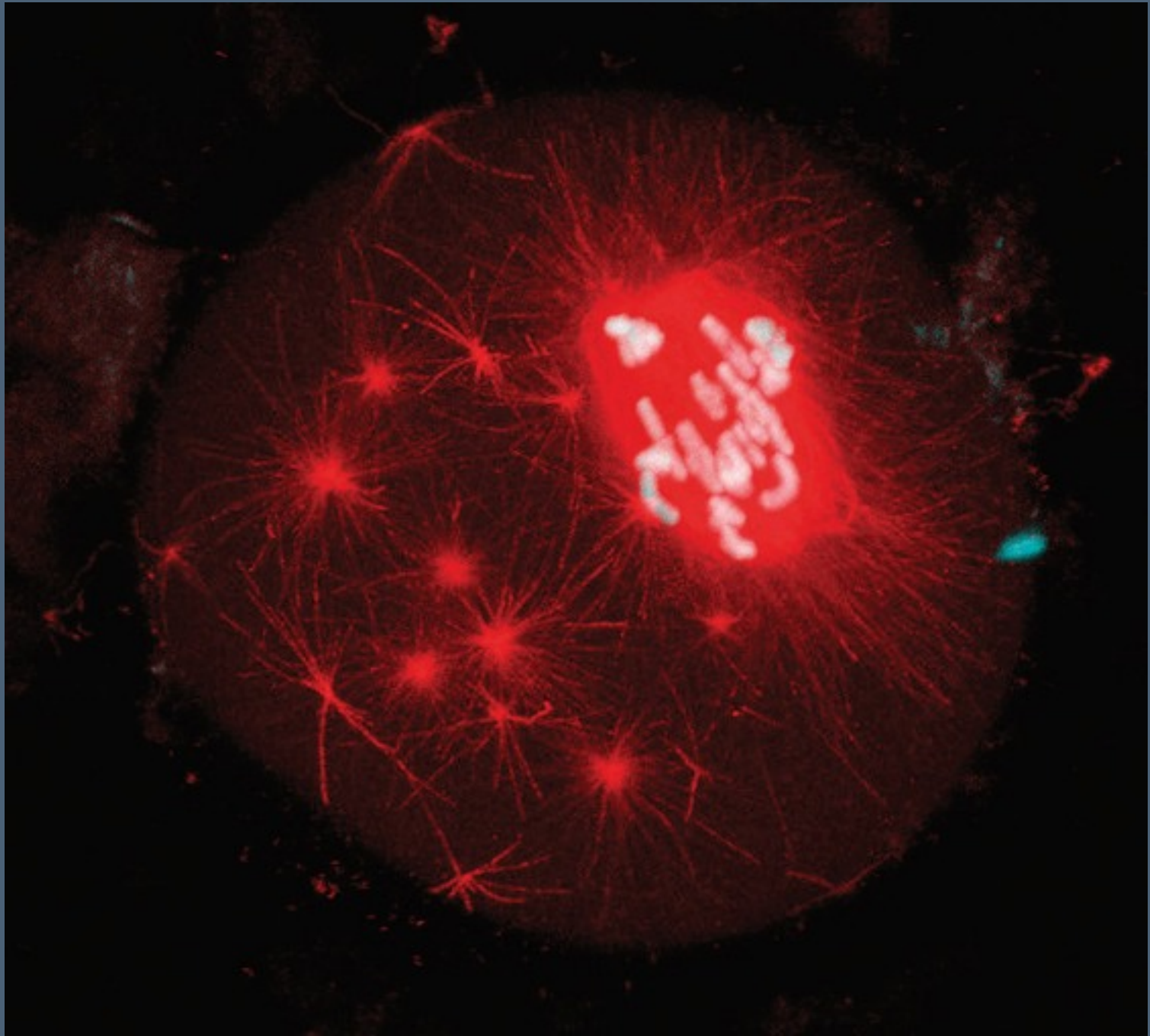
2-photon Microscope



Pulsed excitation laser is then scanned across the sample.

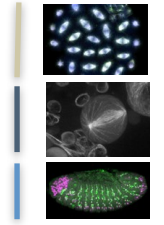
Longer wavelengths are scattered to a lesser degree than shorter ones, and penetrate deeper into the sample.

In addition, these lower-energy photons are less likely to cause damage outside the focal volume.



Spindle formation in mouse oocyte, labelled with Hoechst, Alexa 680. M Schuh. EMBL, Heidelberg, Germany

3 Flavours of Microscope



Problem:
Out of Focus
Light

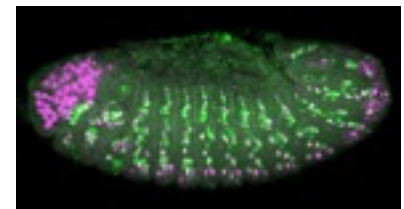
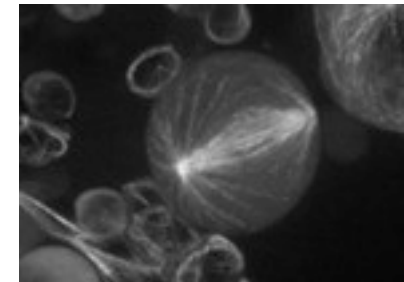
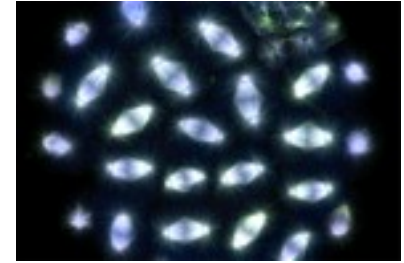


Confocal

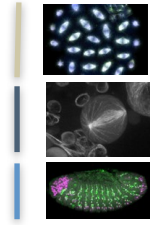
Laser
Scanning

Spinning disc

2-Photon



3 Flavours of Microscope

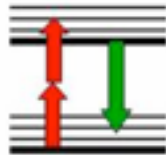
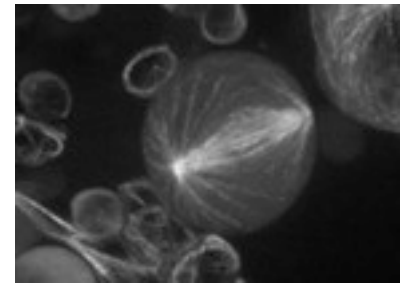
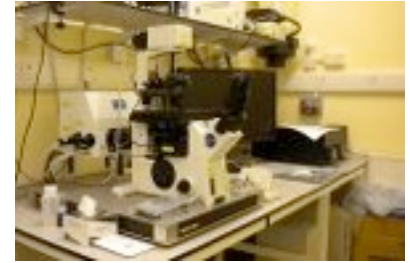


Problem:
Out of Focus
Light

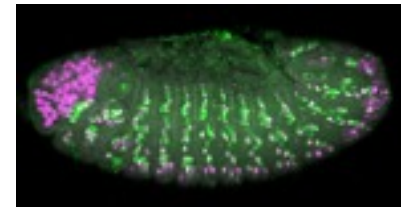


Laser
Scanning

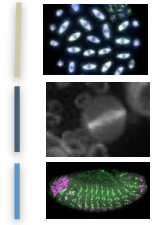
Spinning disc



2-Photon



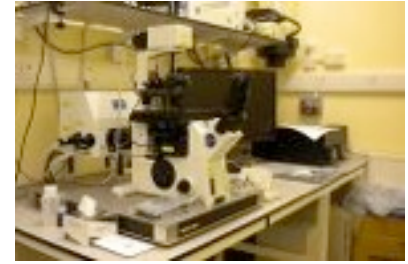
3 Flavours of Microscope



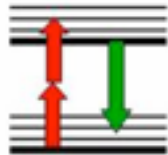
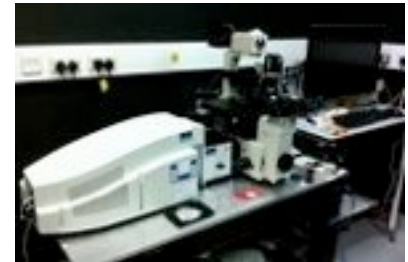
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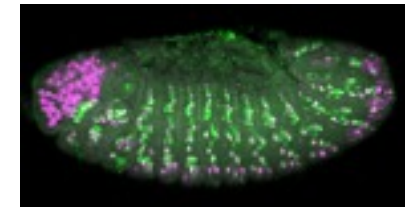
Laser
Scanning



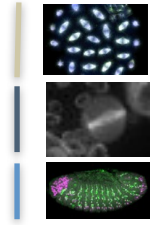
Spinning disc



2-Photon



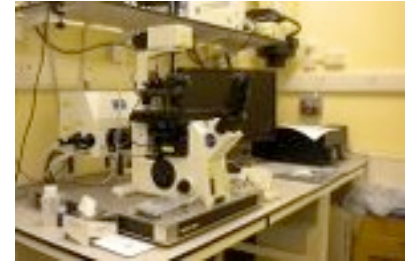
3 Flavours of Microscope



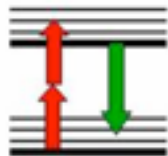
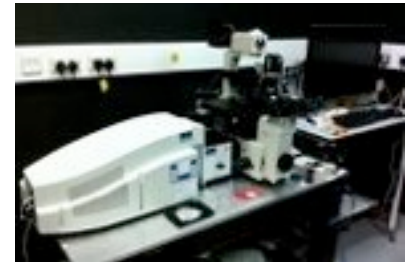
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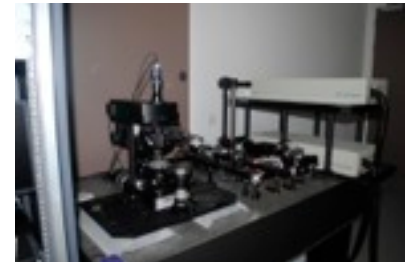
Laser
Scanning



Spinning disc



2-Photon





<http://www.micron.ox.ac.uk>

