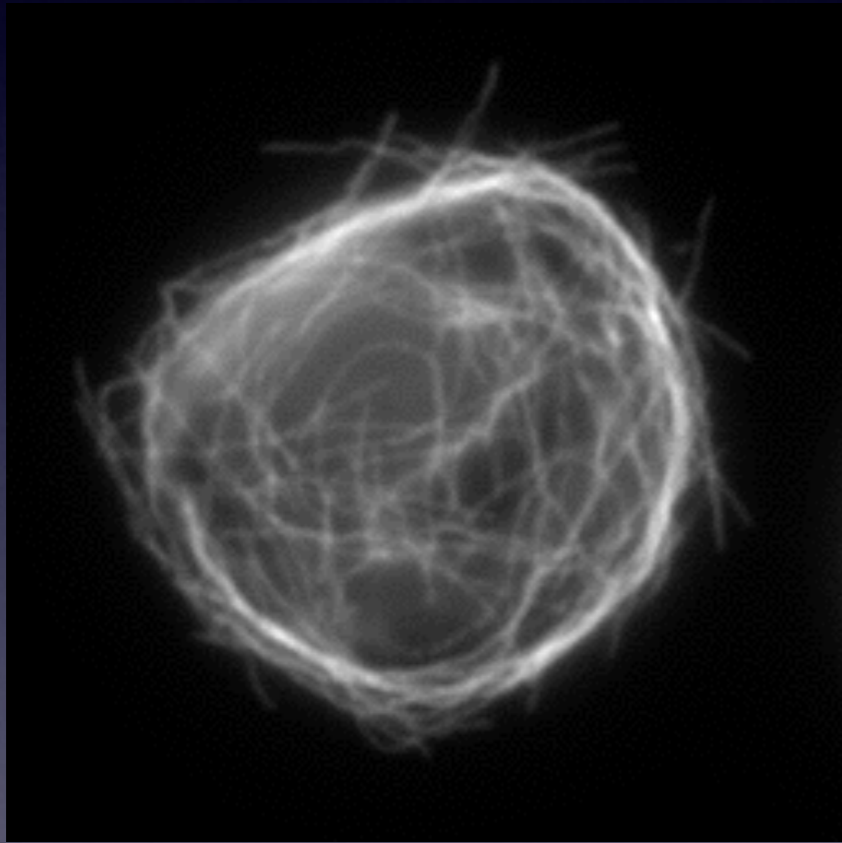
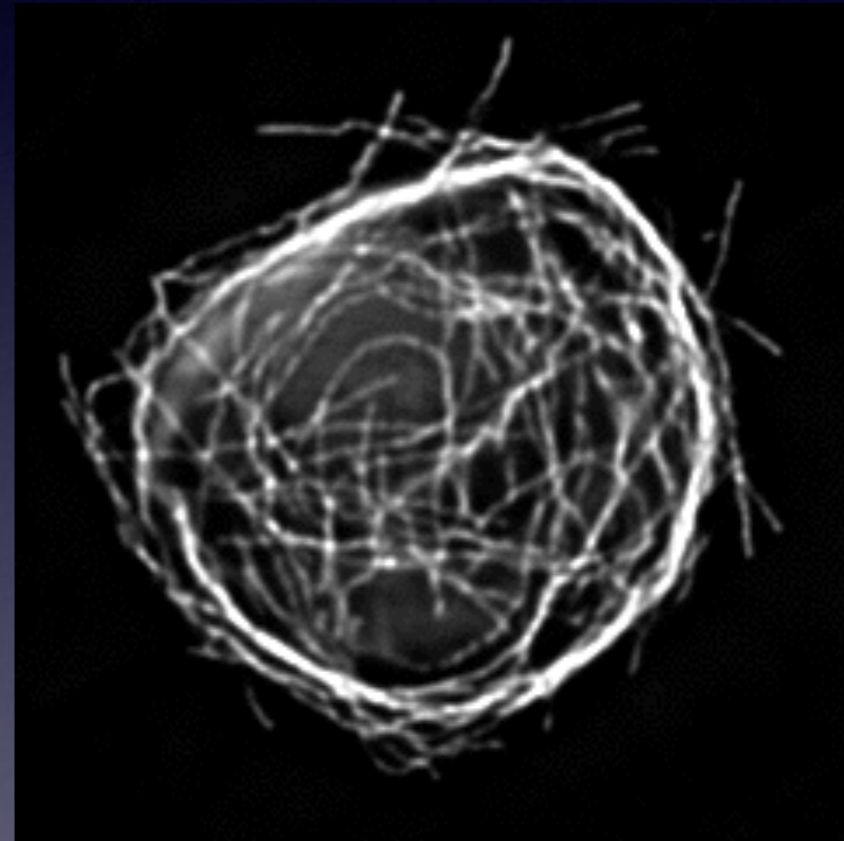


# Lecture 9

## Advanced Widefield Microscopy & Bespoke Microscopes



Widefield



Deconvolved Widefield

# Lecture 9

## Advanced Widefield Microscopy & Bespoke Microscopes

Ian Dobbie  
x13323

# Overview

- Image formation and airy rings
- Beads and spherical aberration
- How deconvolution works
- Super fast acquisition
- Bespoke microscope design - pro's and cons

# What is a microscope image

- The microscope produces a magnified, but also distorted, image
- Record the light intensity on a camera.

# Microscopic imaging in mathematical terms.

- Take your sample
- Multiply it at every point by the imaging process in the microscope (convolve the PSF with the object).
- Produce the image.

# The most important things to think about.

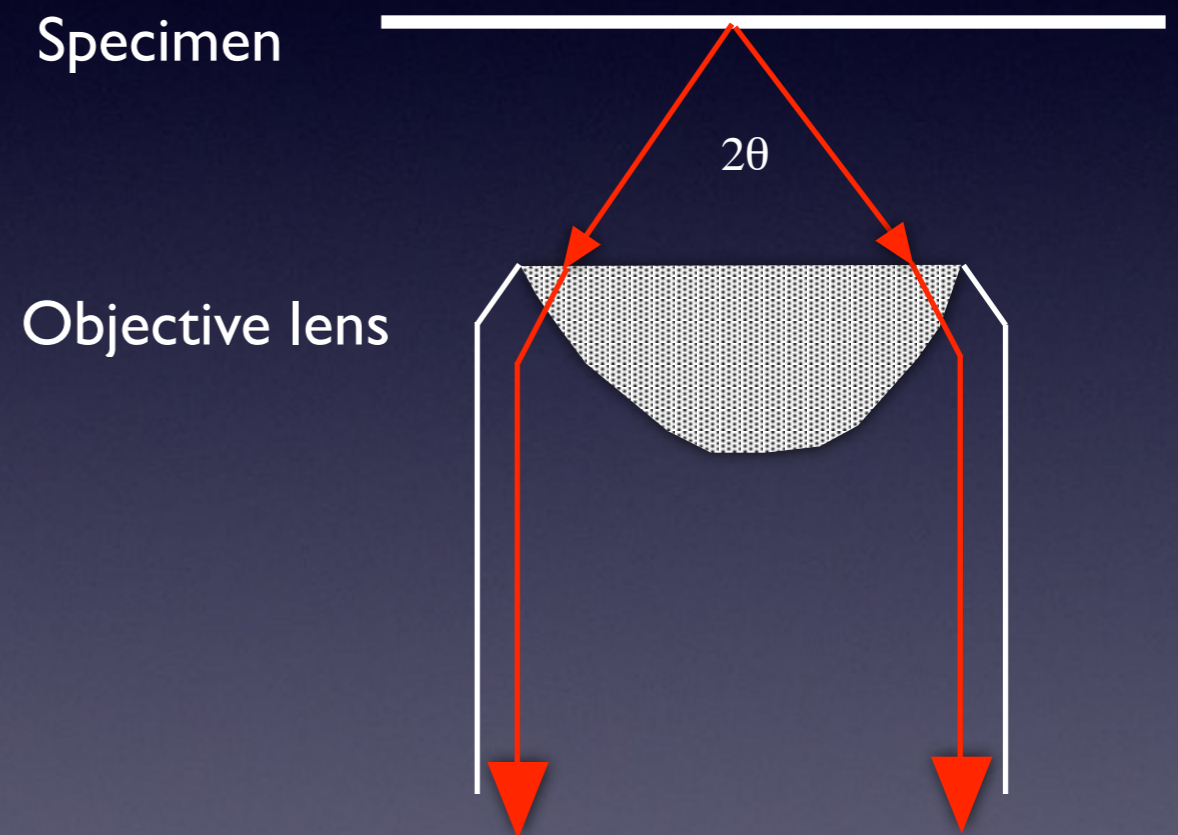
Contrast :- What is the difference between what you want to see and everything else?

Resolution :- How small things can you see?

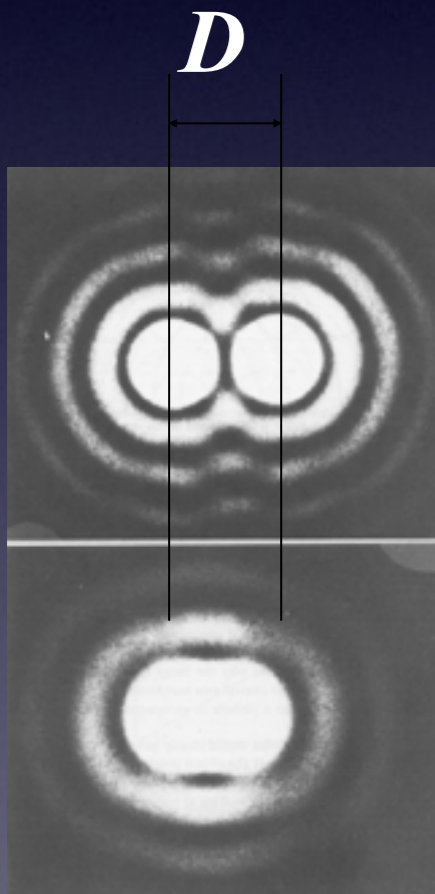
Nothing else

# Microscope Resolution

- No lens has perfect resolution, even in theory
- Resolution depends on the angle ( $\theta$ ) of the cone of light that the objective can collect from the specimen.
- Rule of thumb:  
Resolution limit  $\sim \lambda/2$



# Resolution: A technical definition, the Rayleigh Criterion



$D$ , the distance of two  
closest points that can be  
distinguished

$$D = 1.22 \lambda / (NA_{\text{obj}} + NA_{\text{cond}})$$

Epi-Fluorescence:  $NA_{\text{cond}} = NA_{\text{obj}}$   
so  $D = 1.22 \lambda / 2NA$

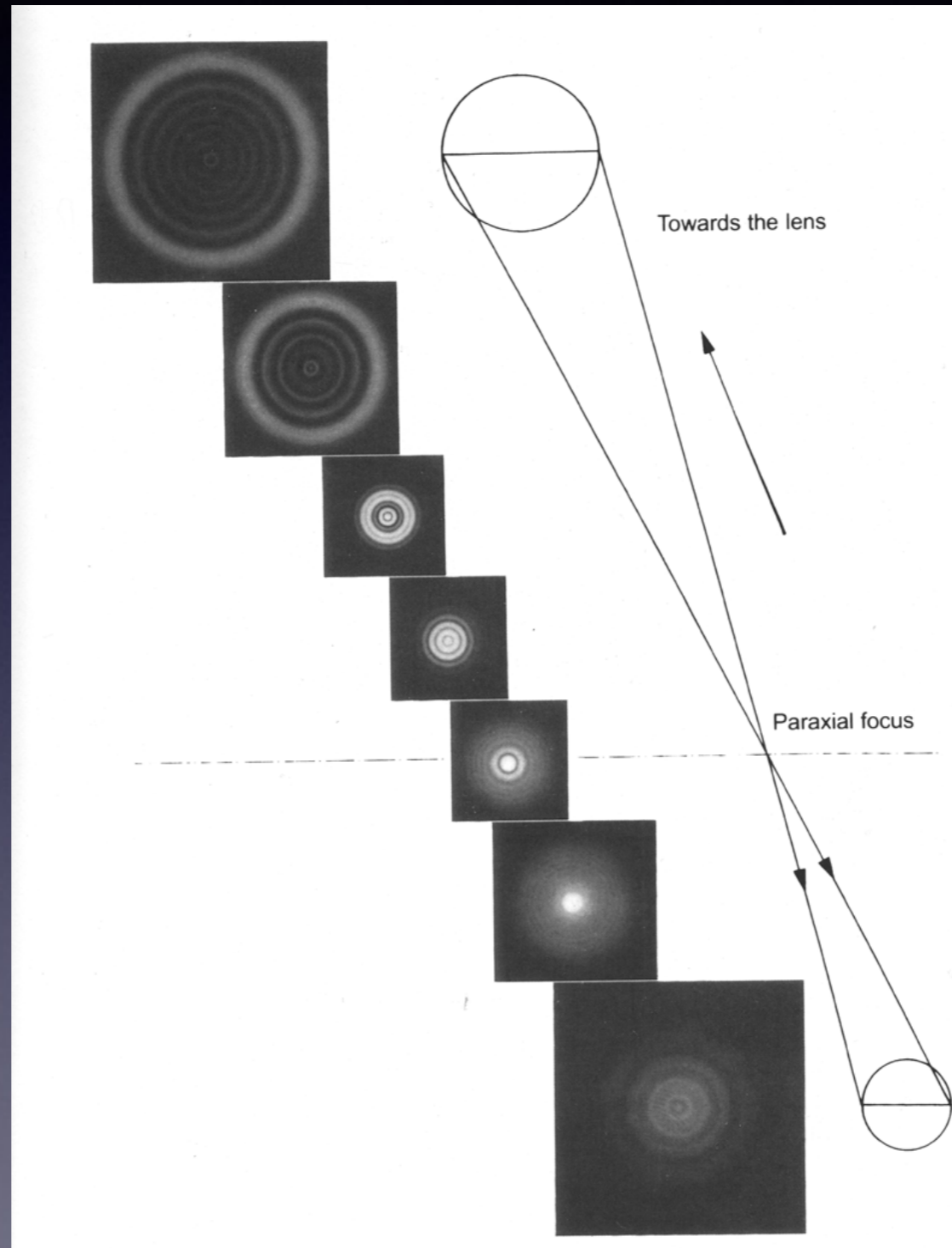
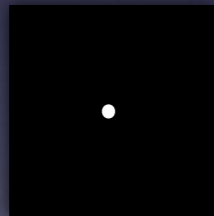


# The Point Spread Function - PSF

- The image of an infinitely small point.
- Limited by resolution
- 3D structure also very important.

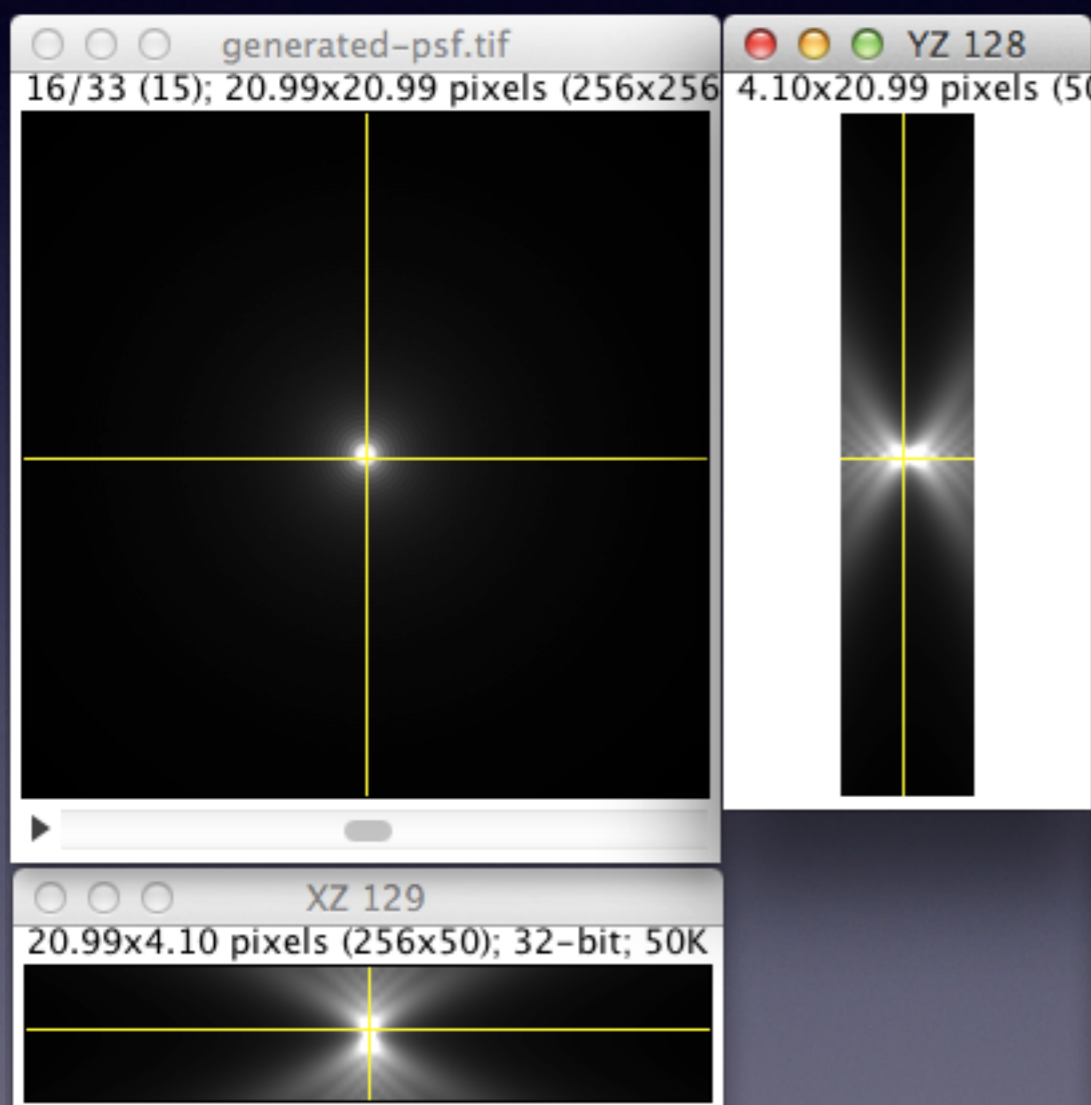
# Image quality- the problem of "out-of-focus light" point spread function and airy rings

Sample object: a "sub-resolution"  
fluorescent bead

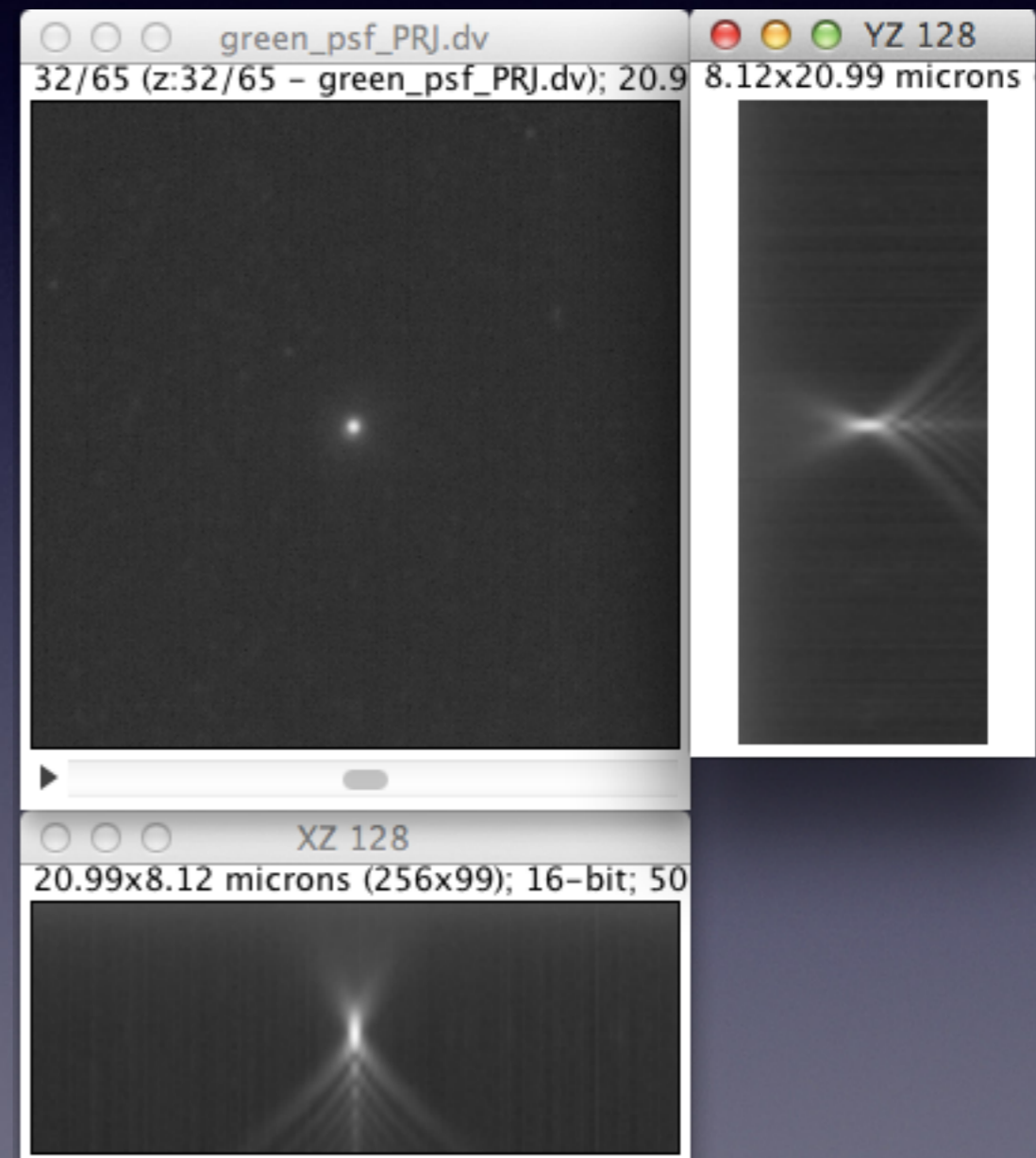


# Theoretical and measured PSF

## Orthogonal views

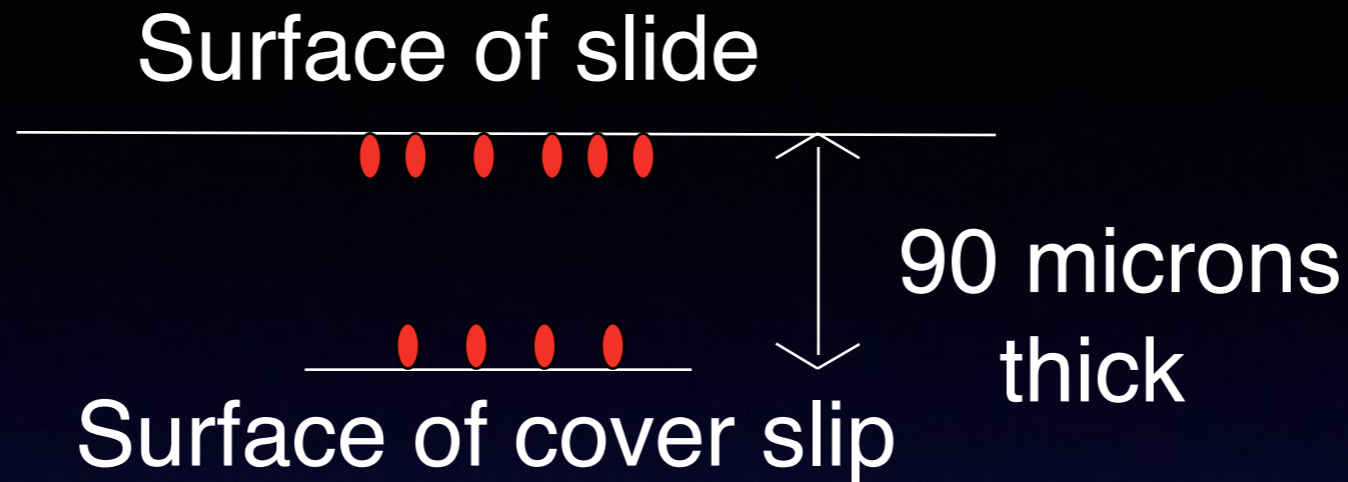


Generated PSF



Real PSF

# Bead slide



**Tetraspeck beads:** chromatic registration  
DAPI/FITC/Rhodamine/Cy5

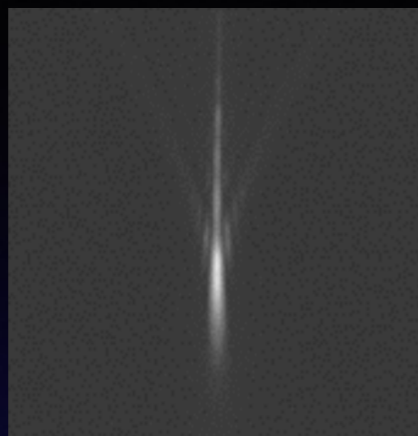
**Beads (PS Spec):** Single fluorochrome  
Brighter -better for generating  
point spread functions for deconvolution

**Inspeck Intensity beads:** Measure dynamic range

# Affects of deep imaging ( $90\mu\text{m}$ ) and collar settings on spherical aberration and psf of 60X/NA1.2<sub>w</sub>



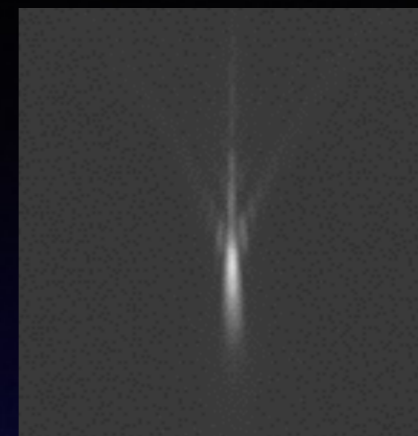
0.13 surf



0.13 deep



0.15 surf



0.15 deep



0.17 surf



0.17 deep

Data from  
Alejandra Clark



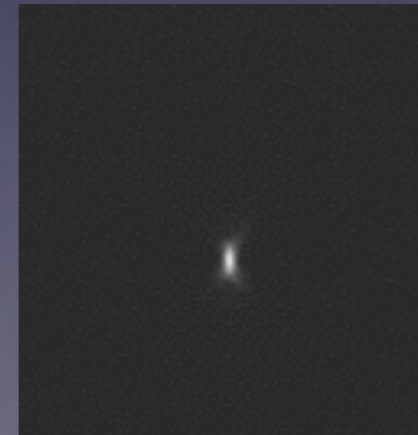
0.19 surf



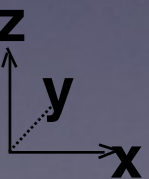
0.19 deep



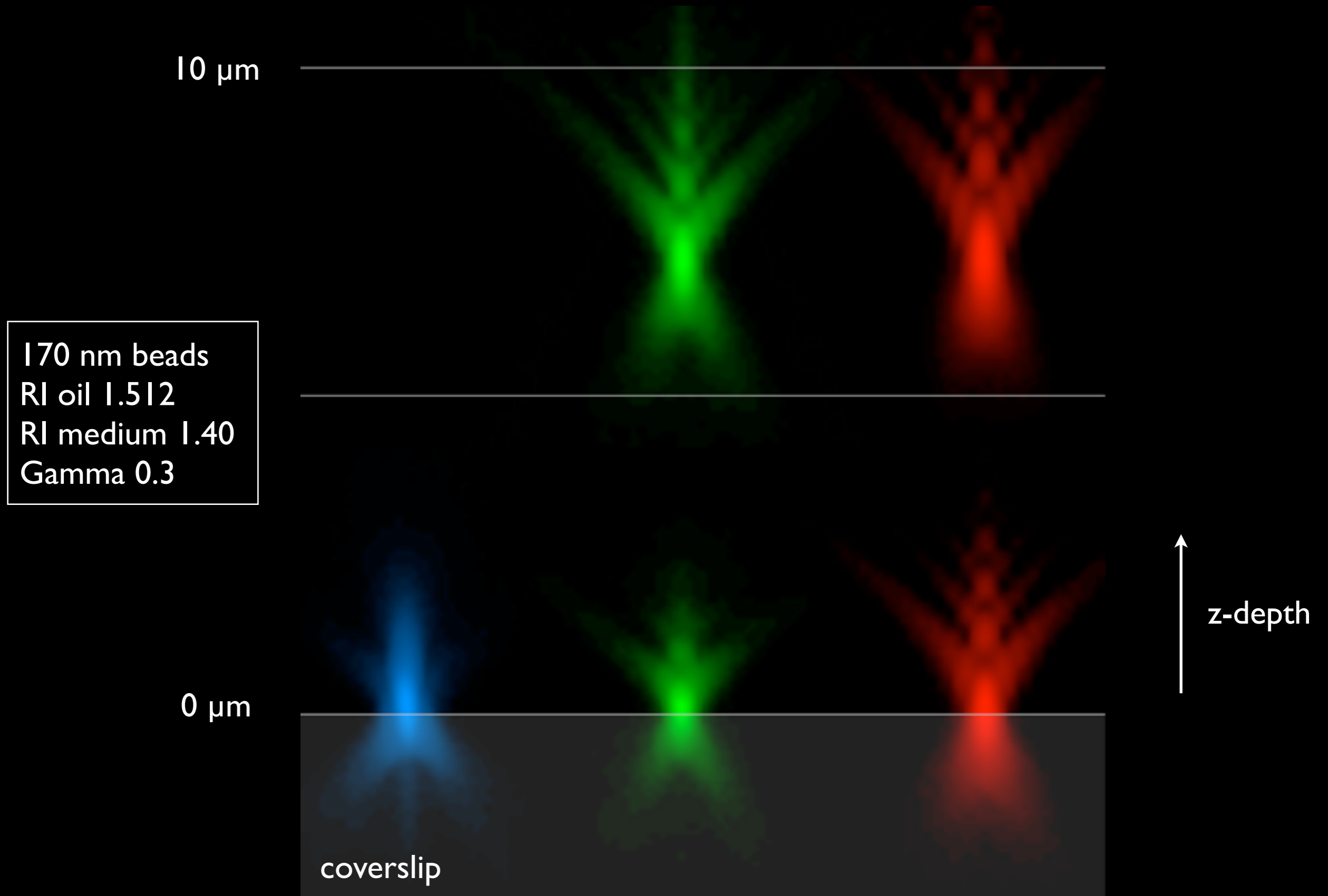
0.21 surf



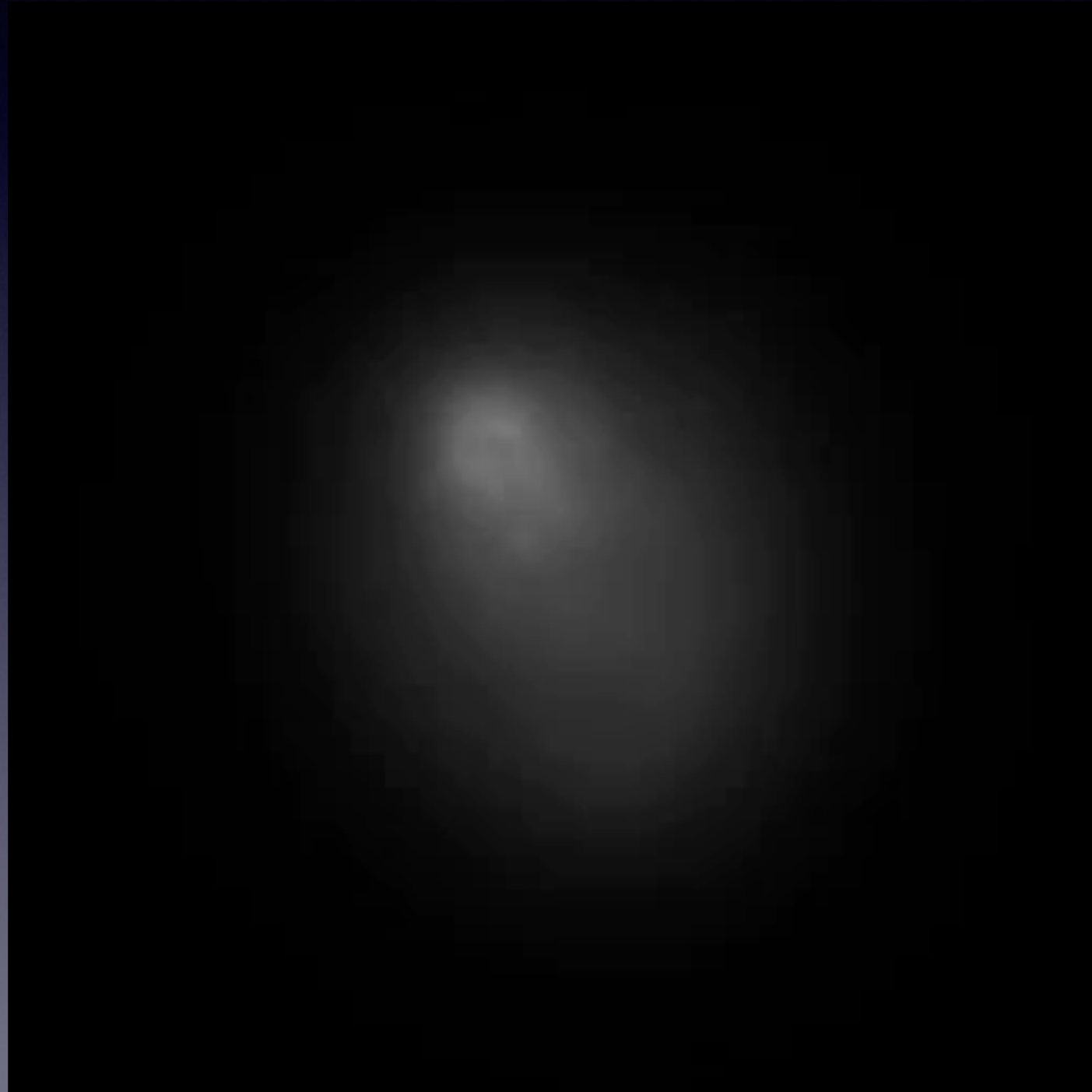
0.21 deep



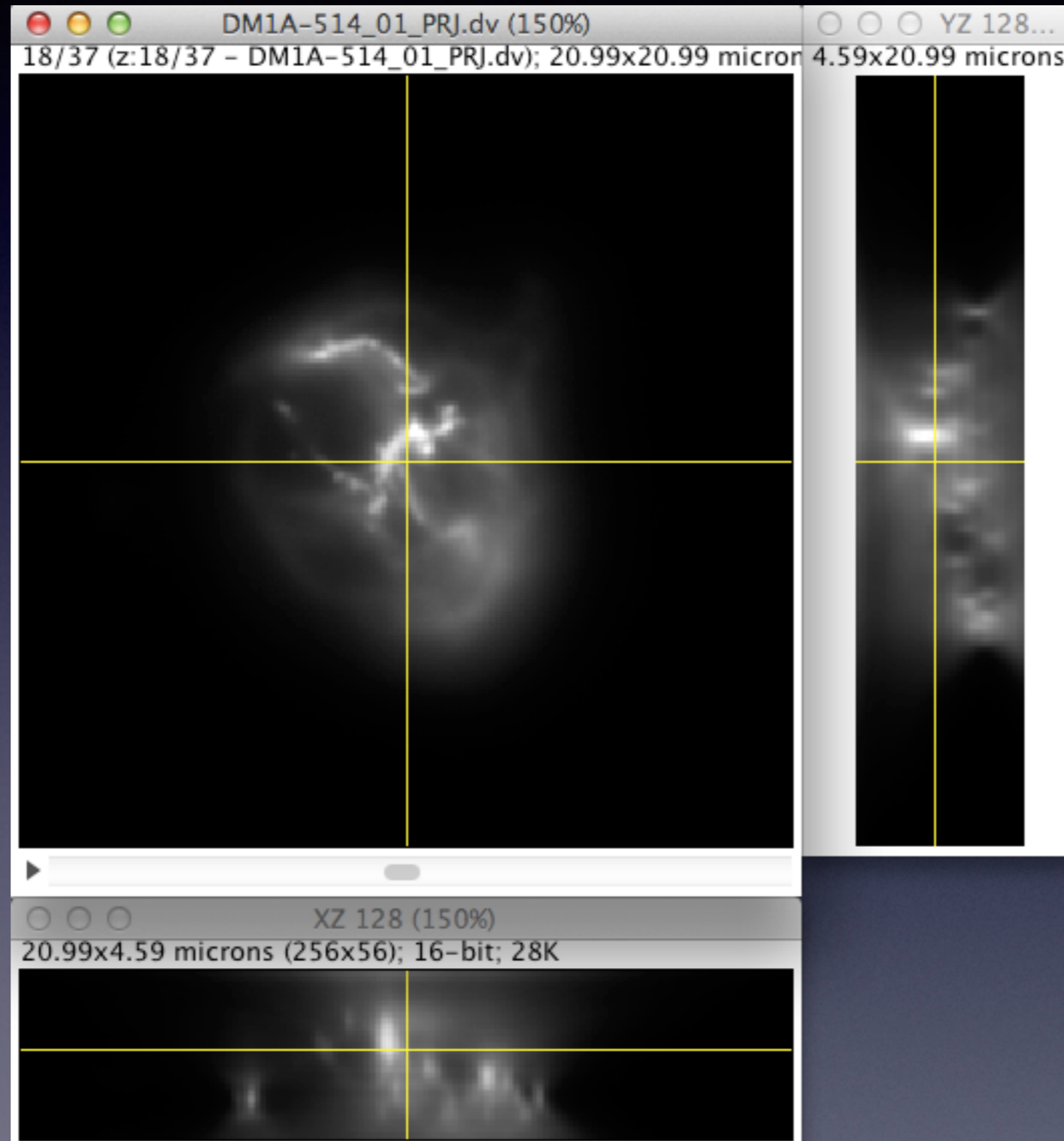
# Spherical aberration dependent on wavelength, depth, RI



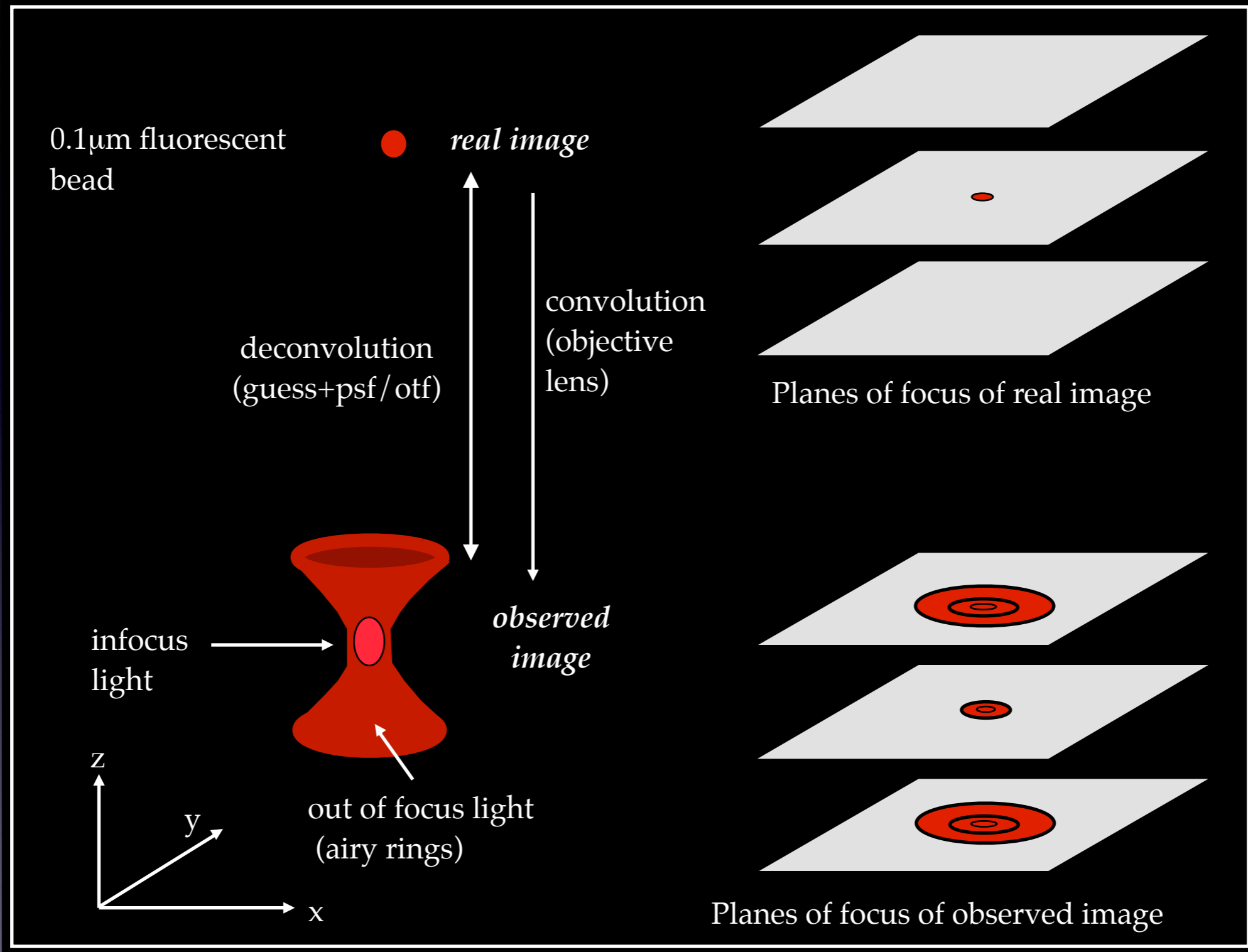
# Conventional Epi- Fluorescence Image



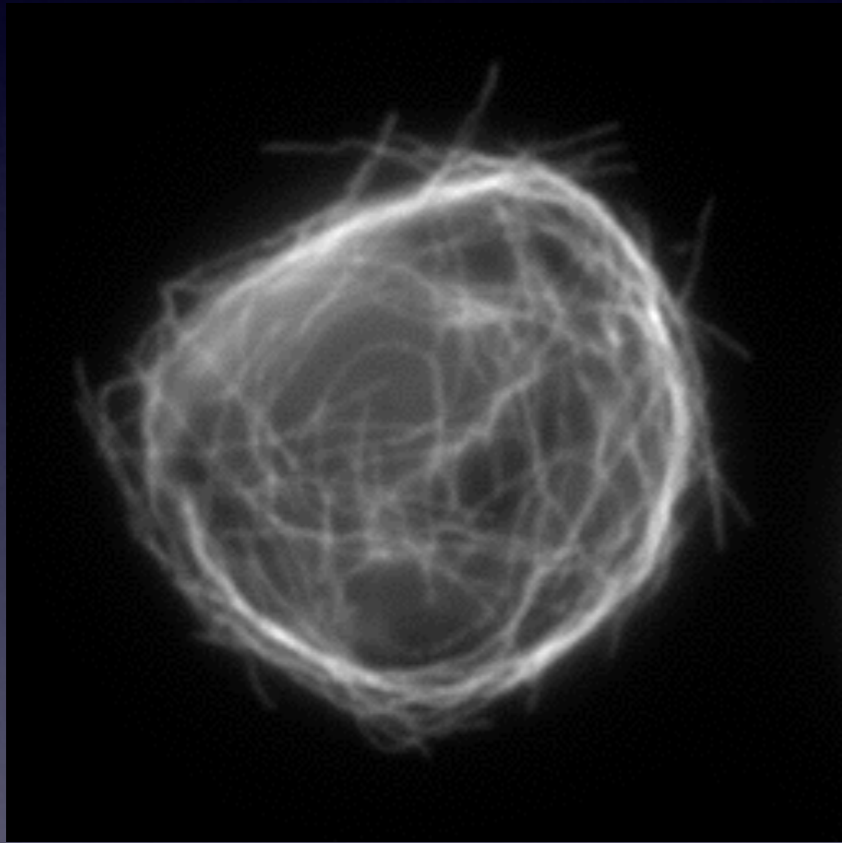
# Orthogonal views



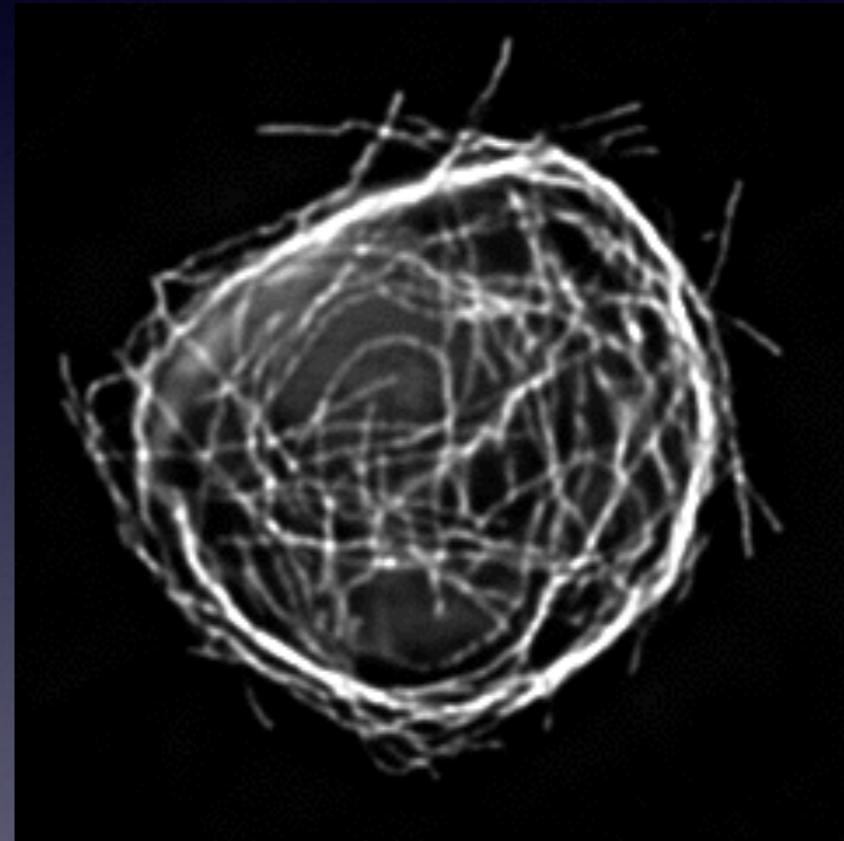




# Original Image versus Deconvolved image.



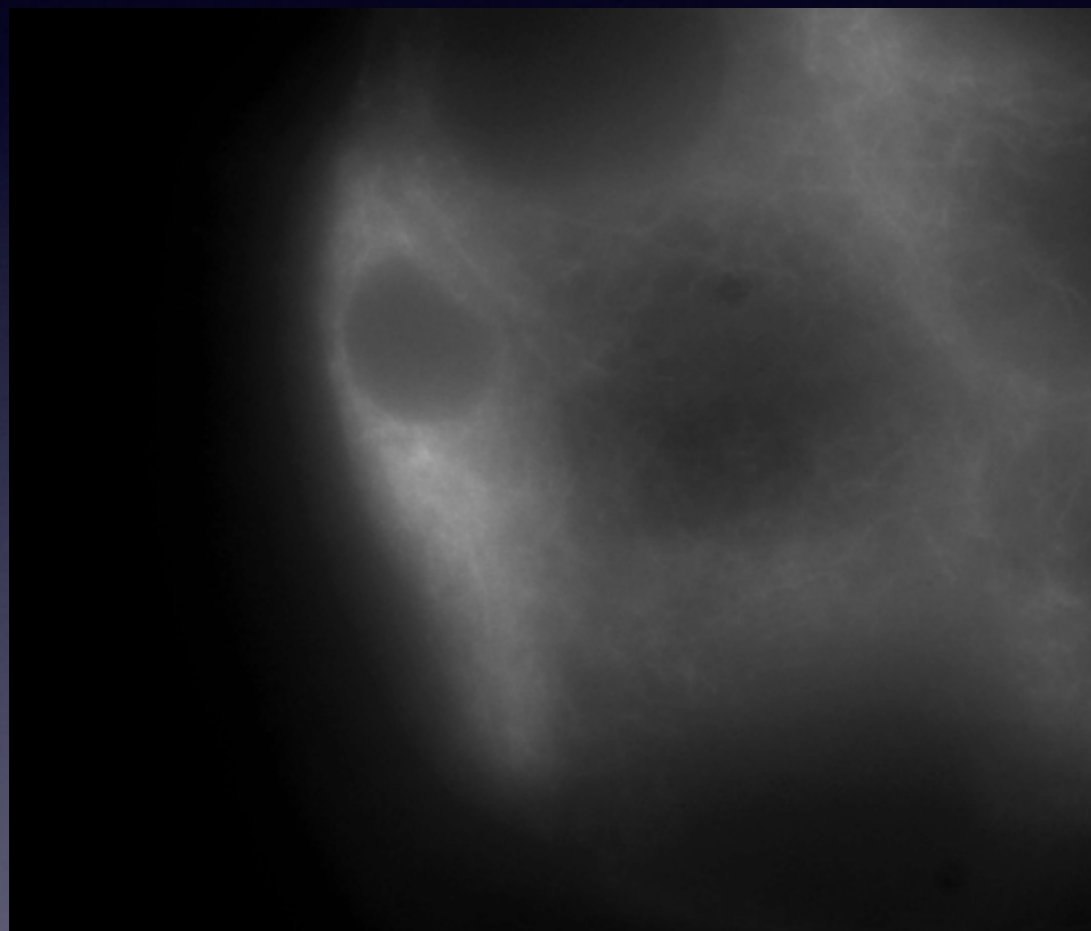
Widefield



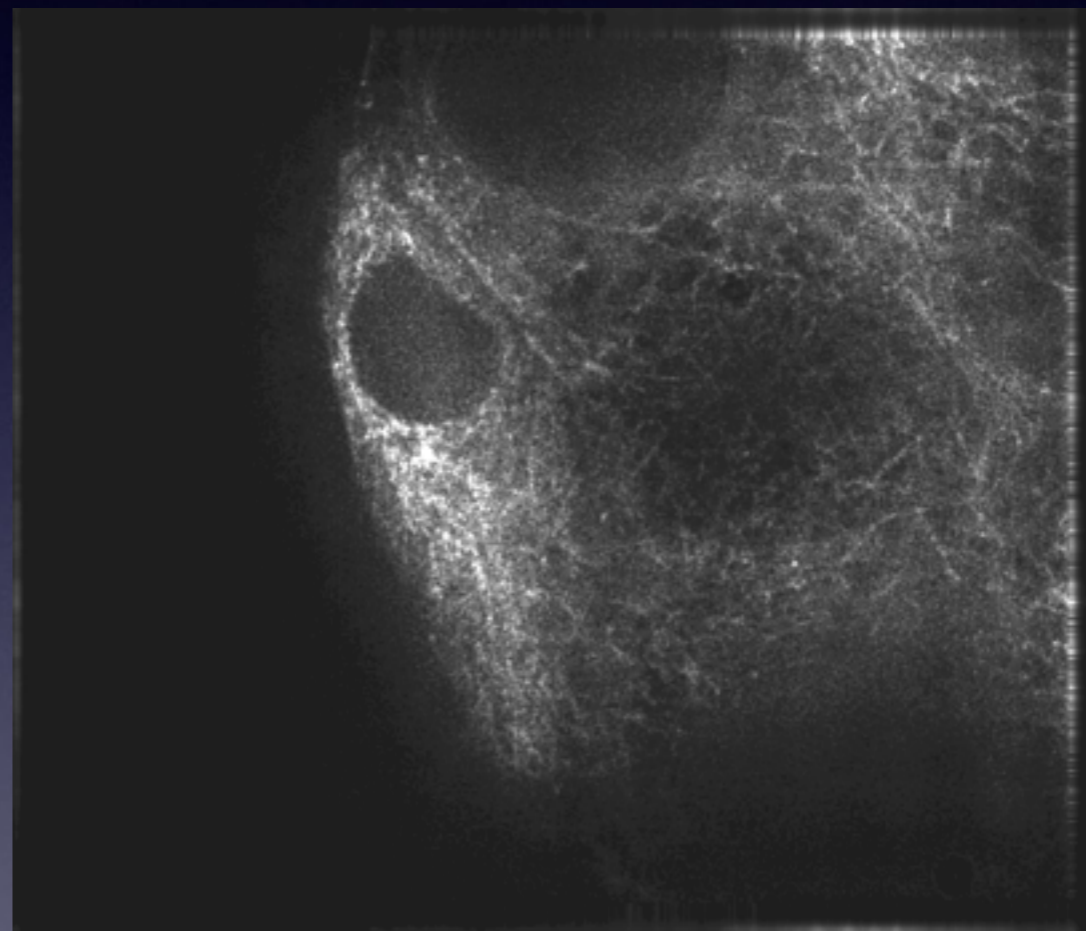
Deconvolved Widefield

# A real example of deconvolution

Before deconvolution



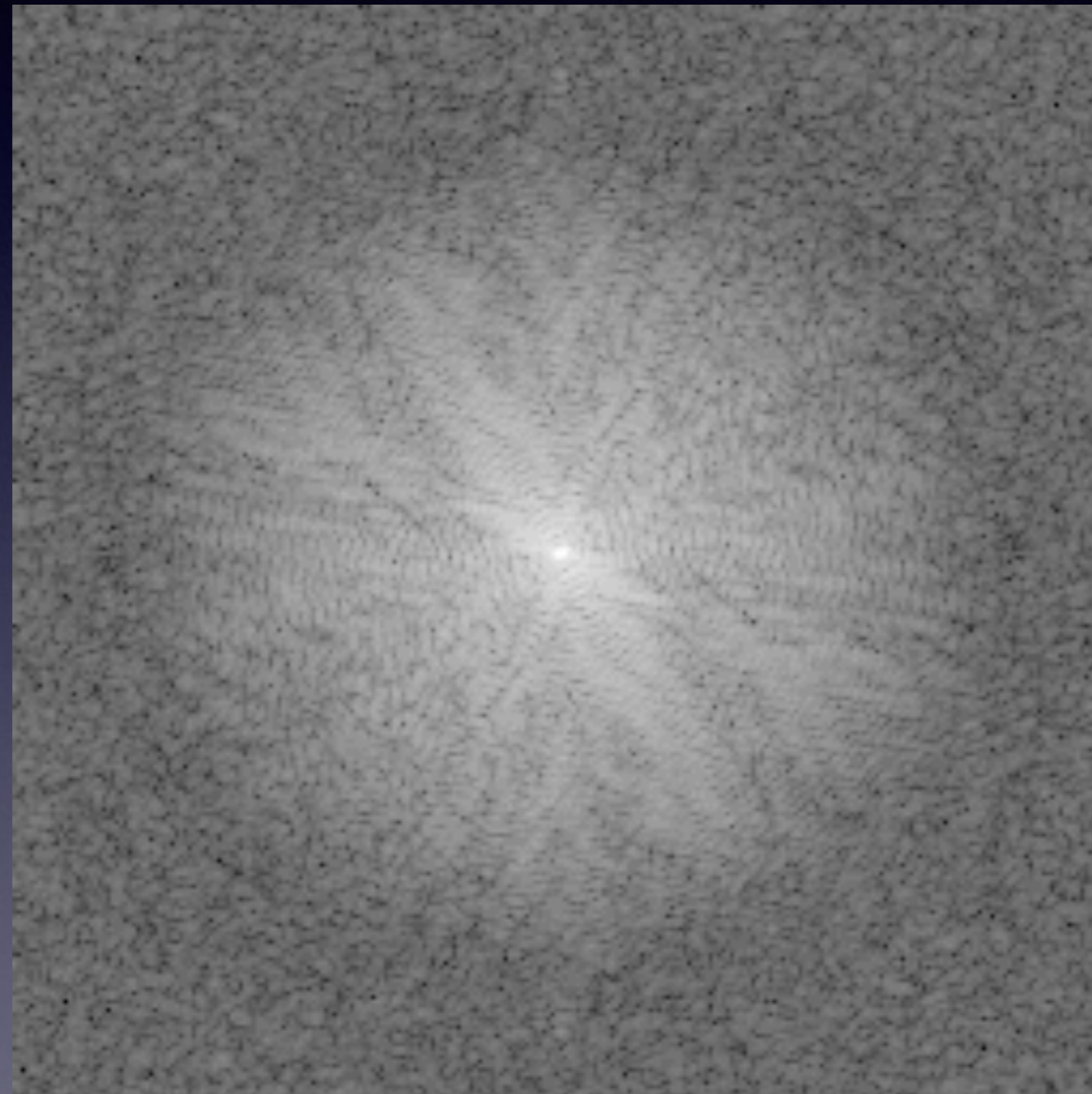
After deconvolution



# Improvements in Deconvolution

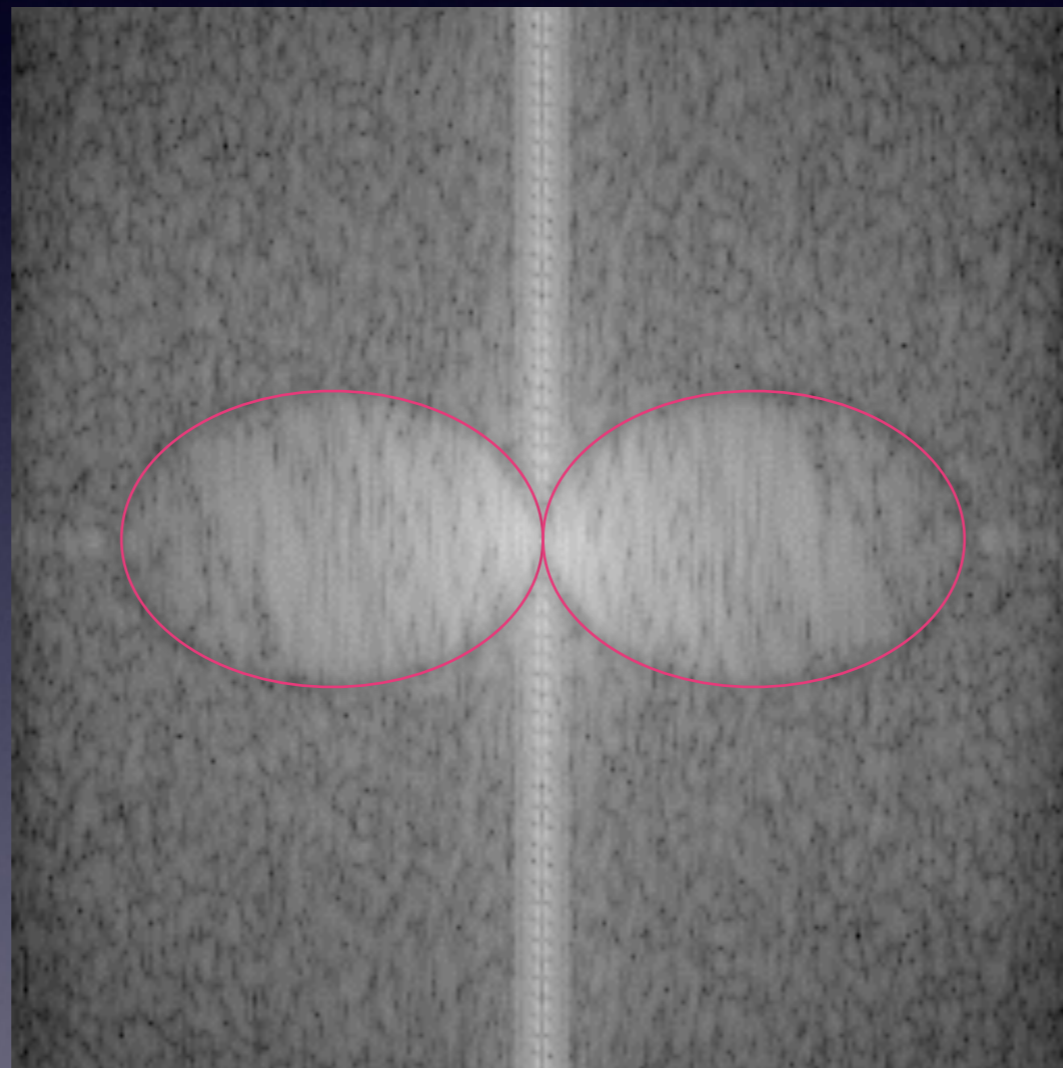
- Increases contrast as out of focus background is removed
- Reduces signal spread, hence increases resolution

# Fourier Transform



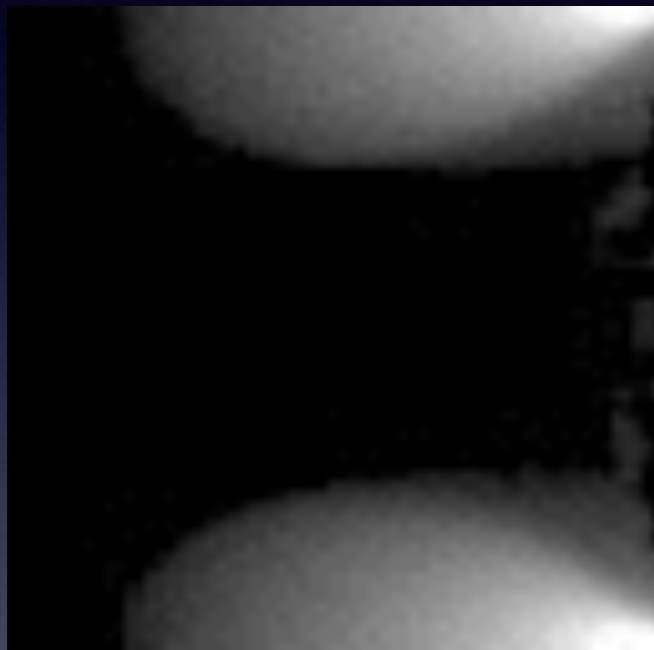
FFT of a single slice (Z plane) of  
image stack

# Fourier Transforms in XZ plane

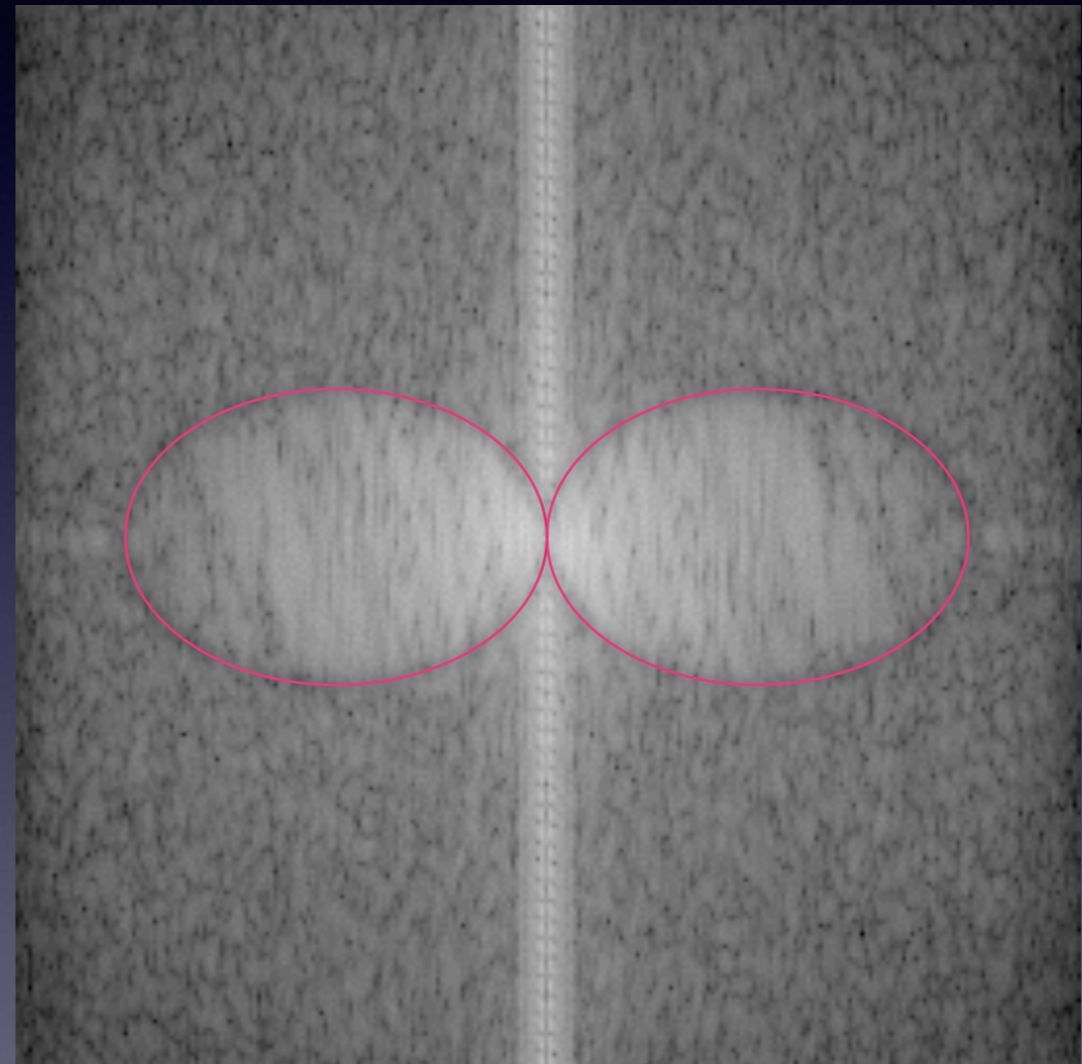


FFT of an orthogonal slice of  
image stack

# Applying the PSF in practice

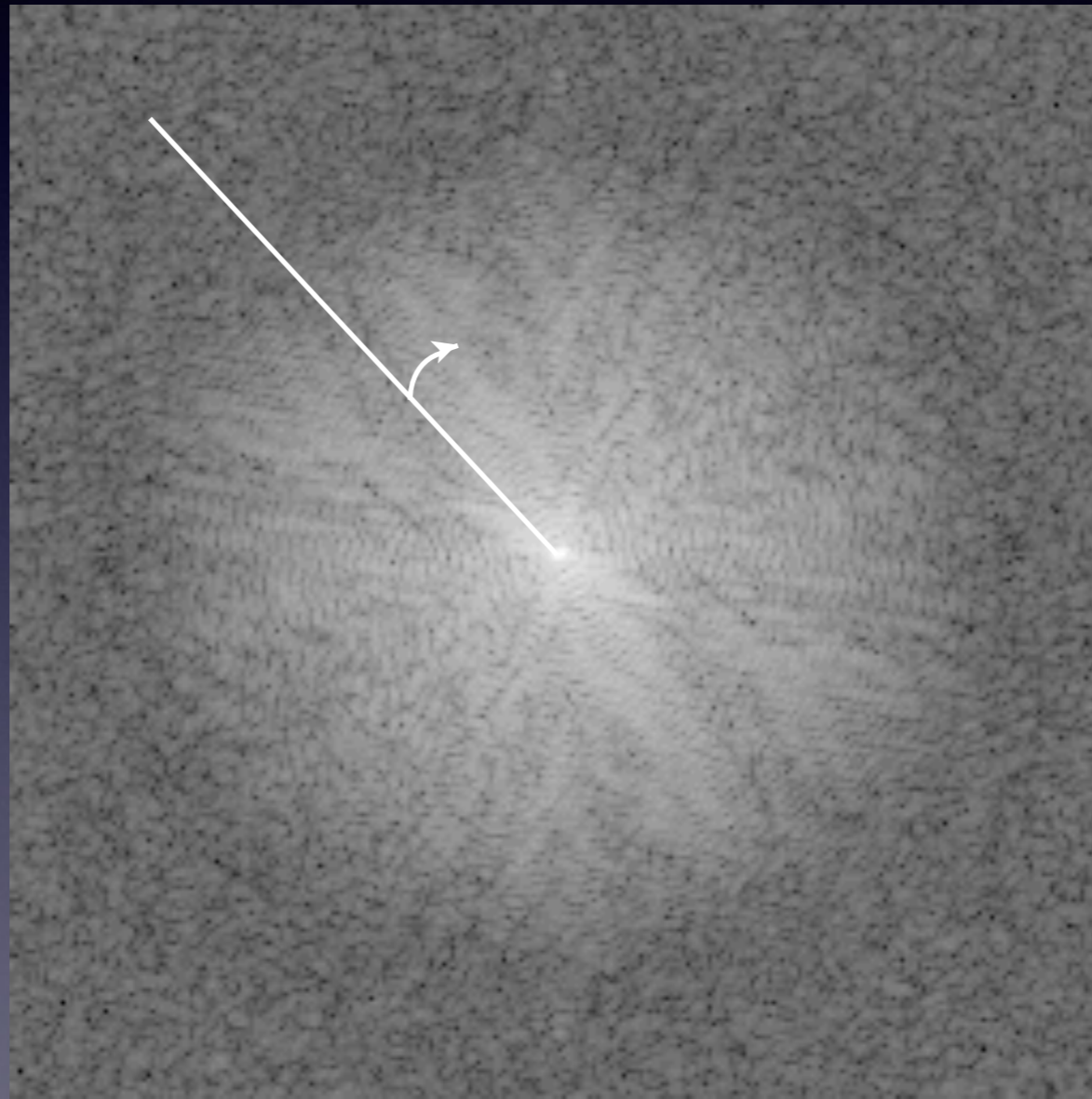


Optical Transfer Function  
OTF - FFT of the PSF



FFT of an XZ section of  
the real image

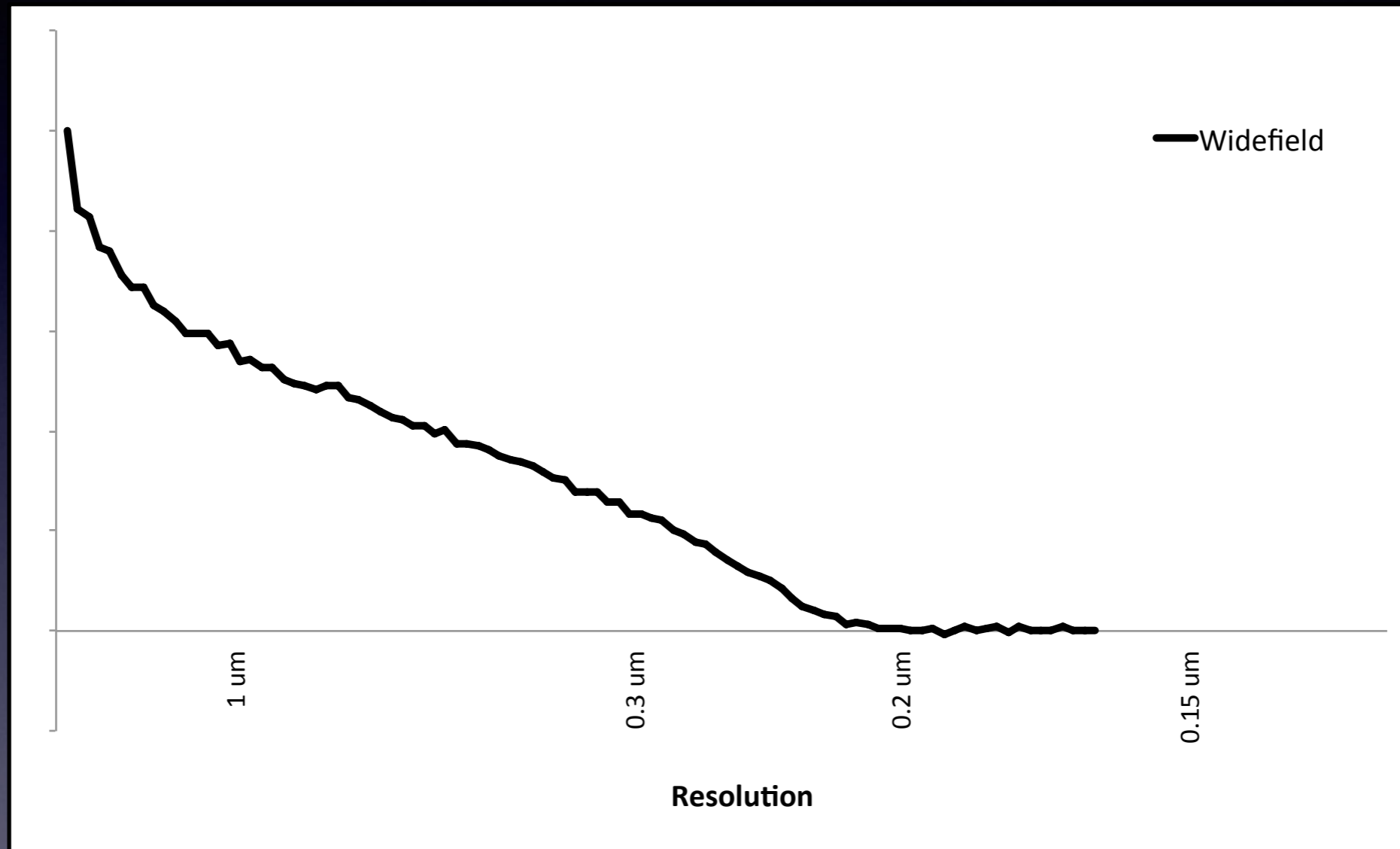
# Fourier Transforms to Assess Resolution



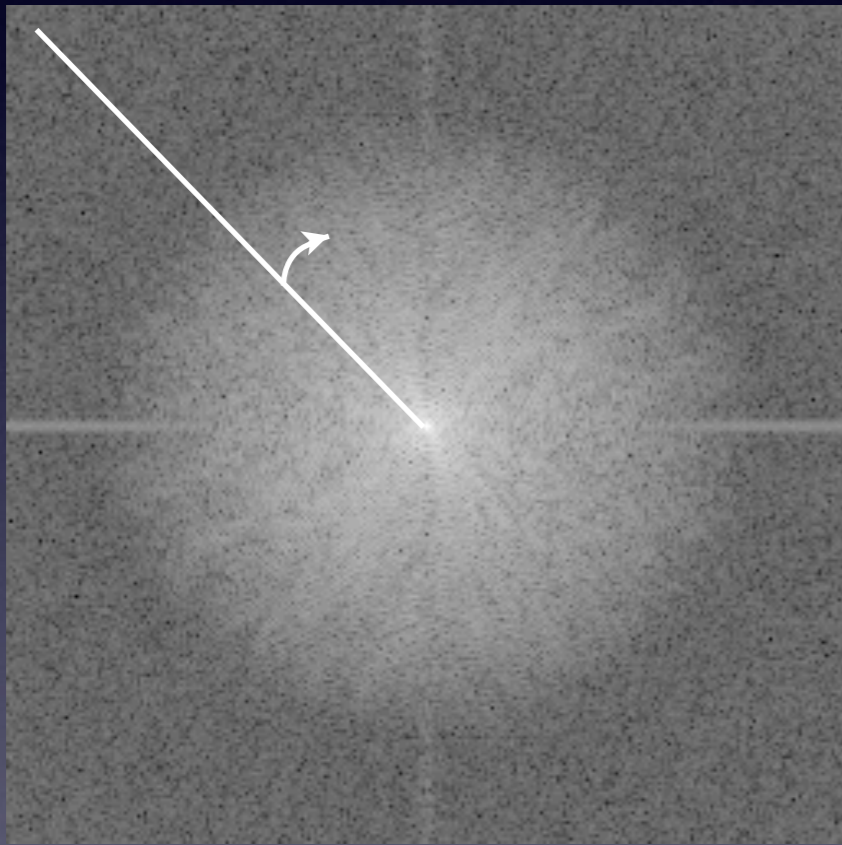
FFT of a single slice (Z plane) of  
image stack



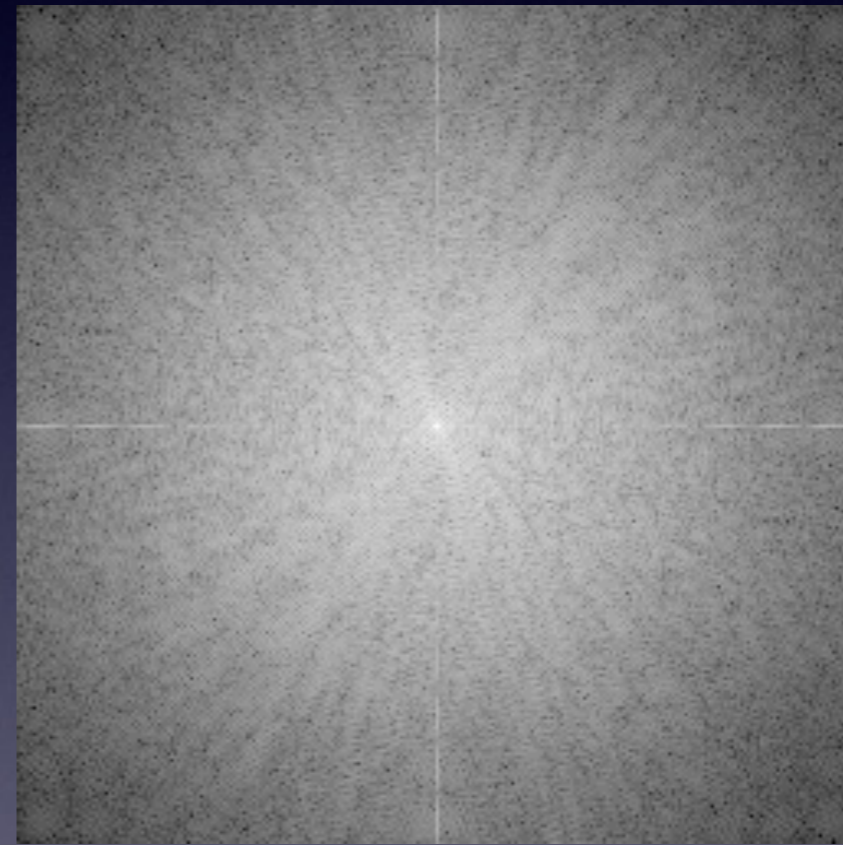
# Radial Integrals of FTs



# Fourier Transforms to Assess Resolution



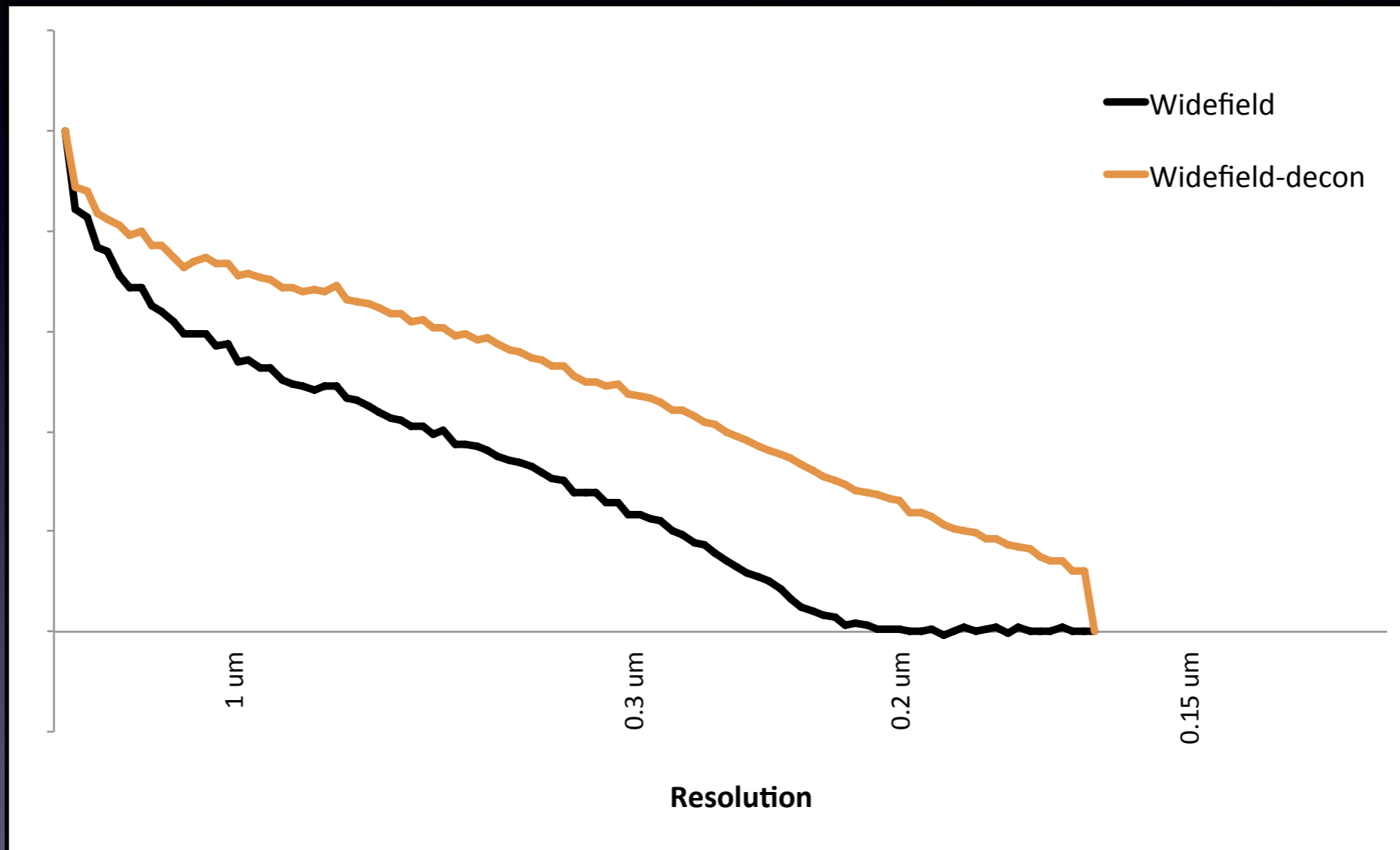
FFT of Widefield



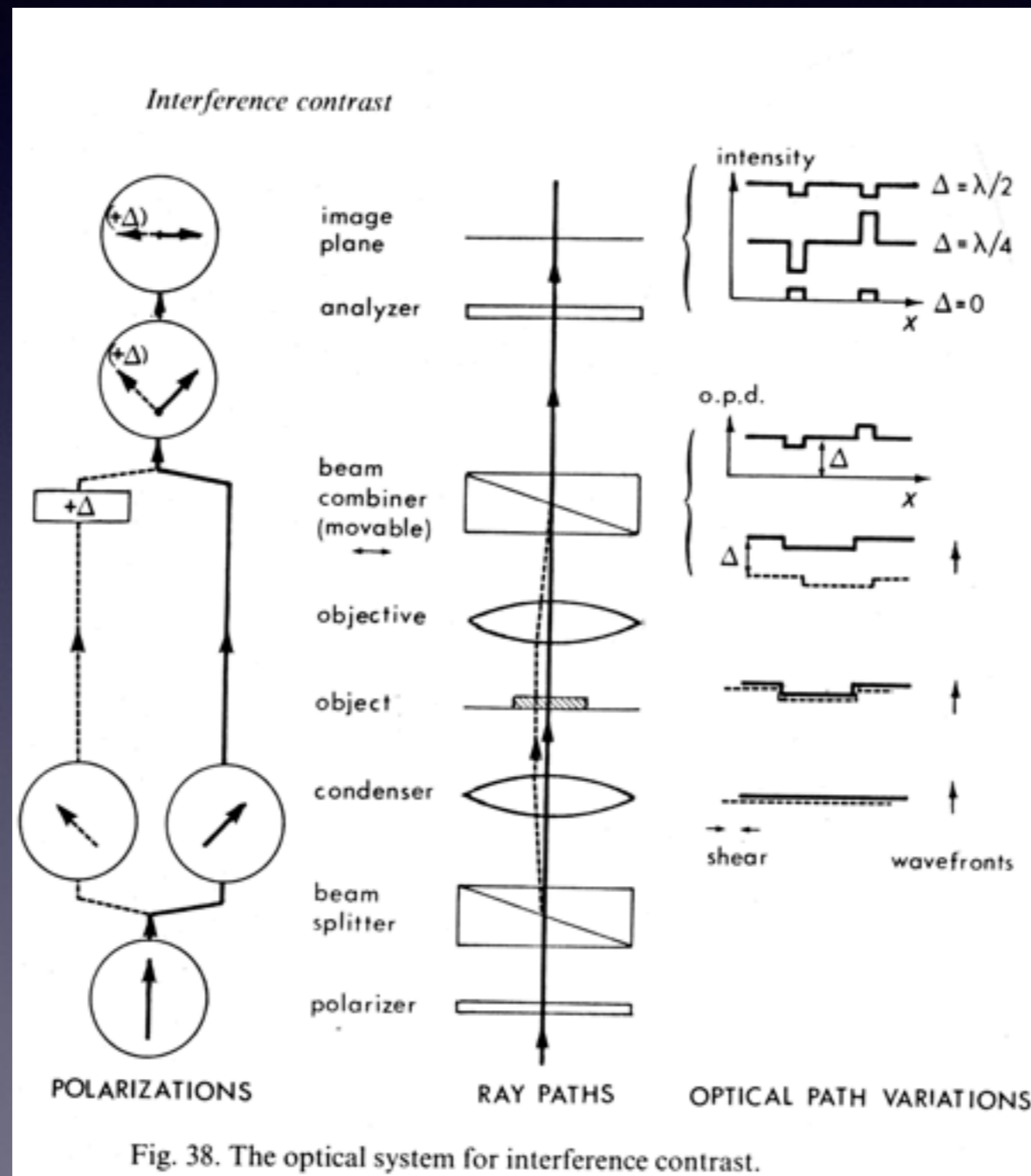
FFT of deconvolved

Widefield  
FTs of Microtubule images  
at equivalent scale

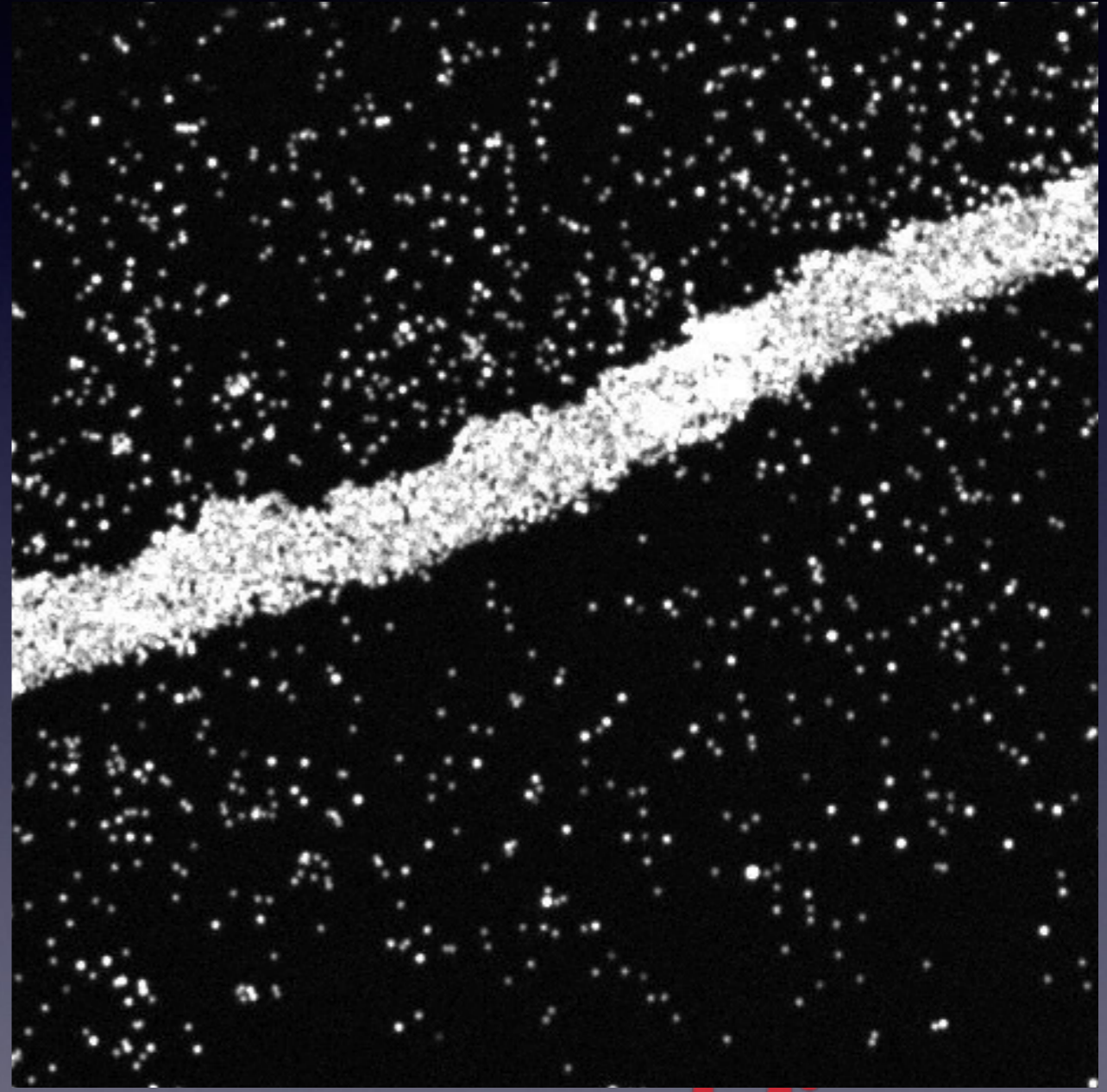
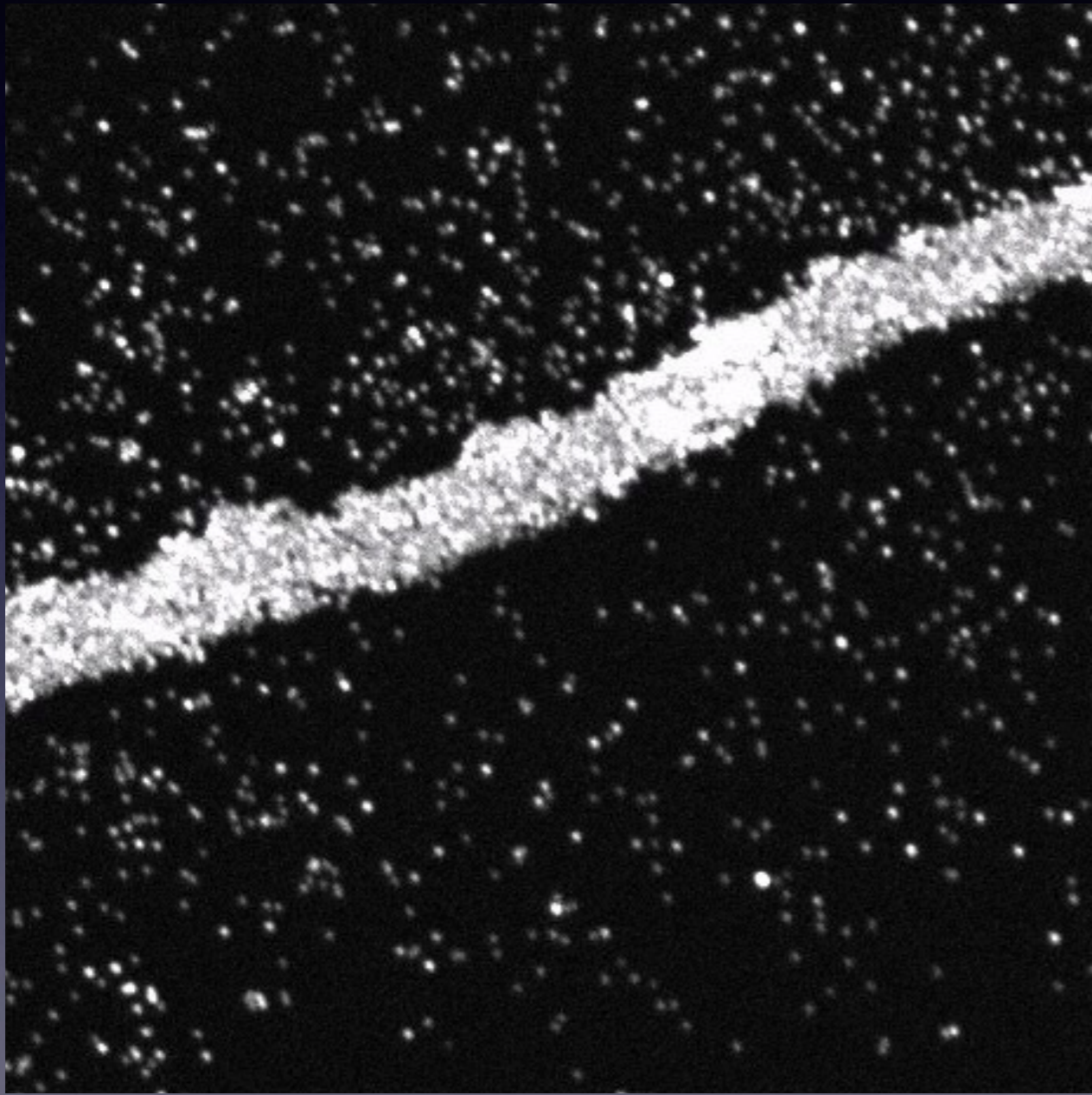
# Radial Integrals of FTs



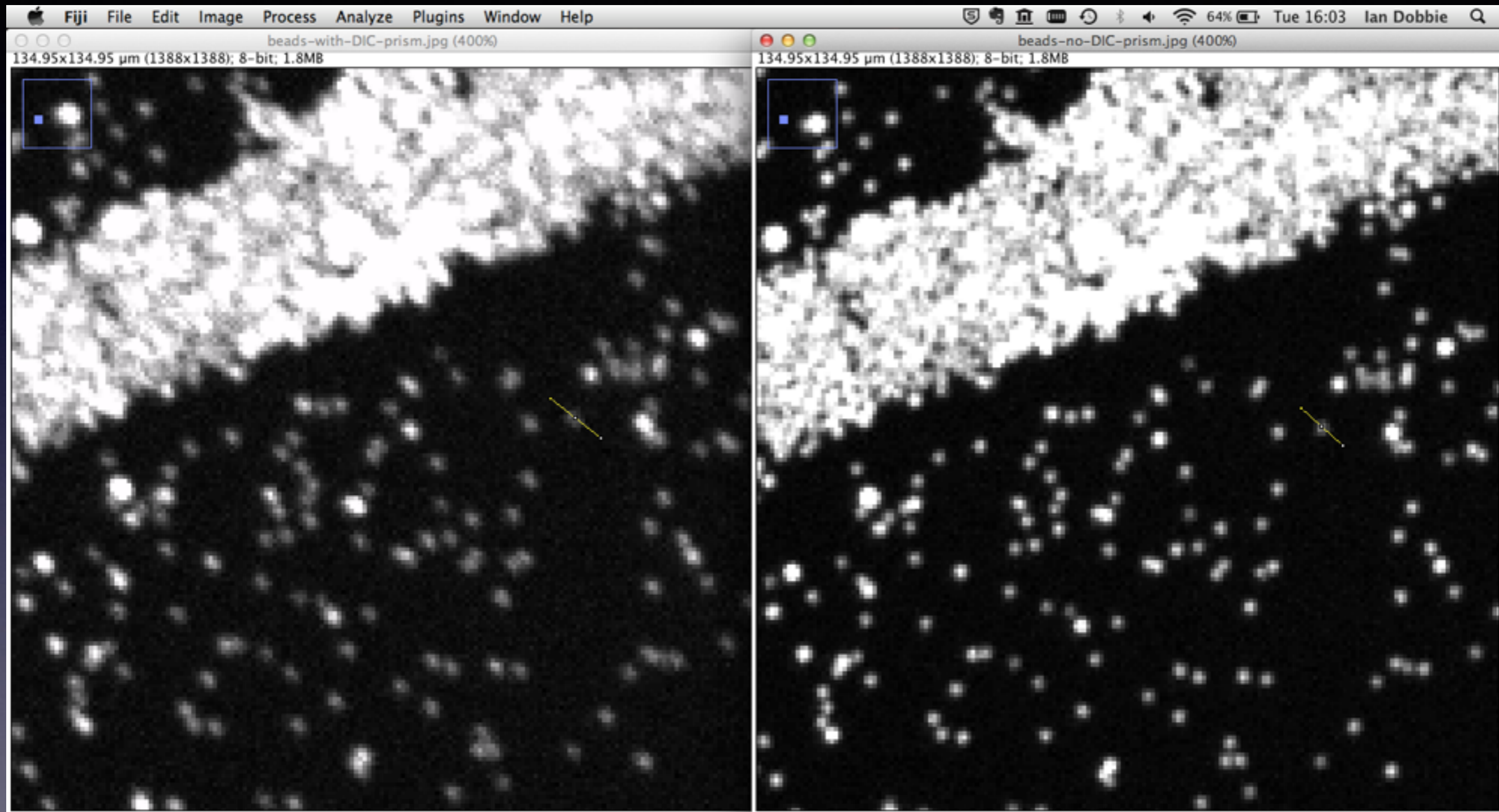
# How a DIC prism effects fluorescence imaging



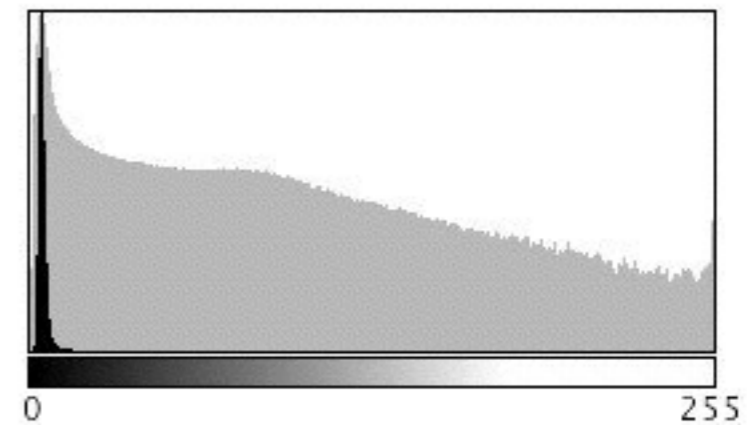
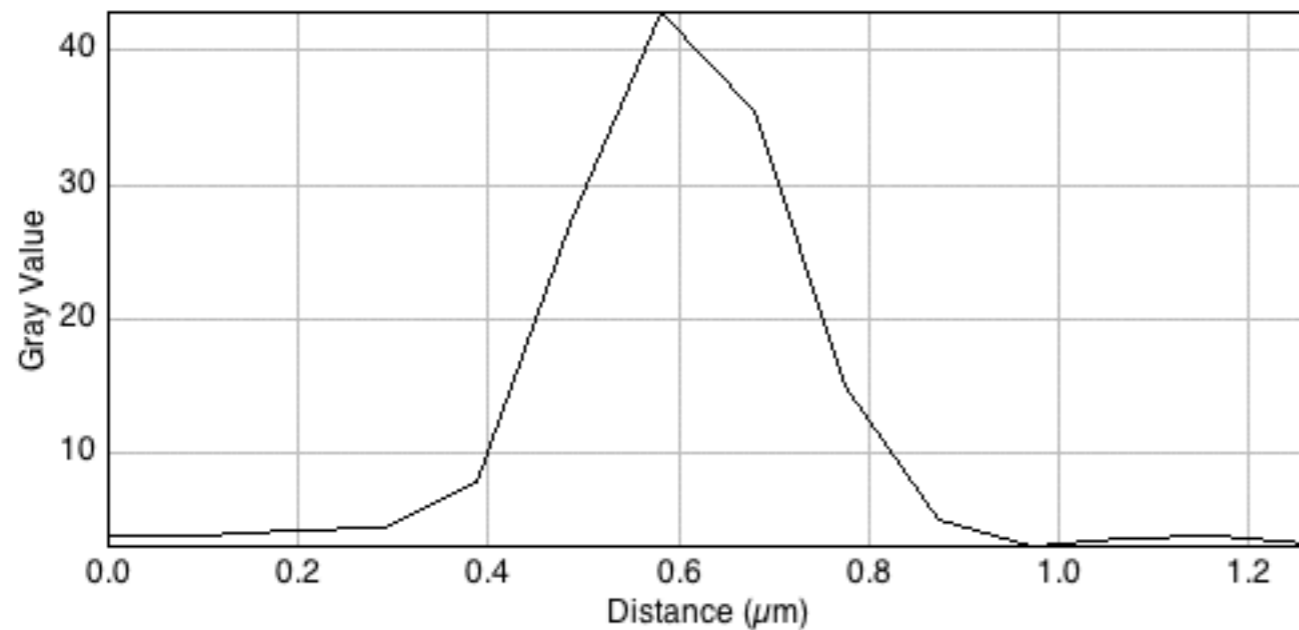
# With/without DIC prism



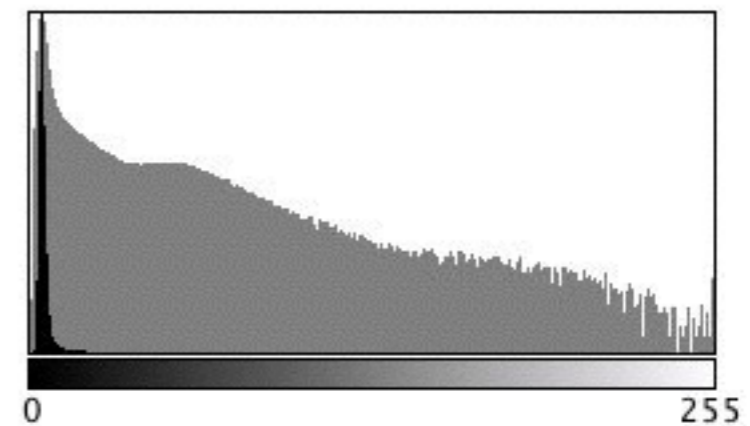
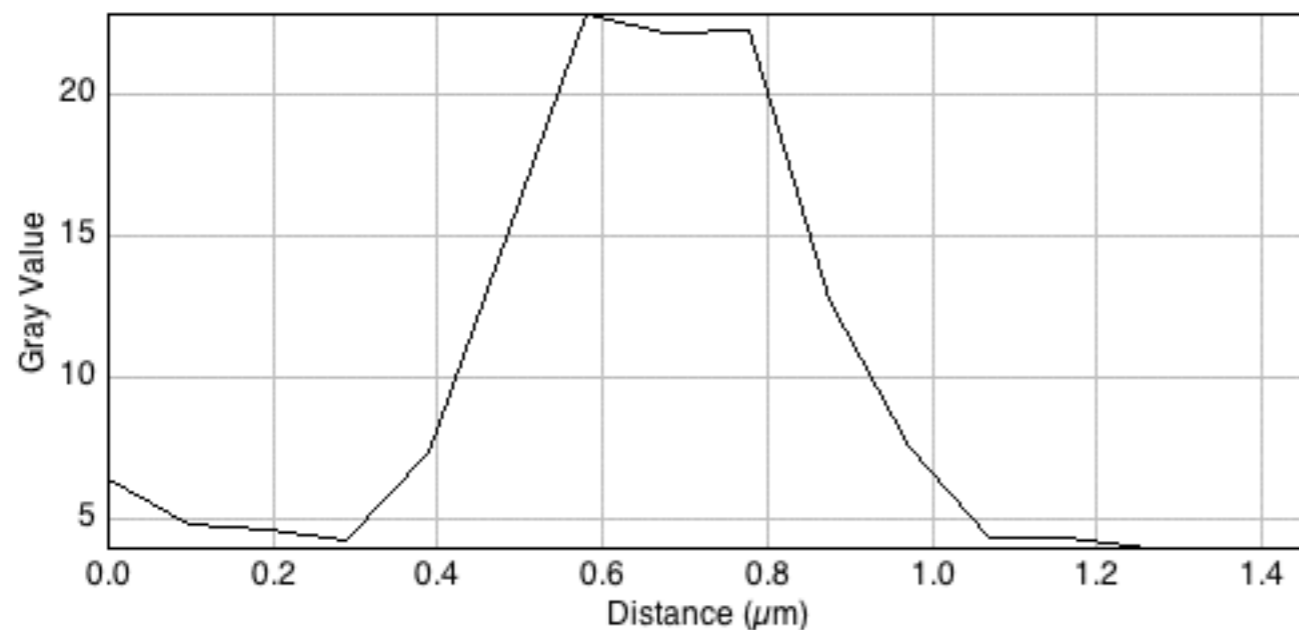
# With/without DIC prism



# Line scans and histograms

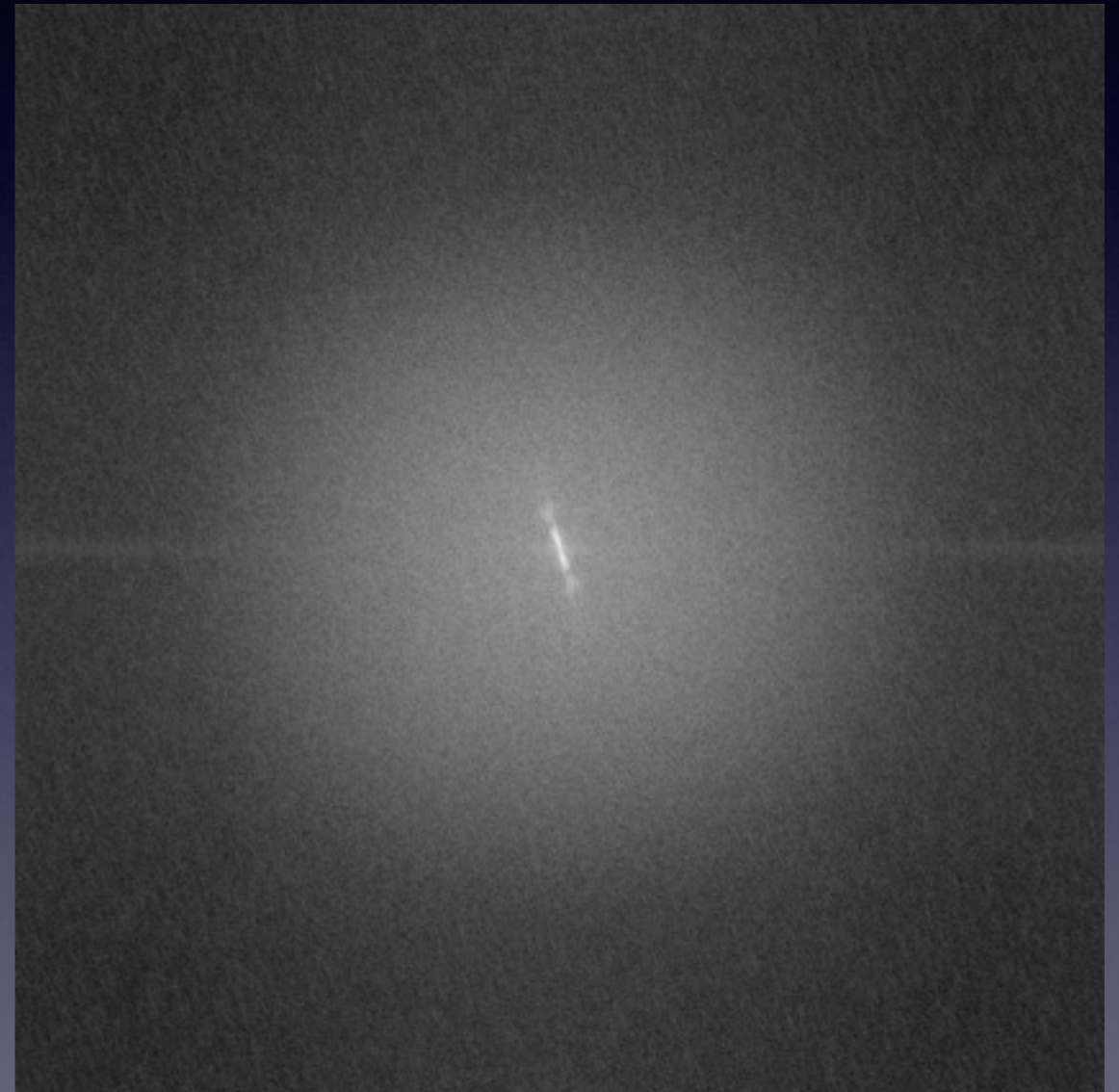
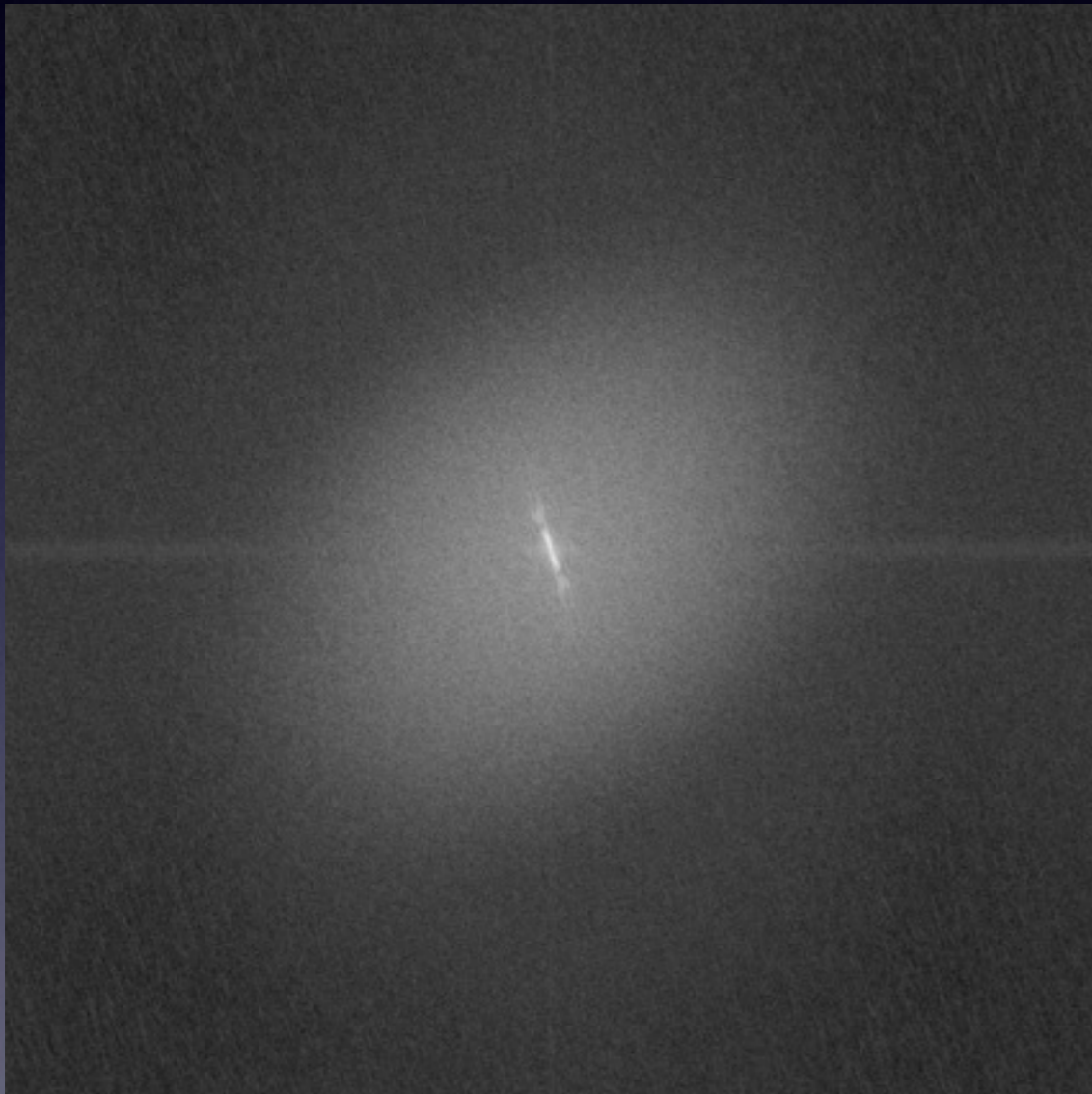


Count: 1926544    Min: 0  
Mean: 8.969        Max: 255  
StdDev: 20.096    Mode: 4 (538941)



Count: 1926544    Min: 0  
Mean: 7.461        Max: 255  
StdDev: 13.722    Mode: 4 (547814)

# FFTs with/without DIC prism

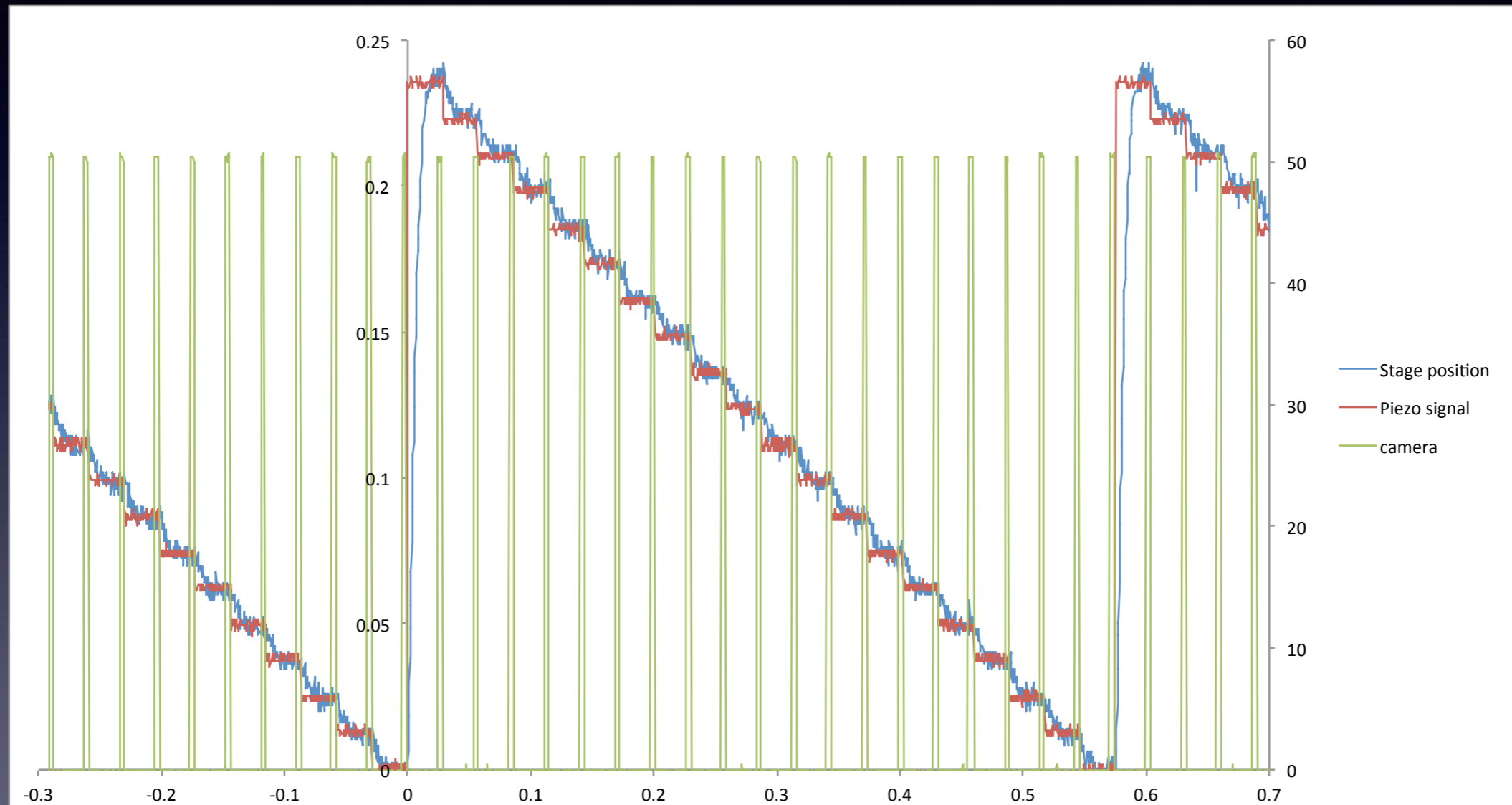




# Super Fast Acquisition (FastZ)

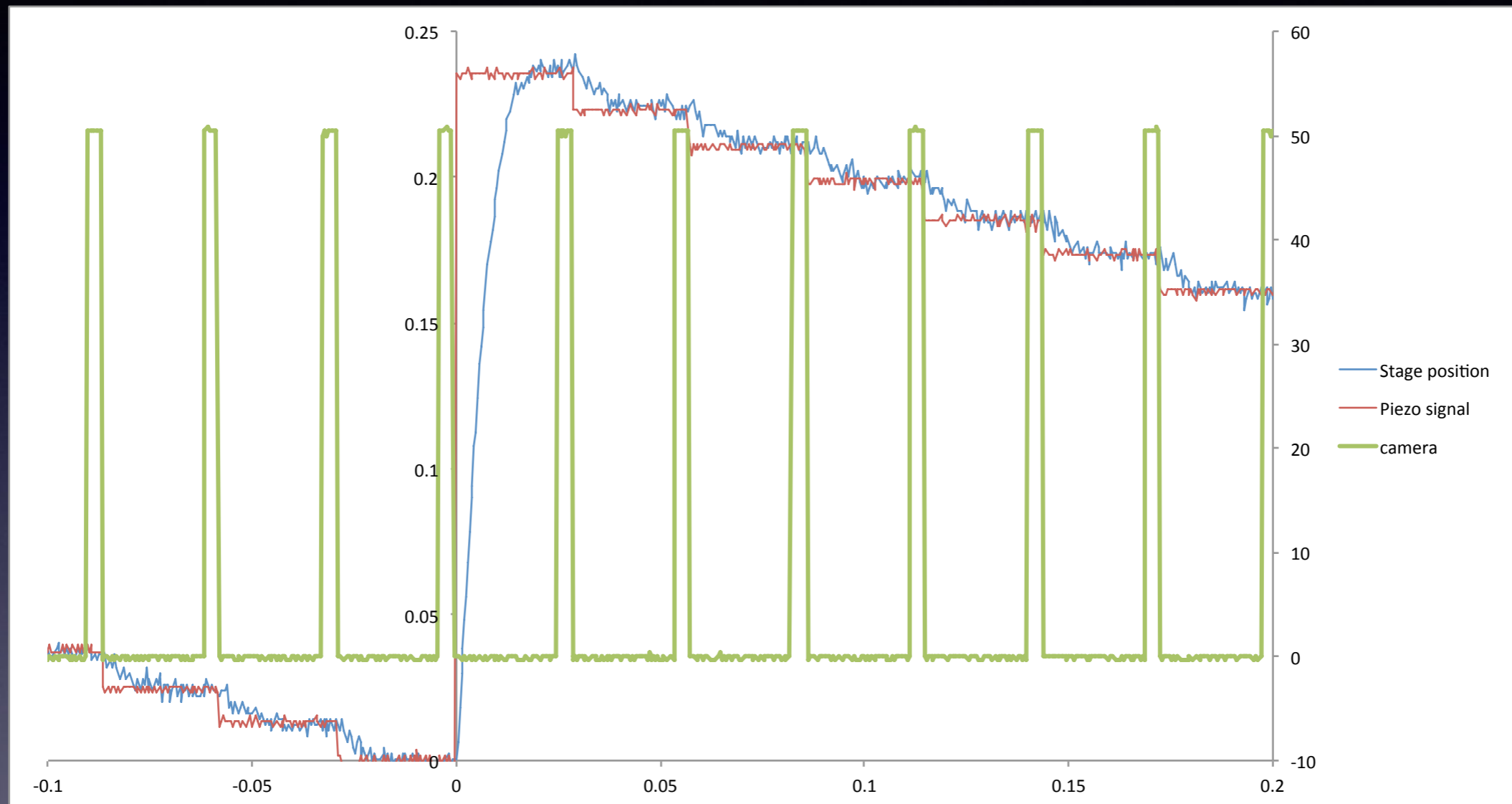
- Ramp the Z position instead of stepping it
- Take images as fast as possible during ramp
- Delay between stacks to allow stage to return to initial position

# Conventional widefield Z stack

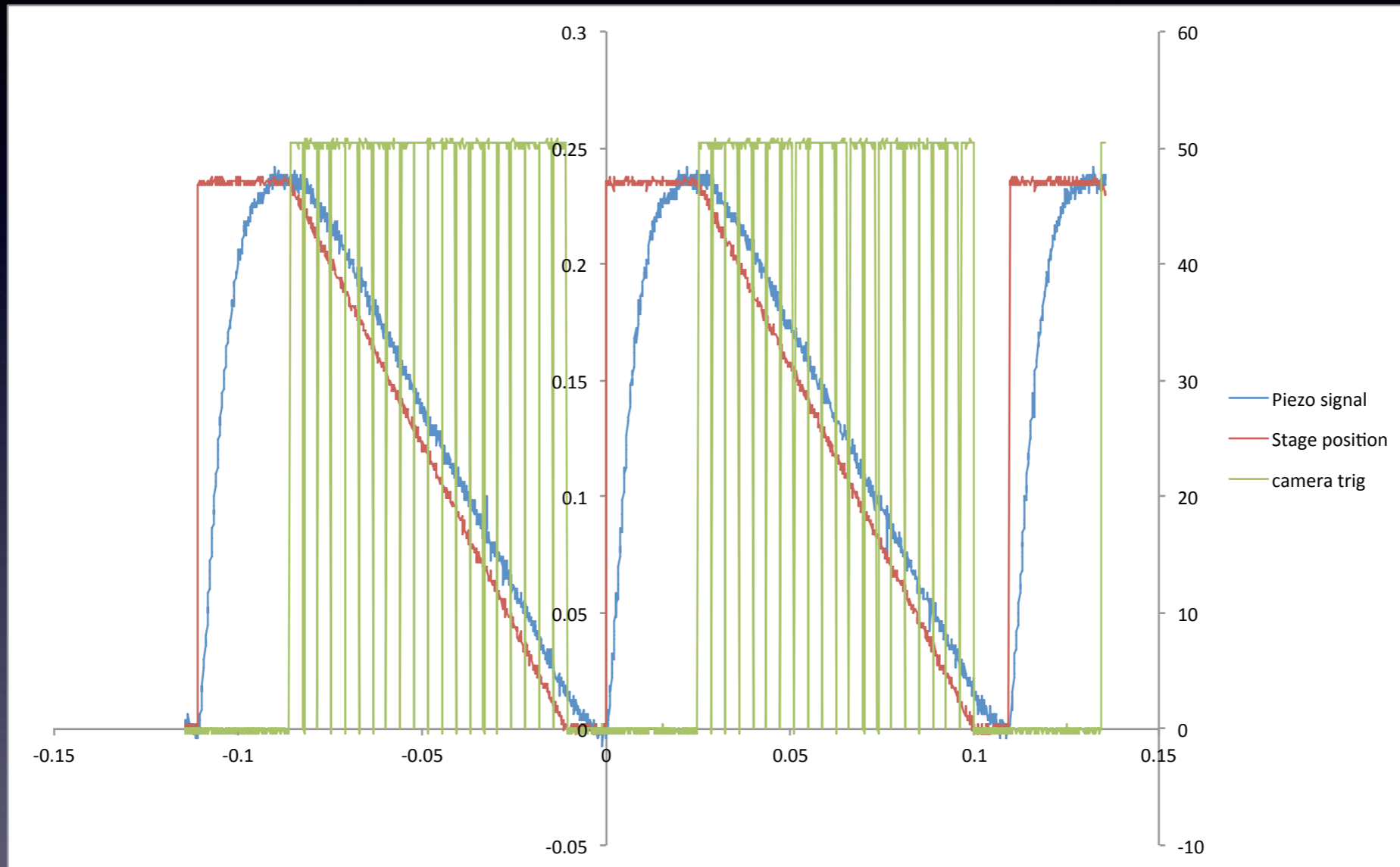


20 Z planes as fast as possible

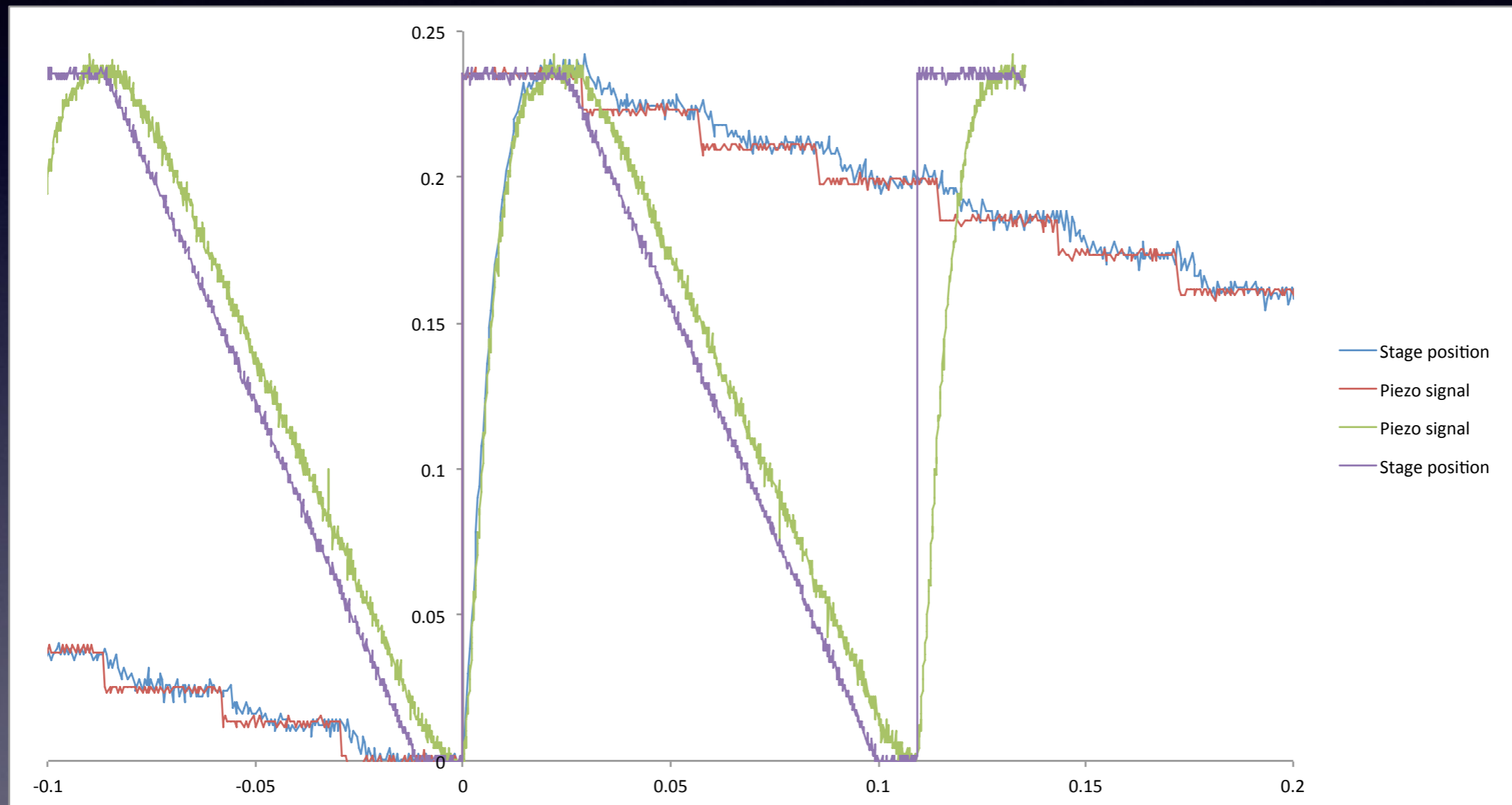
# Coventional Z stack



# Ramp Z stack



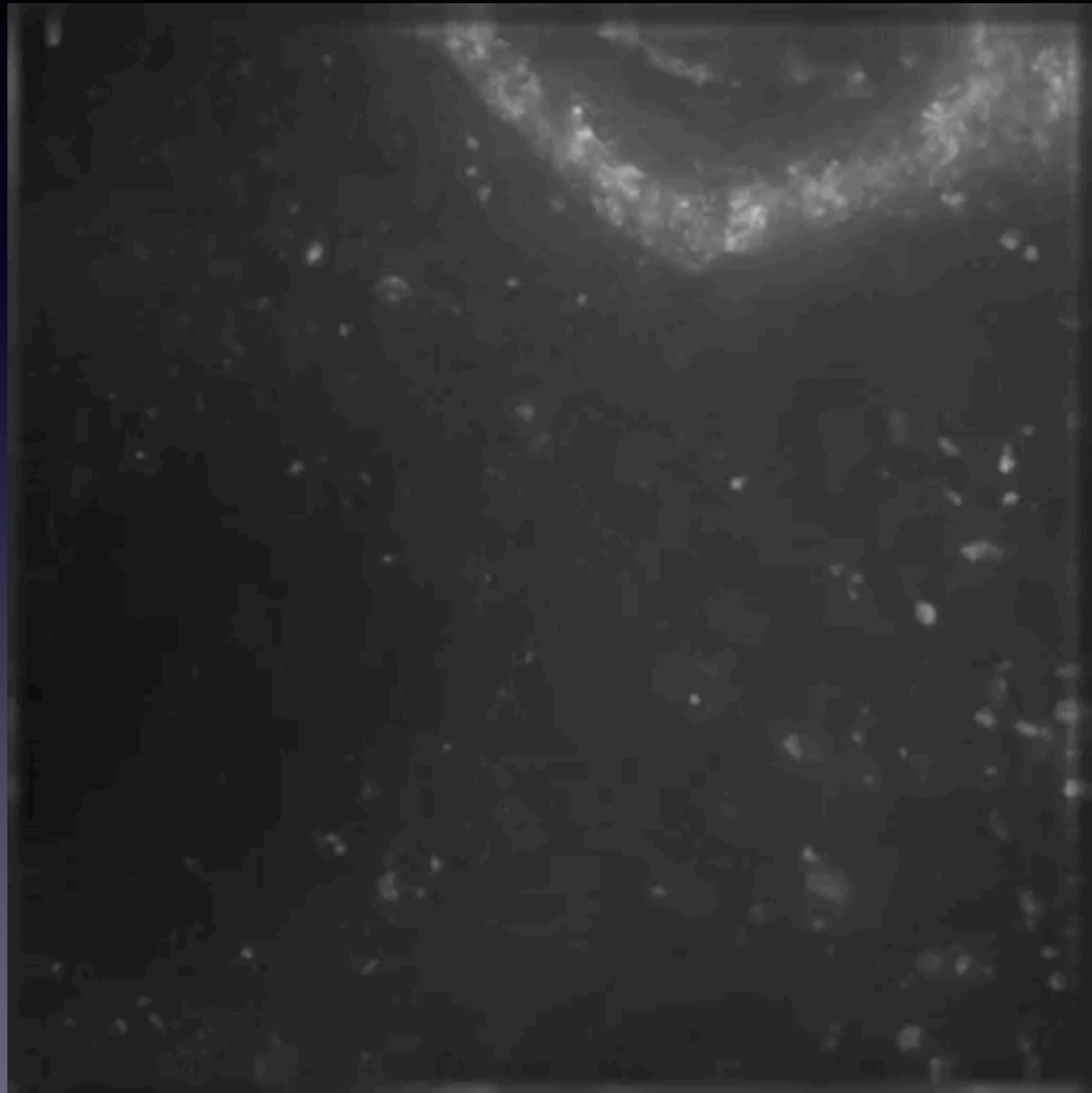
# Comparison: FastZ to normal



# Speed increases

- Depends on stack height, image size, exposure time.
- Test sample, 512x512 pixel images, 1 ms exposure 20 Z slices of 200 nm.
- Conventional cycle time = 575 ms
- FastZ cycle time = 109 ms

# FastZ - Results

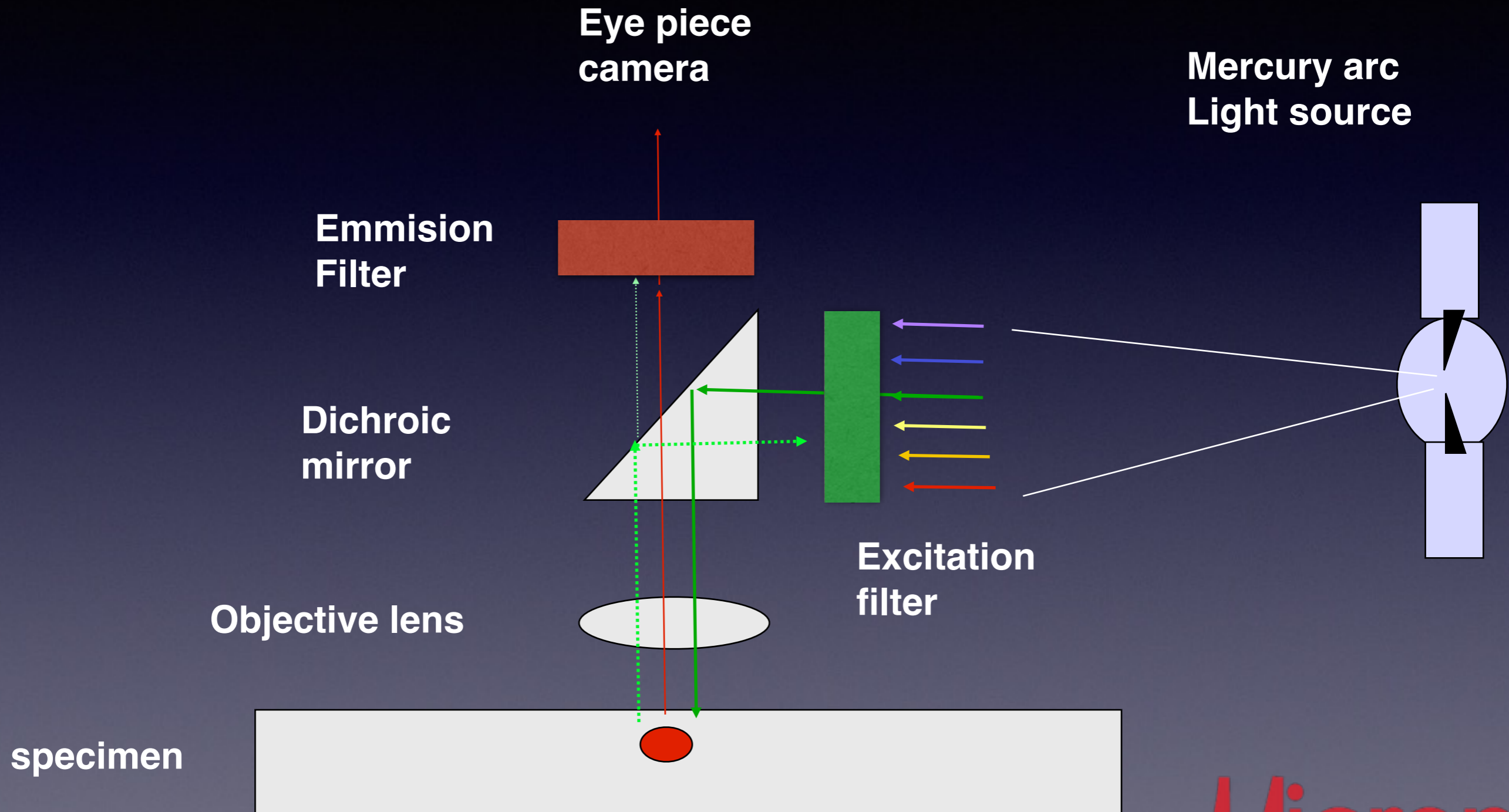


Me3IB-GFP Drosophila oocyte  
25-slices, 8 stacks/s - 200 frames/s

**Micron**  
OXFORD

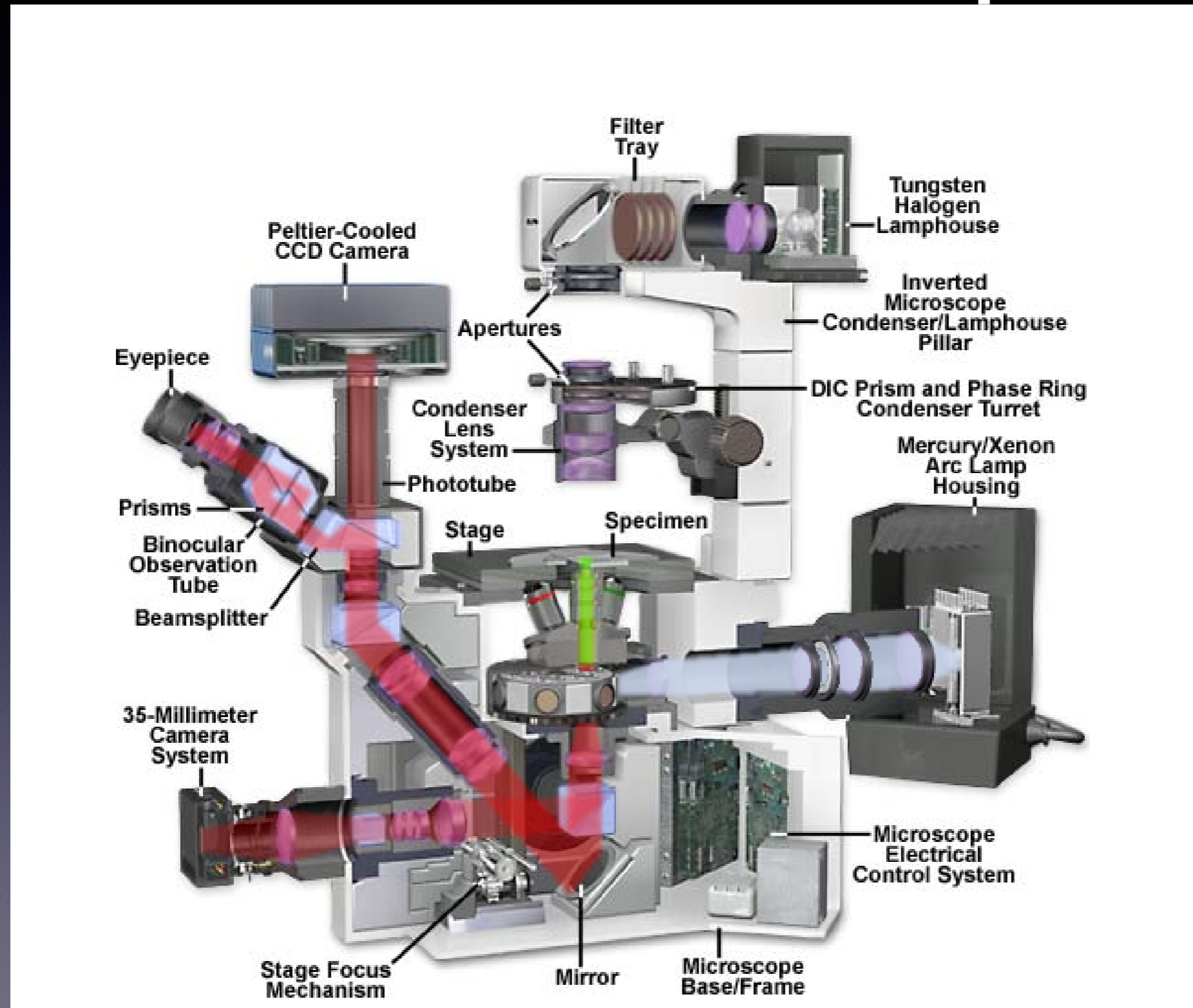
# Reminder

## How do fluorescence microscopes work ?





# Problem: the design of all conventional microscope stands



**How can we improve the basic design of widefield microscopes?**

**By dispensing with the normal microscope stand and building your own microscope from optical components on a breadboard**

# The solution -build your own bespoke microscope



**Mark Leake  
with the  
Slimfield  
TIRF  
microscope  
(Biophysics  
prize)**

# Bespoke Microscopes

**Why bother?**

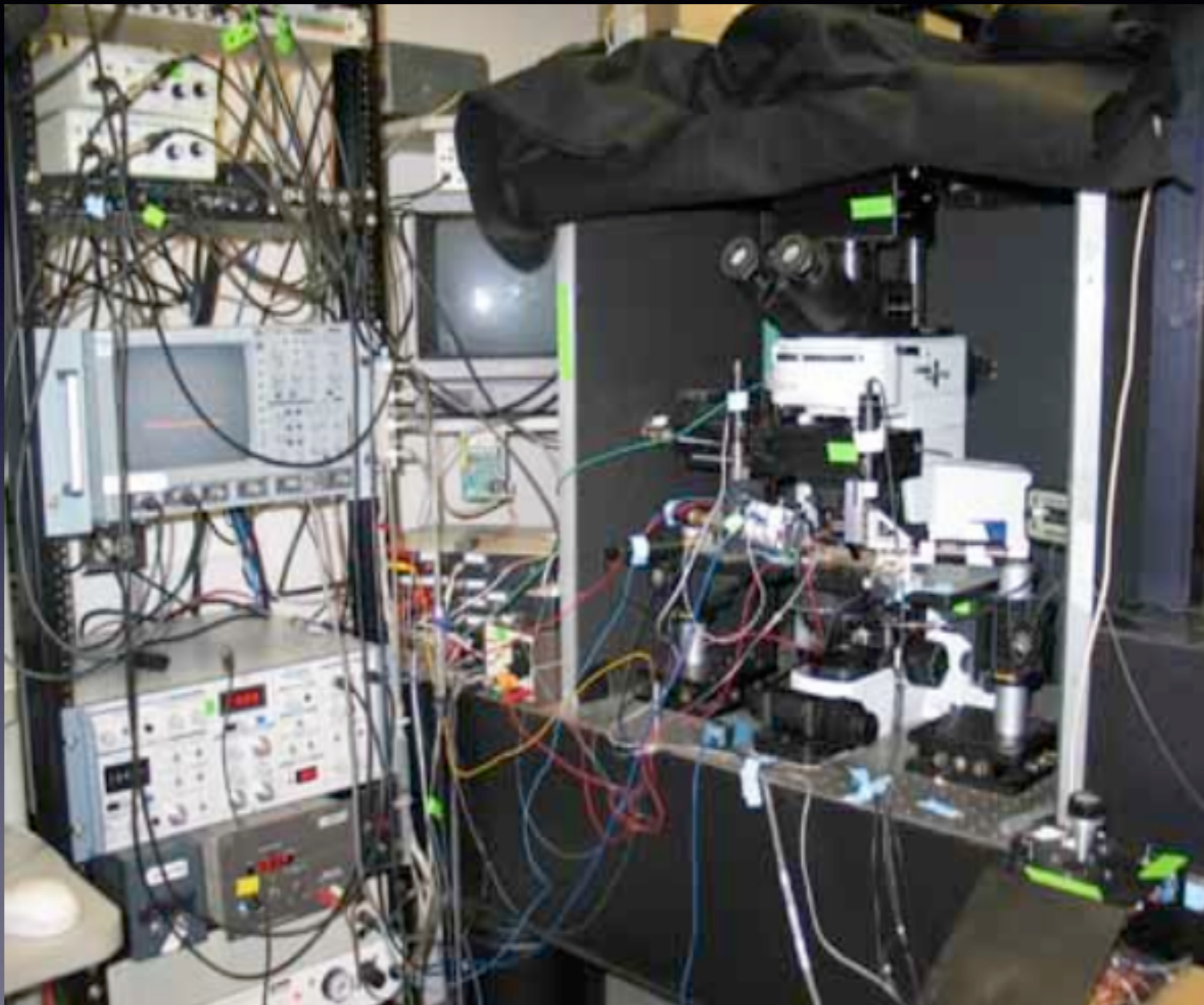
**Specific applications -better than commercial  
microscopes**

**Flexibility**

**Cost**

# Popular bespoke microscope

## Multiphoton for neuroscience work



# Bespoke Microscopes

## Why NOT to bother?

- Salary of physicist/engineer required
- Long building time required (it's hard)
- Not supported by a company  
(repairs are costly and lengthy)
- Not always easy to use by biologists

# Example of Bespoke Microscopes

## OMX-T microscope

Designed and built by John Sedat and Dave Agard,  
UCSF

## Live PALM microscope

Designed and built by Stephan Uphoff and  
Achillefs Kapanidis, Micron Oxford

## WOSM

Designed and built by Nick Carter and Rob Cross,  
Warwick University

## Openspim

Designed and built by Pavel Tamacek and his team  
at Dresden MPI

## Holographic microscope

Irwin Said and Richard Berry, Micron Oxford

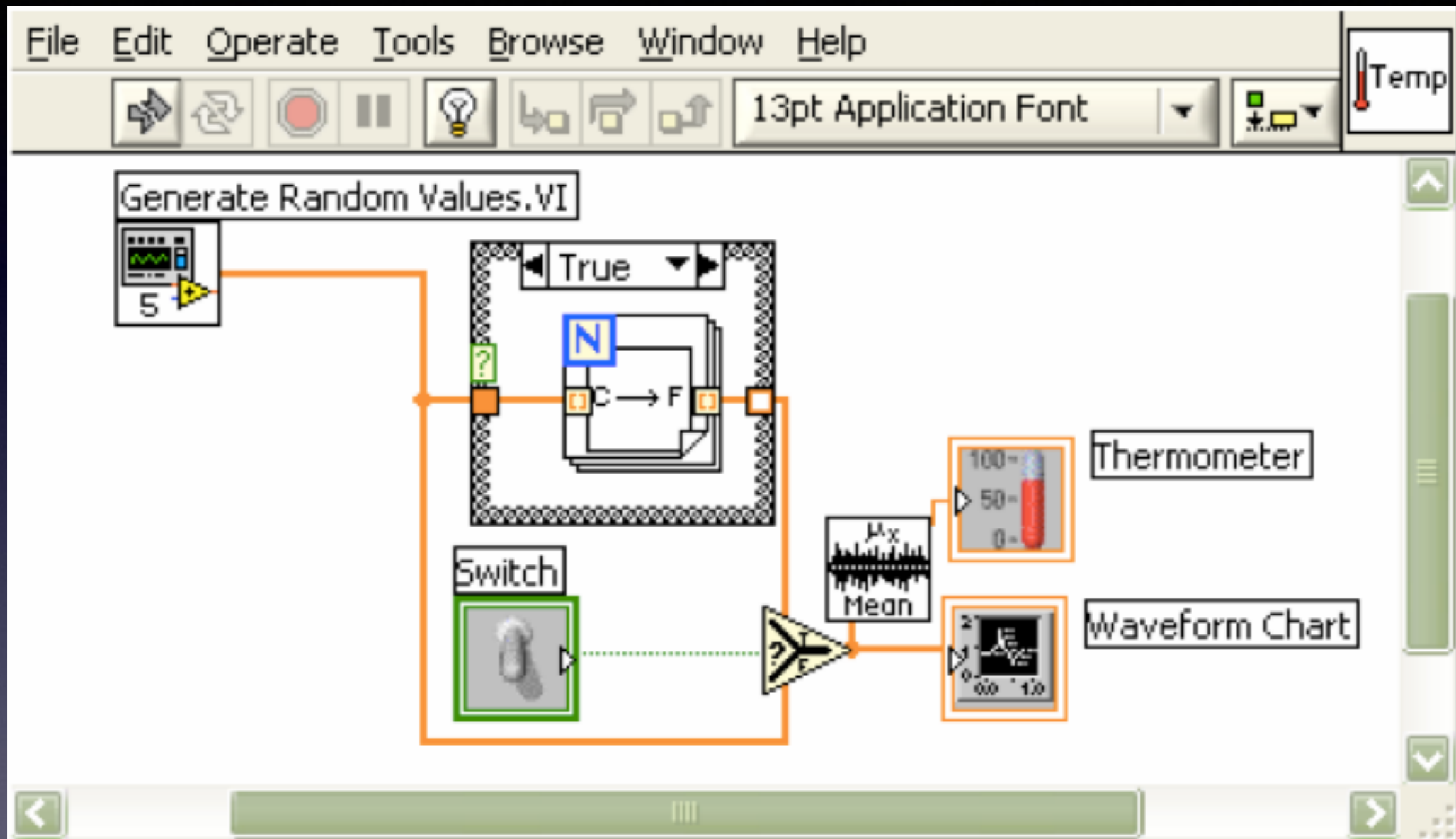


# Software options

- Lab view
- Micromanager
- DIY: SDKs - C++, Python, Visual basic



# Lab view example



# Micromanager

<http://valelab.ucsf.edu/~MM/MMwiki/>

**µManager**  
THE OPEN SOURCE MICROSCOPY SOFTWARE

OVERVIEW · DOWNLOADS · DOCUMENTATION · DEVICES · PROGRAMMING · SUPPORT · EVENTS · CREDITS · LOG IN

welcome to micro-manager!



News

- ▶ Micro-Manager Programmer Job Opening!
- ▶ ImageJ Conference
- ▶ [Open SPIM]
- ▶ Micro-Manager 1.4 Released
- ▶ Recap of Micro-Manager at 2011 ASCB meeting
- ▶ New Getting Started ScreenCast
- ▶ [Watch Micro-Manager in Action]
- ▶ Support for Nikon and Canon DSLRs

## Micro-Manager Open Source Microscopy Software



µManager is a software package for control of automated microscopes. Together with the image processing application **ImageJ**, µManager provides a comprehensive, freely available, imaging solution.

**Download** the most recent version (1.4) from our website. Also check out our **ScreenCast** for a quick tour on getting started.



µManager has a simple and clean user interface, through which it lets you execute common microscope image acquisition strategies such as time-lapses, multi-channel imaging, z-stacks, and combinations thereof. µManager works with microscopes from all four major manufacturers (Leica, Nikon, Olympus and Zeiss), most scientific-grade cameras and many scientific-grade lenses.

# Some rules of thumb

- Clean and dust free environment
- Oscilloscope and soldering iron - you will need them!
- Good tools and spare parts
- Important to think about user interface
- Important to think about continuity of the project and workflow of experiments
- Important to think about data analysis

# Justification for Bespoke Systems

- Often necessary for specific specialised problems.
- Easily optimised for several parameters, speed, sensitivity etc...
- Can provide extremely flexible systems

**BUT** think hard as it is likely to be harder, longer and more expensive than at first thought.

# How expensive is it?

**Building costs**

**Hardware ~£100-250k**

**Salaries 1-3 years (~£50-£150)**

**Total cost ~£150-350k**

**Commercial OMX system ~£750k**

# Summary

- Recap on image formation
- Fluorescent beads showing aberrations
- How deconvolution works
- Bespoke microscope building projects pro's and cons.