

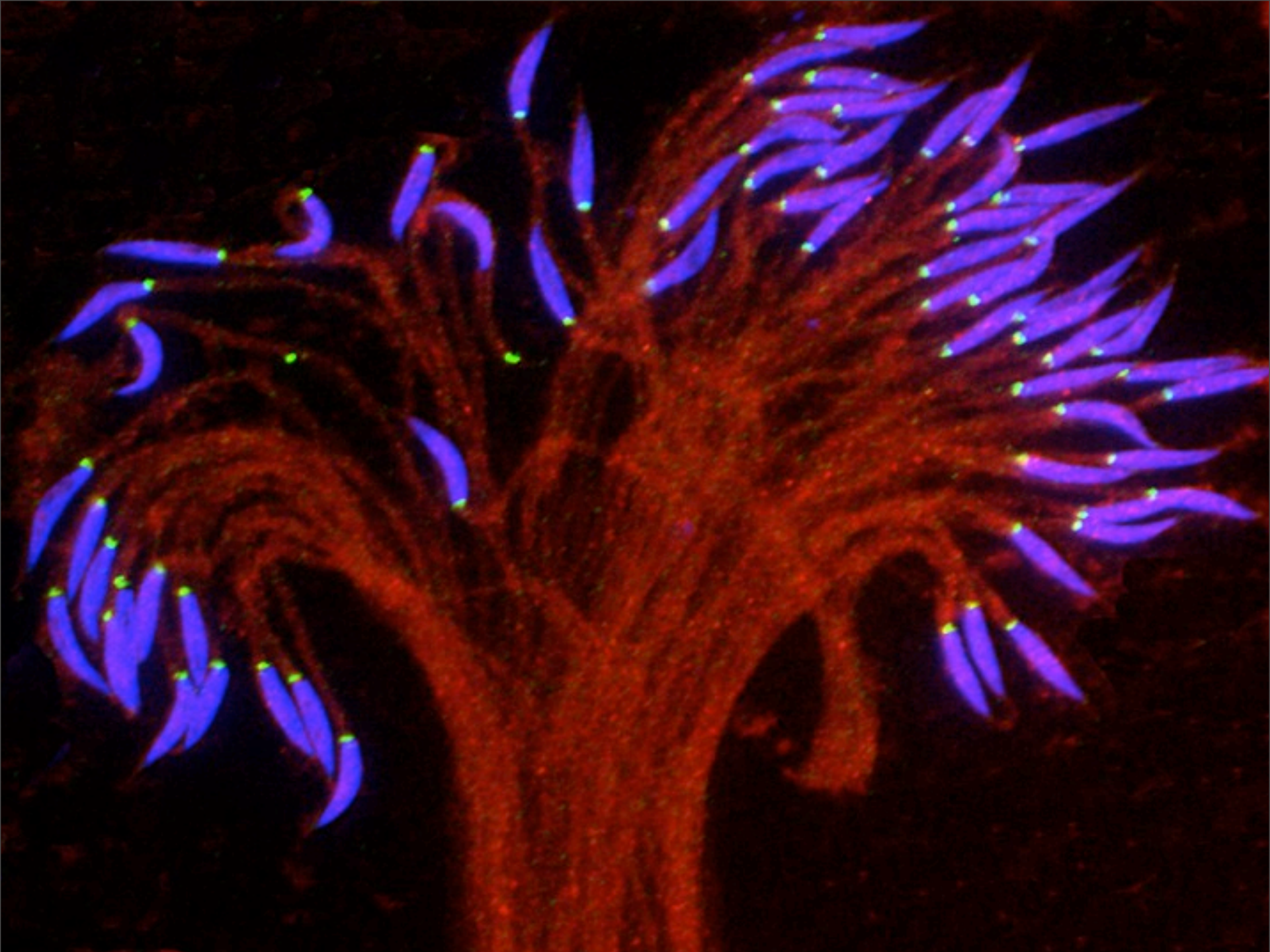
A confocal microscopy image showing several cells. The cytoskeleton is stained red, forming a dense network of filaments. The nuclei are stained green, appearing as bright spots. Purple, elongated structures are visible within the cells, possibly representing specific organelles or protein complexes. The background is dark, highlighting the cellular structures.

# Confocal Microscopy

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The Dunn School of Pathology

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Monday, 12 March 12

# Outline of Talk

1. Confocal microscopy - some history

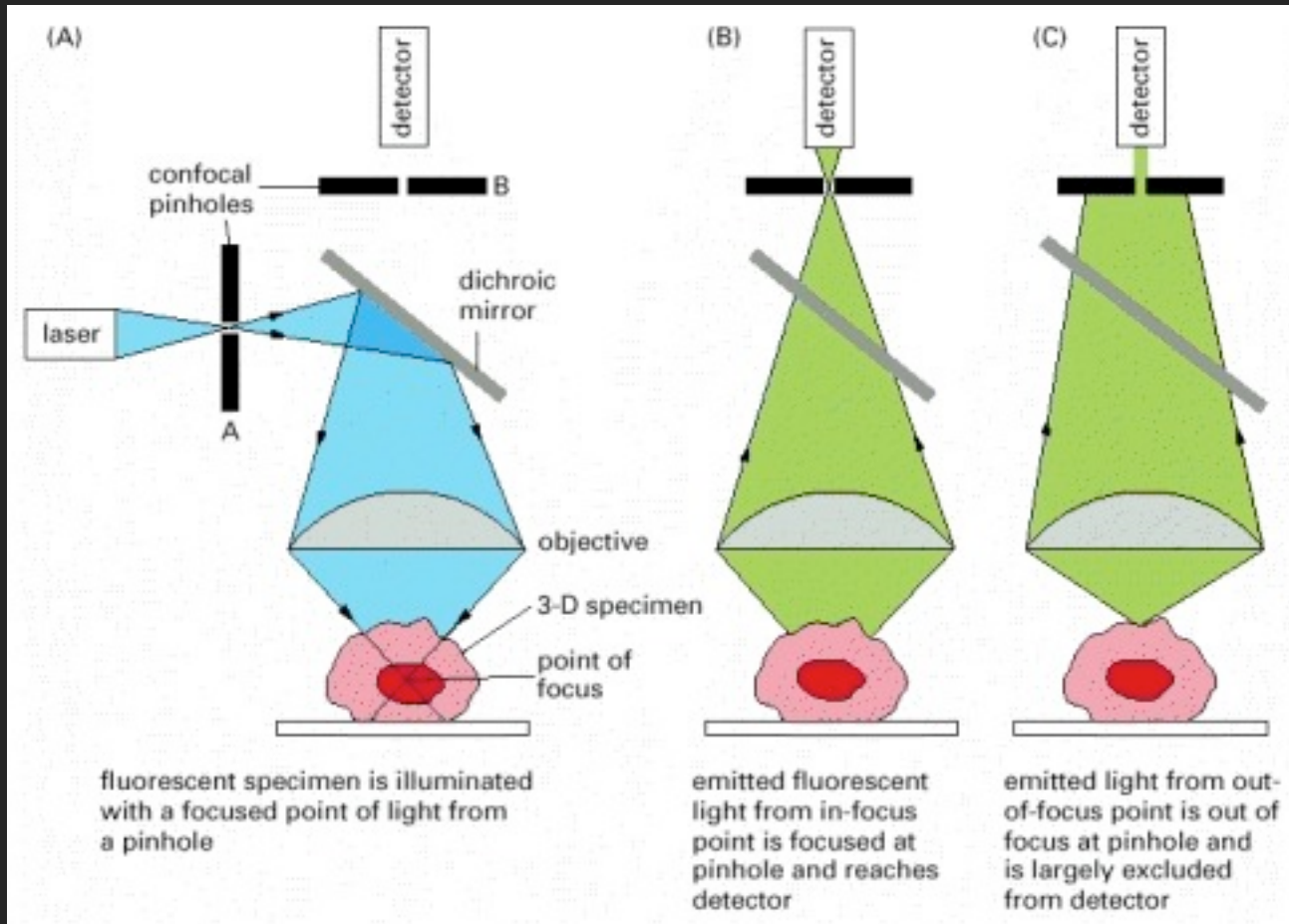
2. Laser Scanning Confocal Microscopy (LSCM)

3. Spinning Disk Confocal Microscopy

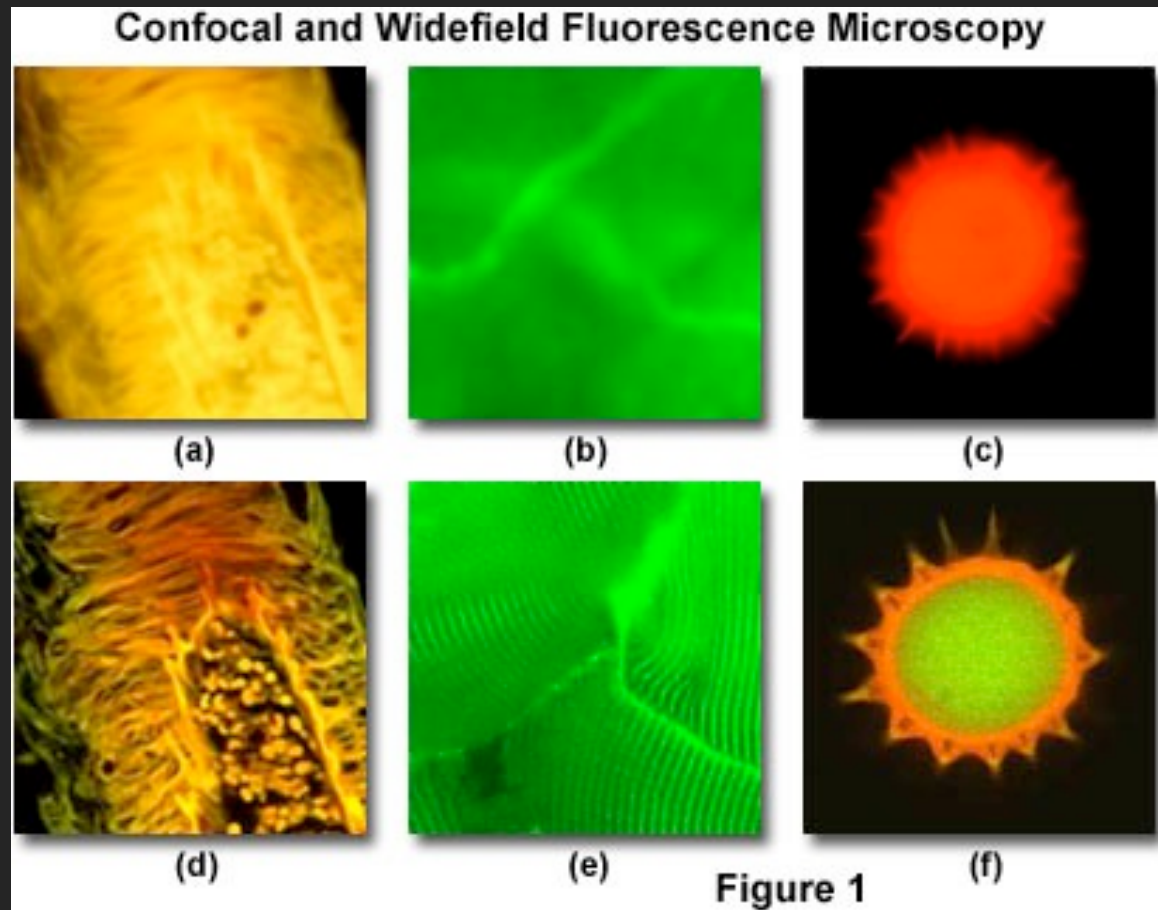
4. Widefield vs. LSCM vs. Spinning Disc

Murray et al., Evaluating performance in 3D  
fluorescence microscopy. *J. Microscopy* (2007)

# The principle of the confocal microscope



# Comparison of Confocal and Widefield Microscopy



# Short History of the Confocal Microscope

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Confocal “concept” patented by Marvin Minsky in 1957.

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Brakenhoff, Stelzer developed “stage” scanning confocal in late 1970s.

White, Amos and Wilson developed MRC 500 point scanning confocal - marketed commercially in 1987.

Amos and White, Biol. Cell 2003

# Simplified View of a “Point-Scanning” Confocal

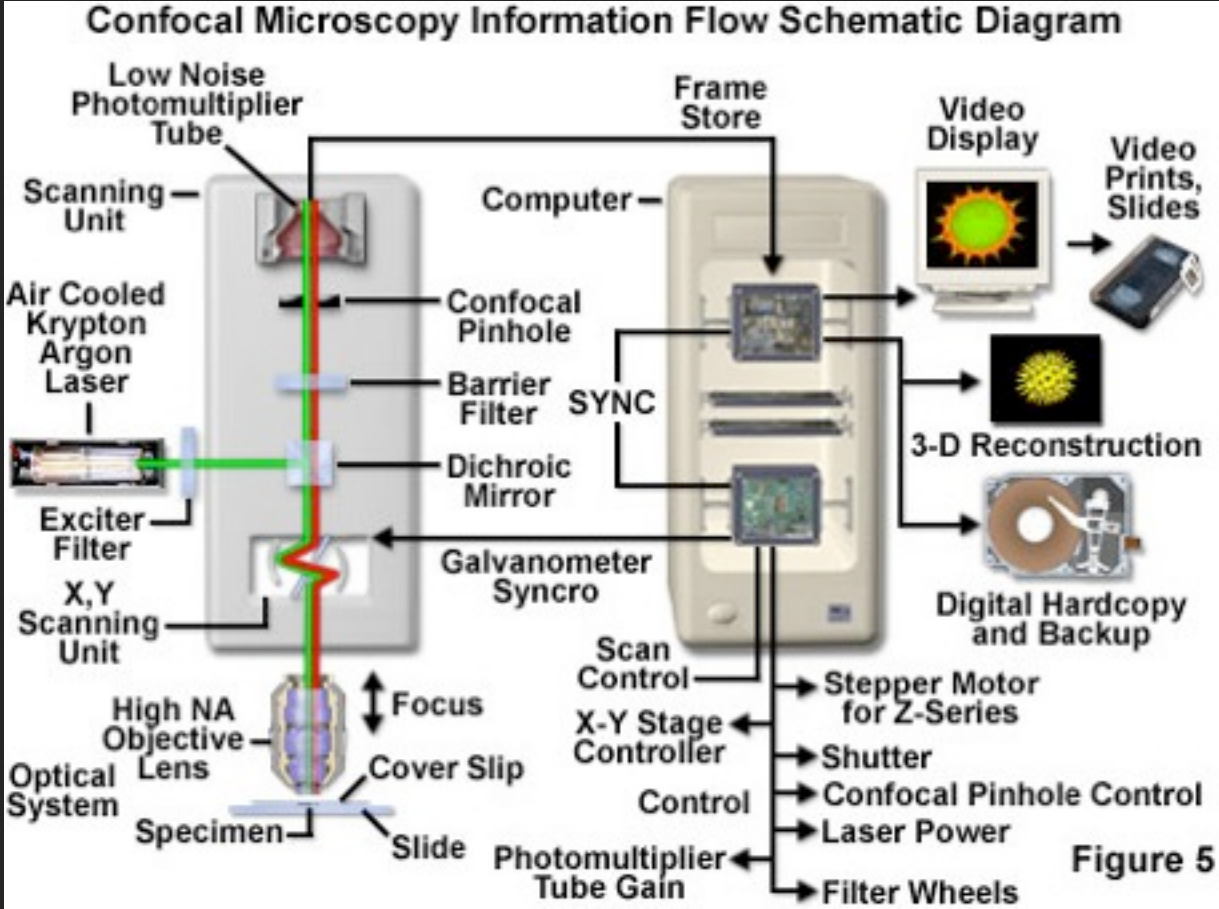
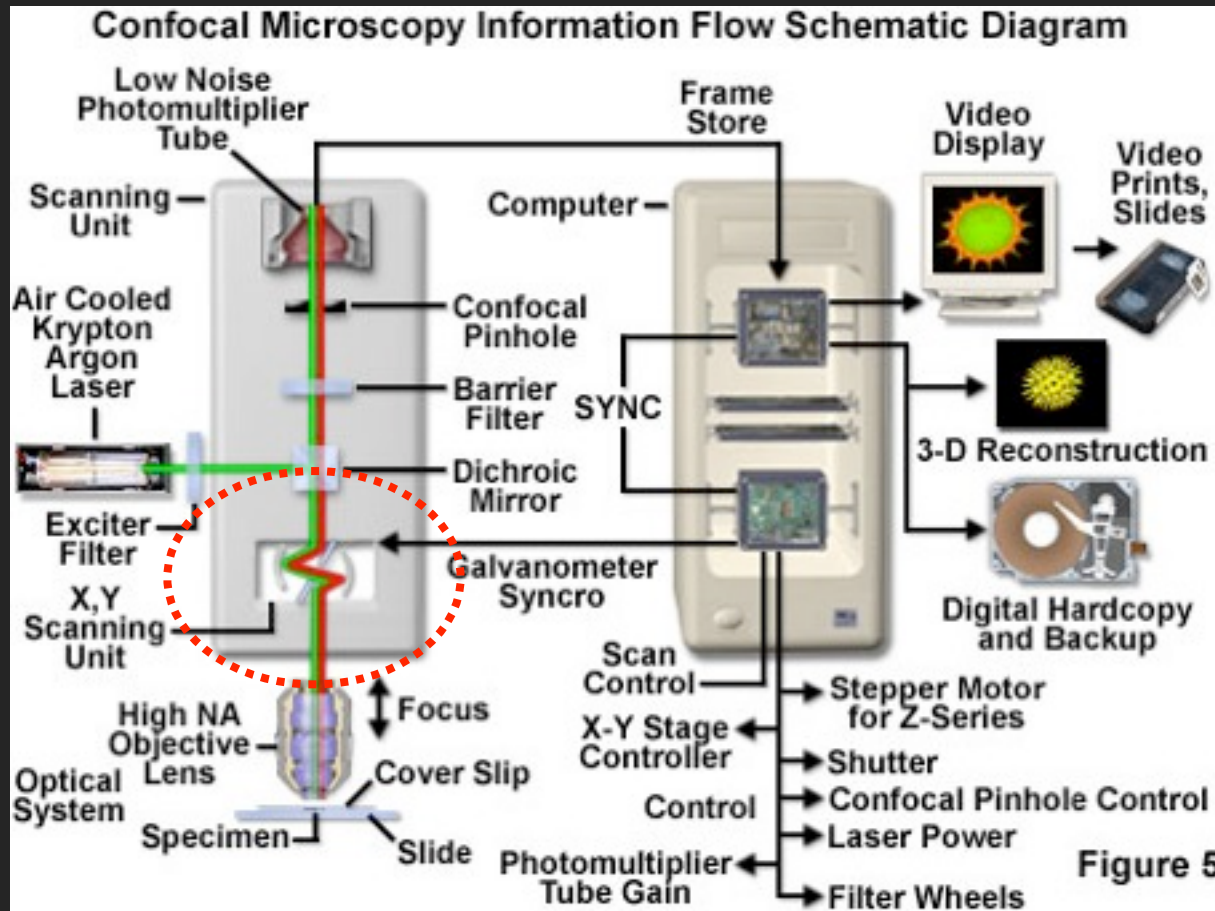
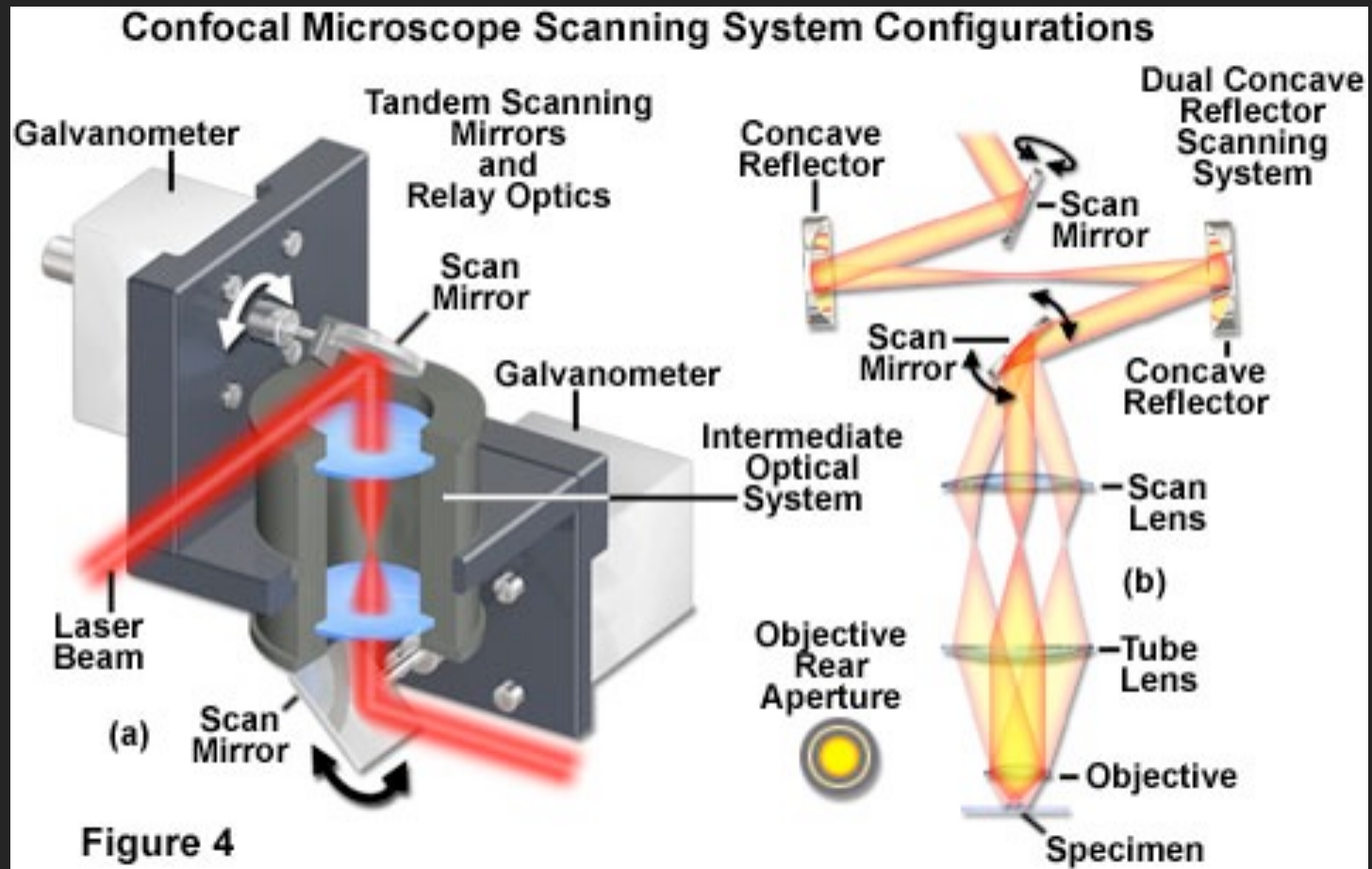


Figure 5

# Simplified View of a “Point-Scanning” Confocal



# The Galvanometer/Mirror Scanning System



<http://www.olympusmicro.com/primer/java/galvanometerscanning/index.html>

# Simplified View of a “Point-Scanning” Confocal

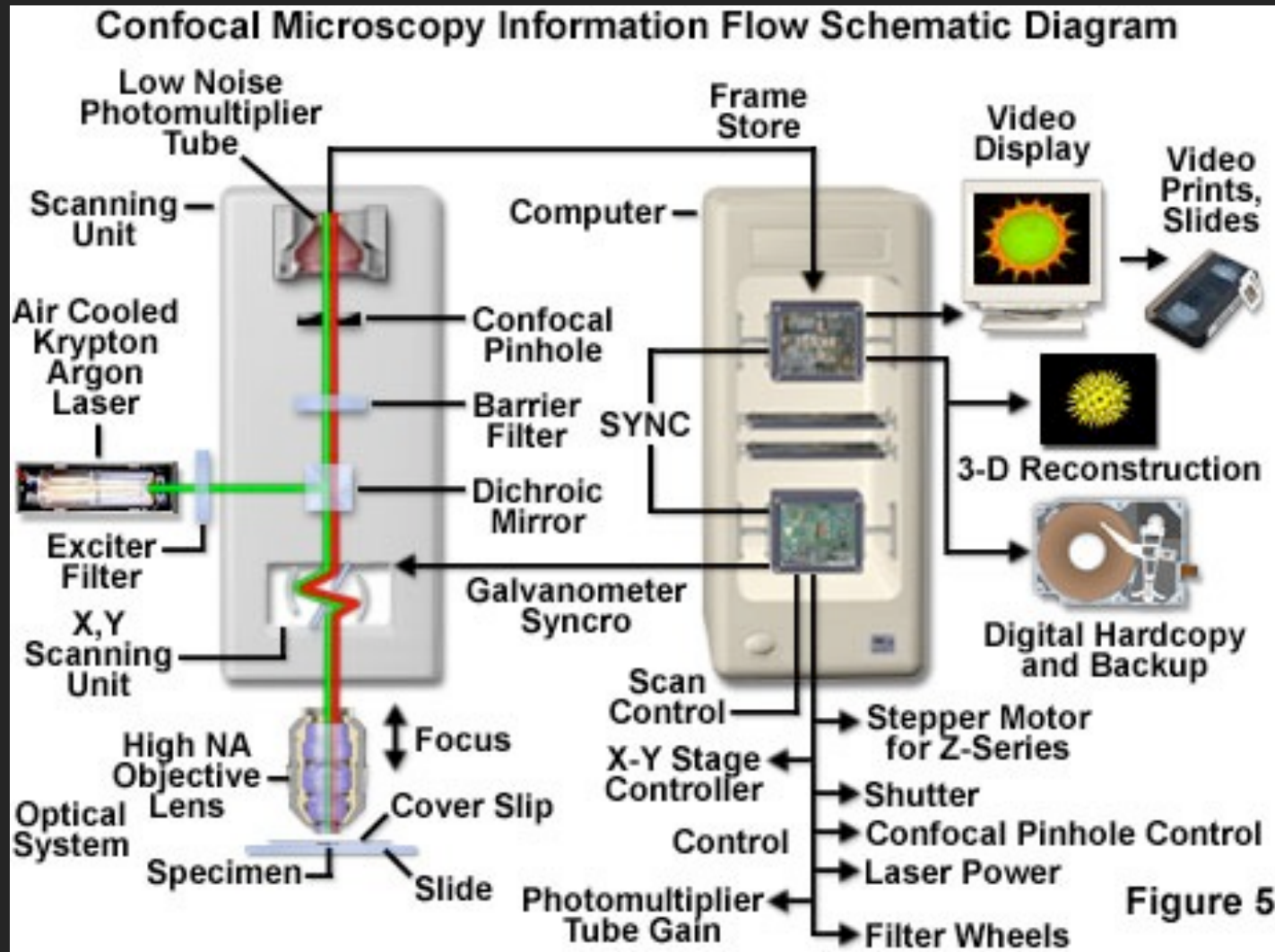


Figure 5

# Simplified View of a “Point-Scanning” Confocal

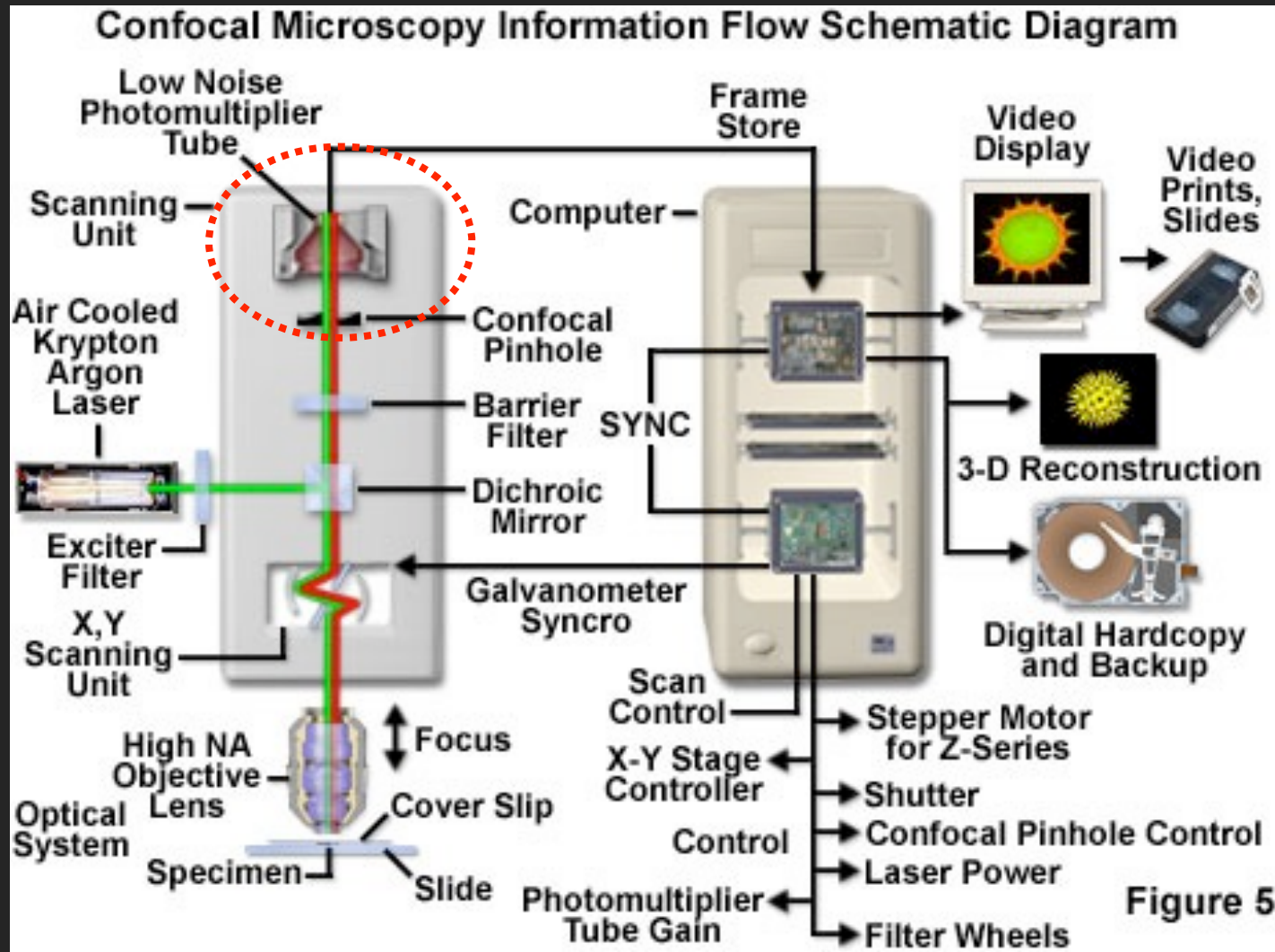
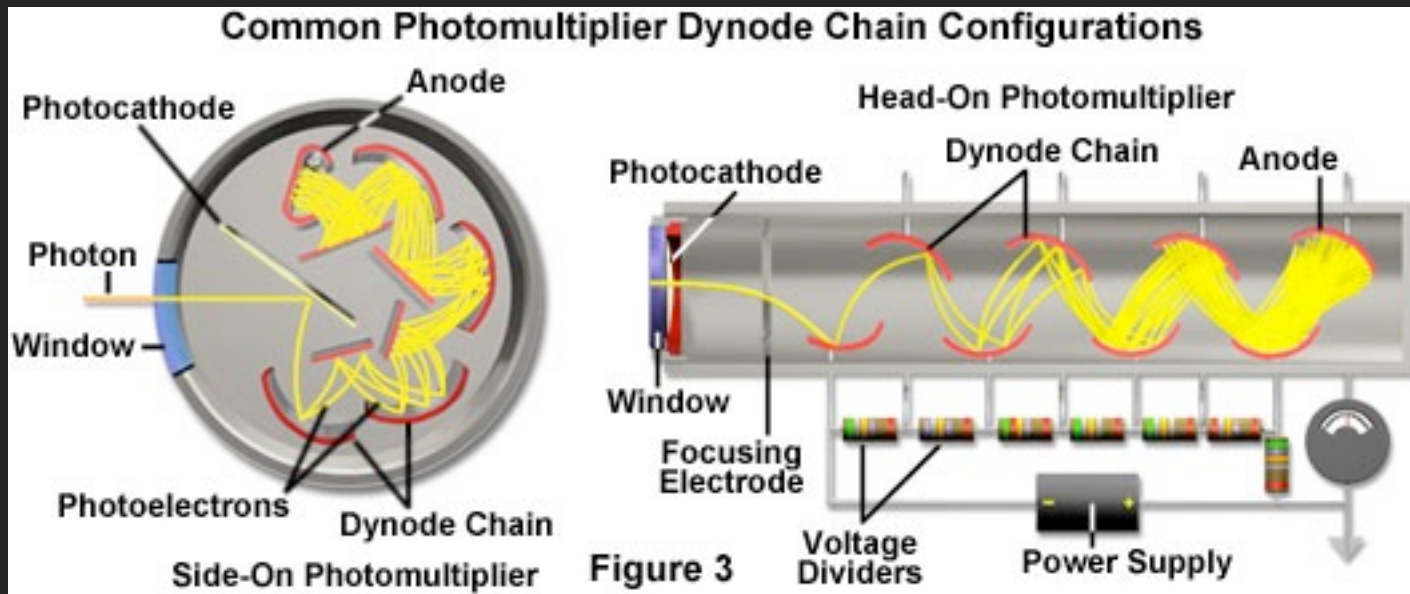


Figure 5

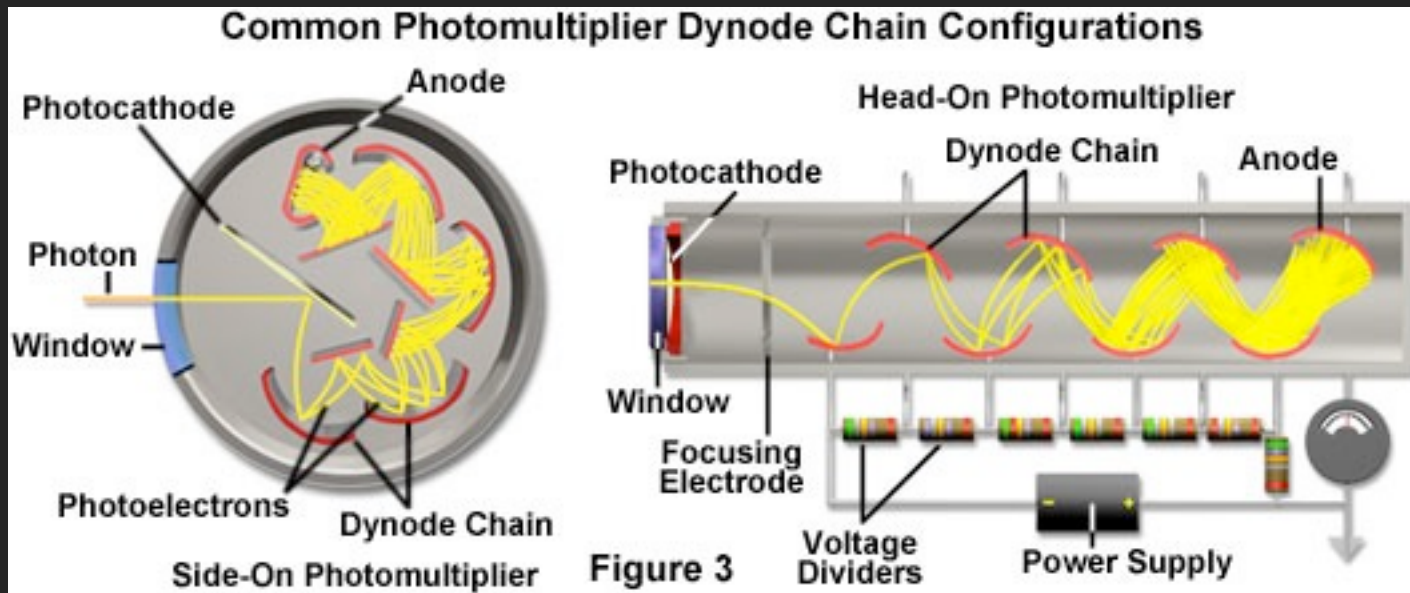
# The Photomultiplier (PMT)



<http://www.olympusmicro.com/primer/java/channelpmt/index.html>

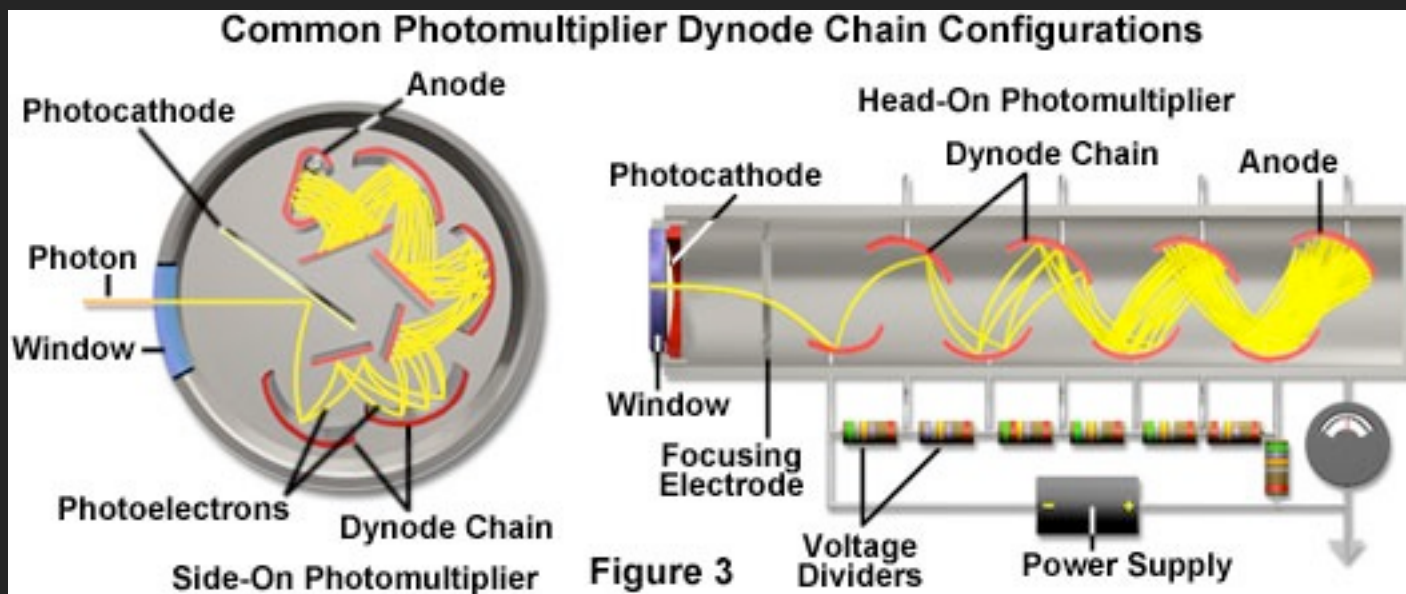


# The Photomultiplier (PMT)



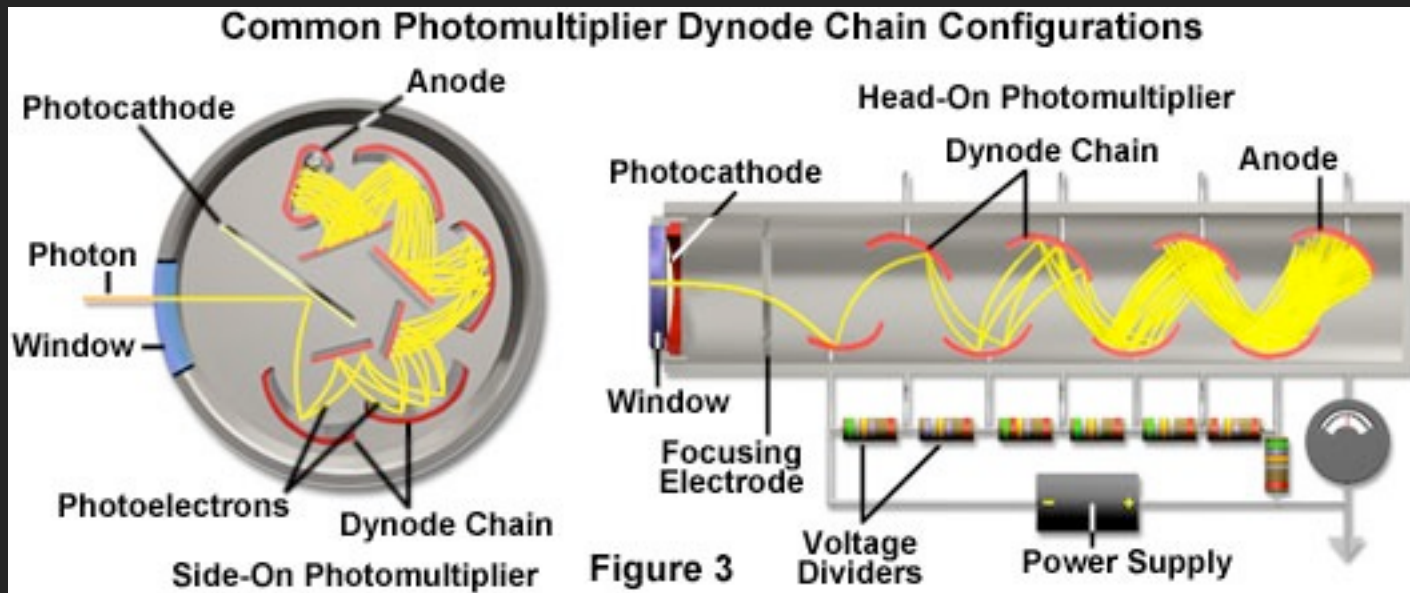
Very Low Noise (even without too much cooling)  
Very Rapid Response  
Huge Potential for Signal Amplification ( $\sim 1 \times 10^7$ )

# The Photomultiplier (PMT)



<http://www.olympusmicro.com/primer/java/channelpmt/index.html>

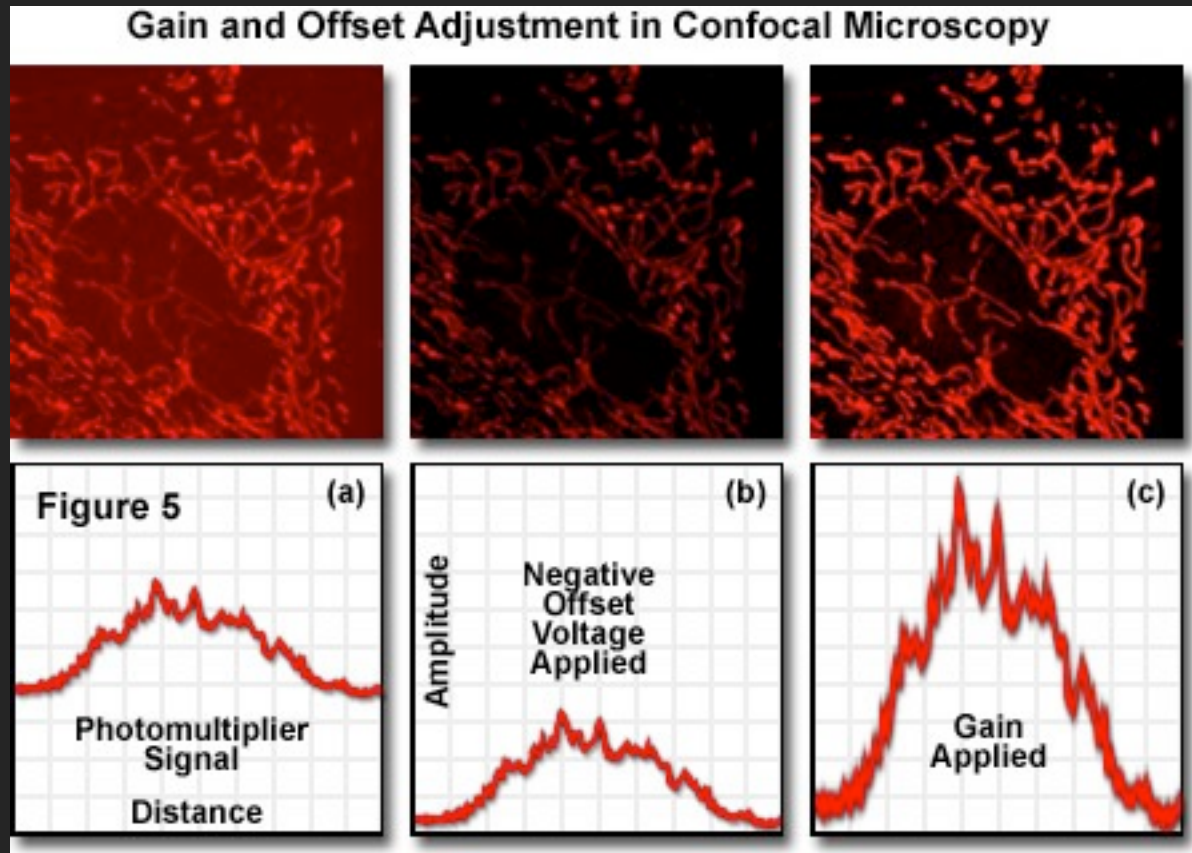
# The Photomultiplier (PMT)



Low Quantum Efficiency (QE) (~15-40%)  
GaAsP!

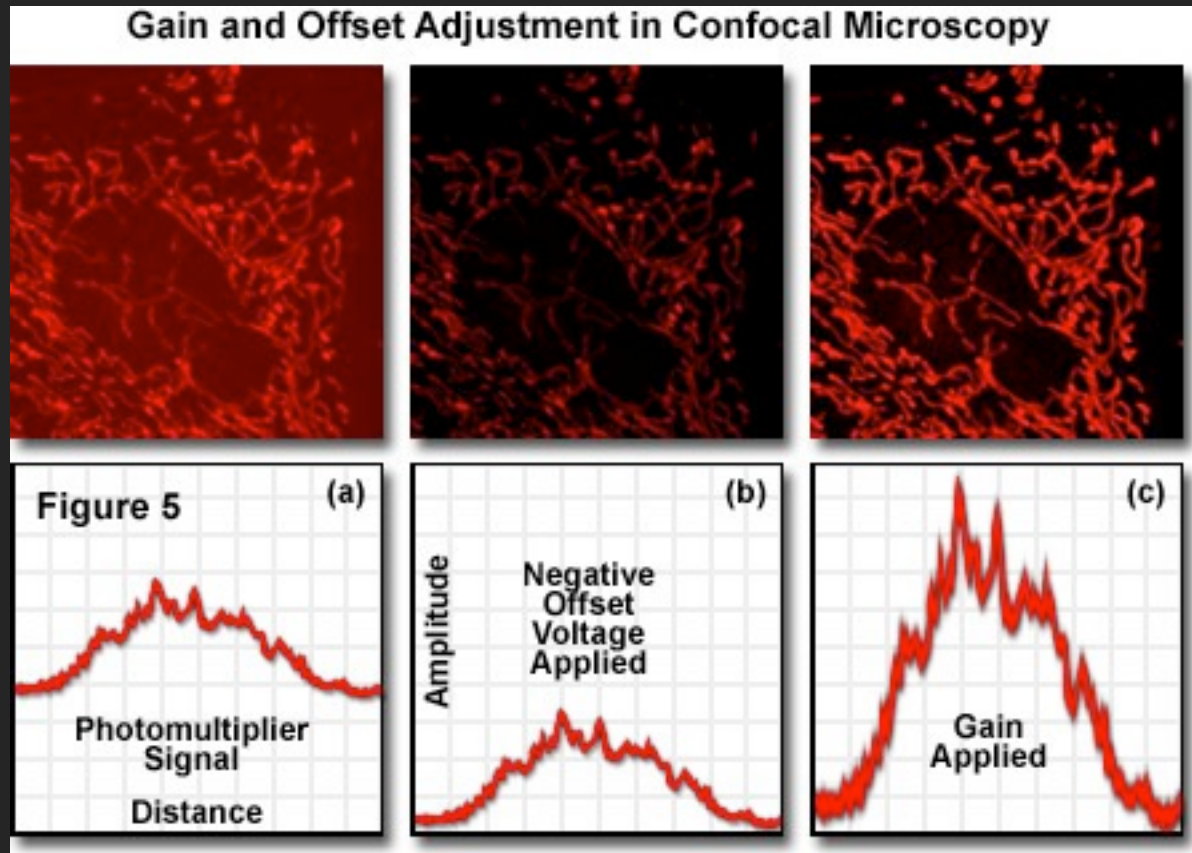
<http://www.olympusmicro.com/primer/java/channelpmt/index.html>

# Adjusting Offset and Gain of the PMT



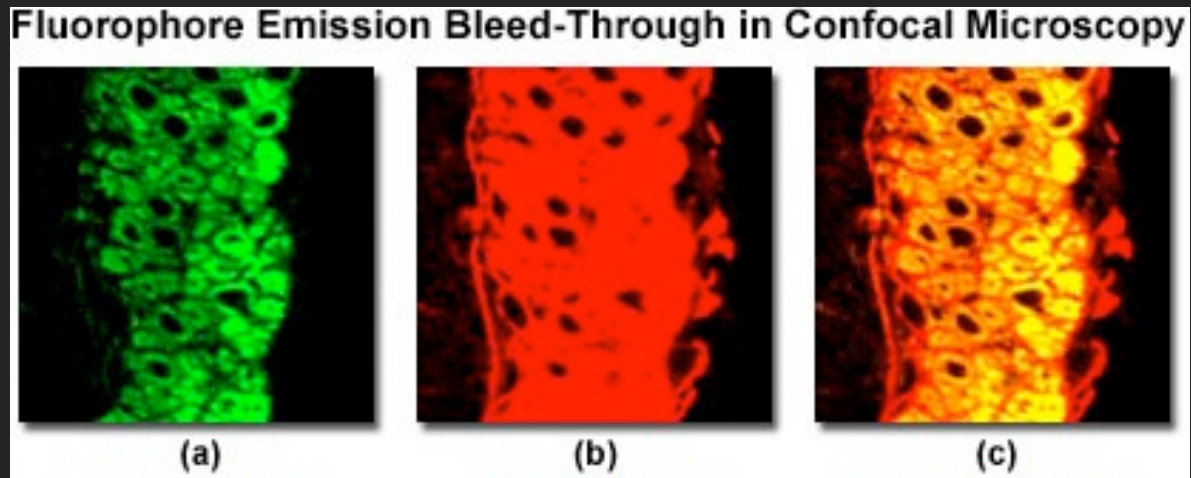
<http://www.olympusmicro.com/primer/java/confocalsimulator/index.html>

# Adjusting Offset and Gain of the PMT



**Beware - this is how your image will be saved!**

# The Problem of “Bleedthrough”



<http://www.olympusmicro.com/primer/java/confocalsimulator/index.html>

# The Problem of “Bleedthrough”

Fluorophore Emission Bleed-Through in Confocal Microscopy

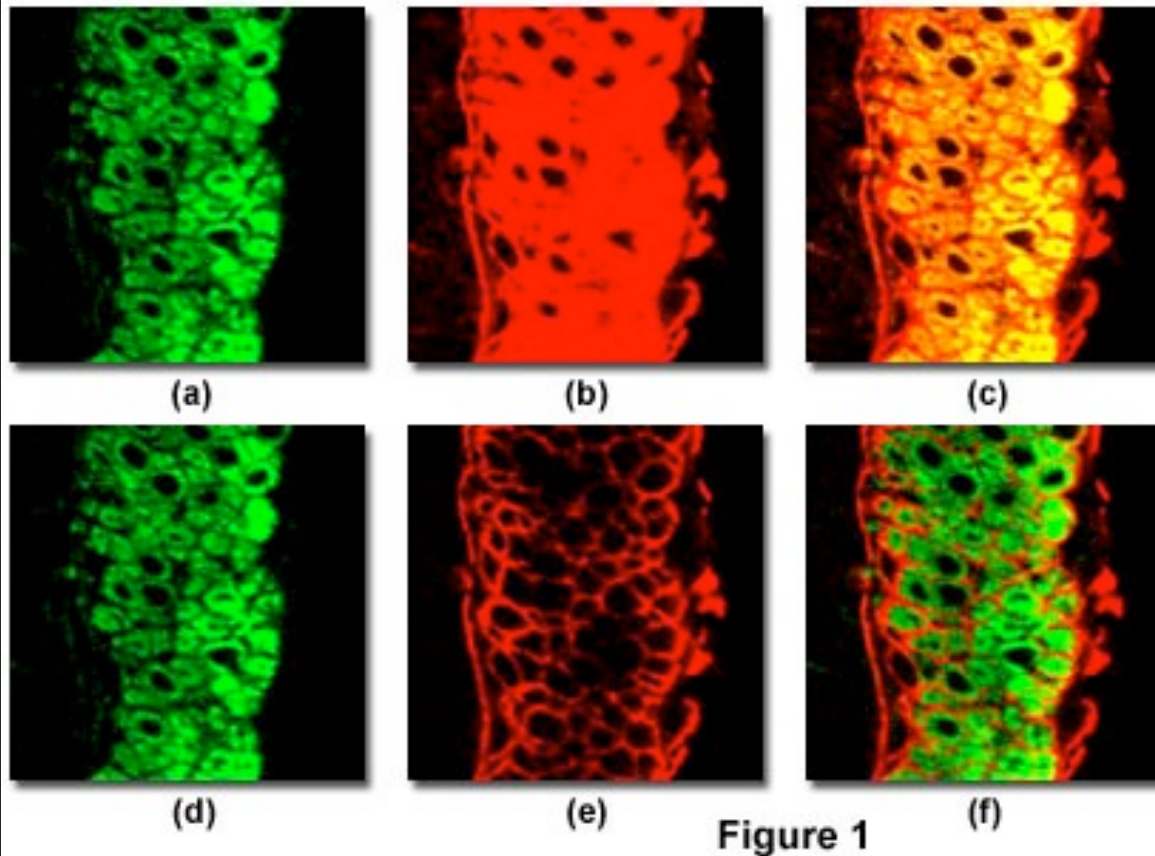
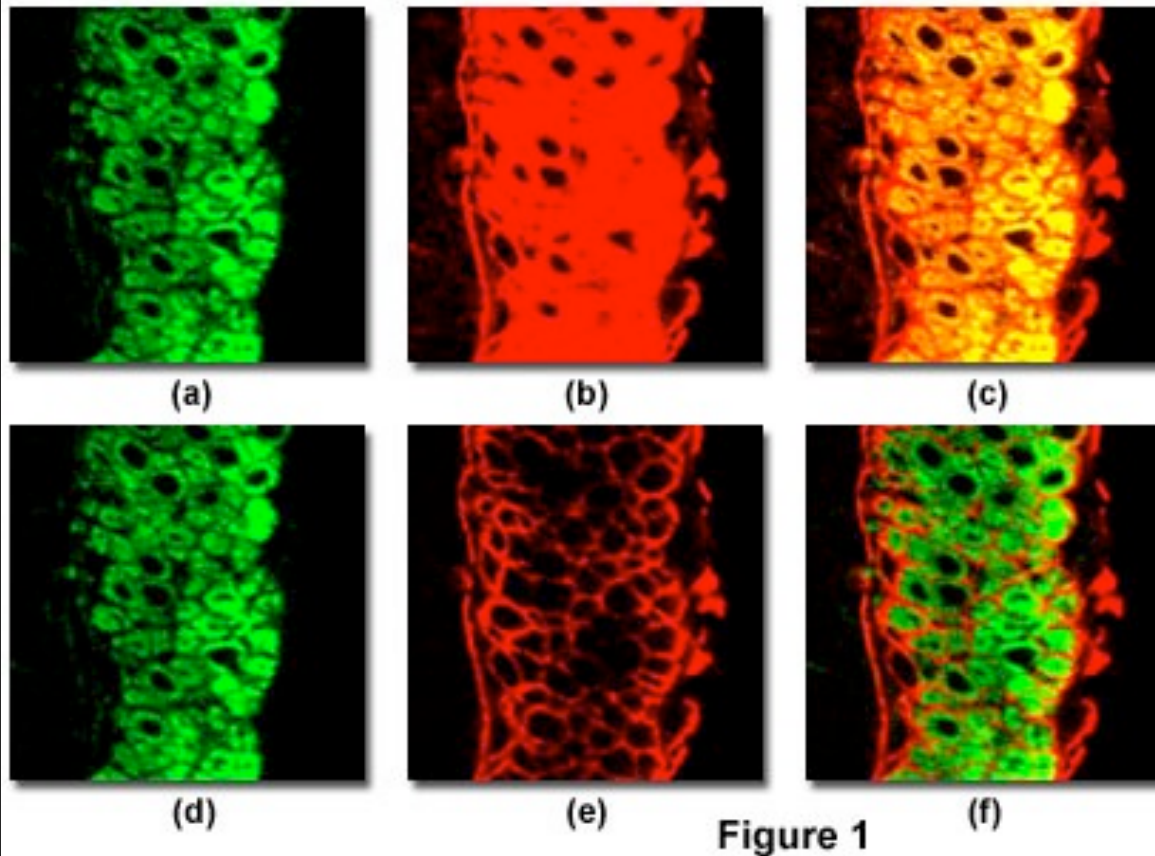


Figure 1

# The Problem of “Bleedthrough”

Fluorophore Emission Bleed-Through in Confocal Microscopy



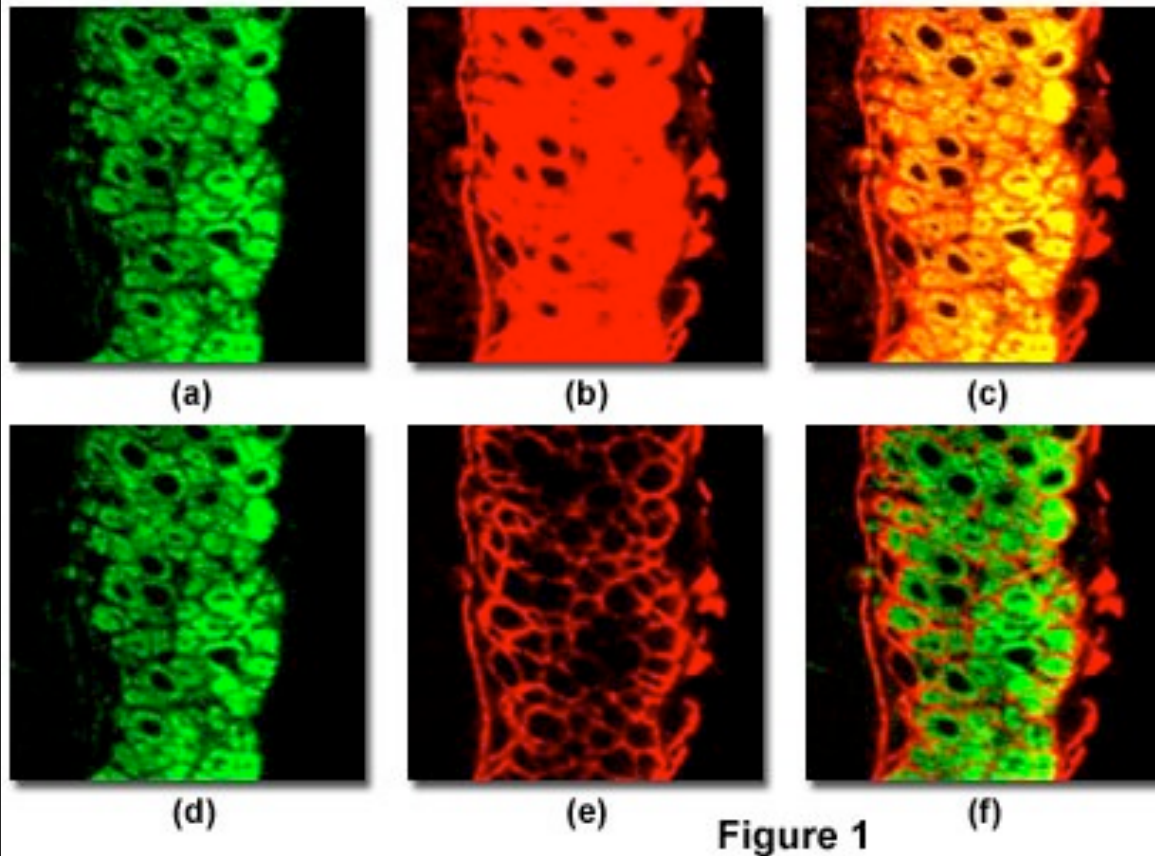
Sequential scanning

<http://www.olympusmicro.com/primer/java/confocalsimulator/index.html>



# The Problem of “Bleedthrough”

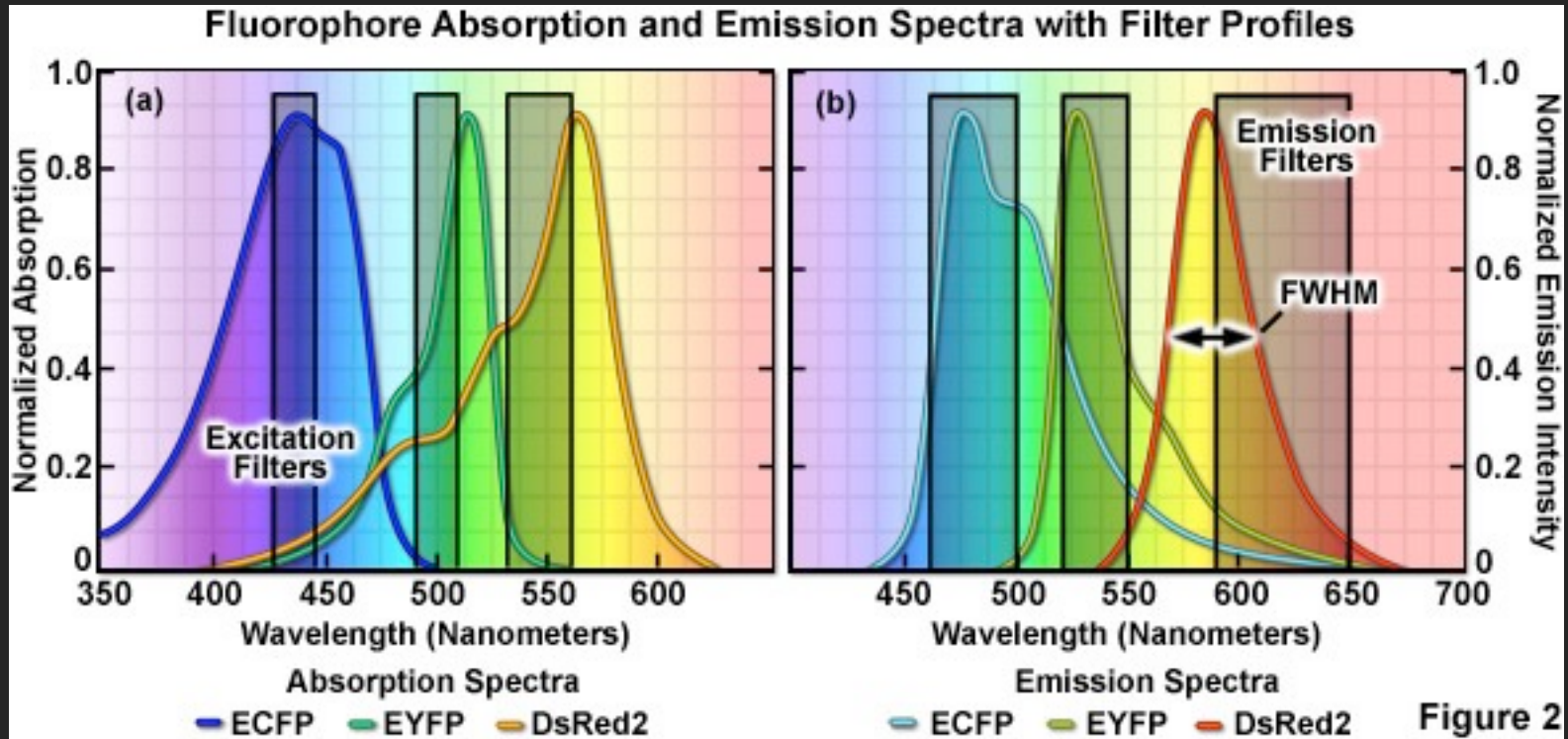
Fluorophore Emission Bleed-Through in Confocal Microscopy



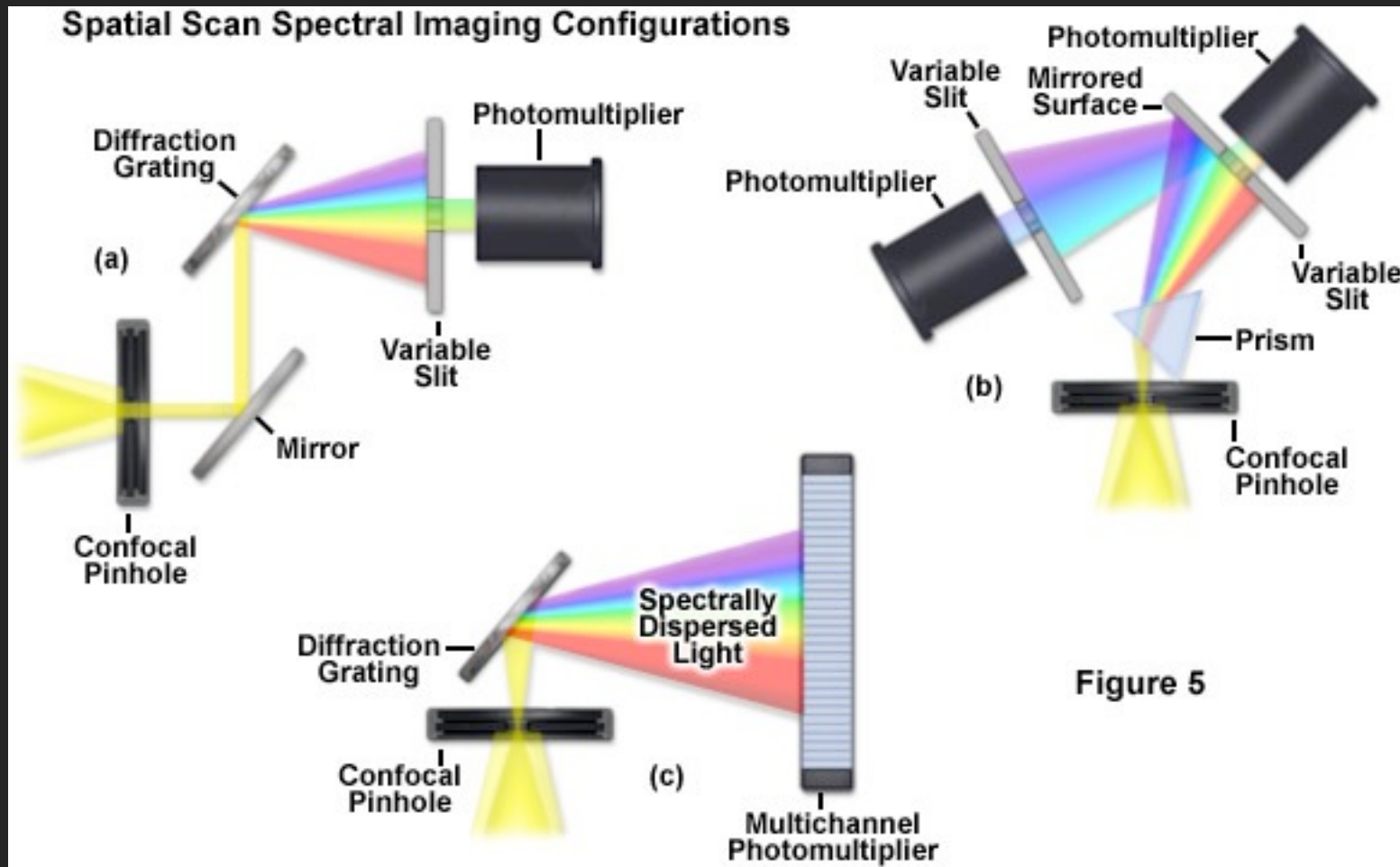
Sequential scanning  
- but a problem for live cells

<http://www.olympusmicro.com/primer/java/confocalsimulator/index.html>

# Minimising “Bleedthrough”



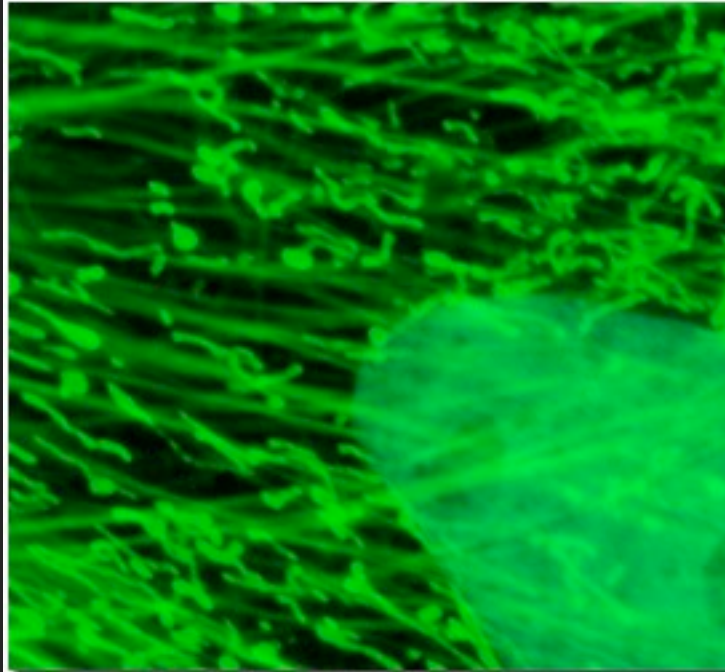
# Spectral Unmixing



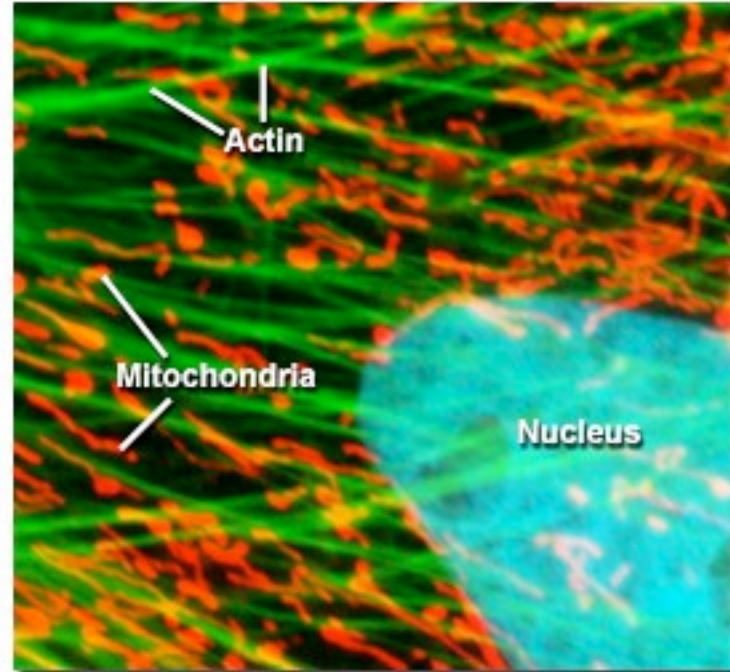
<http://zeiss-campus.magnet.fsu.edu/tutorials/spectralimaging/linearunmixing/index.html>

# Spectral Unmixing

Spectral Imaging and Linear Unmixing of Fixed Cells with Synthetic Dyes



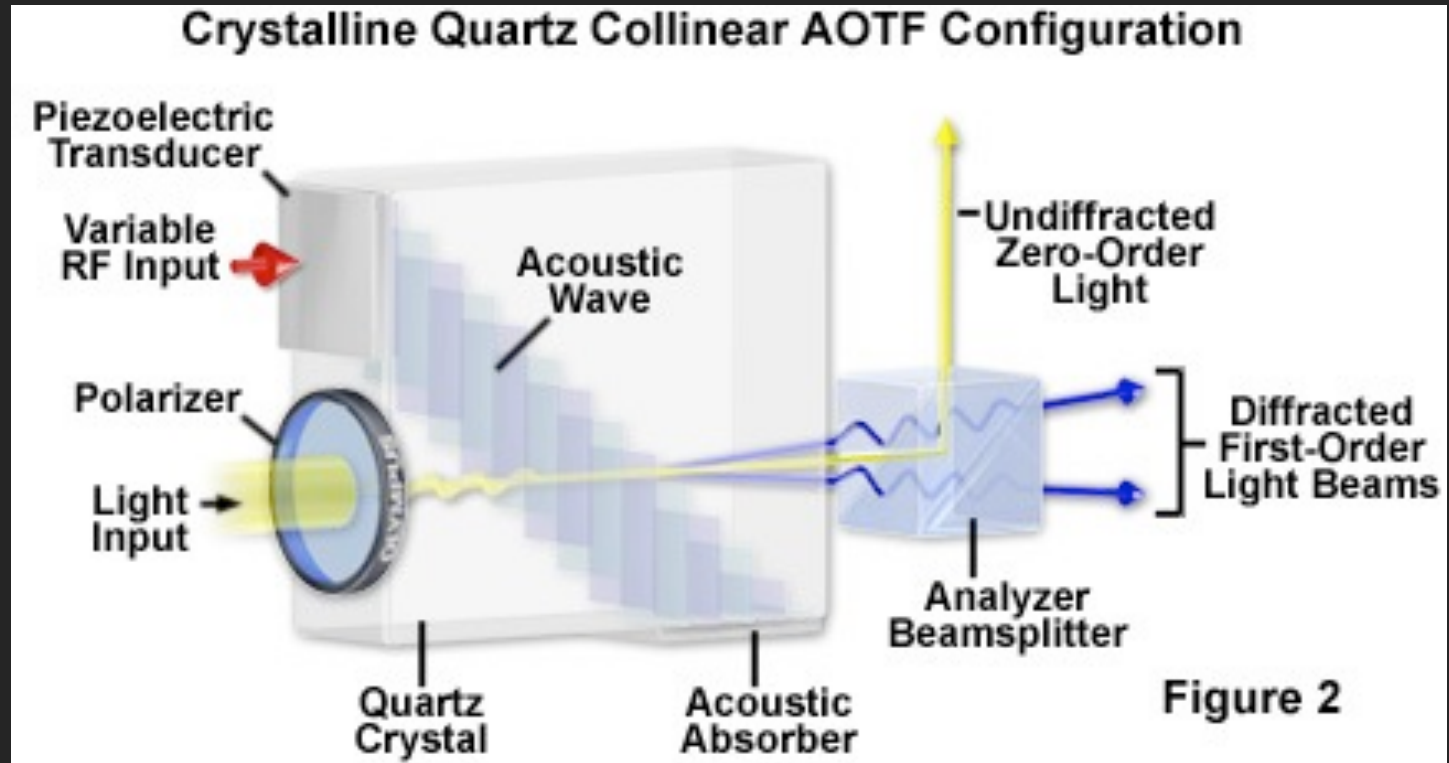
(a)



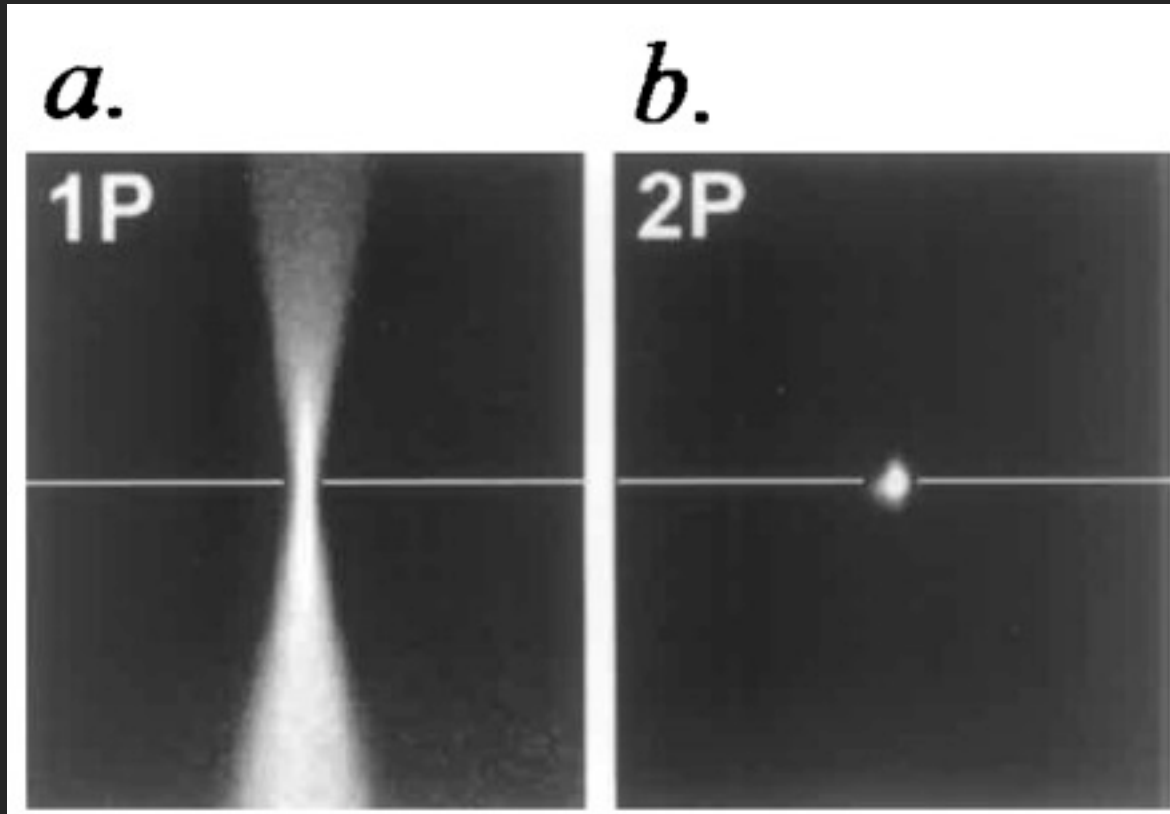
(b)

Figure 1

# Acousto Optic Tunable Filter (AOTF)

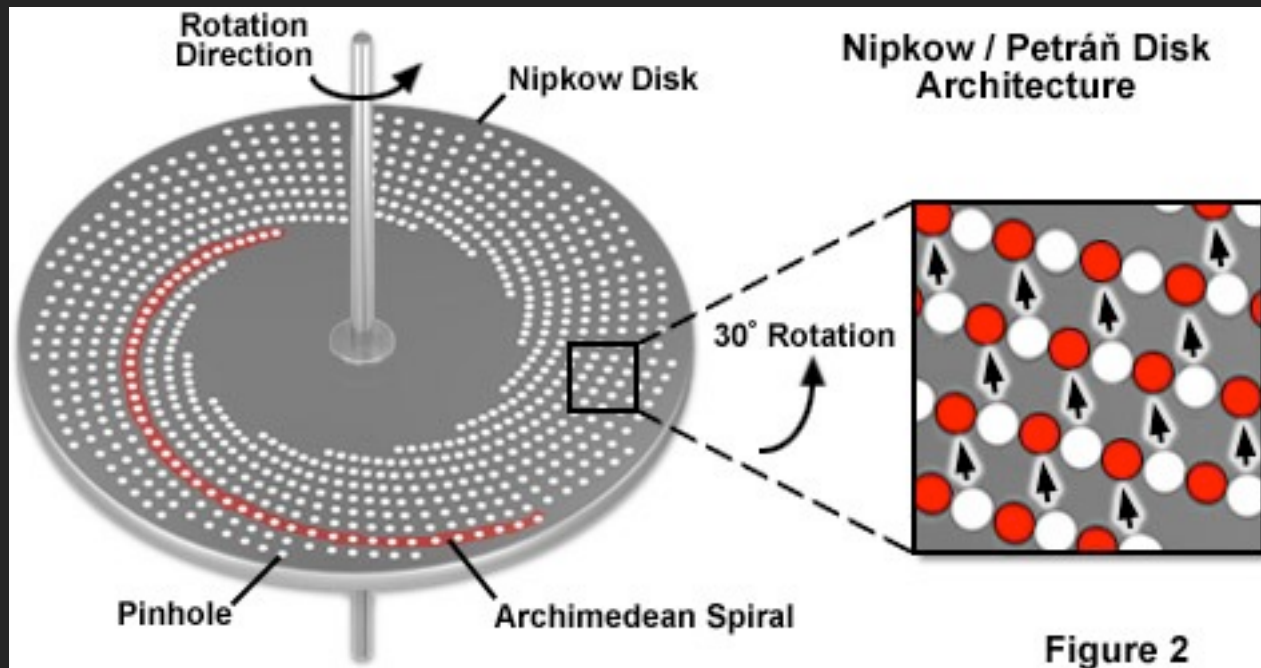


# Multiphoton Confocal Microscopy



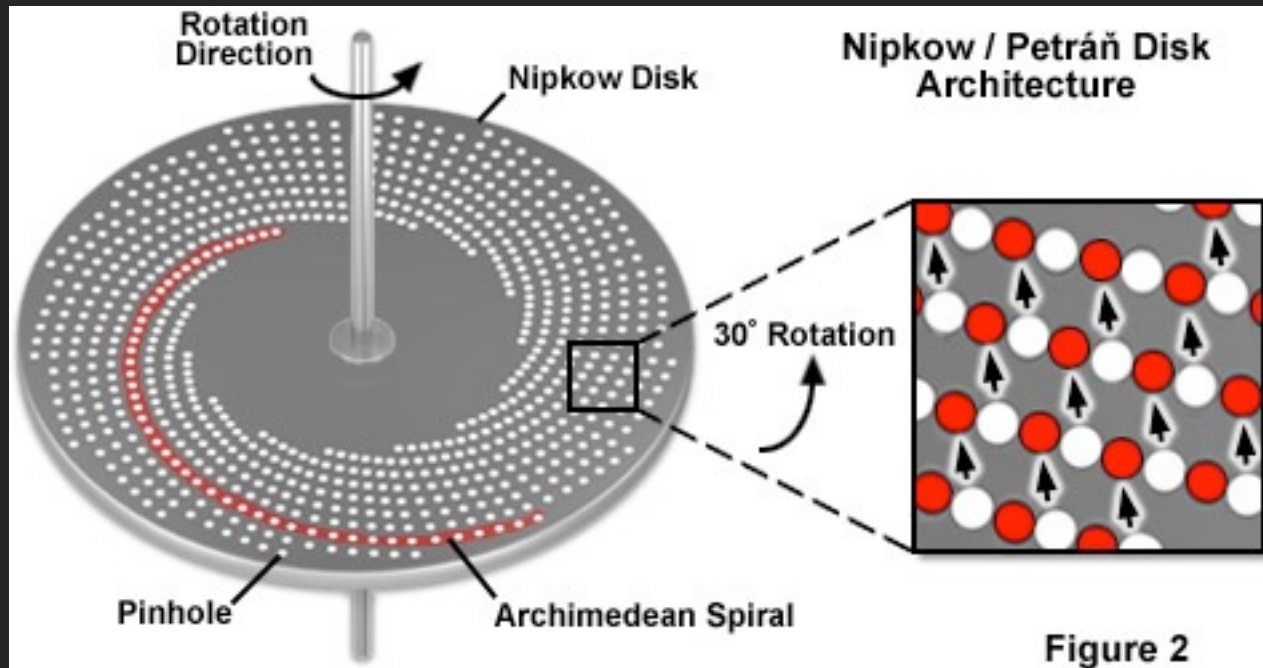
<http://www.olympusmicro.com/primer/java/multiphoton/jablonski/index.html>

# Spinning Disk Confocal Microscopy



**The Nipkow Disk**  
Paul Nipkow, 1884  
Eggar and Petran, 1967

# Spinning Disk Confocal Microscopy

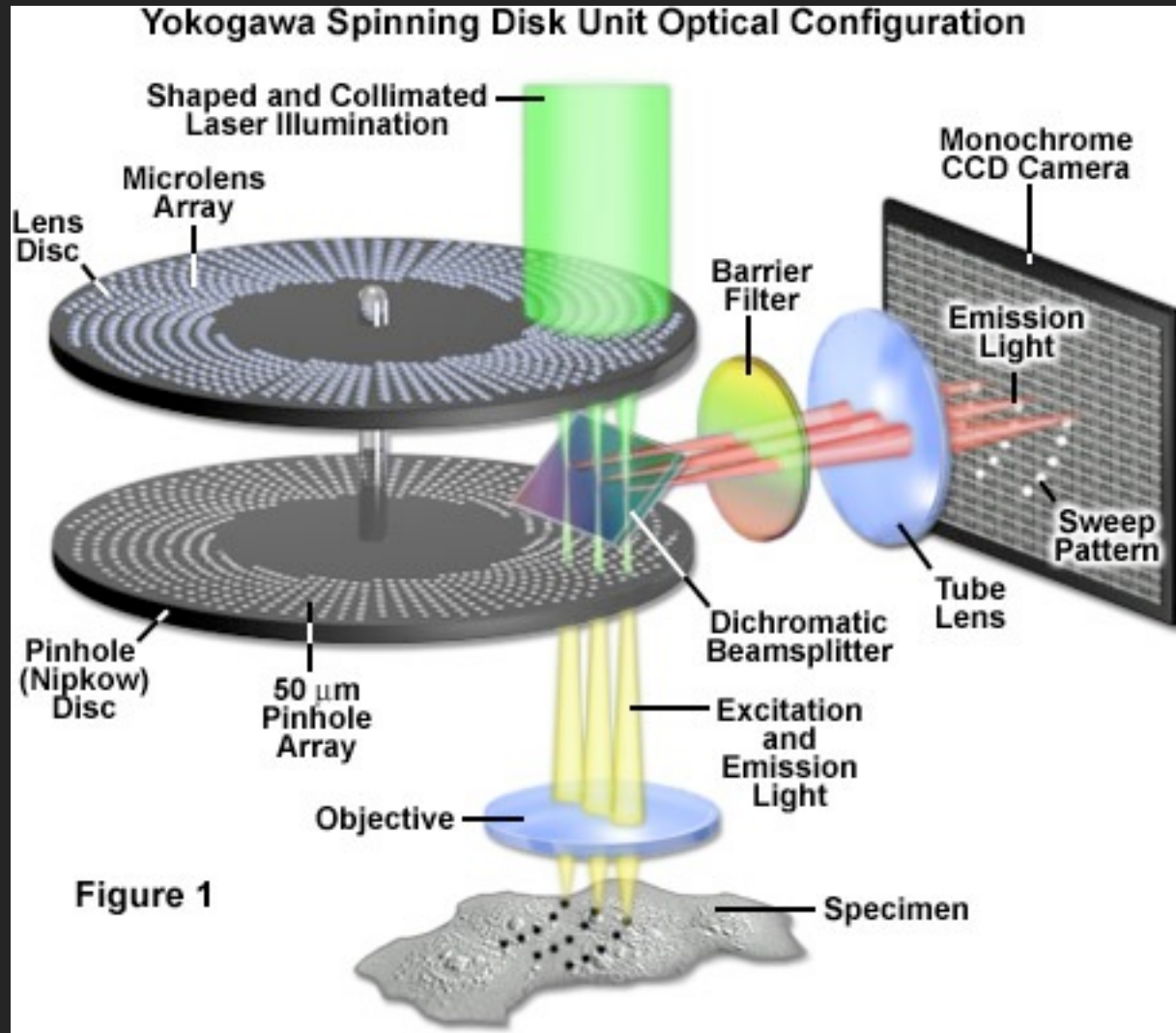


## Constant Battle:

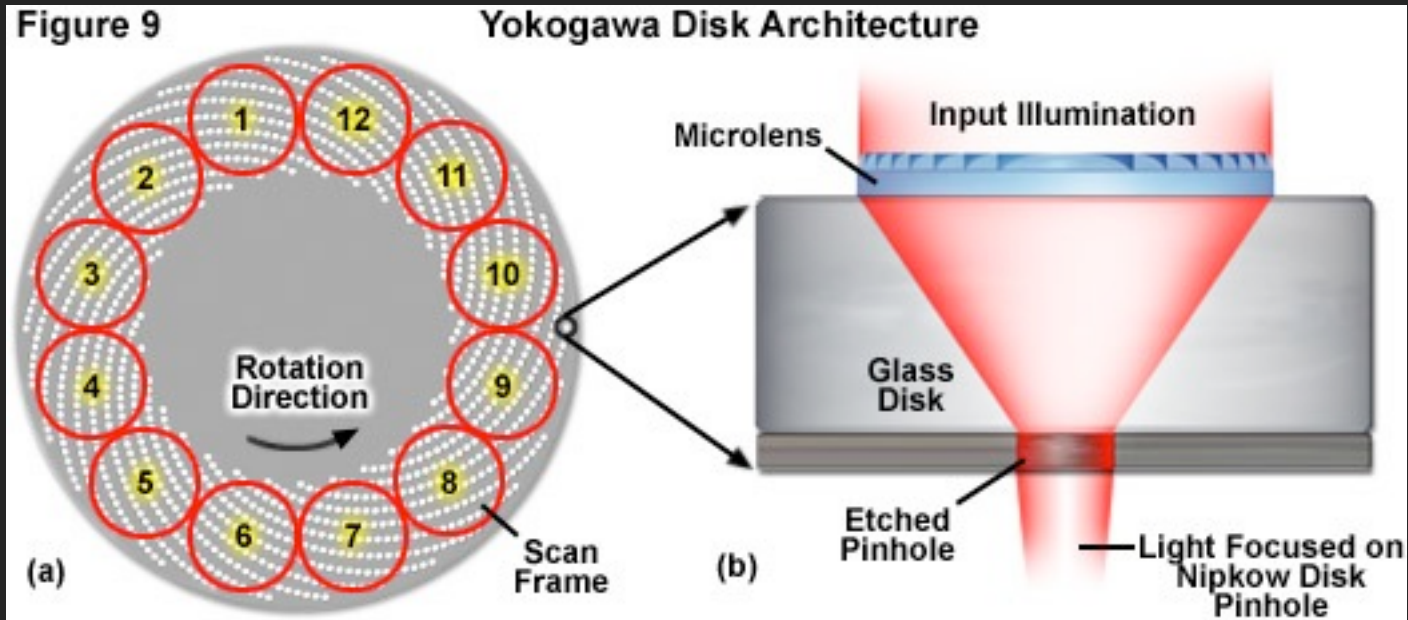
Larger pinholes - brighter image, but less “confocal”  
Smaller spacing - more light gets through, but “crosstalk”



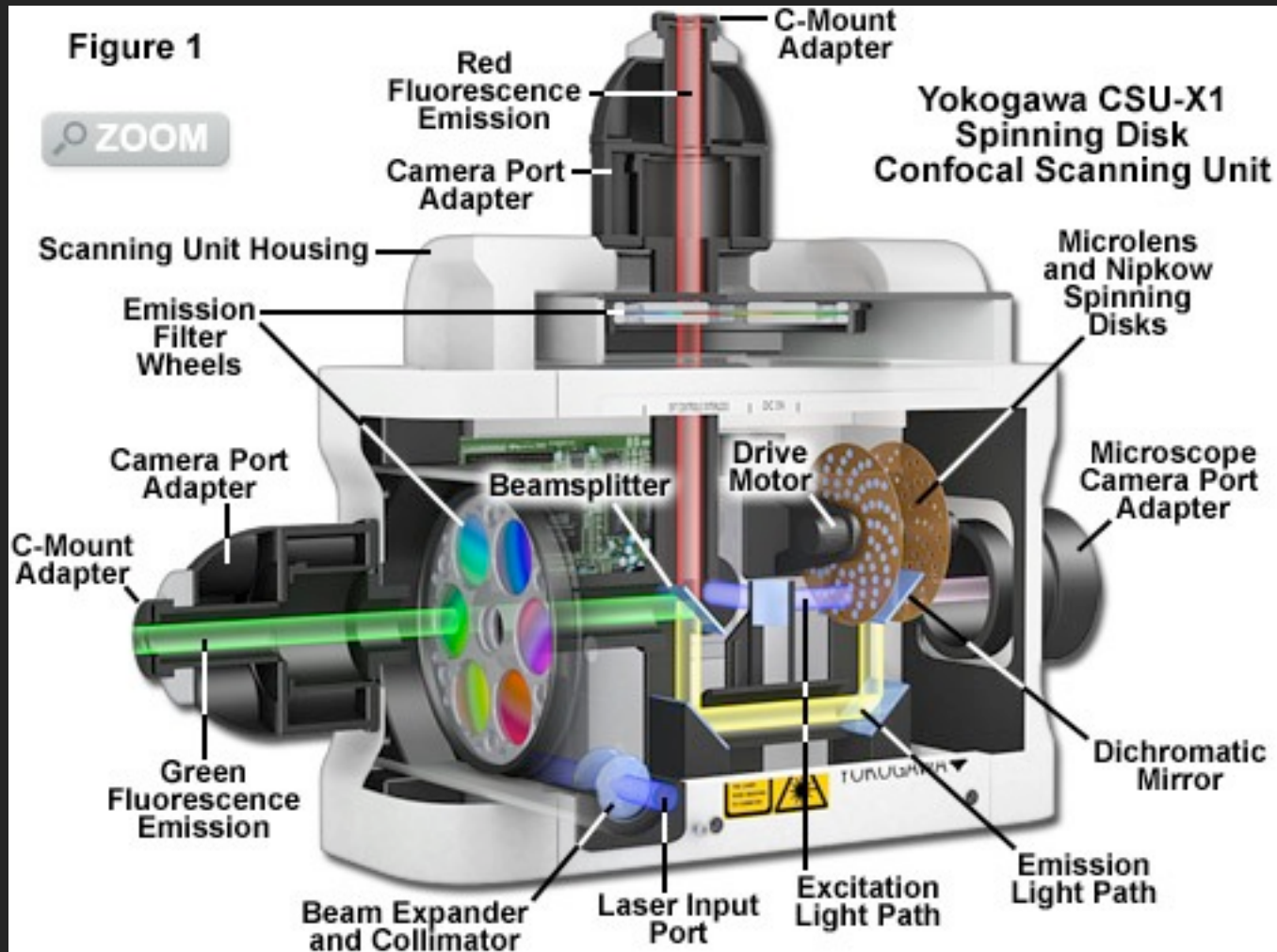
# The Yokogawa Spinning Disk



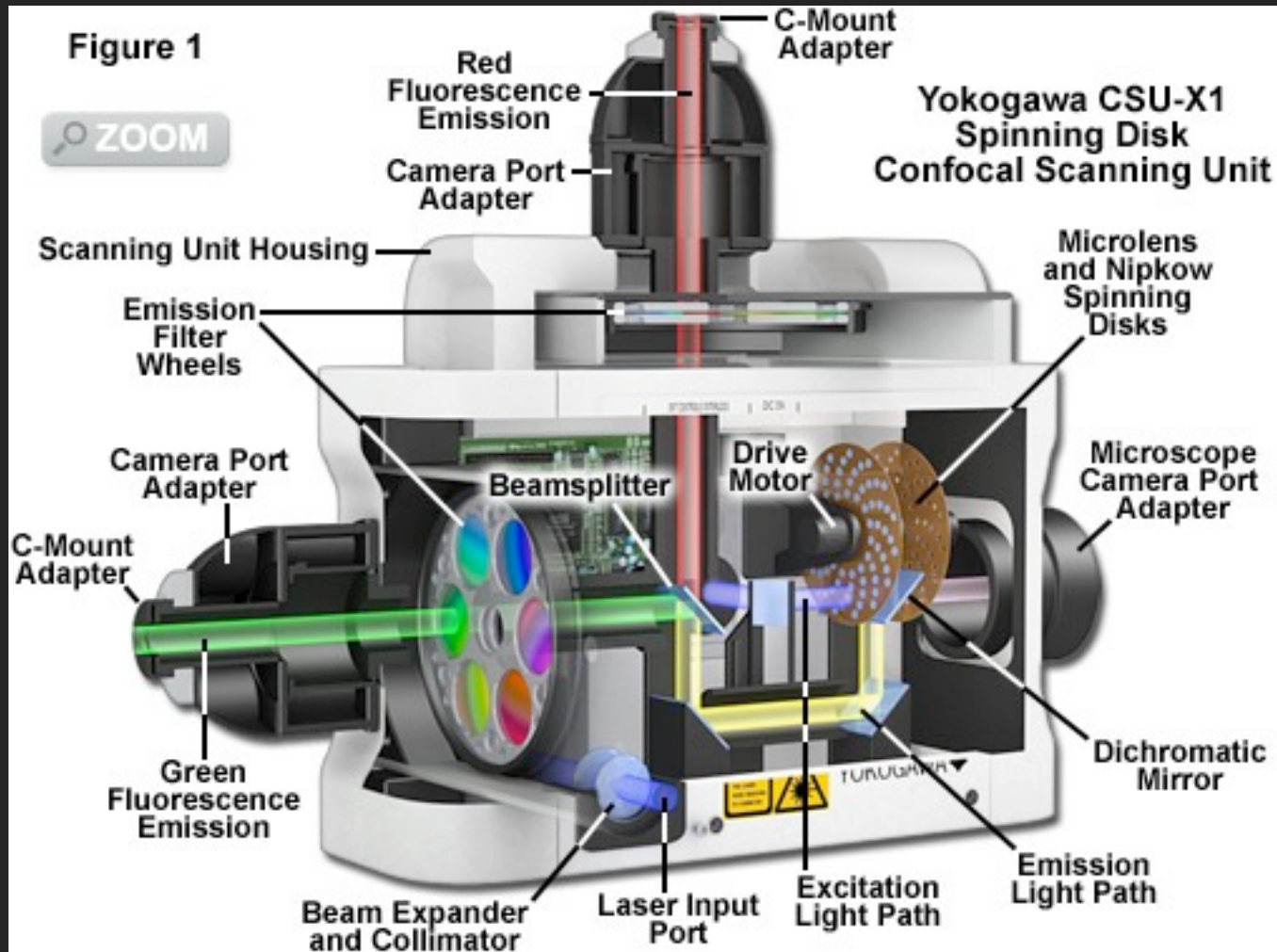
# The Yokogawa Spinning Disk



# The Yokogawa CSU-X1 Spinning Disk

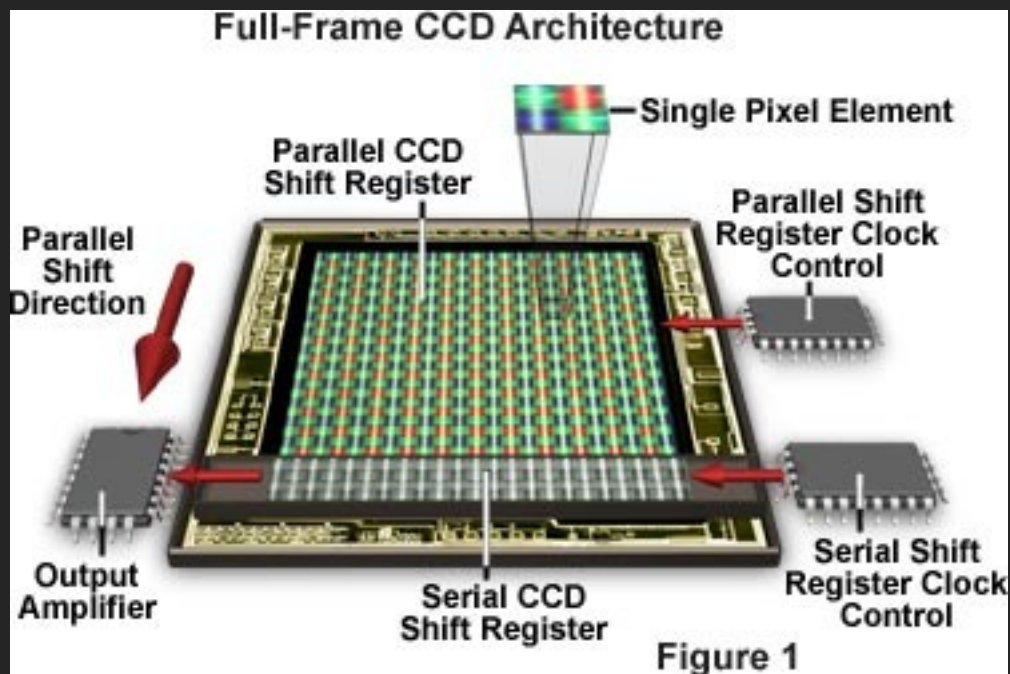


# The Yokogawa CSU-X1 Spinning Disk

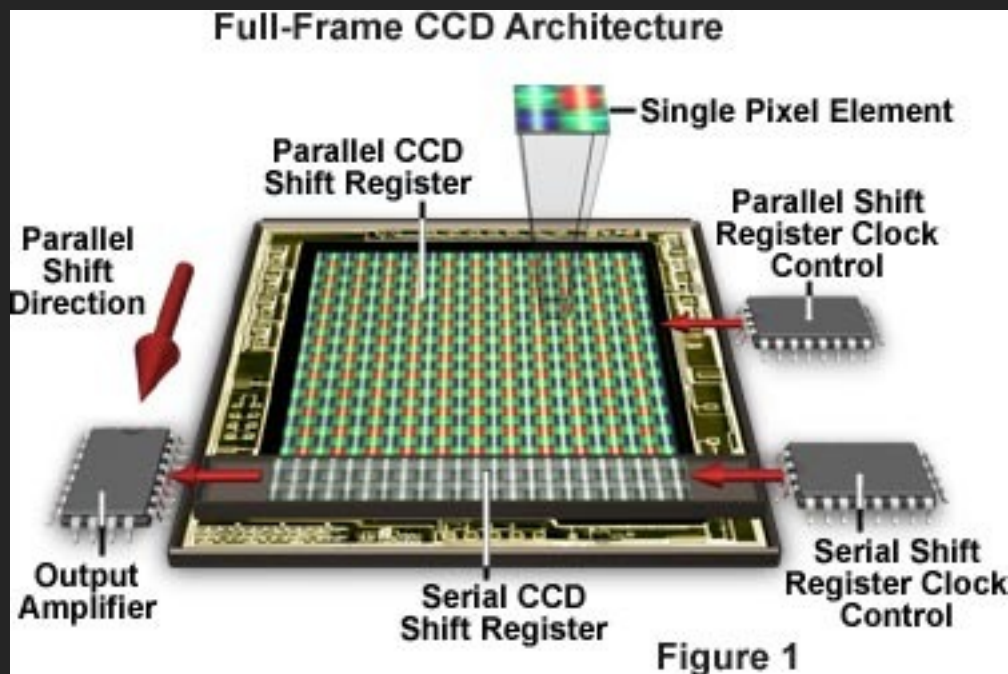


Can collect 2000 images per second

# The Charged Couple Device (CCD) Camera

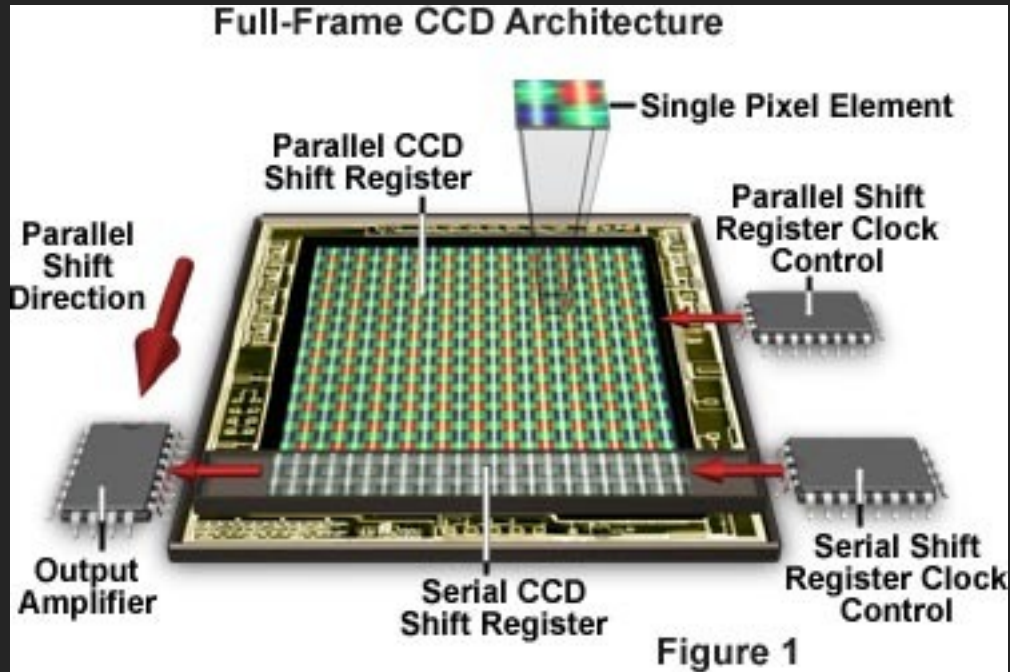


# The Charged Couple Device (CCD) Camera

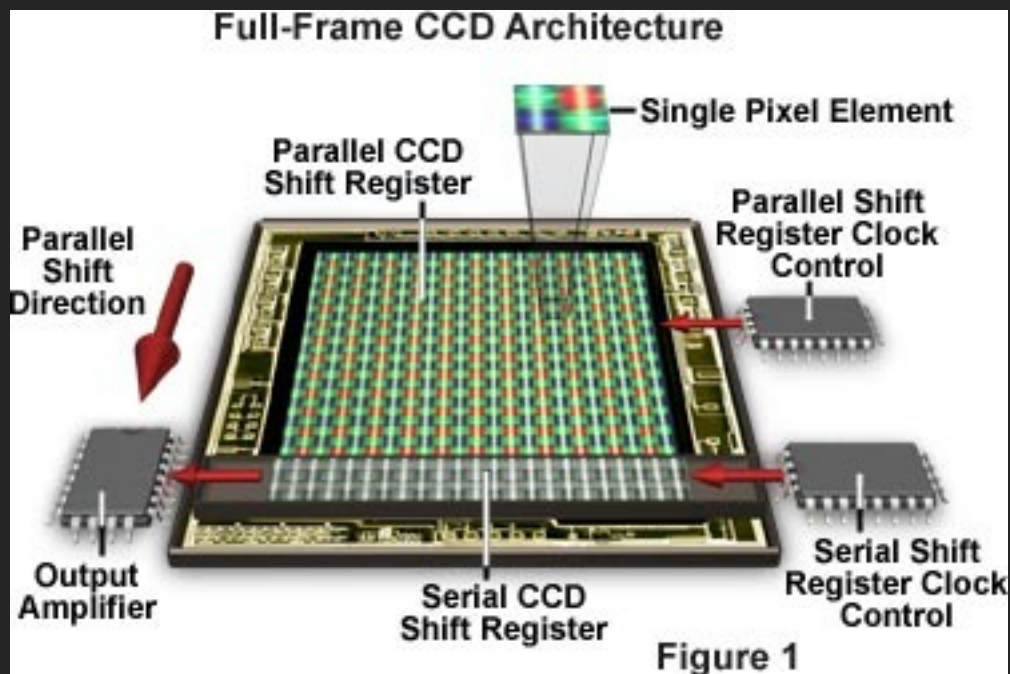


Can get very high QE - up to 95%  
Can be very fast

# The Charged Couple Device (CCD) Camera



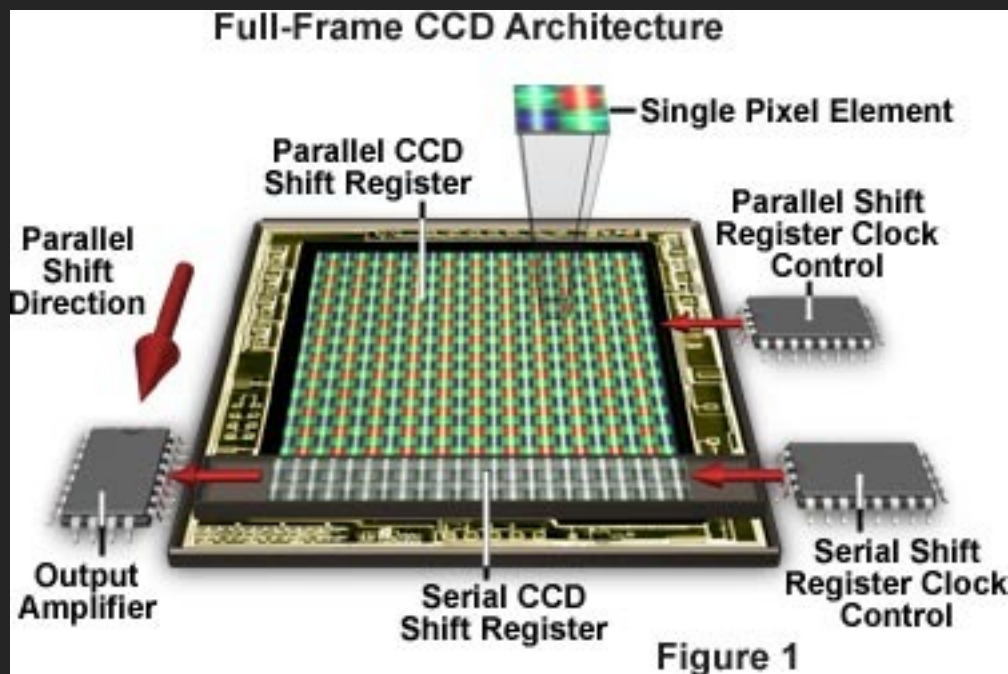
# The Charged Couple Device (CCD) Camera



Noise!



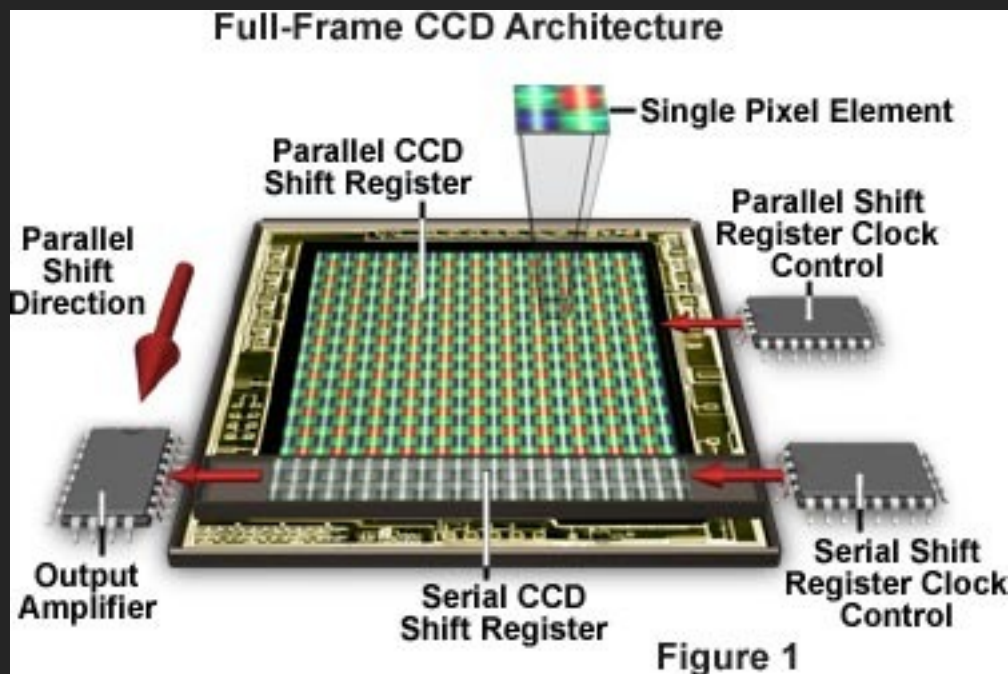
# The Charged Couple Device (CCD) Camera



Noise!

Dark Noise - less of a problem at low temperature

# The Charged Couple Device (CCD) Camera

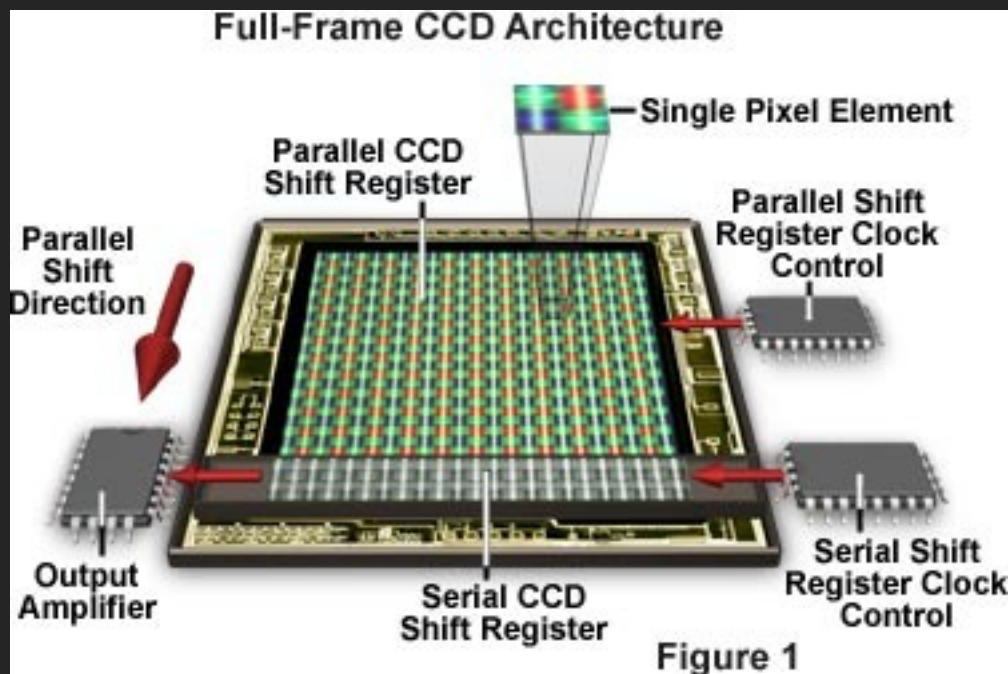


Noise!

Dark Noise - less of a problem at low temperature

Read Noise - inherent to camera, but worse at high speed

# The Charged Couple Device (CCD) Camera



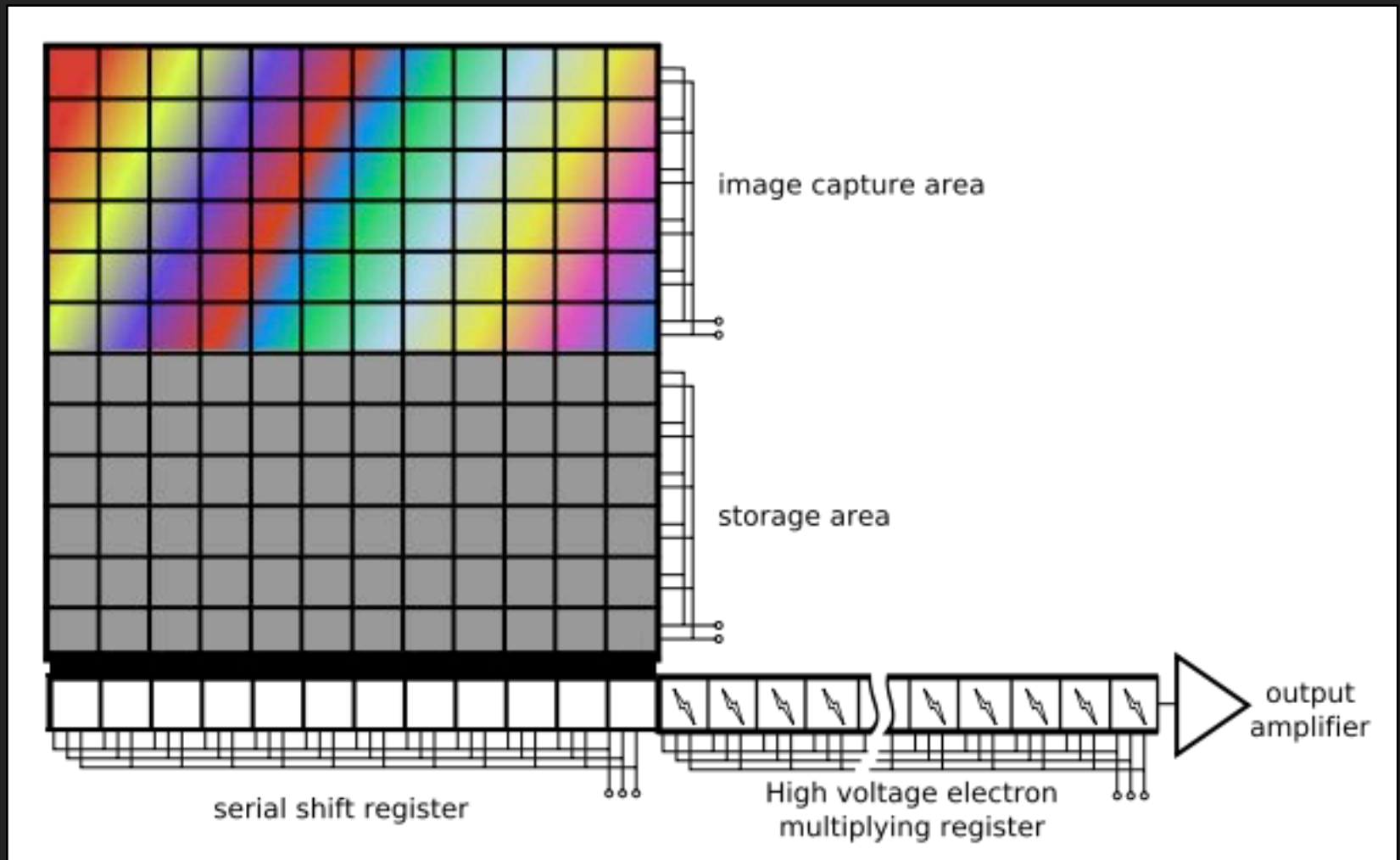
## Noise!

Dark Noise - less of a problem at low temperature

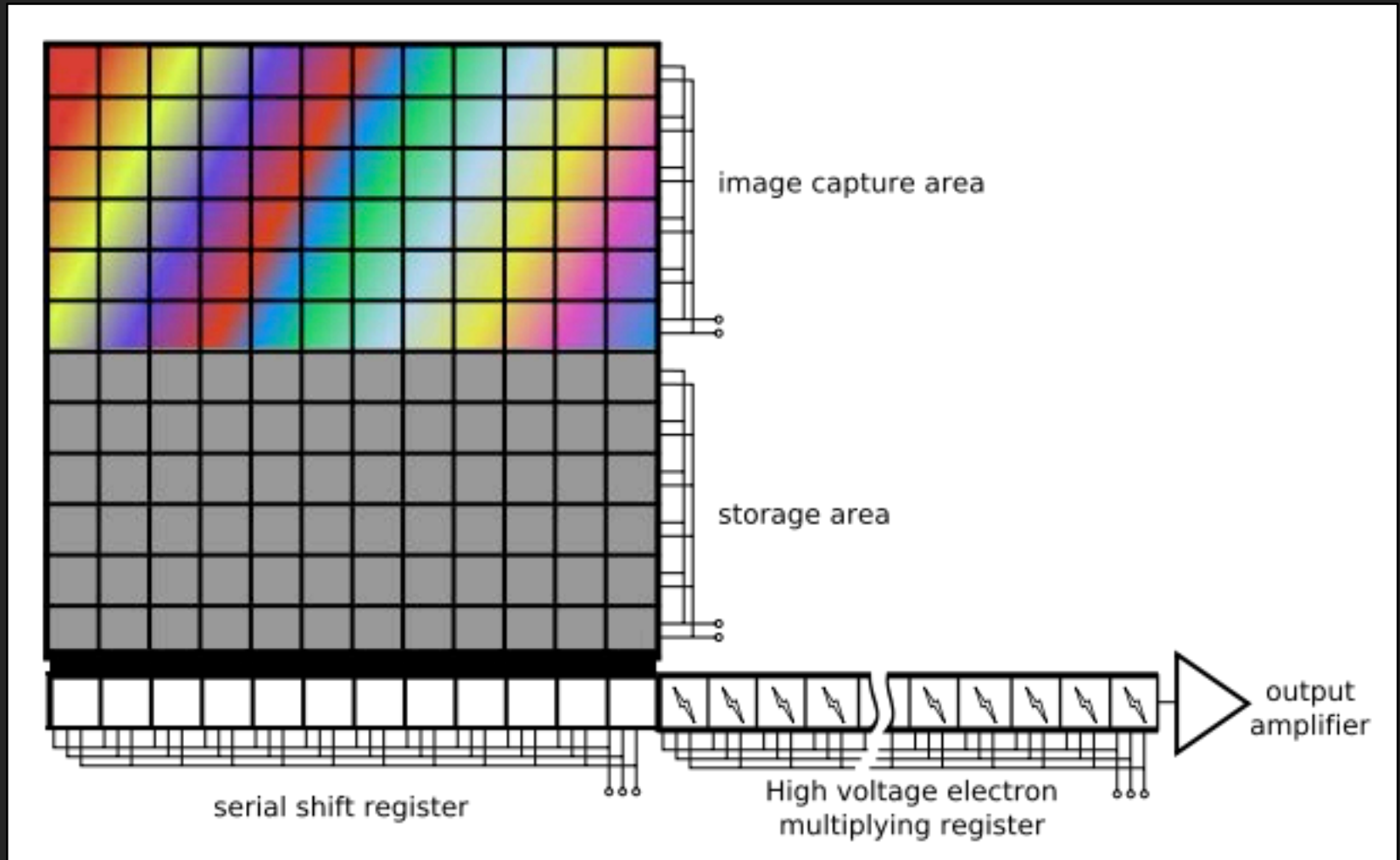
Read Noise - inherent to camera, but worse at high speed

Shot Noise - due to stochastic nature of fluorescence

# The Electron Multiplying Charged Couple Device (EMCCD) Camera

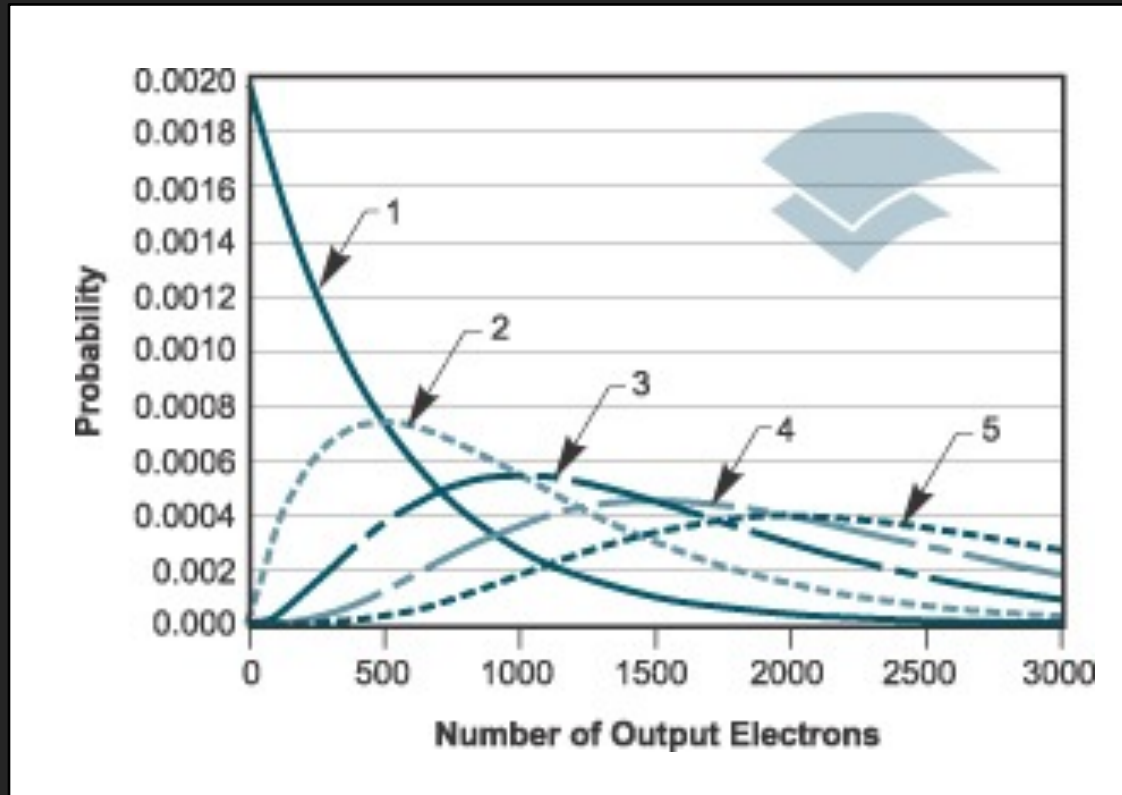


# The Electron Multiplying Charged Couple Device (EMCCD) Camera



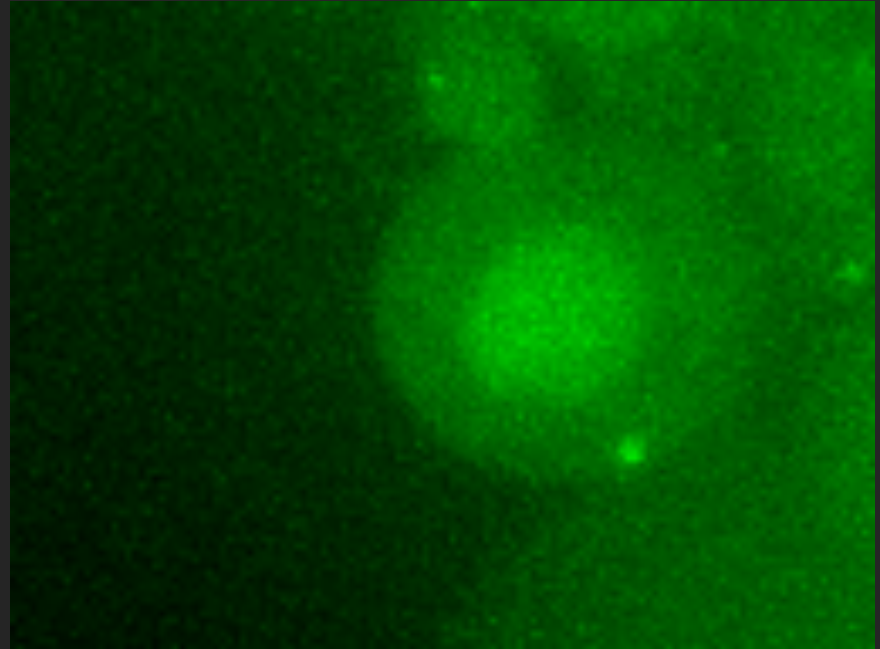
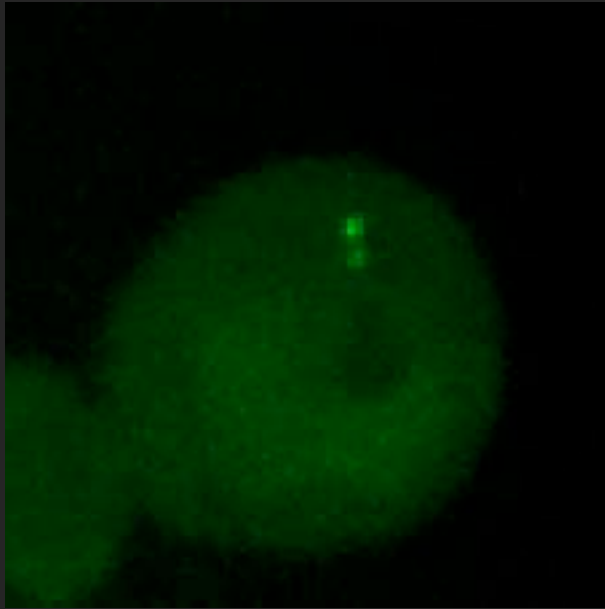
**Beware!!**

# How quantitative is an EMCCD Camera?



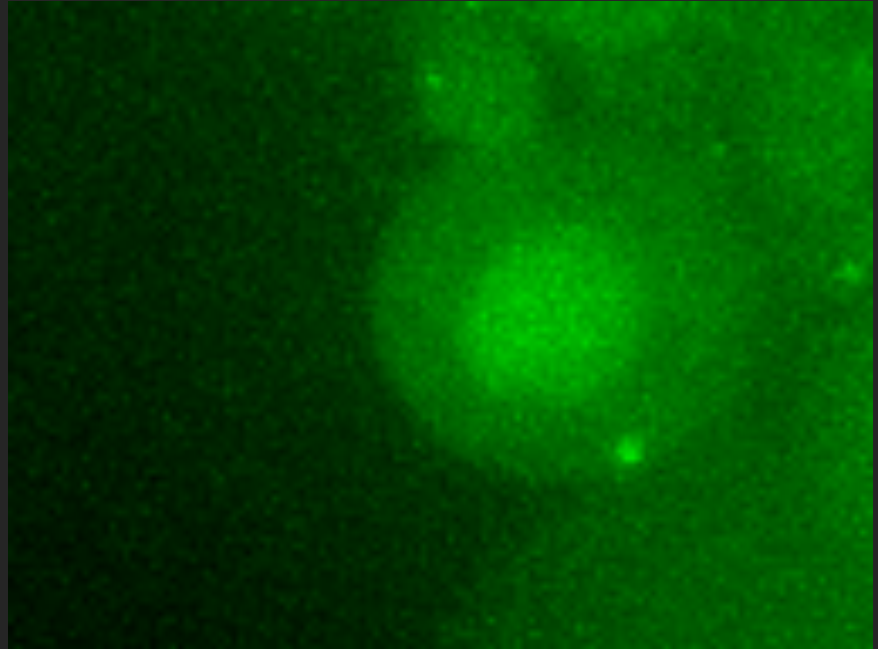
# The Electron Multiplying Charged Couple Device (EMCCD) Camera

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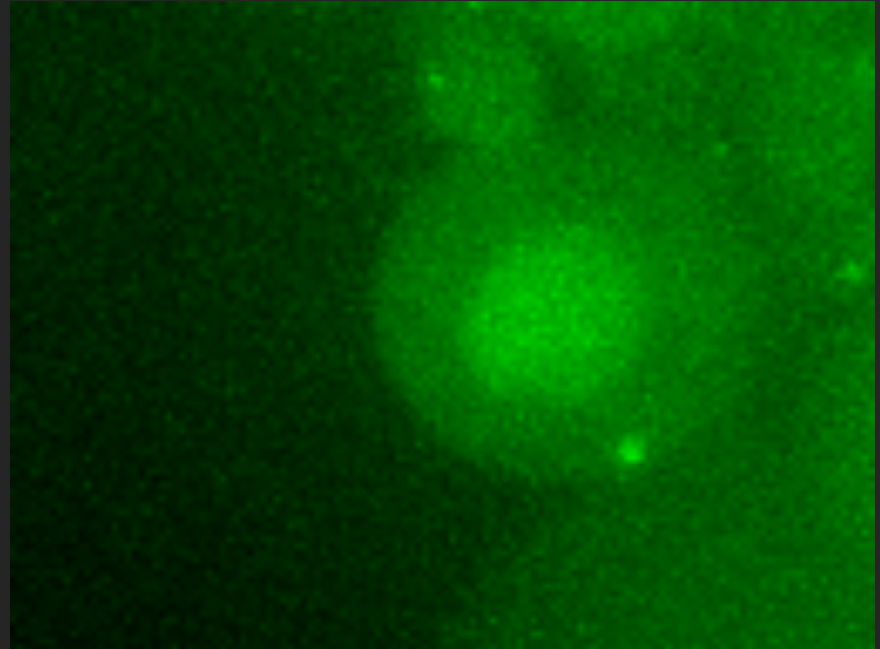
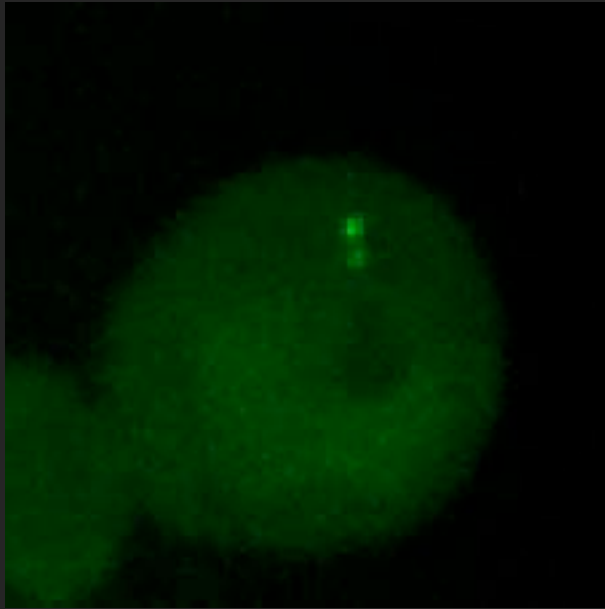




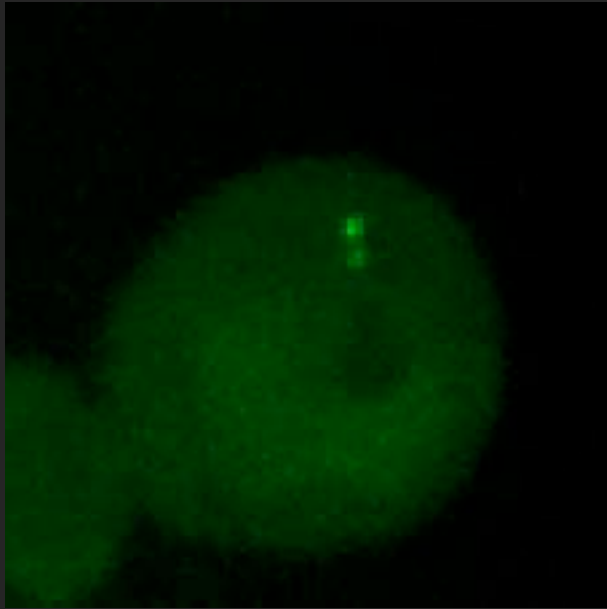
# The Electron Multiplying Charged Couple Device (EMCCD) Camera



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# The Electron Multiplying Charged Couple Device (EMCCD) Camera



# Understanding Zoom!



# Point Scanning vs. Spinning Disk

# Point Scanning vs. Spinning Disk

Speed

Slow (secs)

Fast (100msec)

# Point Scanning vs. Spinning Disk

Speed

Slow (secs)

Fast (100msec)

Sensitivity

OK

OK

# Point Scanning vs. Spinning Disk

Speed	Slow (secs)	Fast (100msec)
Sensitivity	OK	OK
Flexibility	Good	Poor



# Point Scanning vs. Spinning Disk

Speed	Slow (secs)	Fast (100msec)
Sensitivity	OK	OK
Flexibility	Good	Poor
Bleaching	Poor	Good

# Point Scanning vs. Spinning Disk

Speed	Slow (secs)	Fast (100msec)
Sensitivity	OK	OK
Flexibility	Good	Poor
Bleaching	Poor	Good
Pretty Pictures	Unbeatable!	Pretty damn good!

# Point Scanning vs. Spinning Disk

Speed	Slow (secs)	Fast (100msec)
Sensitivity	OK	OK
Flexibility	Good	Poor
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Pretty Pictures	Unbeatable!	Pretty damn good!
Pretty Movies	Good if process slow	Unbeatable!

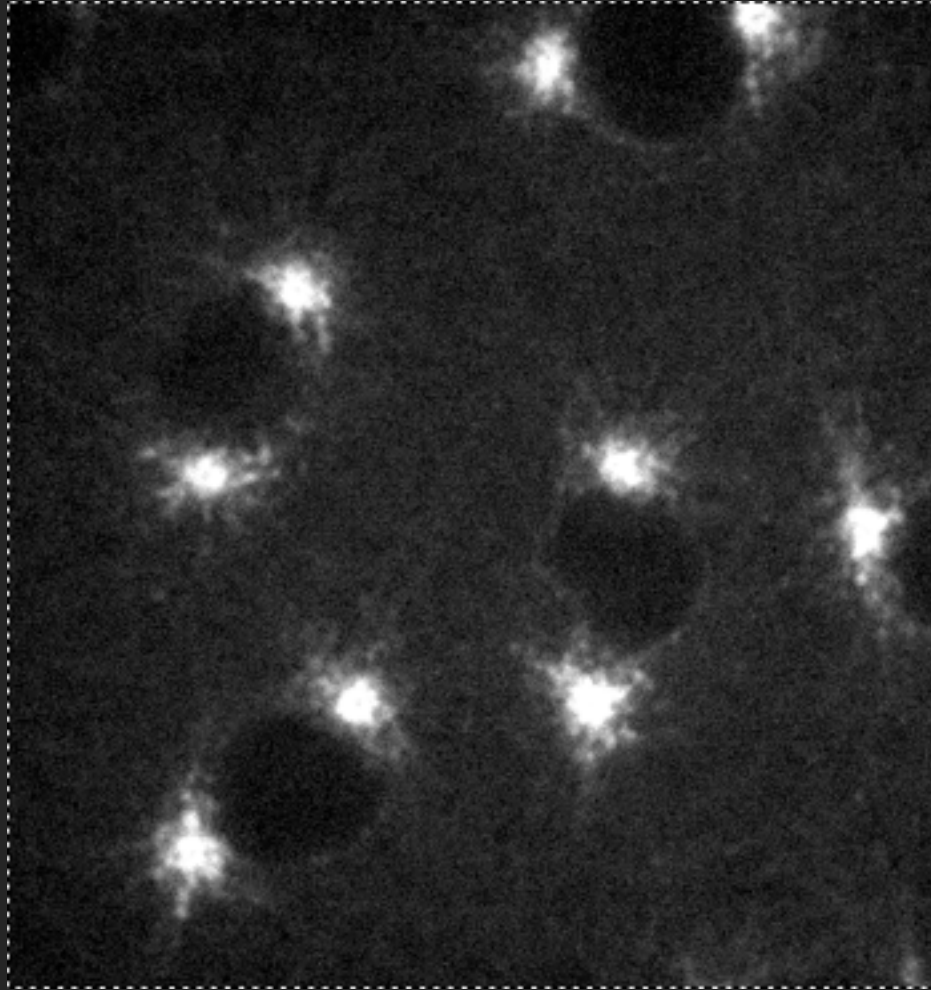
# Point Scanning vs. Spinning Disk

Speed	Slow (secs)	Fast (100msec)
Sensitivity	OK	OK
Flexibility	Good	Poor
Bleaching	Poor	Good
Pretty Pictures	Unbeatable!	Pretty damn good!
Pretty Movies	Good if process slow	Unbeatable!

Murray et al., Evaluating performance in 3D  
fluorescence microscopy. J. Microscopy (2007)

# Example of fast imaging - single plane

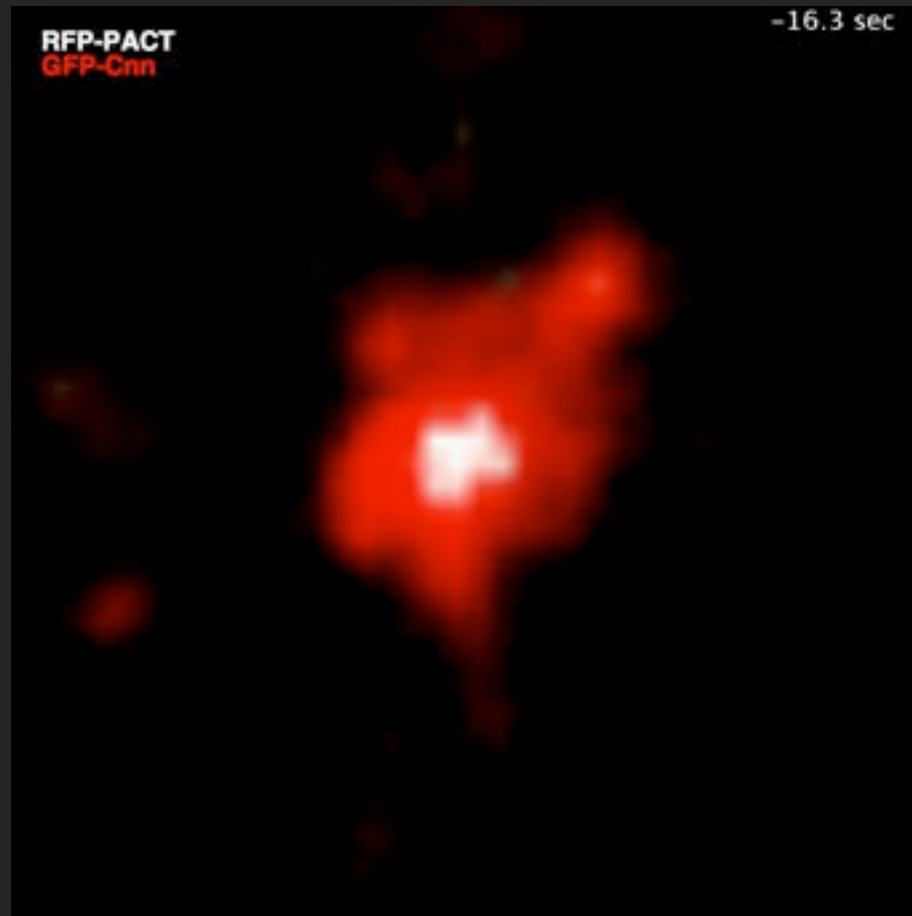
# Example of fast imaging - single plane



# Example of FRAP on Spinning Disk

**Centrioles**  
**Cnn**

# Example of FRAP on Spinning Disk



Centrioles  
Cnn

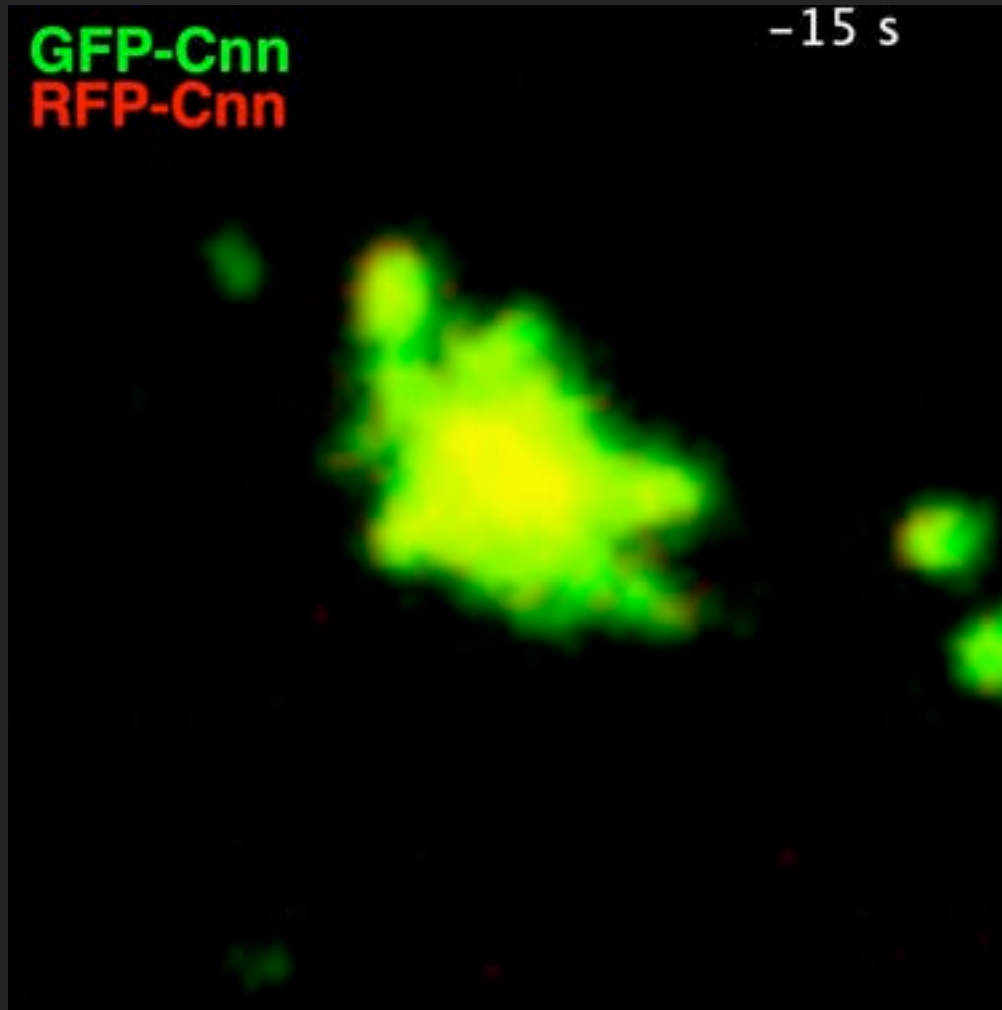


# Example of FRAP on Spinning Disk

**Centrioles**  
**Cnn**

# Another example of FRAP on Spinning Disk

# Another example of FRAP on Spinning Disk

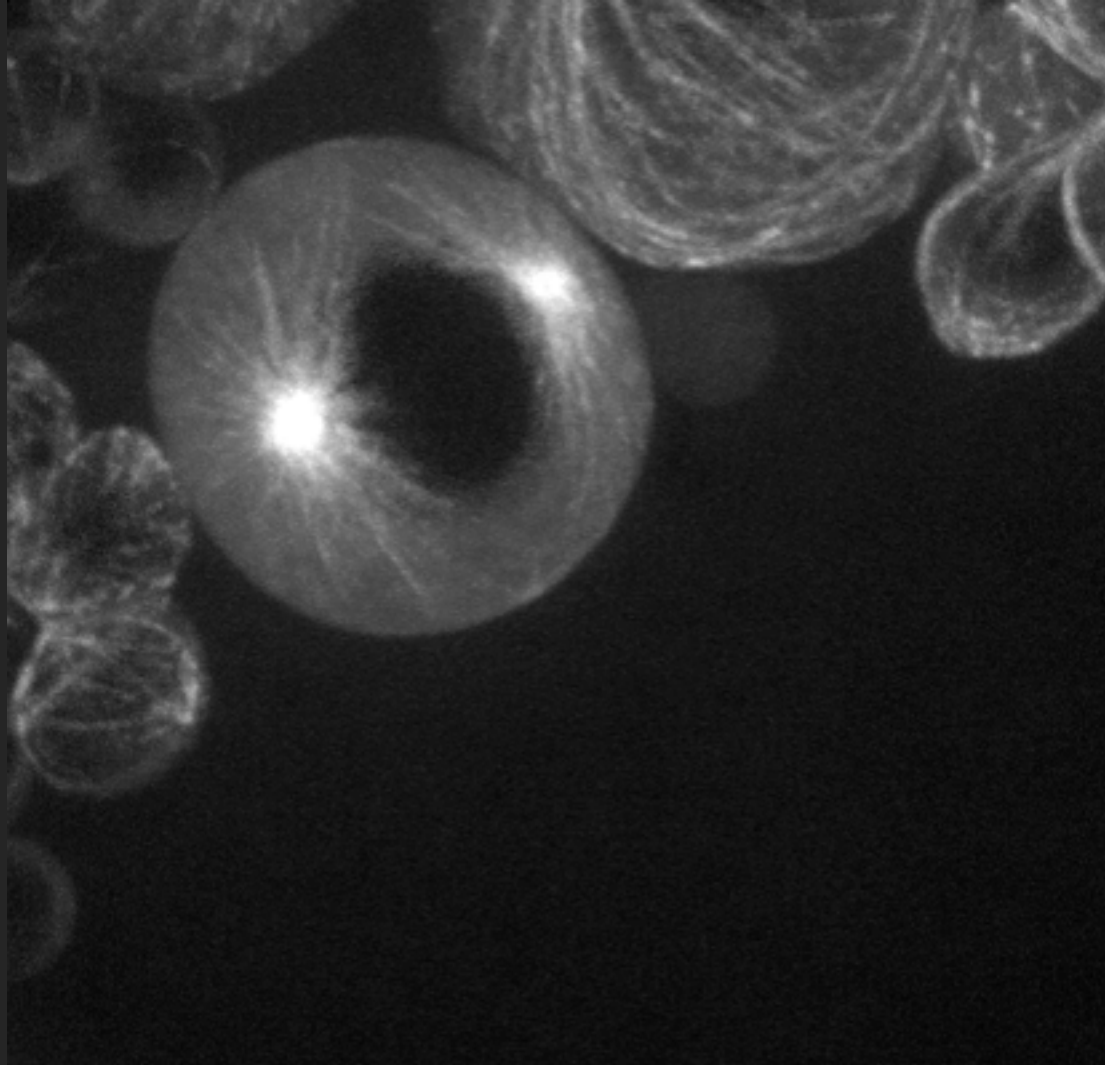


# Another example of FRAP on Spinning Disk

# Cell division in brain stem cells (neuroblasts)

Microtubules

# Cell division in brain stem cells (neuroblasts)



Microtubules

# Cell division in brain stem cells (neuroblasts)

Microtubules

# Identifying proteins required for centrosome duplication

Centrosomes

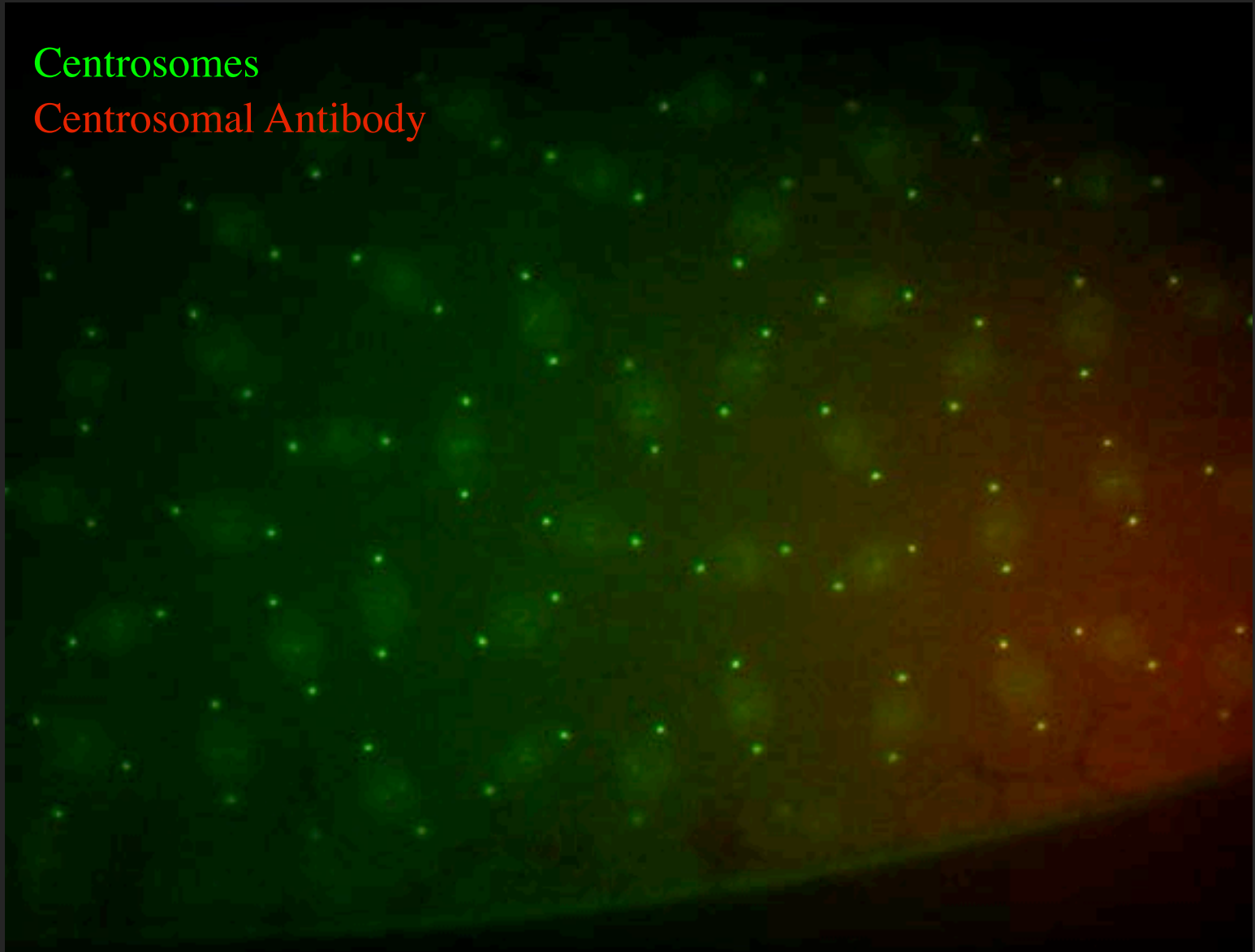
Centrosomal Antibody

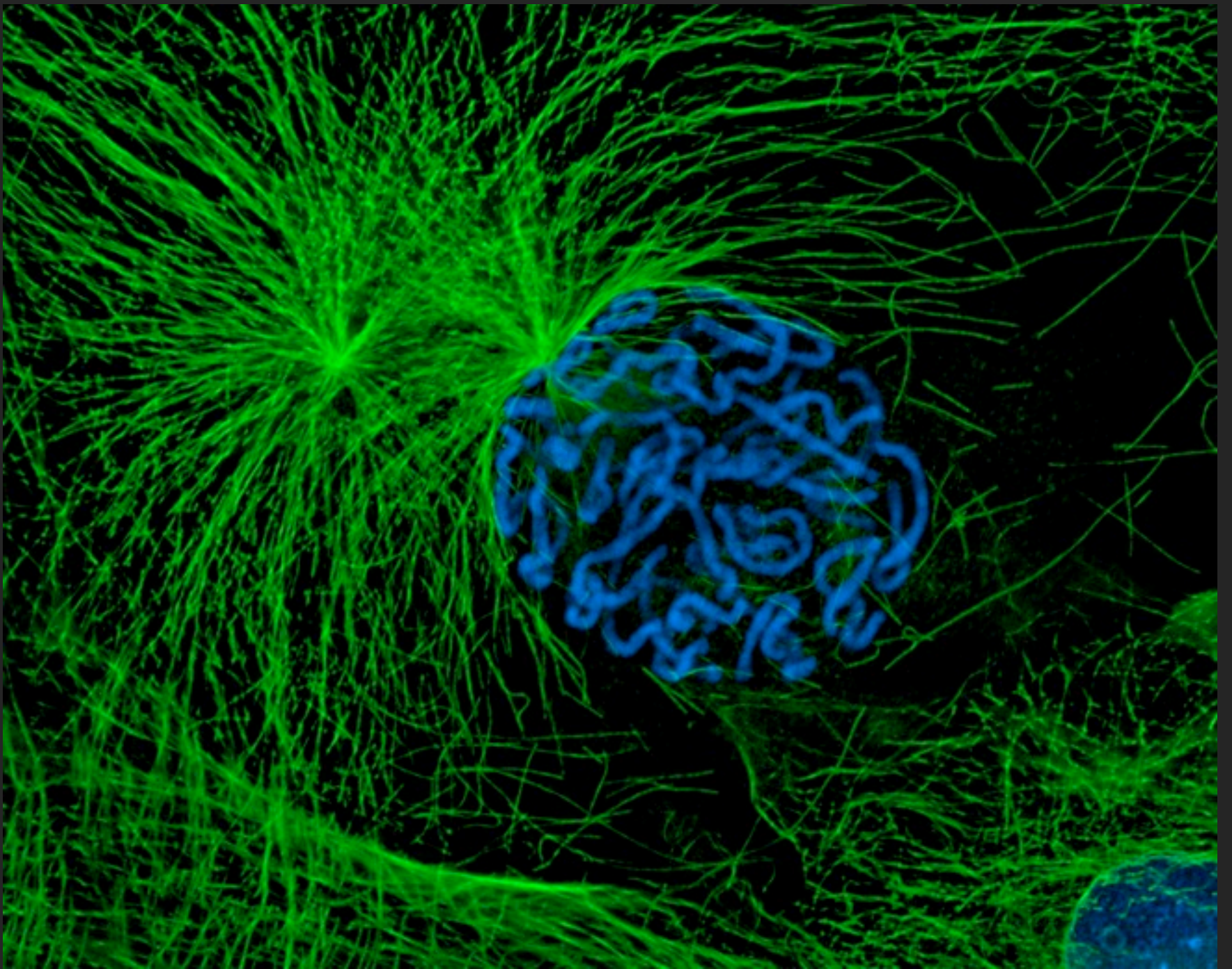


# Identifying proteins required for centrosome duplication

Centrosomes

Centrosomal Antibody





Monday, 12 March 12