

A fluorescence microscopy image of a cell. The nucleus is stained pink, and the cytoskeleton is stained blue. The background is black.

# Dyes and Fluorescent Proteins

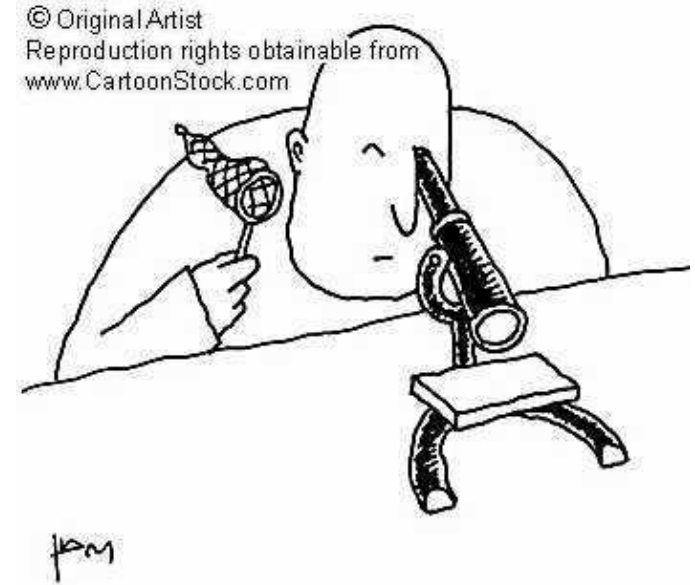
Mark Howarth

Lecturer in Bionanotechnology

Department of Biochemistry

# Overview

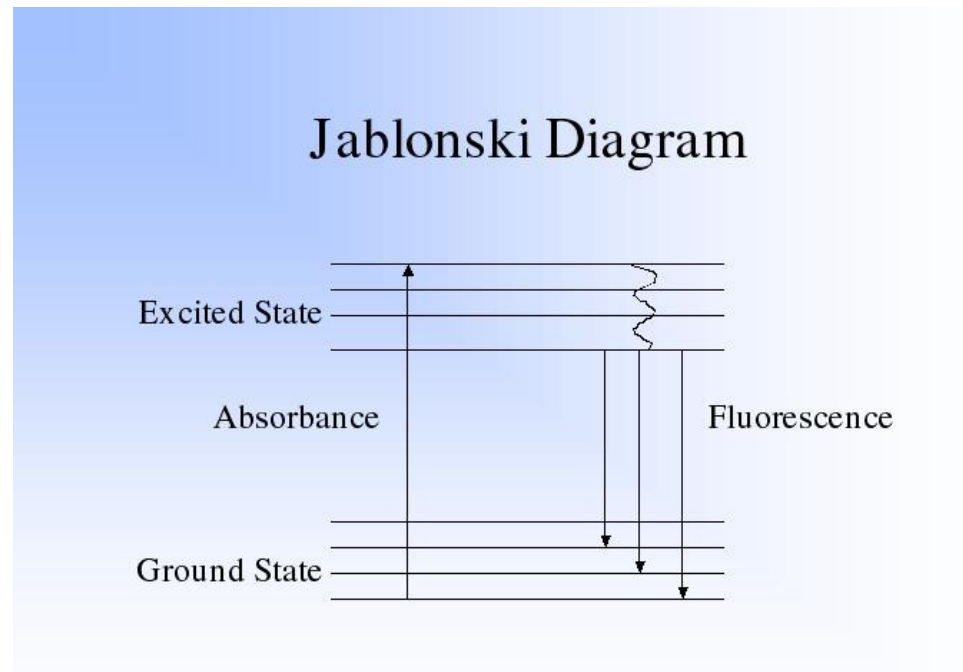
1. What is fluorescence
2. What kind of structures are fluorescent
3. How to make and target fluorescent probes
4. Fluorescent probes for cellular structure and function



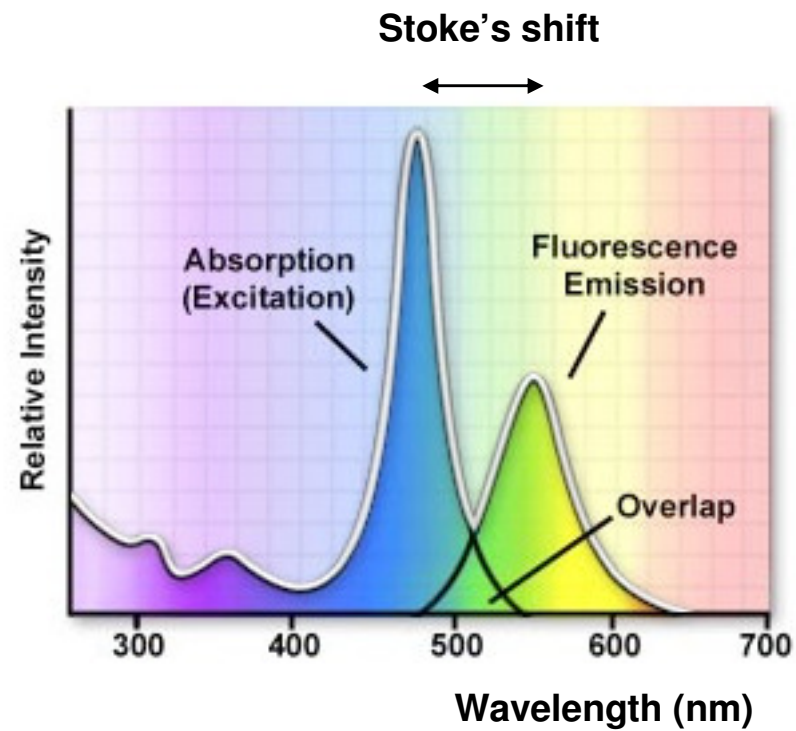
# What is fluorescence?

Absorption of a photon leading to emission of a photon of a longer wavelength

Energy levels?

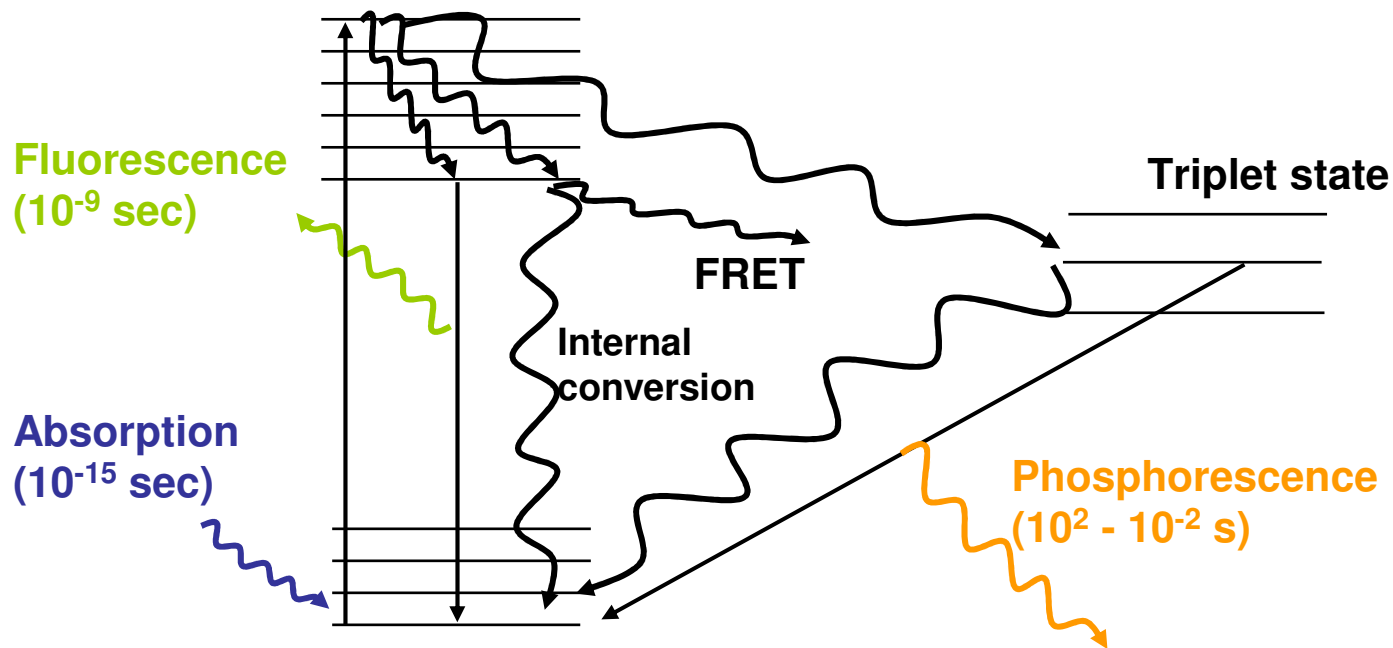


# Fluorescence spectrum



for fluorescein

# Not all energy emitted as fluorescence



$$\text{Quantum yield} = \frac{\text{no. of fluorescent photons emitted}}{\text{no. of photons absorbed}}$$

e.g. EGFP QY=0.6 For every 10 photons absorbed, 6 are emitted.  
(at optimal temp, pH etc.)

# Overview

1. What is fluorescence

2. What kind of structures are fluorescent

3. How to make and target fluorescent probes

4. Fluorescent probes for cellular structure

# What sort of molecules are fluorescent?

## **Organic fluorophores**

especially

1. Intrinsic fluorophores (source of autofluorescence)
2. Dyes
3. Fluorescent proteins

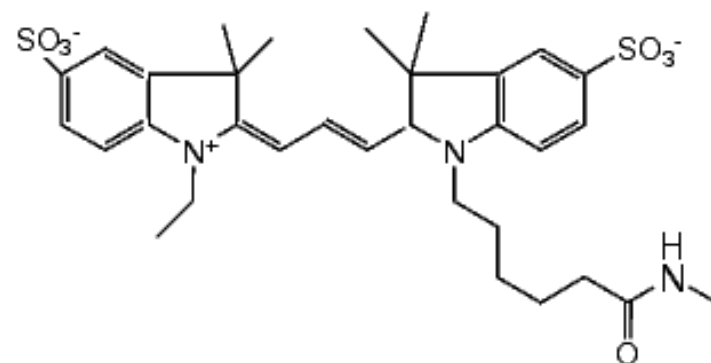
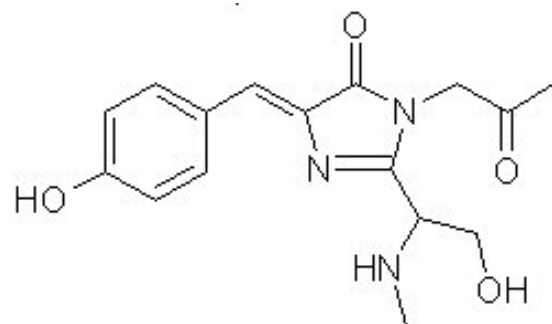
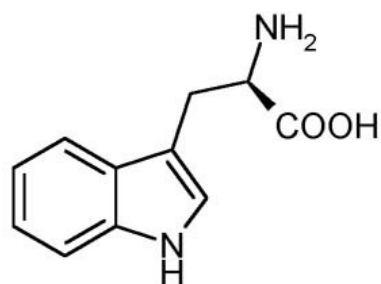
## **Inorganic fluorophores**

especially

1. Lanthanides
2. Quantum dots

# What sort of molecules are fluorescent?

## 1. Organic fluorophores

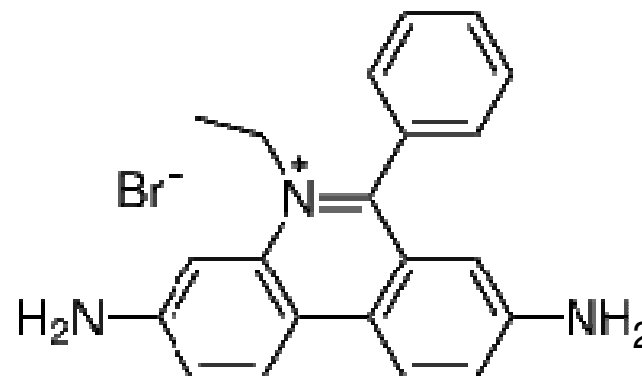
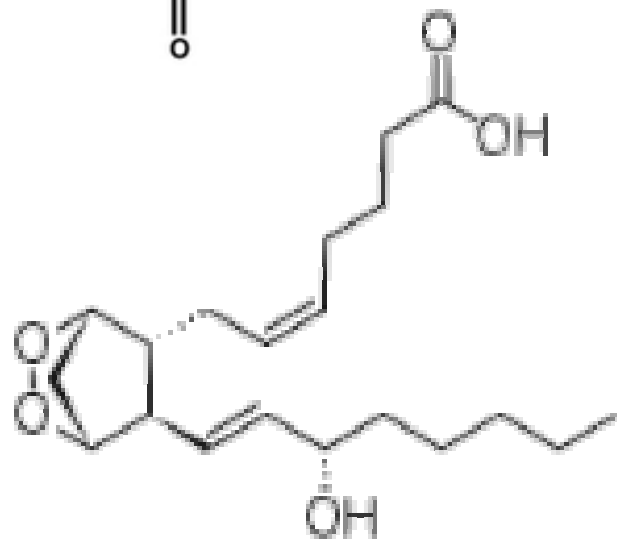
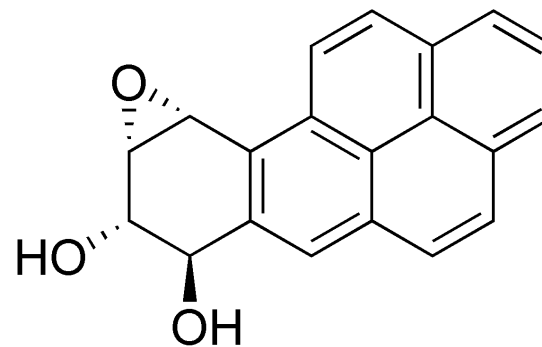
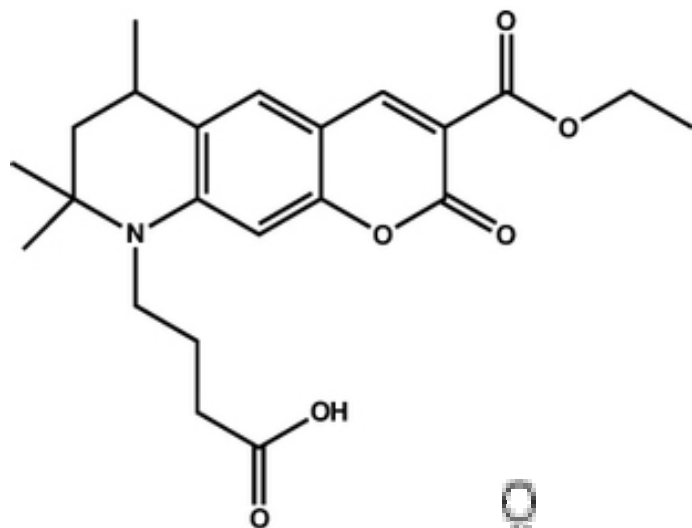


### Chemical features:

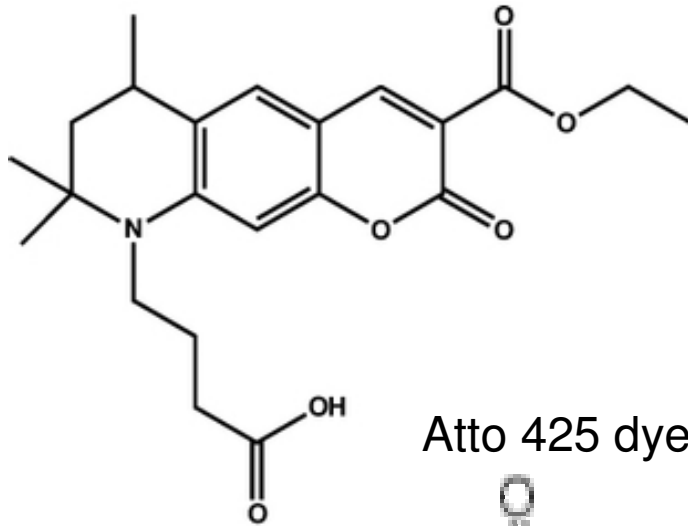
1. Conjugation
2. Rigidity especially fused aromatic rings
3. Heteroatoms



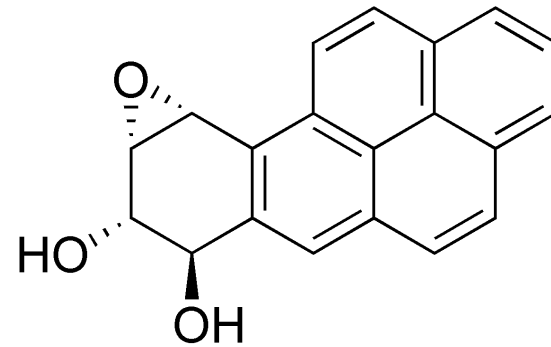
Please rank these in order of fluorescence



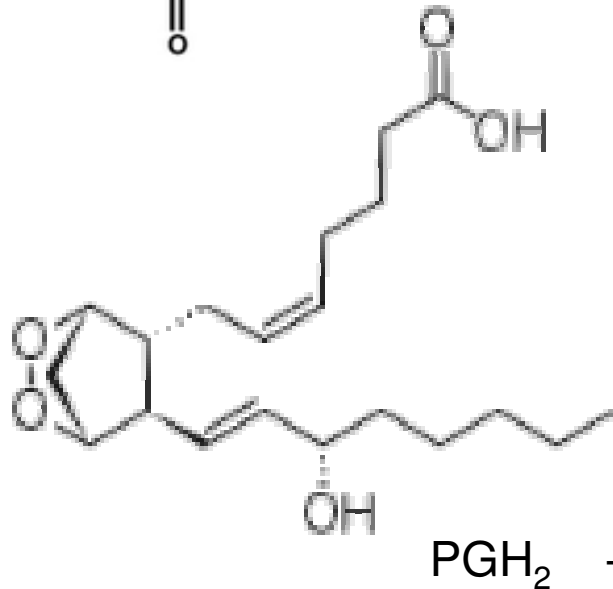
# Rank these in order of fluorescence-answers



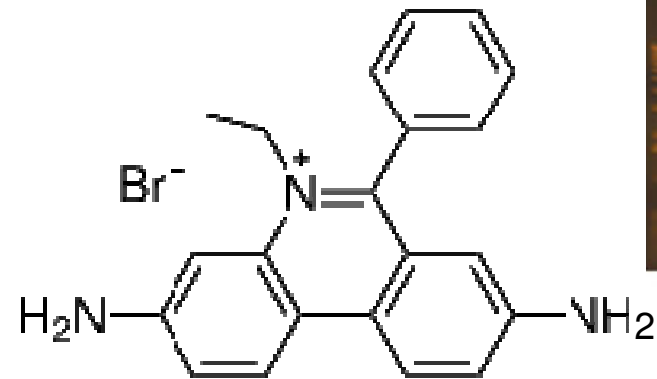
Atto 425 dye +++



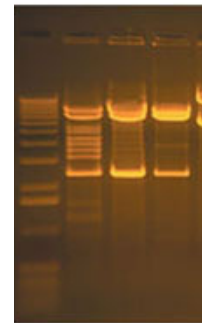
benzopyrene diol epoxide +?



PGH<sub>2</sub> -

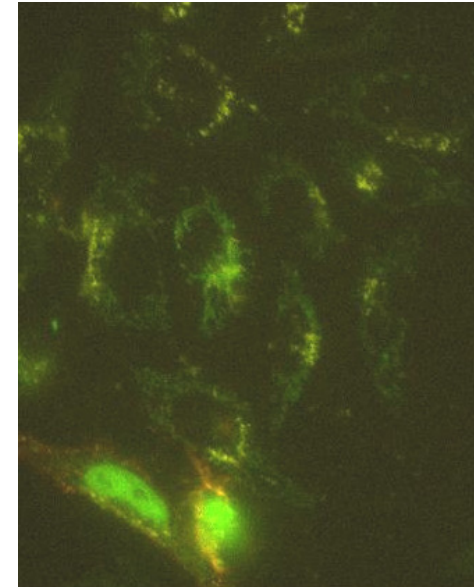
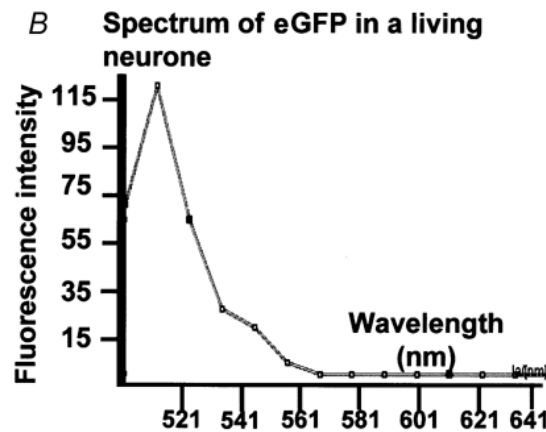
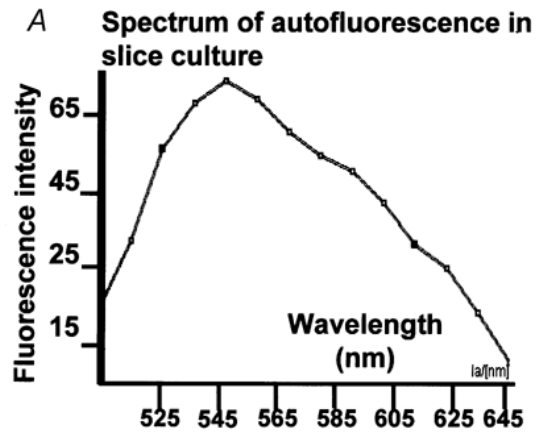


ethidium bromide ++



# What sort of molecules are fluorescent?

## 1. Endogenous organic fluorophores



Most common autofluorescent molecules:

Flavins, NADH, NADPH, elastin, collagen, lipofuscin

Avoiding autofluorescence:

choose dye emitting in red with big Stokes shift

add quencher (Crystal violet)

add reducing agent to react with autofluorescent molecules

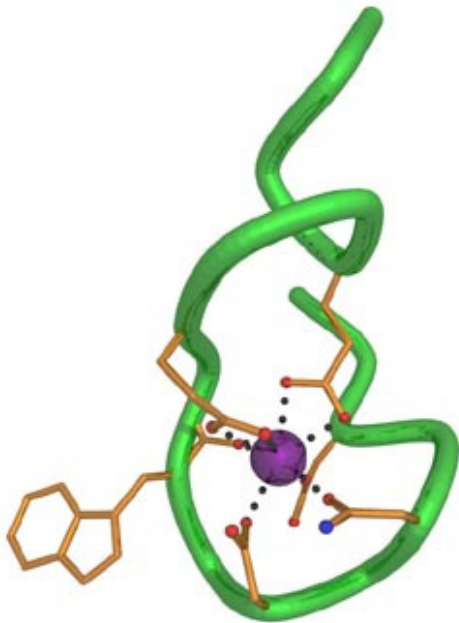
time-gate fluorescence

# What sort of molecules are fluorescent?

## 2. Inorganic fluorophores

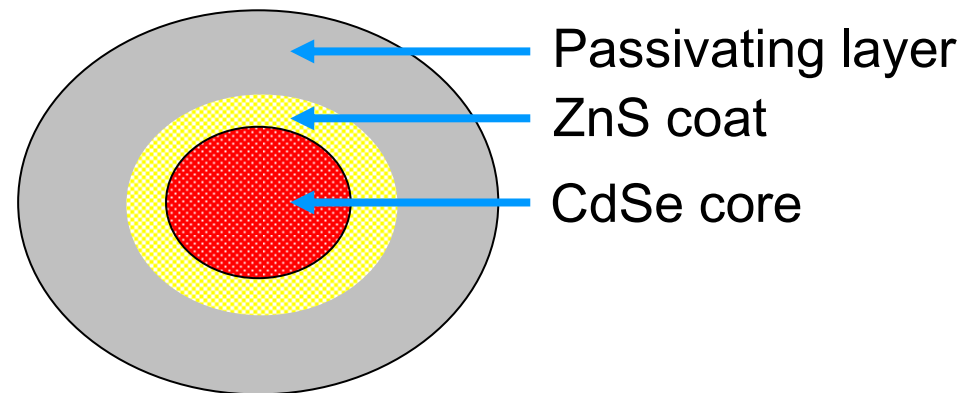
### Lanthanides

Peptide sequence binds  $Tb^{3+}$  and protects from quenching by water



Curr Opin Chem Biol. 2010;14(2):247-54.  
Lanthanide-tagged proteins--an illuminating partnership. Allen KN, Imperiali B.

### Quantum dots



Michalet X, et al. Quantum dots for live cells, in vivo imaging, and diagnostics. Science. 2005 307(5709):538-44.

# How good is a fluorophore?

## 1. Excitation and emission appropriate

background worse in UV + with small Stokes shift  
good match to filters on your microscope  
look at other fluorophores at same time

## 2. Bright

see small numbers of fluorophores,  
low self-quenching, high QY and absorbance

## 3. Stable to photobleaching

exciting light damages fluorophore

## 4. Non-toxic

## 5. Environment-insensitive (especially to pH)

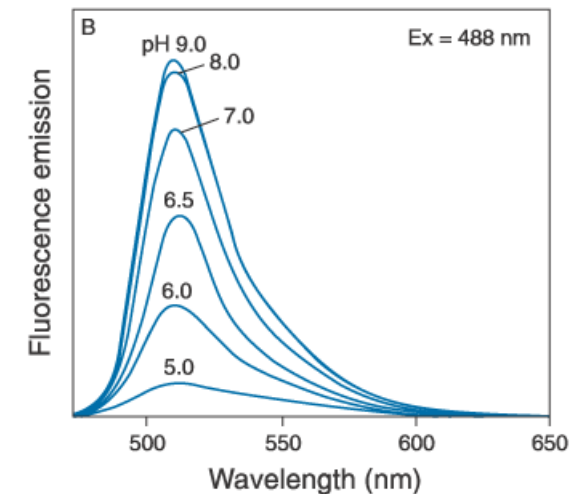
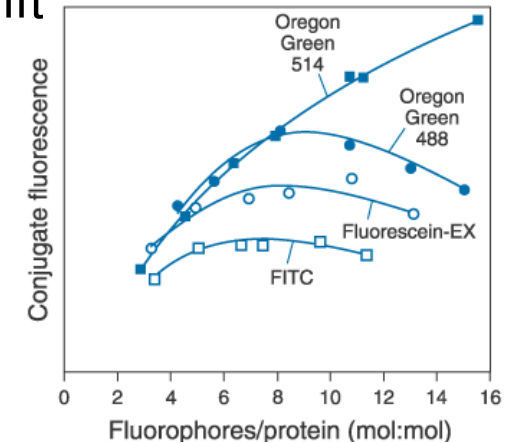
## 6. Little non-specific binding

## 7. Small

## 8. Little blinking

## (9. Cost)

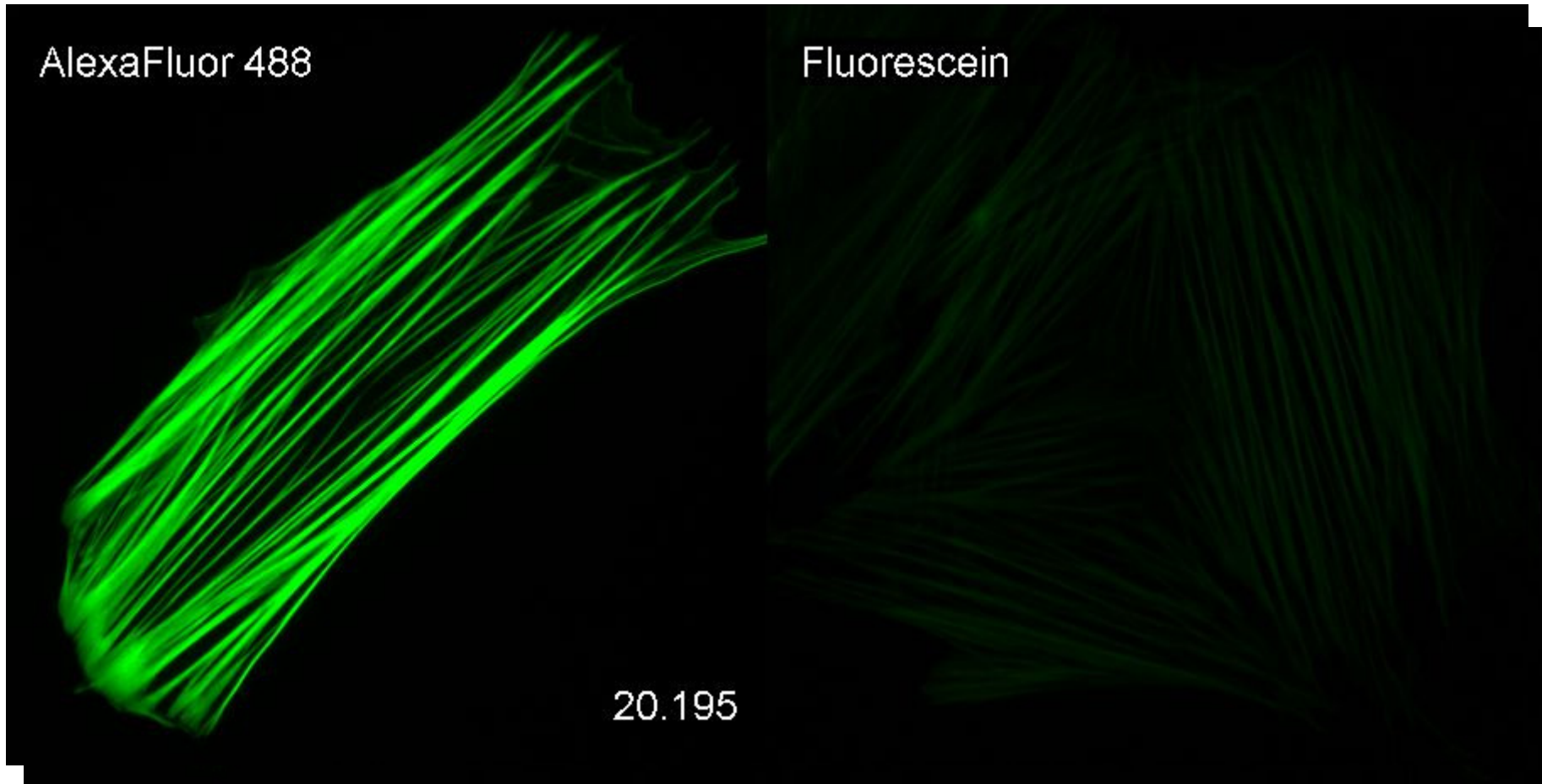
Green dye  
self-quenching



Fluorescein pH sensitivity 13

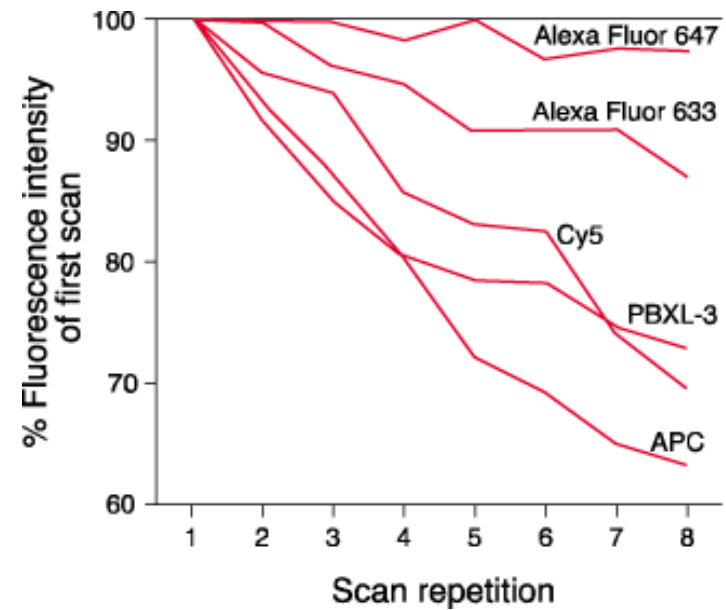
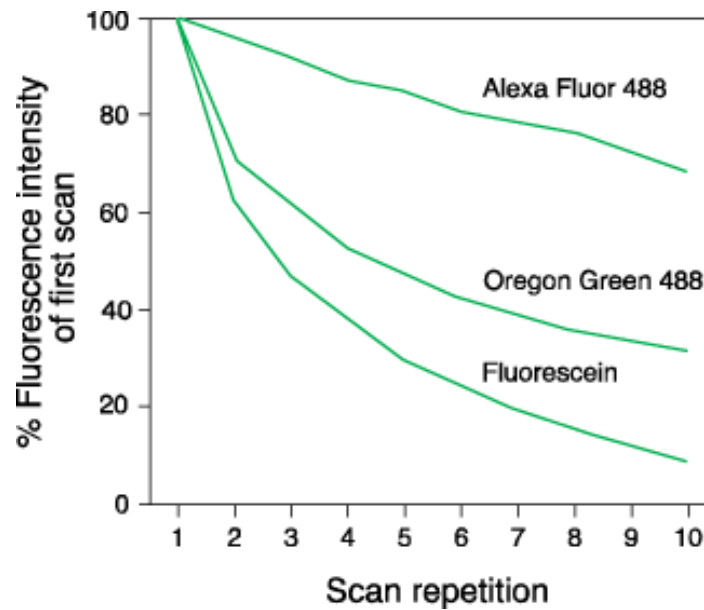
# *Alexa Fluor 488 vs Fluorescein Bleaching*

2x Real Time



# Alexa Fluor Dyes – Photostability

Laser-scanning  
cytometry  
EL4 cells  
biotin-anti-CD44  
+ streptavidin  
conjugates



Fluorescein is the commonest dye  
but has poor photostability.

# Protecting the fluorescence signal - Antifade Reagents for fixed cells

Scavenge and prevent reactive oxygen species from forming.

For fixed cells:

**Home made:** 0.3% p-phenylene-diamine (Sigma)  
or Propyl Gallate

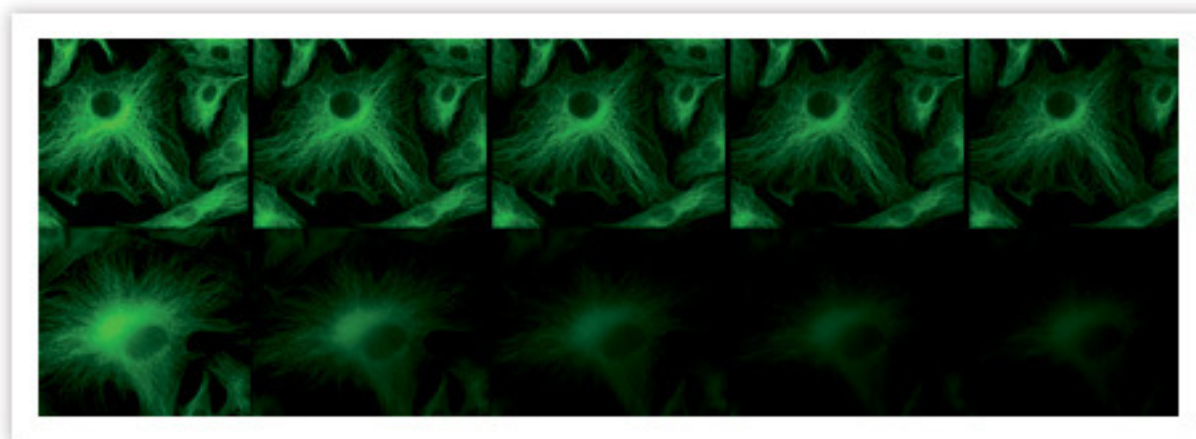
**Vectashield:** Proprietary, very effective all round, affects psf

**Dabco**

**Prolong Gold®**

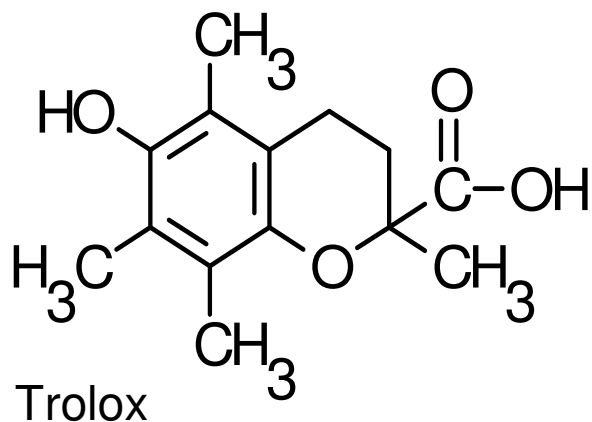
+ Prolong Gold

Untreated

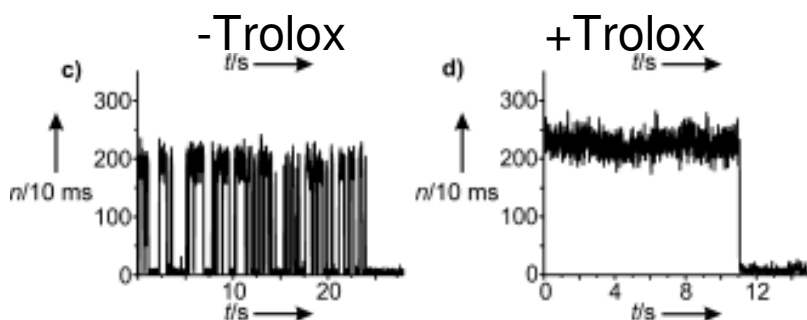




# Antifade Reagents for Live Cells



- Trolox is an antioxidant that can reduce bleaching  
compatible with live specimens  
water-soluble  
working conc.  $\sim 100 \mu\text{M}$
- Ascorbic acid is an alternative antioxidant
- Depleting oxygen (especially used for some single molecule experiments) with Glucose Oxidase and Catalase greatly reduces bleaching.
- Can stop not only bleaching but also blinking

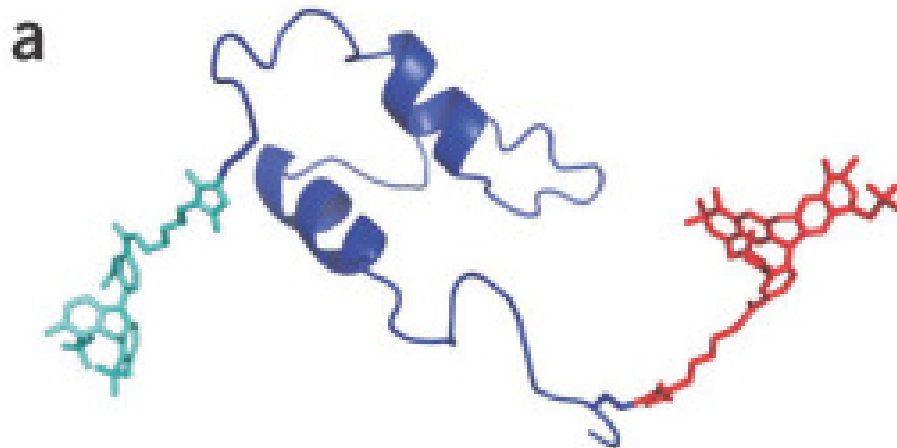
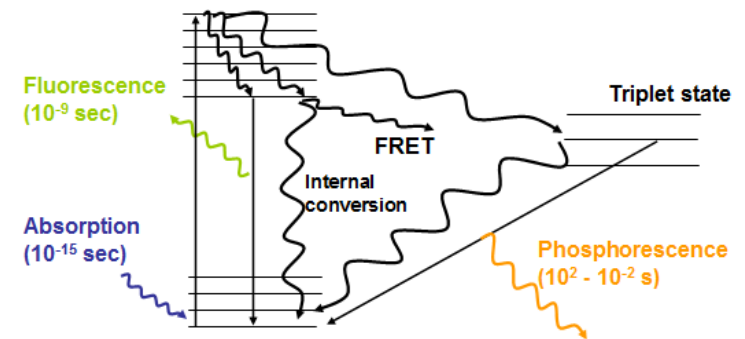


Blinking of single molecule of Atto647N on DNA,  
Vogelsang Tinnefeld Ang Chem 2008

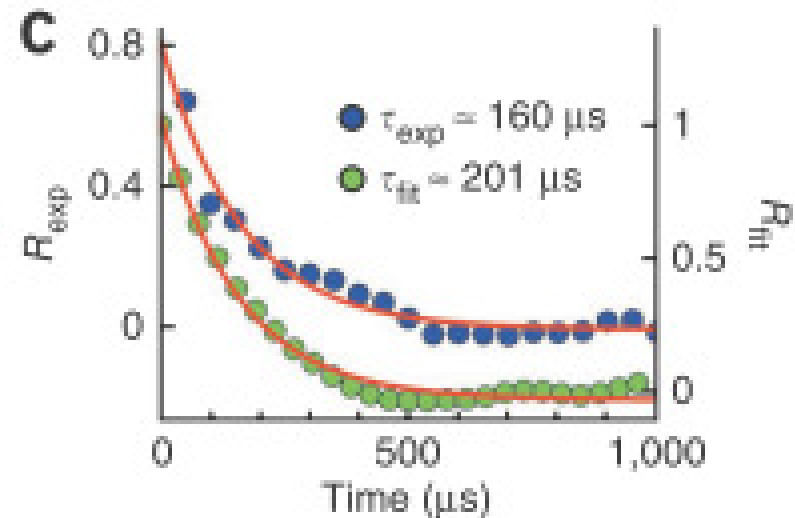
# Microsecond! fluorescent measurements with Trolox + cysteamine

Oxygen helps stop triplet-state build-up  
BUT oxygen promotes photobleaching  
For rapid photon cycling-

1. leave oxygen in
2. add Trolox to further quench triplet state
3. include cysteamine (a thiol) to protect from singlet oxygen and hydroxyl radicals

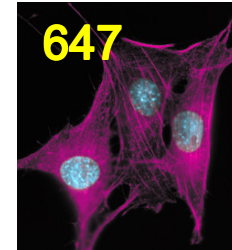
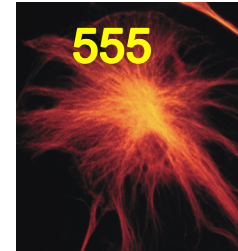
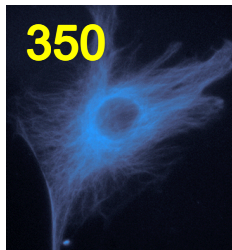


Green/Red Alexa dye FRET  
on rapid folding protein



# Multiplexing- four main colours

Excitation wavelengths:



Emission wavelengths:  
350

Blue  
400

green  
450 500

orange/red  
550 600

far red  
650

700

DAPI/UV	FITC	TRITC	FAR RED
<p><b>Alexa Fluor® 350</b> Coumarin, AMCA</p>	<p><b>Alexa Fluor® 488</b> Fluorescein (FITC) Cy2</p>	<p><b>Alexa Fluor® 555</b> Rhodamine, TAMRA, TRITC Cy3</p>	<p><b>Alexa Fluor® 647</b> Cy5, APC</p>
			<p><b>Alexa Fluor® 594</b> Texas Red, Cy3.5</p>

Colour Selection	◆	Brightness	◆	Photostability
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# Overview

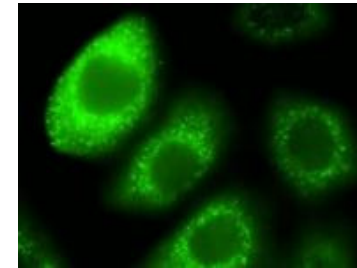
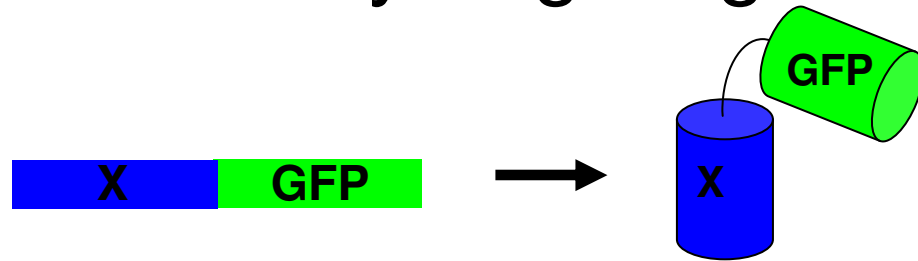
1. What is fluorescence

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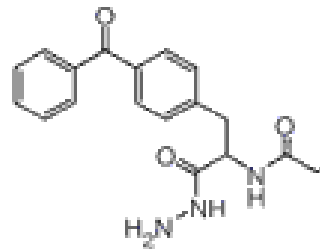
4. Fluorescent probes for cellular structure

# Major bottleneck to using new probes is difficulty targeting them

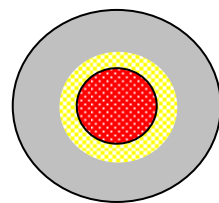


fluorescent proteins  
easy to target

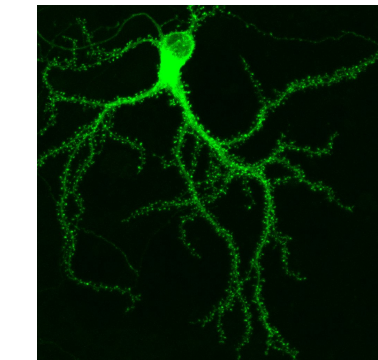
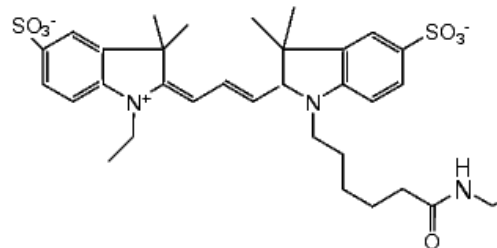
photoaffinity  
probes



quantum  
dots



organic  
fluorophores



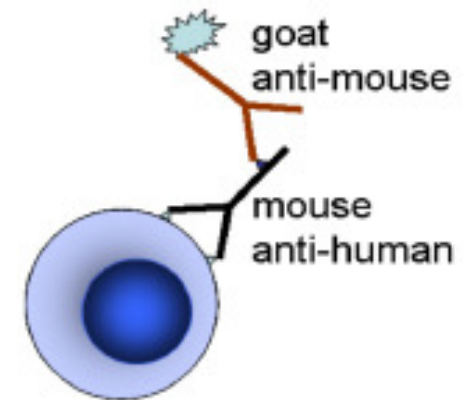
other probes hard  
to target

# Antibodies for cellular imaging

## Live cells

Label plasma membrane and secretory pathway  
Penetrate plasma membrane  
(microinjection, electroporation, pinosome lysis, streptolysin, cell permeable peptides, ester cage)

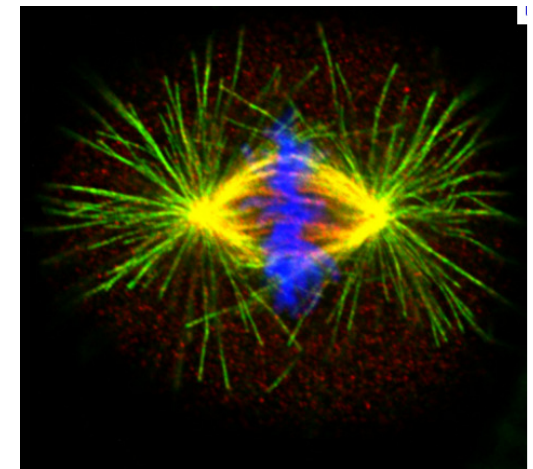
*Get dynamics, avoid fixation artifacts*



## Fixed cells

Permeabilise

*Still can give enormous amount of useful information*



# Not just antibodies for targeting

Other types of targeting agents:

Proteins

(especially antibodies, but also transferrin, insulin, EGF etc.)

Peptides (MHC class I pathway, proteasome function)

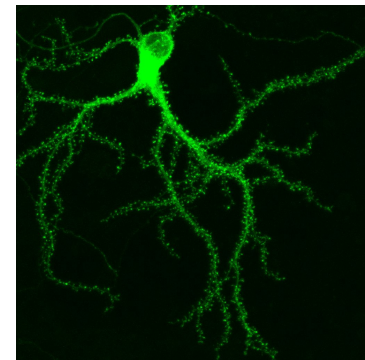
RNA (mRNA, molecular beacons, aptamers, siRNA)

DNA

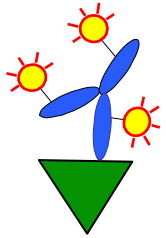
lipids, lipoproteins

drugs

?

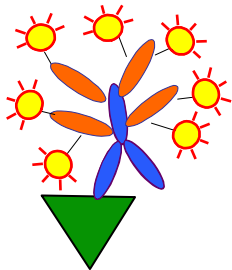


# Getting fluorescence from antibody labelling



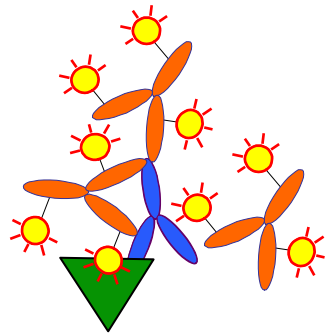
## Directly-labelled Primary Antibody

- Lowest background
- Potentially low signal due to abundance of target or dye
- Dye could affect antigen recognition site



## Zenon Technology

- Brighter Signal
- Dye does NOT affect antigen recognition site

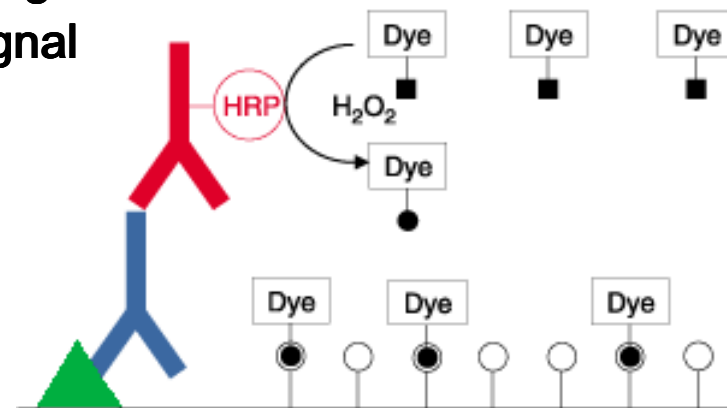


## Indirect-Labeled Secondary Antibody

- Higher Background
- Brighter Signal

## Tyramide Signal Amplification (TSA™)

- Higher background
- Brightest signal





# How to dye: it is easy

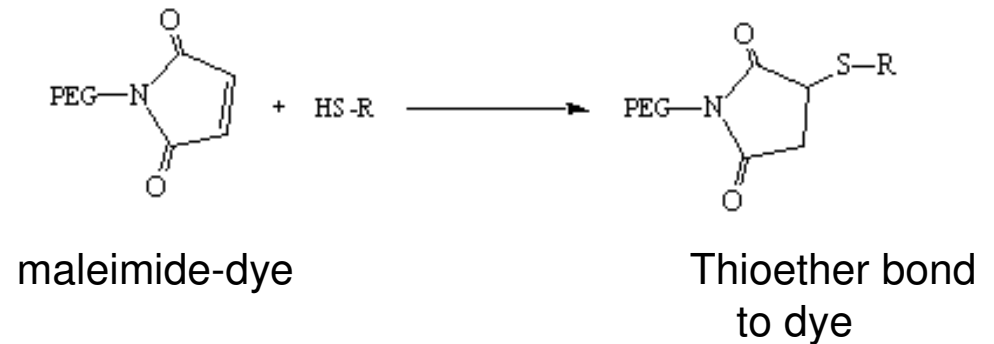
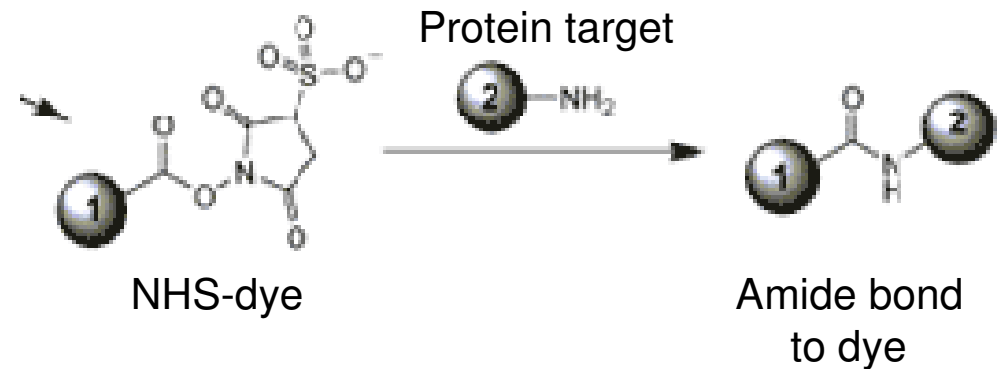
Multiple ways to modify proteins  
(see Molecular Probes catalogue)

Most common ways are to modify:

1. Lysine

or

2. Cysteine

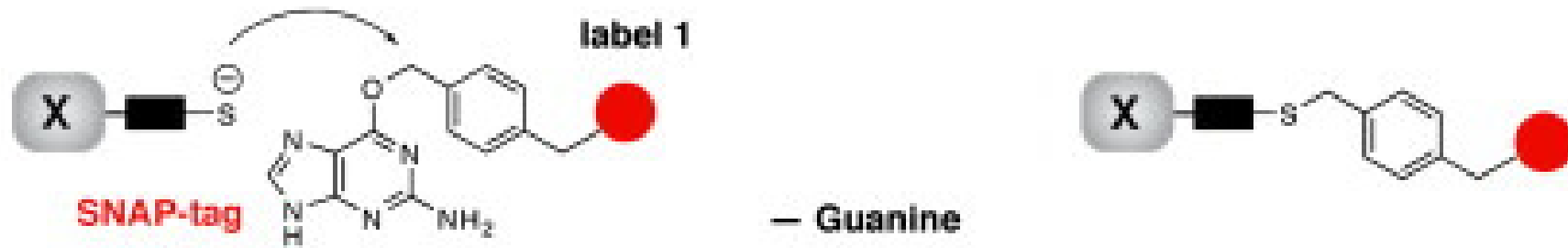


- A Add dye to protein for 3 hr
- B 1cm Sephadex column to remove most free dye (10 min)
- C Dialyse away rest of free dye (24 hr)

# Site-specific protein labelling methods

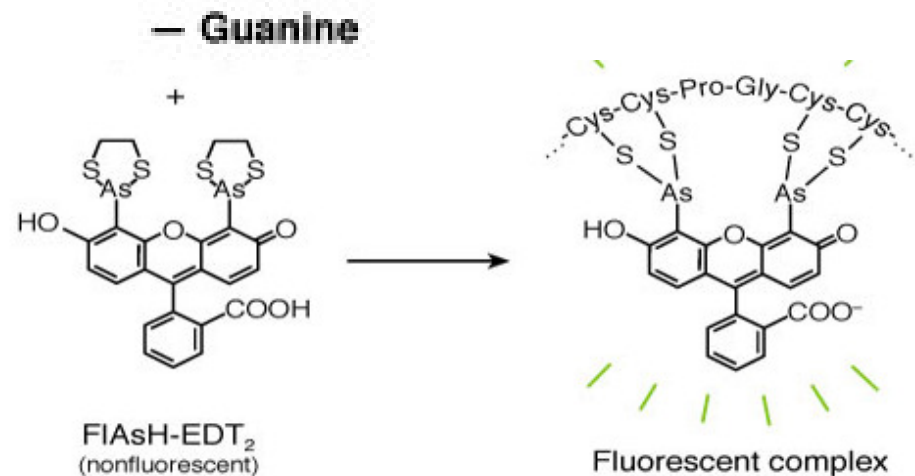
## 1. Binding domain

SNAP-tag (NEB), HaloTag (Promega)



## 2. Binding peptide

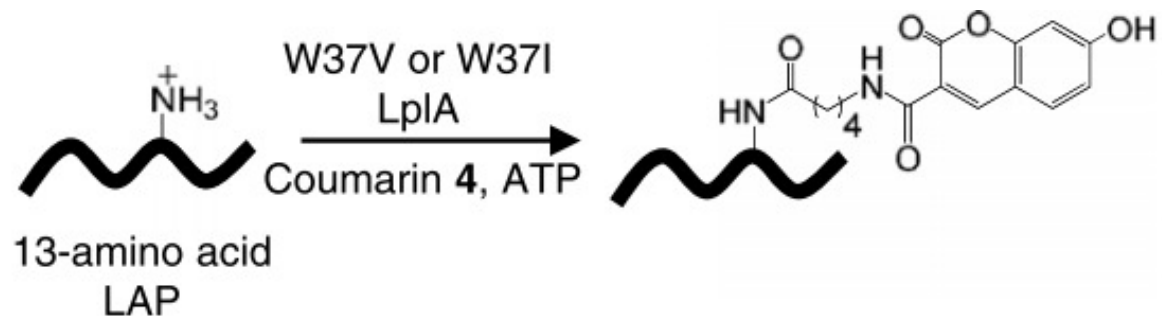
FIAsh (Invitrogen)



## 3. Enzymatic ligation to peptide

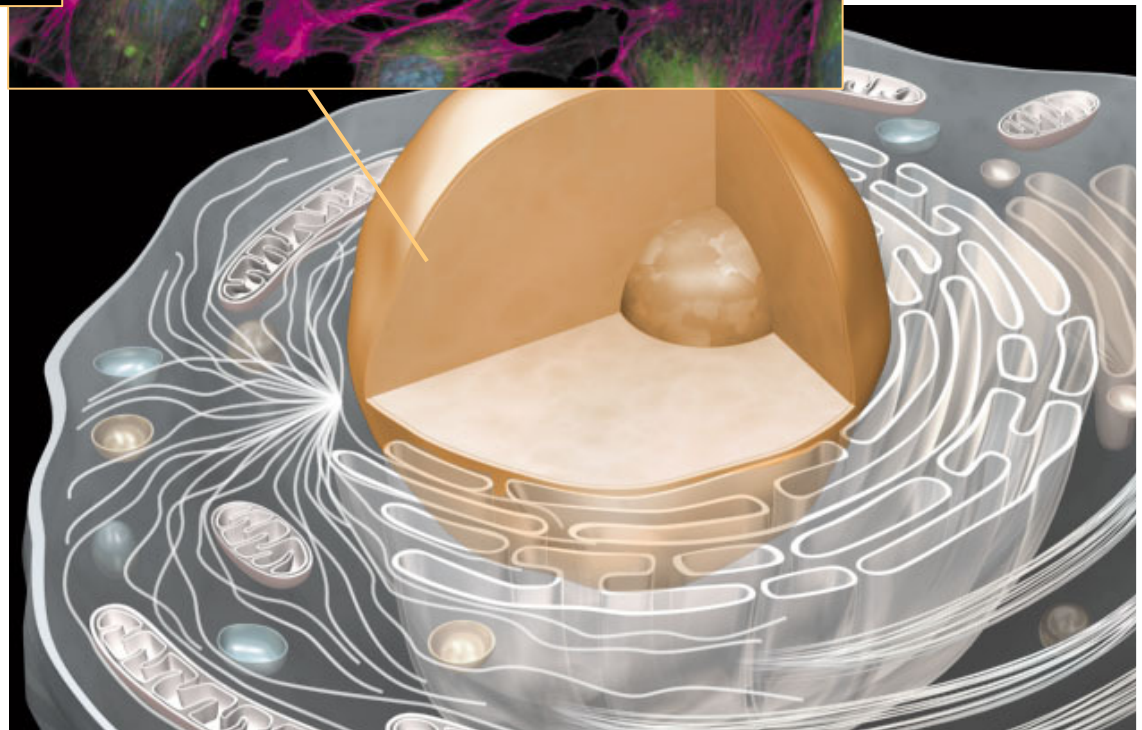
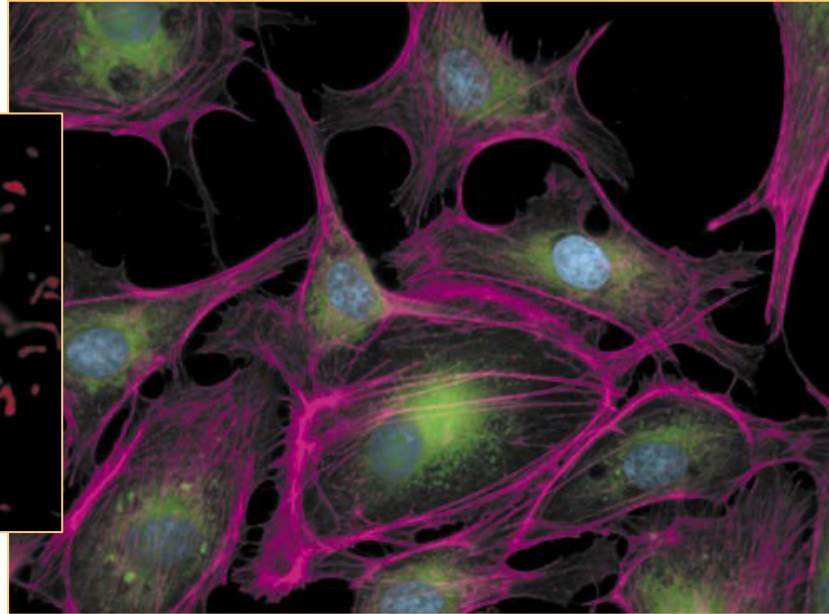
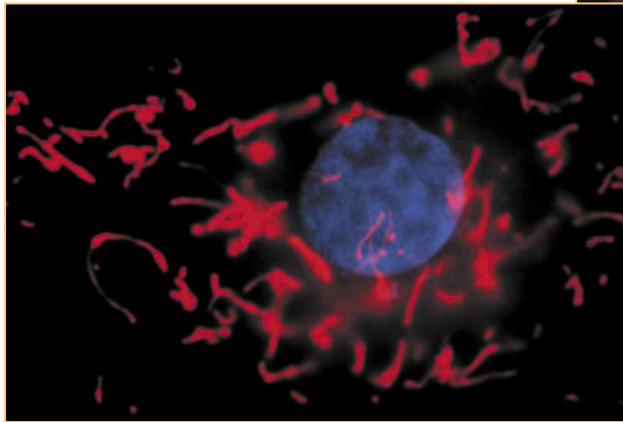
PRIME

AY Ting PNAS 2010



# Overview

1. What is fluorescence
2. What kind of structures are fluorescent
3. How to make and target fluorescent probes
4. Fluorescent probes for cellular structure



## Nucleus

DAPI

(well away from other channels)

Hoechst 33342

Live cells:

usually histone H2B-GFP

(or other monomeric FP)

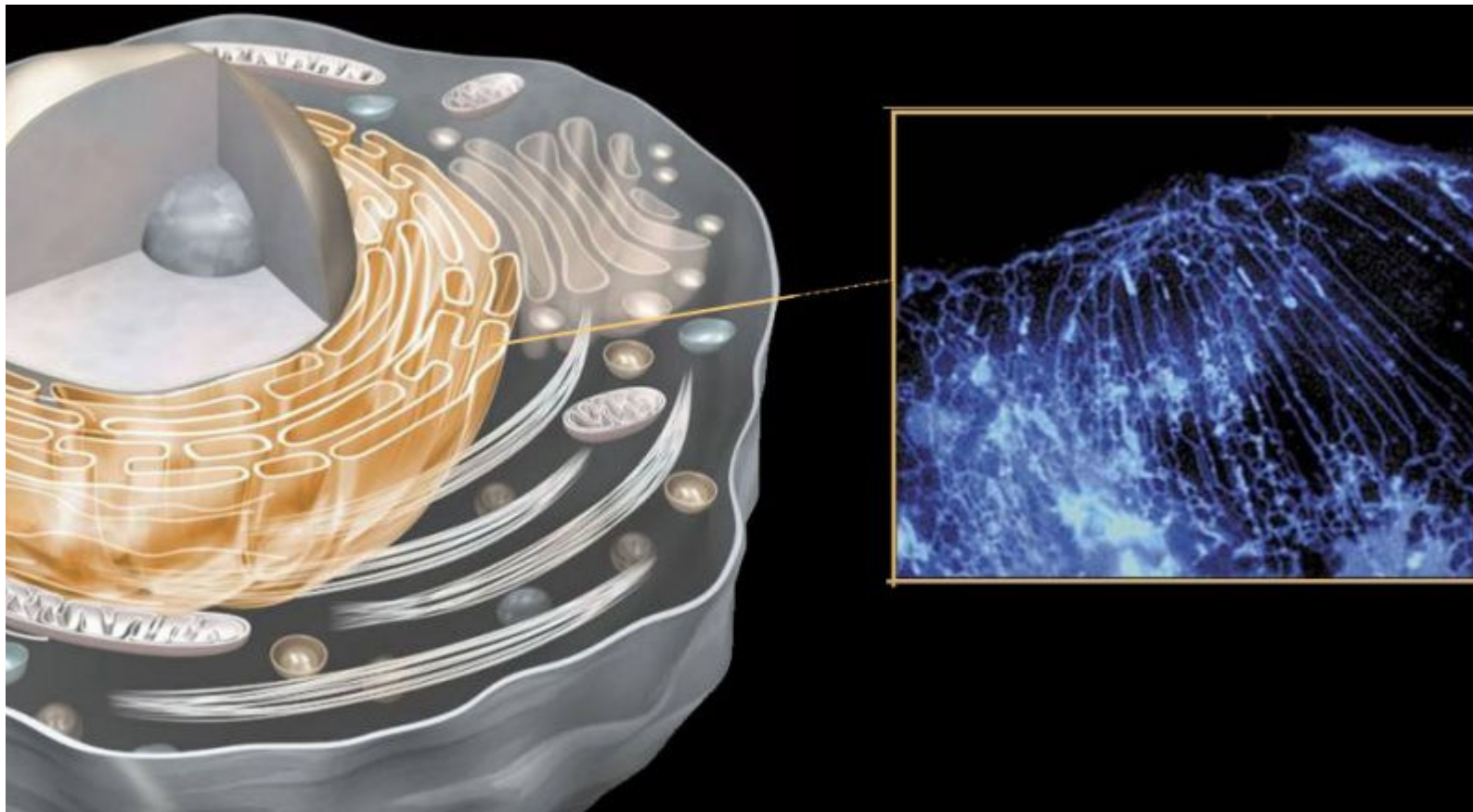
# Endoplasmic Reticulum

ER-Tracker™ Blue-White DPX

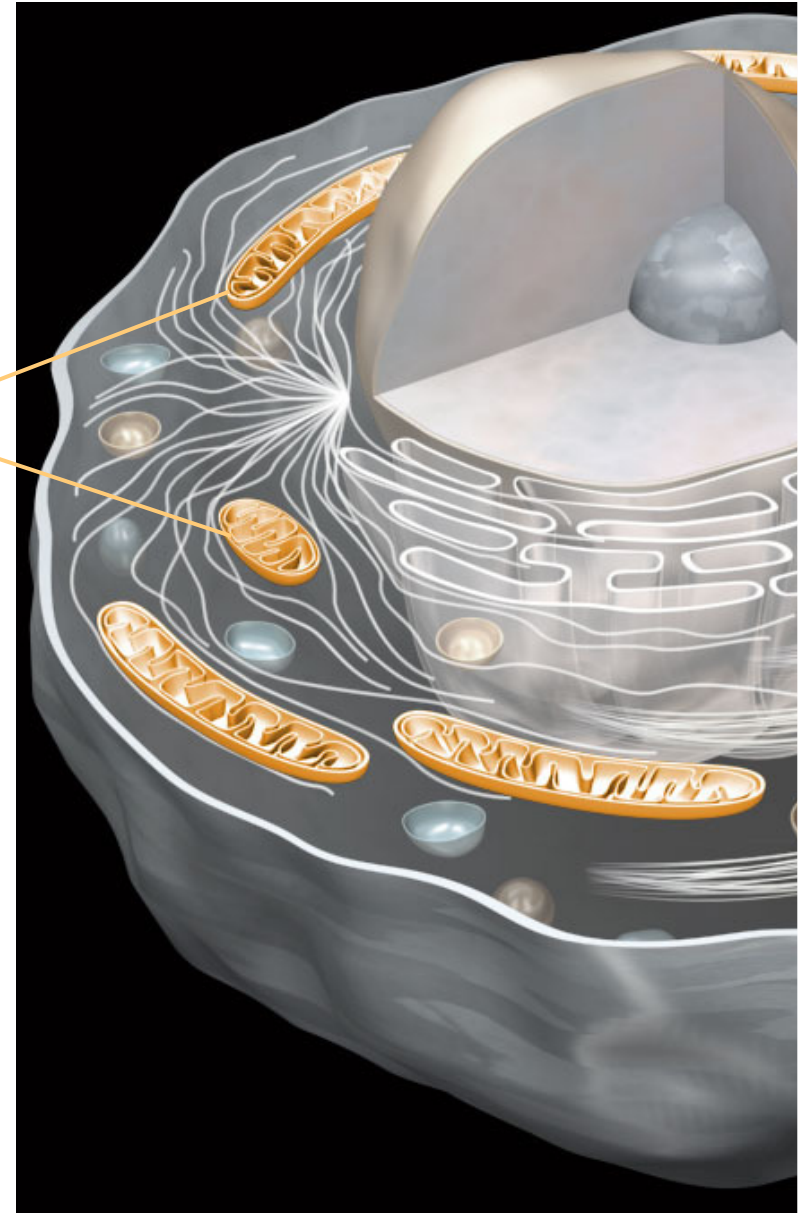
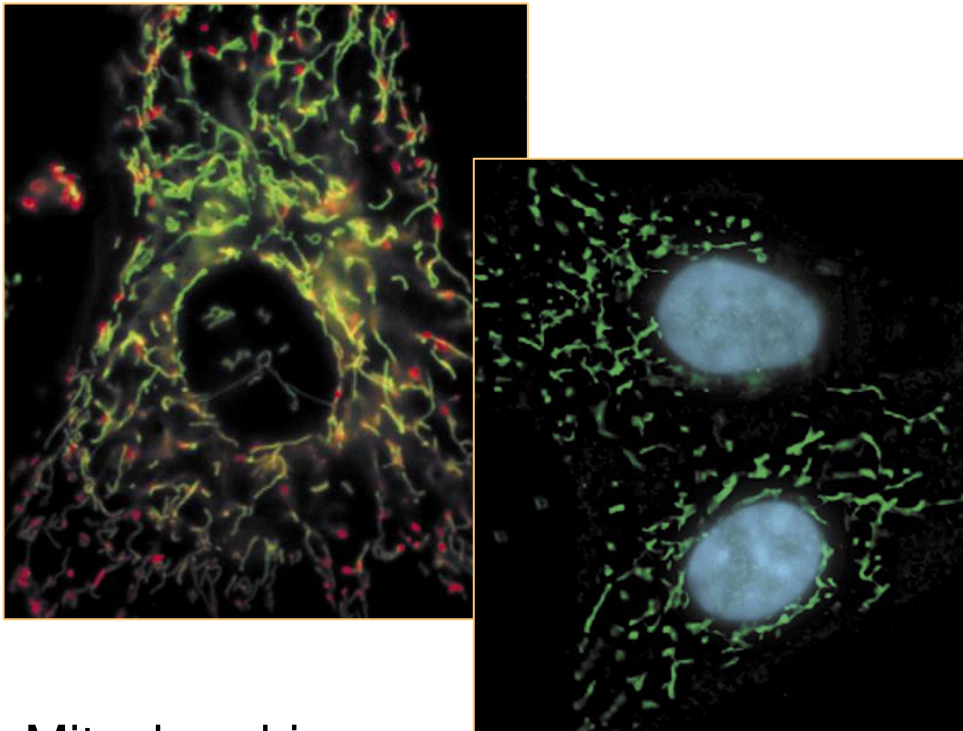
antibody to calnexin

Brefeldin A-BODIPY® 558 conjugate

Live cells: ss-GFP-KDEL







## Mitochondria

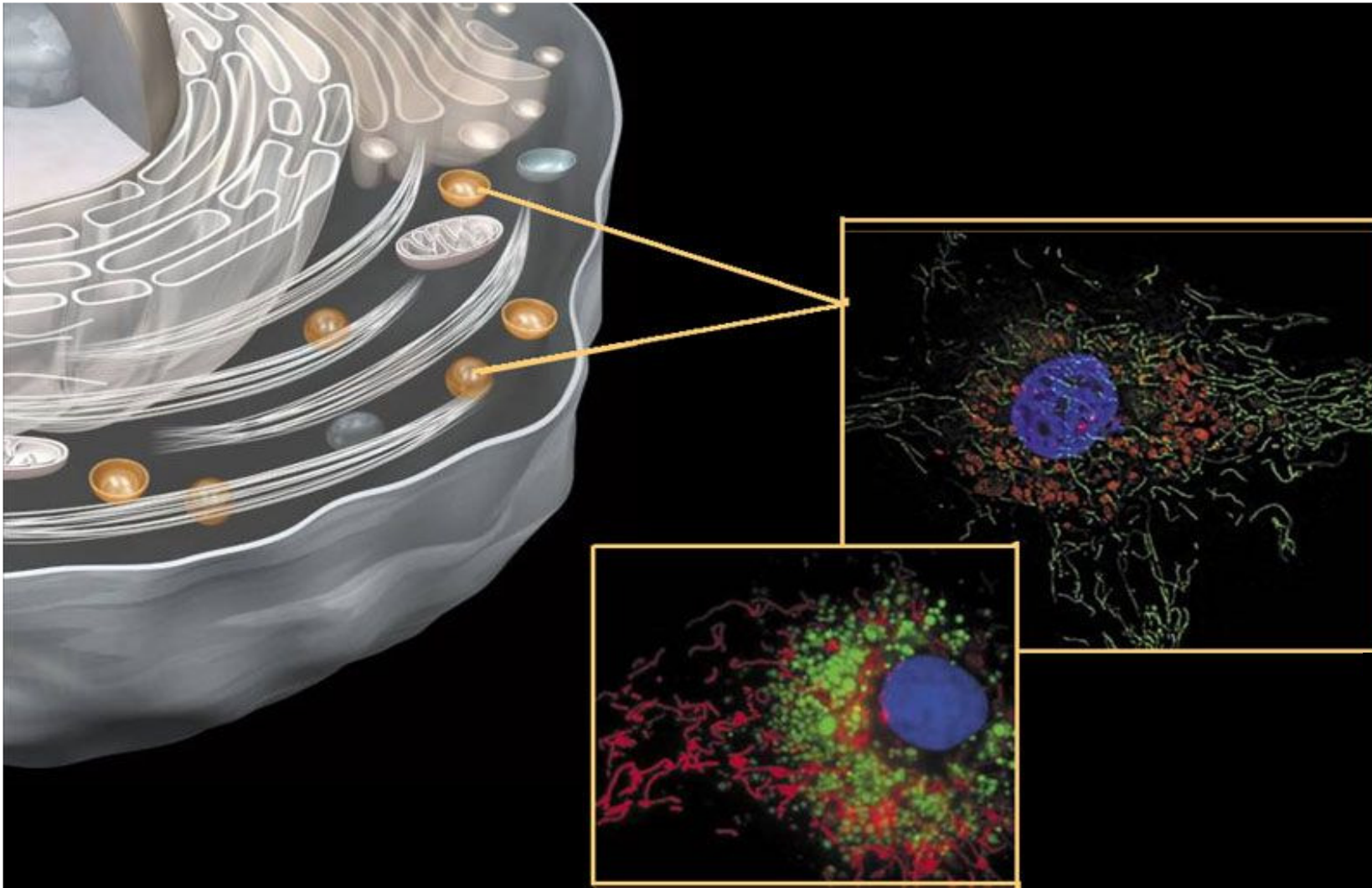
Fixed cells: anti-cytochrome oxidase subunit I Ab

Live cells: MitoTracker® Red/Green/Orange

CMTMRos

JC-1 (red J-aggregates at high conc., red to green depends on membrane potential)

Mitochondrial targeting sequence-GFP

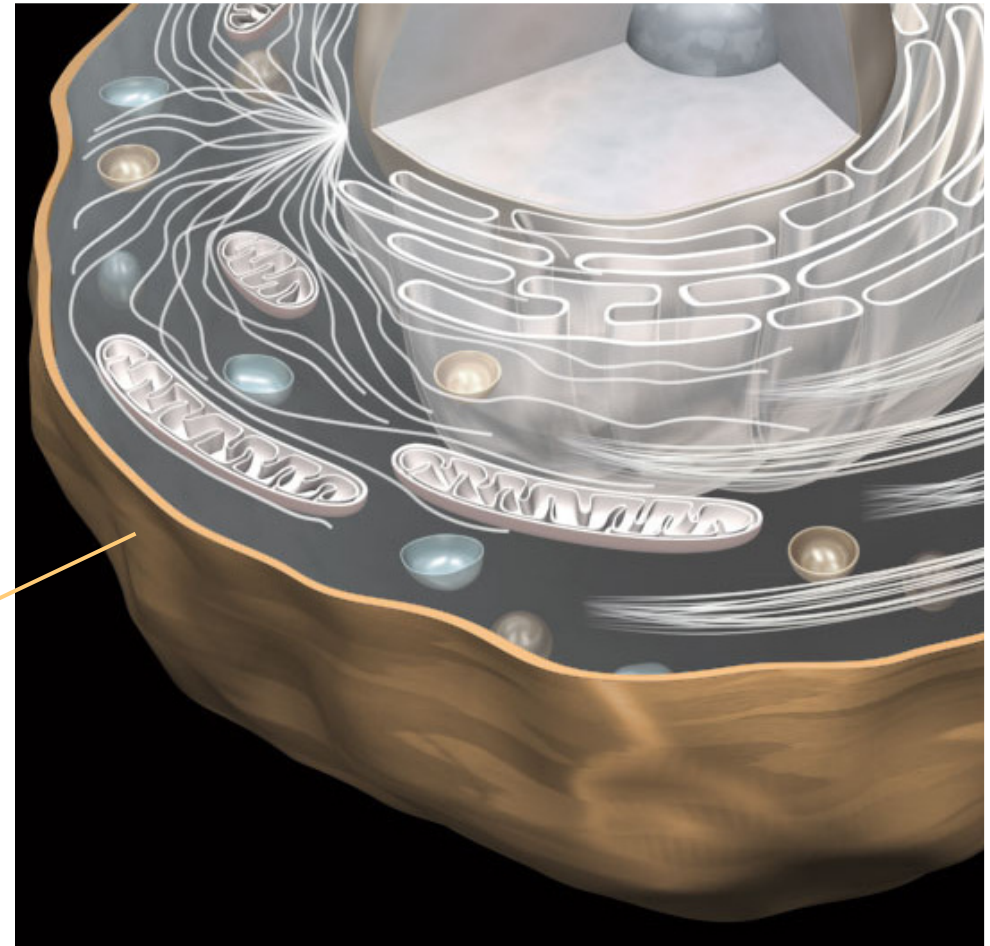
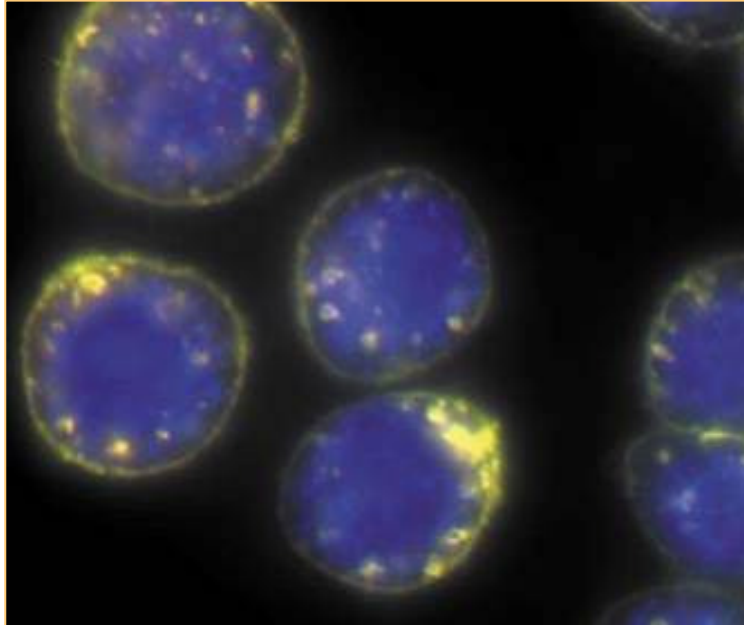


## Lysosomes

Fixed cells: anti-LAMP1

Live cells: LysoTracker® Red /Green (weakly basic amines can accumulate in lysosomes)

LysoSensor™ Yellow/Blue DND-160, LAMP1-GFP

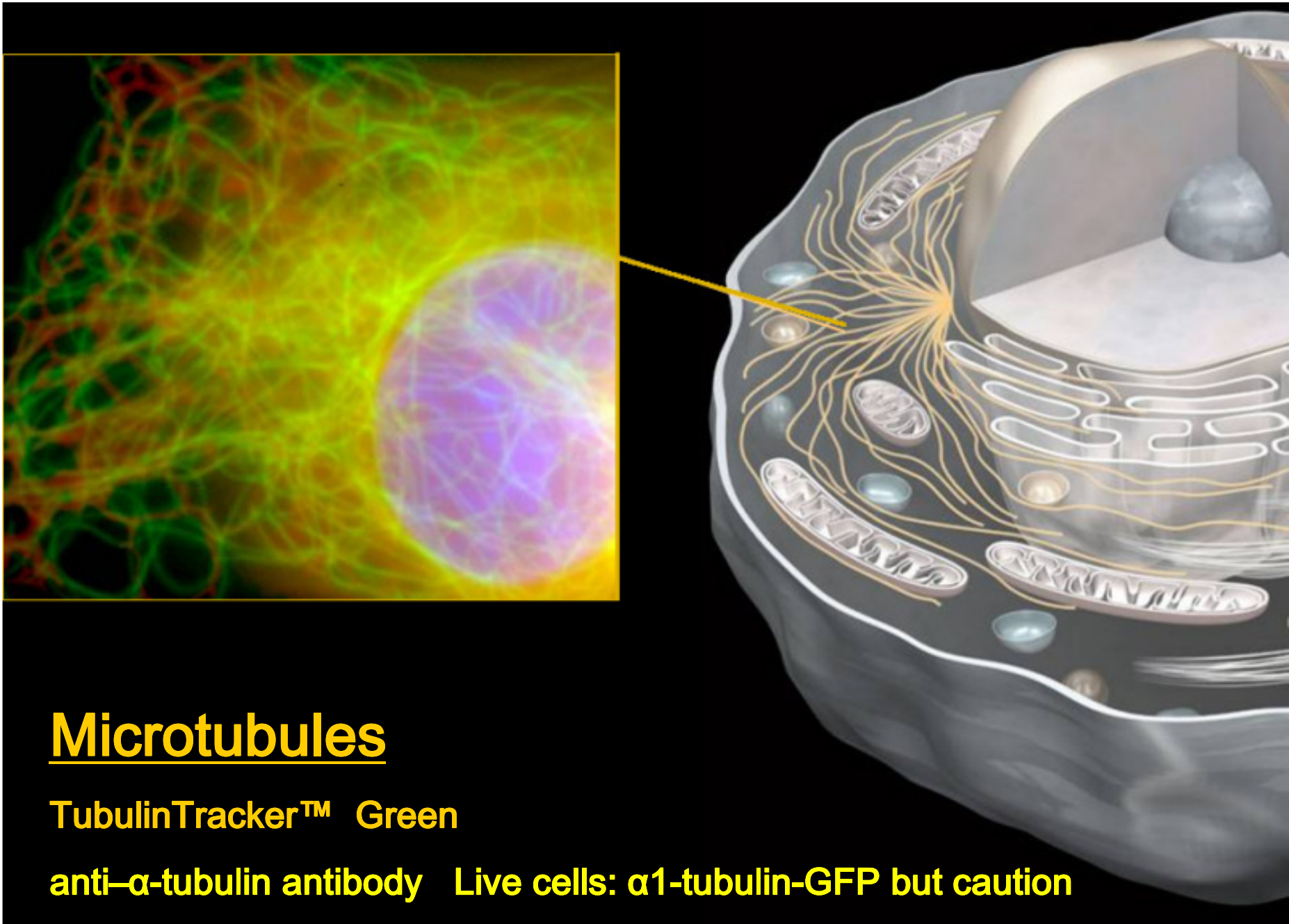


## Lipid Rafts

BODIPY® FL C<sub>5</sub>-ganglioside GM1

Fluorescent Cholera Toxin subunit B (CT-B)

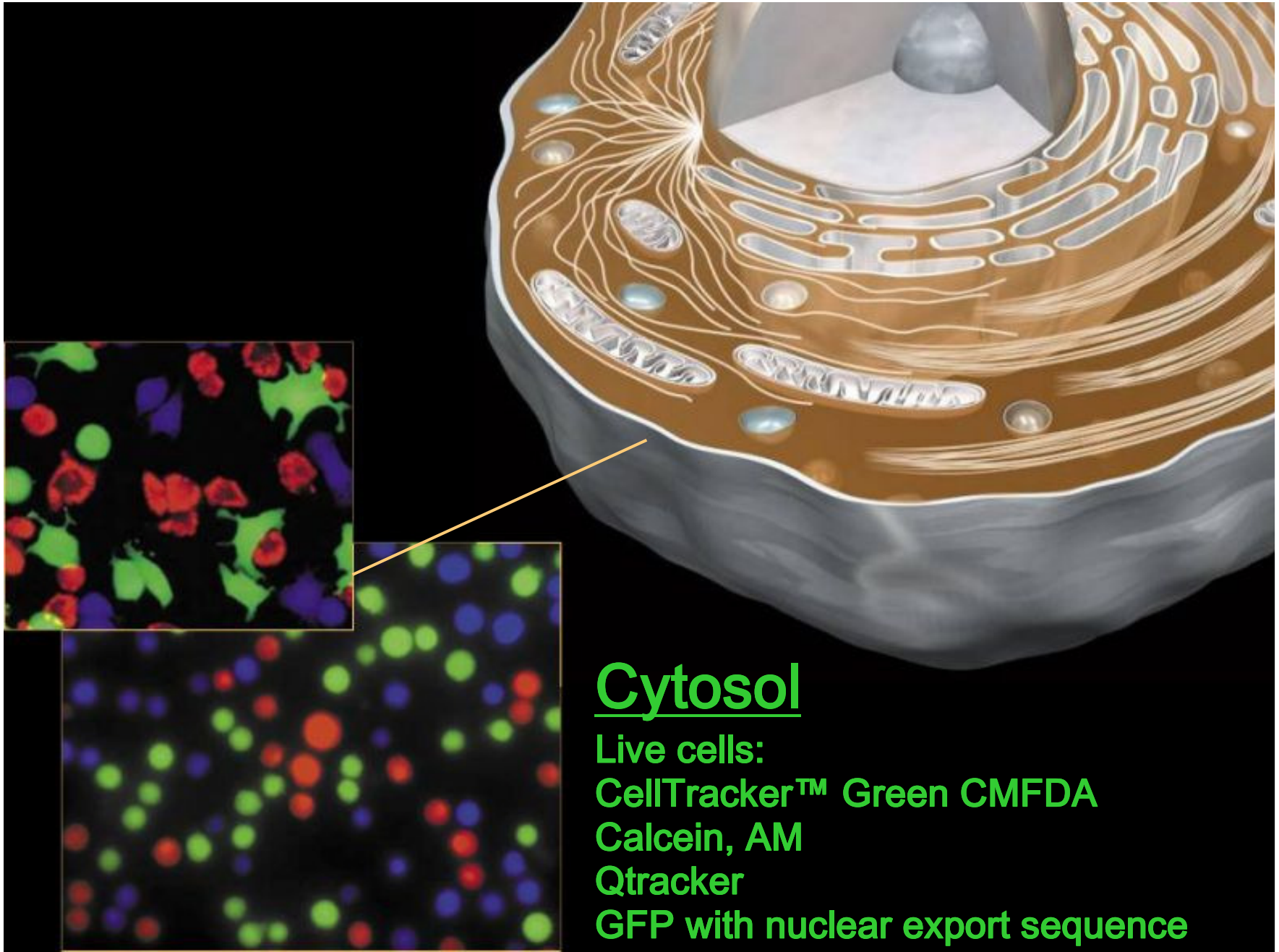




## Microtubules

TubulinTracker™ Green

anti- $\alpha$ -tubulin antibody Live cells:  $\alpha$ 1-tubulin-GFP but caution



# Cytosol

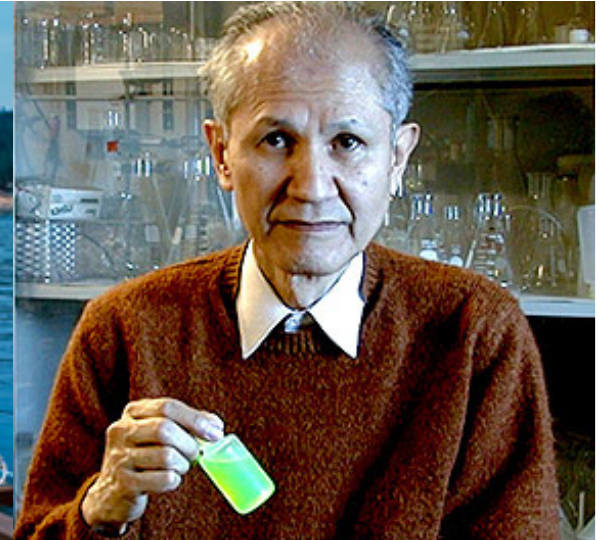
Live cells:  
CellTracker™ Green CMFDA  
Calcein, AM  
Qtracker  
GFP with nuclear export sequence



# The breakthrough of fluorescent proteins from jellyfish

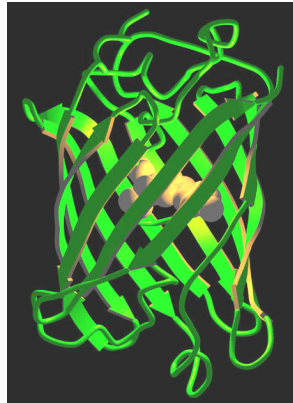


*Aequorea  
victoria*

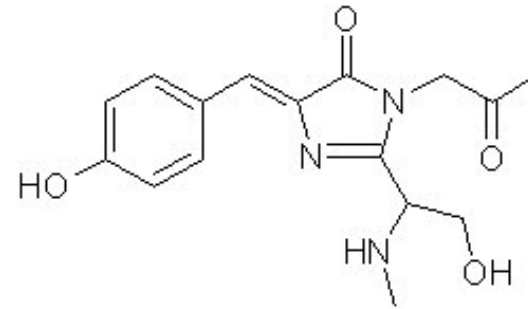


Osamu  
Shimomura

# The breakthrough of fluorescent proteins for live cell imaging



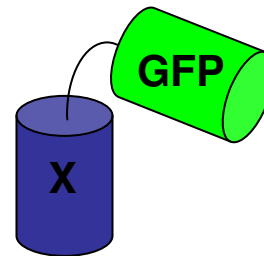
GFP fold  
 $\beta$ -can



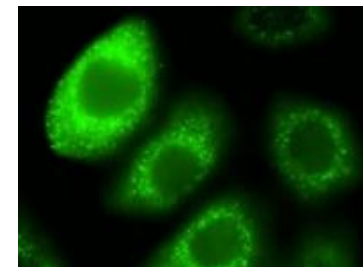
GFP chromophore  
from Ser-Tyr-Gly



Link GFP sequence to gene of your favourite protein

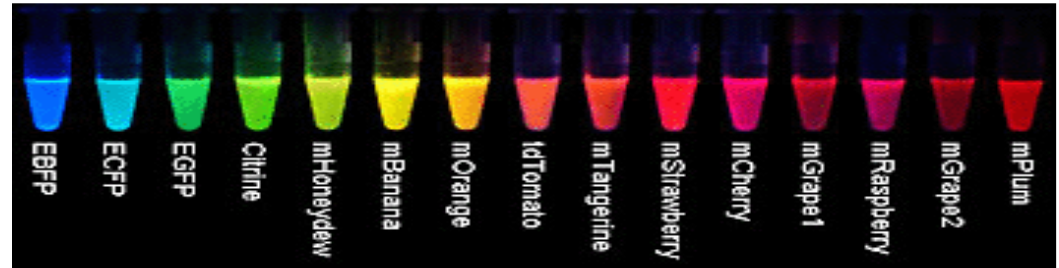


GFP folds and becomes fluorescent



GFP lights up your favourite protein in cell<sub>36</sub>

# Fluorescent proteins are more than just labels



## Photoactivation/Photoswitching

PA-GFP, Dronpa, Eos

## Reporting on environment

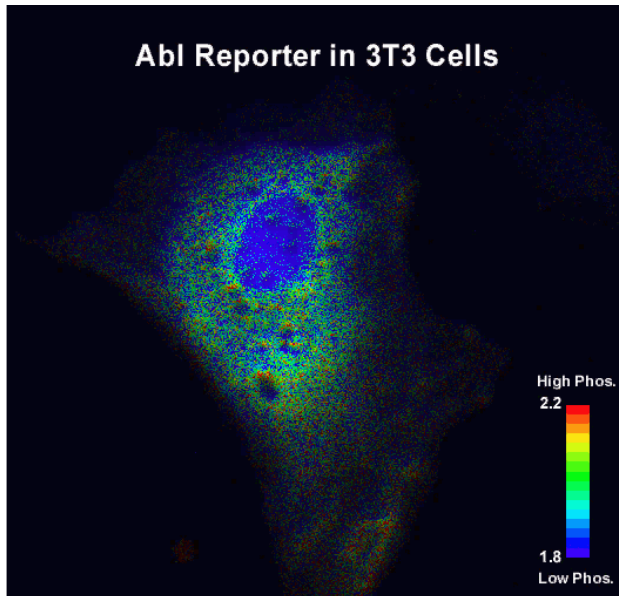
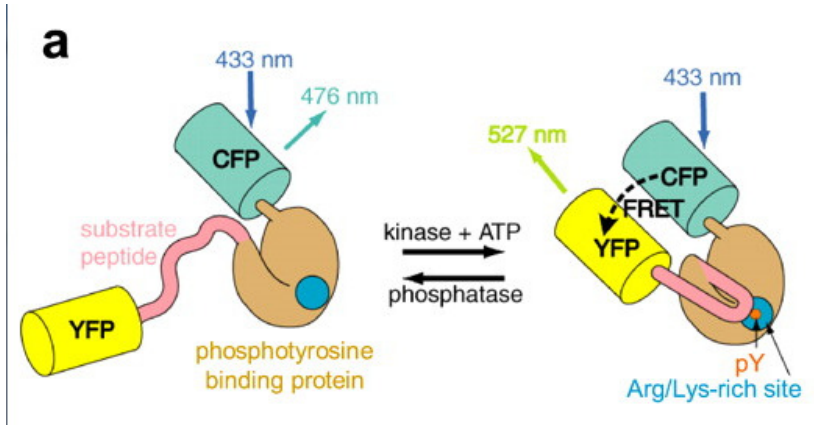
Ca<sup>2+</sup>, phosphorylation, cAMP, cGMP, pH, neurotransmitters, voltage, cell cycle, redox

## Reporting on protein-protein interaction

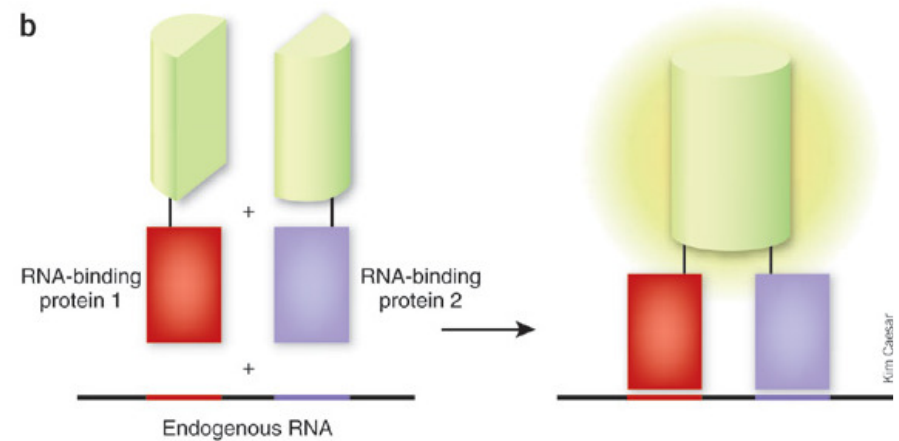
CFP/YFP FRET, split fluorescent proteins

## Modifying environment

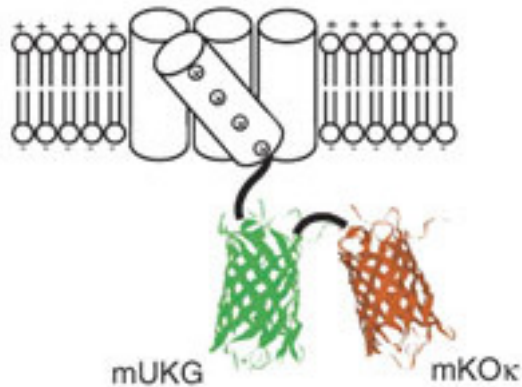
Singlet oxygen generation, Channelrhodopsin



**Targeting advantage to defined compartment, cell-type, developmental stage**



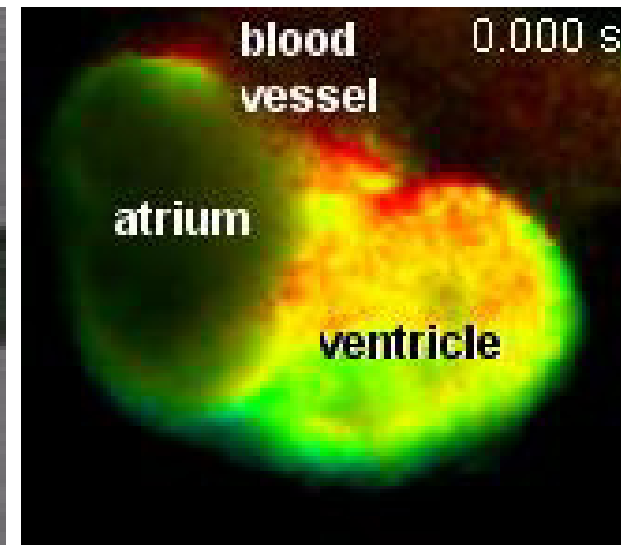
# Sensing voltage with fluorescent protein



Mermaid FRET voltage-sensor  
by FP fusion to voltage-sensing phosphatase

Expressed in zebrafish heart  
Non-invasive testing of mutant phenotypes  
and drug cardiotoxicity.

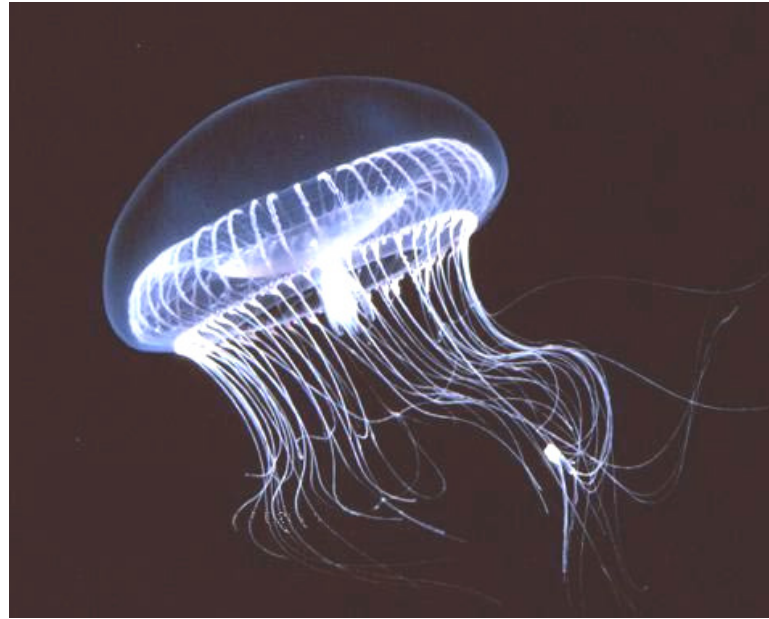
Tsutsui, Miyawaki J Physiol 2010



N.B. FRET sensor ratio crucial

best is YC2.60 cameleon: 600%,  
if <20% then lost in cellular noise

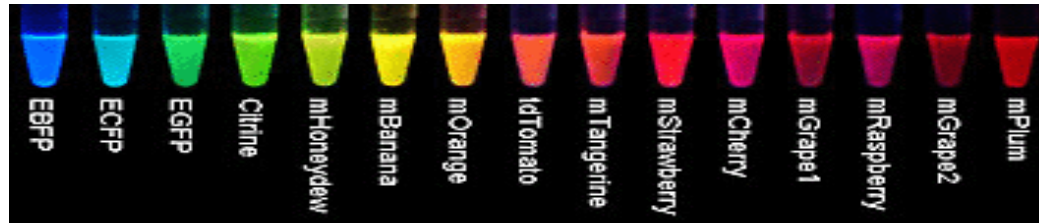
# How good is a fluorescent protein?



*A. victoria* GFP is good for jellyfish,  
but not great for cell biologists!



# How good is a fluorescent protein?



*A. victoria* GFP is terrible!

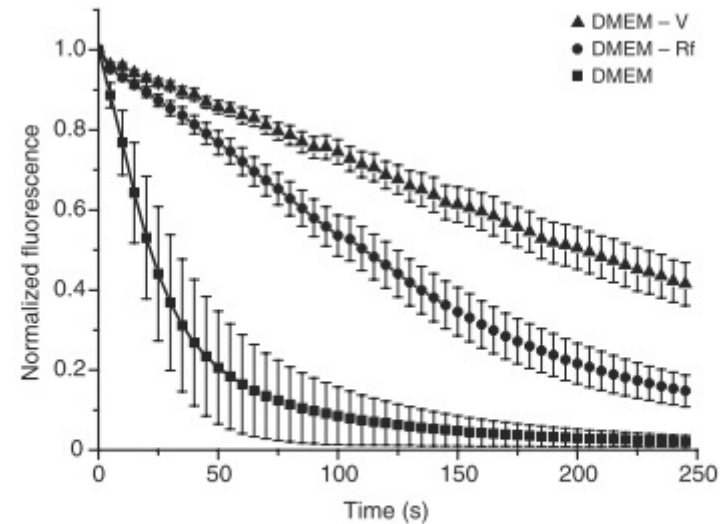
EGFP is OK, but there are now better...

1. Excitation and emission  $\lambda$       good match to filters on your microscope  
look at other fluorophores at same time
2. Bright       $\epsilon \times QY$       YPet 2.5 x EGFP  
TagRFP 2x mCherry
3. Stable to photobleaching      EBFP bad, mCherry and YPet good
4. Non-toxic      attach on right part of your protein  
all make  $H_2O_2$ , FPs can transfer electrons
5. Environment-insensitive      especially to pH, chloride  
CyPet does not fold at  $37^\circ C$ , all need  $O_2$   
Photoactivatable FP did not work in ER
6. Little non-specific binding      fully monomeric, A206K non-dimerising
7. Fast Maturing      Venus 2 min. Red FPs can start off green!  
half-time 40 min mCherry, 100 min TagRFP



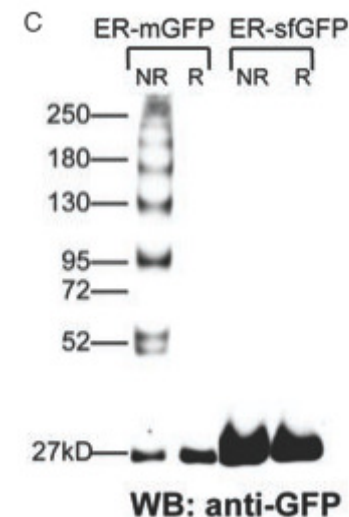
# Problems with GFP in cells

- **GFP with light can donate electrons to different acceptors**  
(FMN, FAD, NAD<sup>+</sup>, cyt. c)  
GFP reddens after transfer:  
photobleaching and phototoxicity  
use DMEM lacking e<sup>-</sup> acceptors  
(riboflavin or all vitamins) for less bleaching  
(HEK 293T happy for 1 week)  
effect for EGFP and PA-GFP, not RFPs  
*Lukyanov Nat Meth 2009*



**Figure 1** | Influence of cell medium on fluorescent protein photostability. Normalized bleaching curves for EGFP in live HEK293T cells maintained in DMEM, DMEM - Rf or DMEM - V. Error bars, s.d. ( $n = 20$  cells).

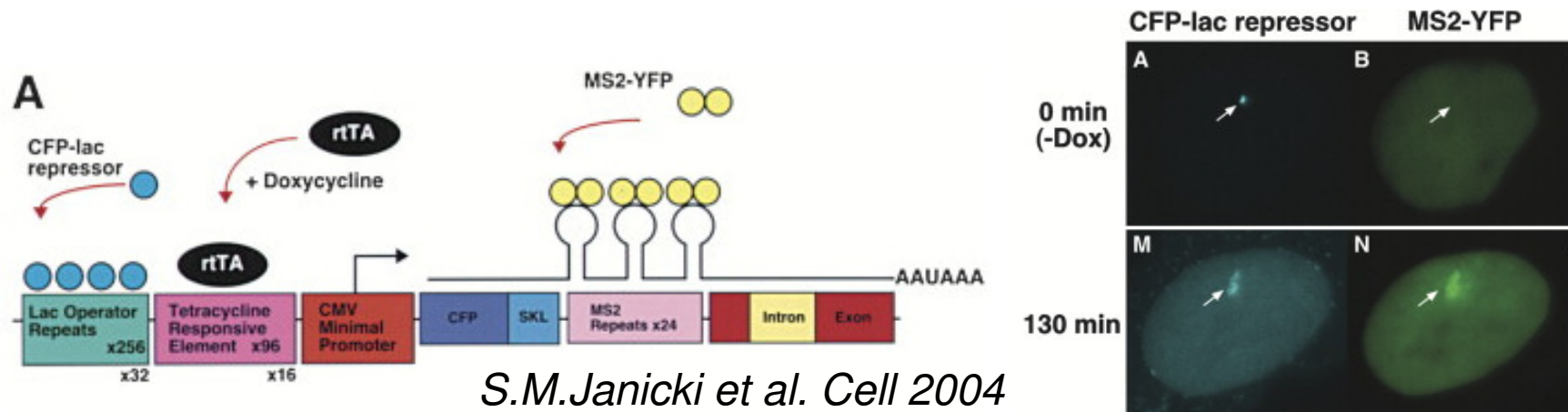
- **EGFP not good in secretory pathway**  
mixed disulfide oligomers in ER and non-fluorescent in *E. coli* periplasm  
(superfolder GFP behaves fine)  
*Erik Snapp, Traffic 2011*



# Fluorescent RNA imaging

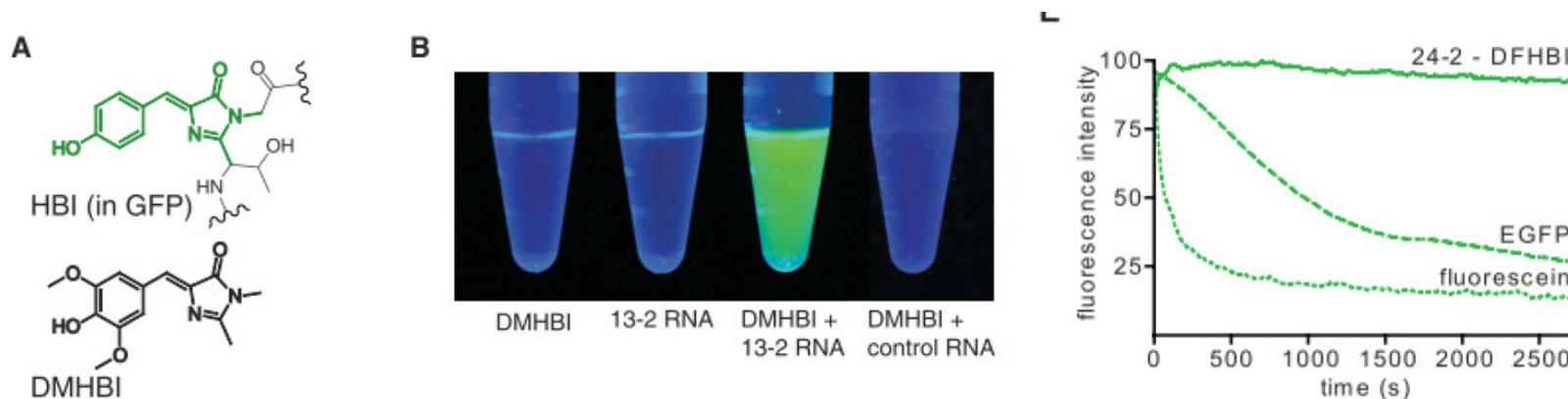
See single mRNA: mRNA stem-loops bound by MS2-YFP

See product of translation: mRNA encodes CFP-SKL which goes to peroxisomes



Spinach RNA 60 nt aptamer binds cell-permeable fluorogenic dye

Photostable. Used to label 5S RNA in HEK cells. *Samie Jaffrey Science 2011*



# Why use light to control biology?

Light control allows extreme temporal and spatial control.

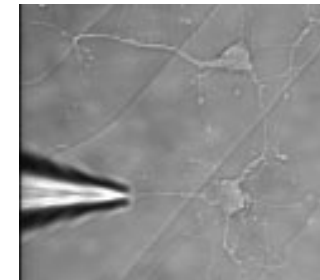


## Temporal control

genes < chemicals < light  
*min-hr*    *s-min*     $\mu\text{s-s}$

## Spatial control

chemicals / genes < light  
*one or many cells*    *1  $\mu\text{m}$  part of cell*



*(note micropipettes for precise small molecule delivery)*

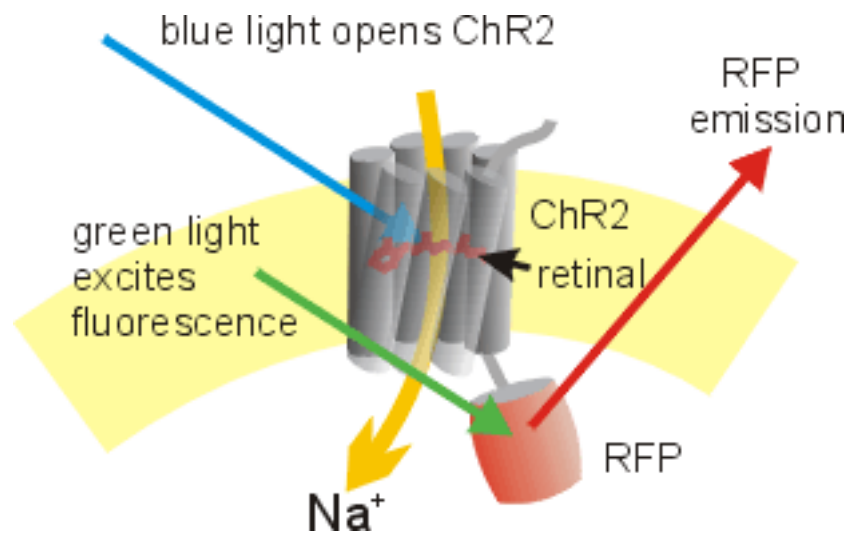
(often combine chemical/light control or gene/light control)  
optogenetics/chemogenetics

Limitations of light? \$\$\$\$\$

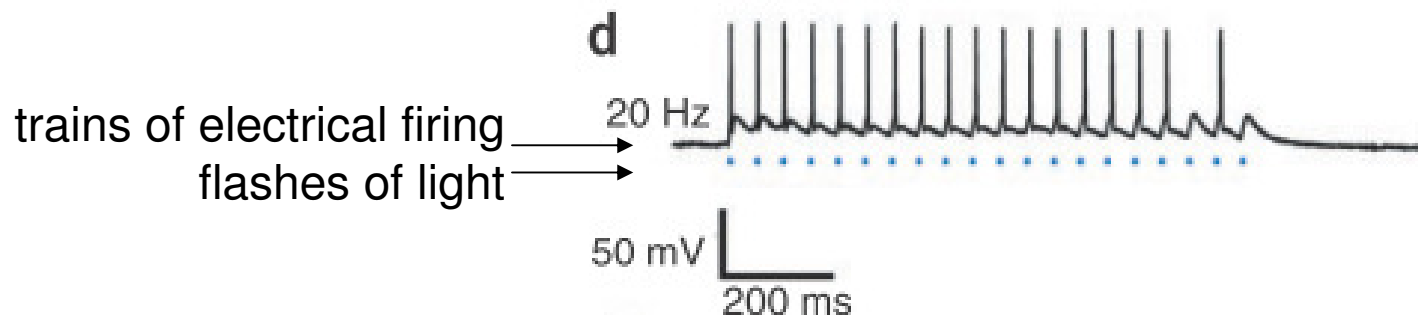
and usually data on one cell at a time

# Controlling biology with light: light-gated ion channels

Channelrhodopsin from an alga, like rhodopsin, undergoes retinal isomerisation in response to light, and changes conformation, but opens a  $\text{Na}^+$  channel. This allows light to control membrane voltage and trigger neuron firing.



to understand neuronal firing patterns  
to control secretion in diabetes  
potentially in fixing neural diseases?  
e.g. damping down overactivity in  
epilepsy

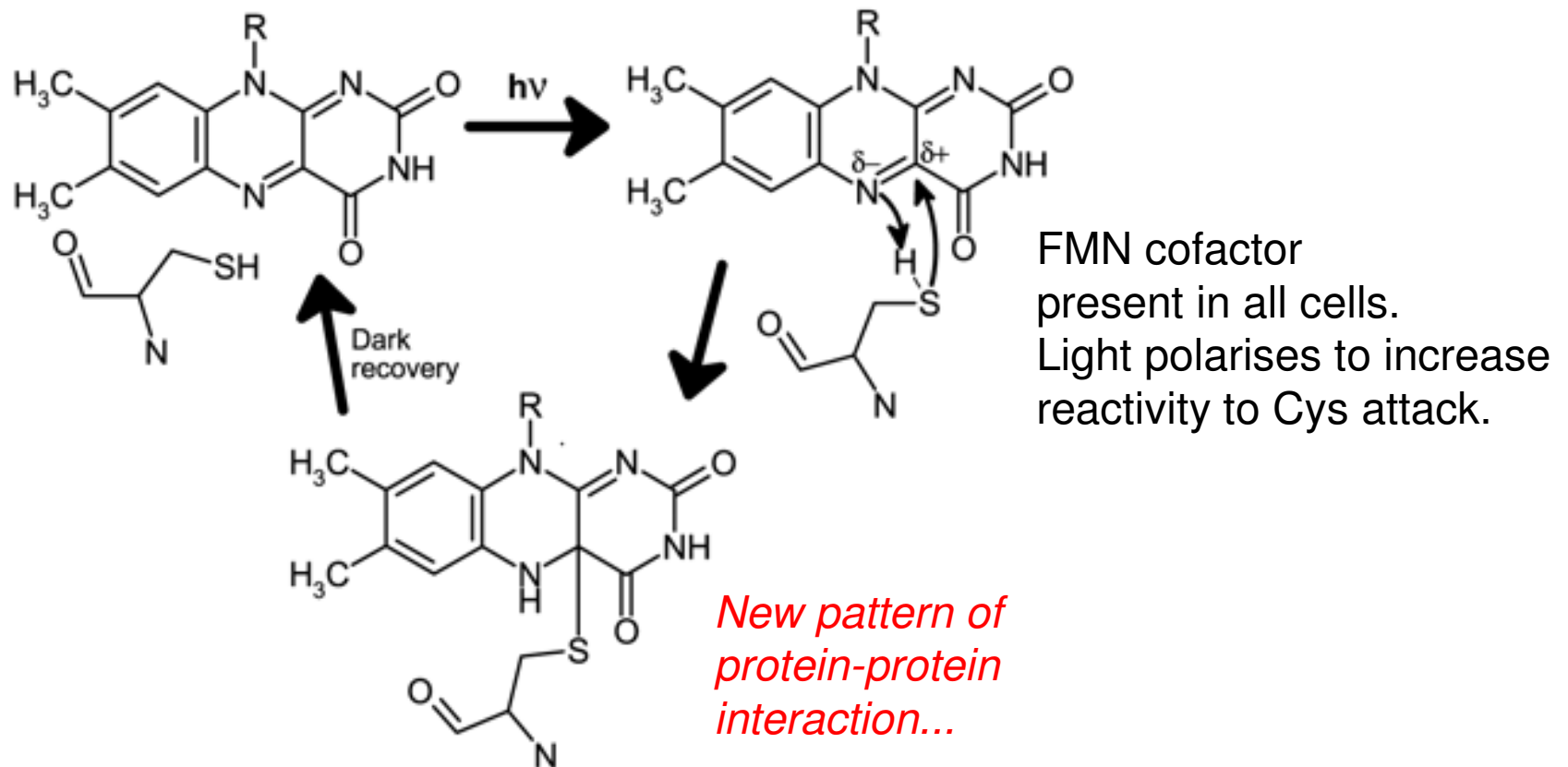


# LOV domains react and switch conformation with light

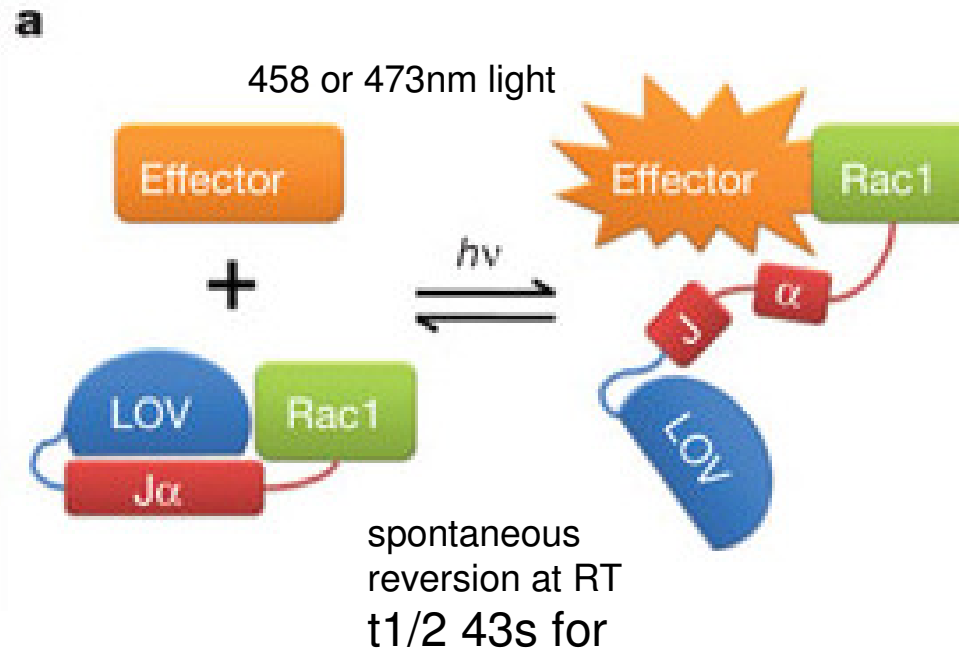
LOV domains:

light, oxygen, voltage responders

ones responding to blue light in bacteria, plants and fungi



# Genetically-encoded photoactivation



1. Constitutively active Rac mutant
  2. Optimise LOV-Rac junction,
  3. knockout GTP hydrolysis and GAP/GNDI/GEF interactions
- $K_d$  for PAK 2  $\mu$ M in dark, 200nM in light 10-fold ratio
- Interaction of Rac with PAK stimulates cell protrusion and migration.



# Correlated Light Microscopy/ Electron Microscopy

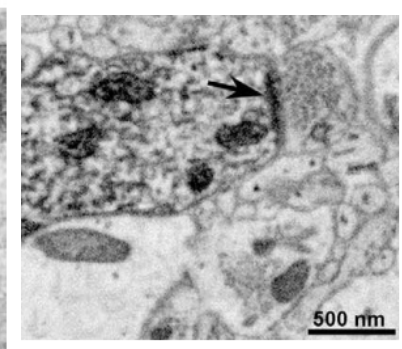
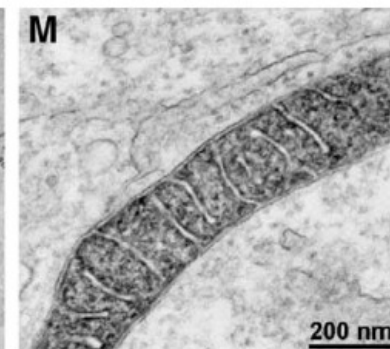
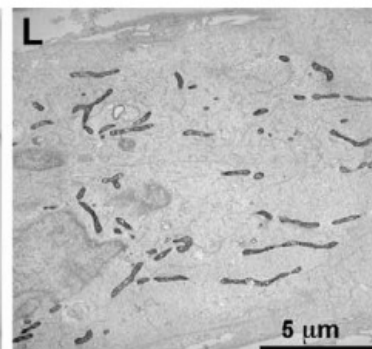
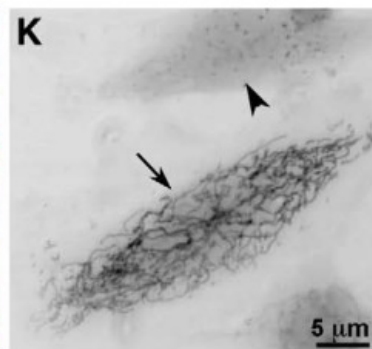
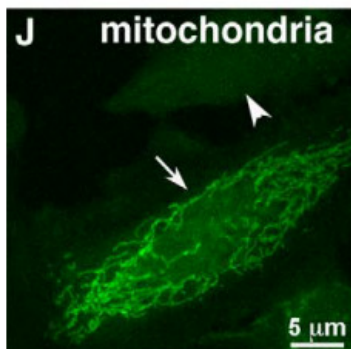
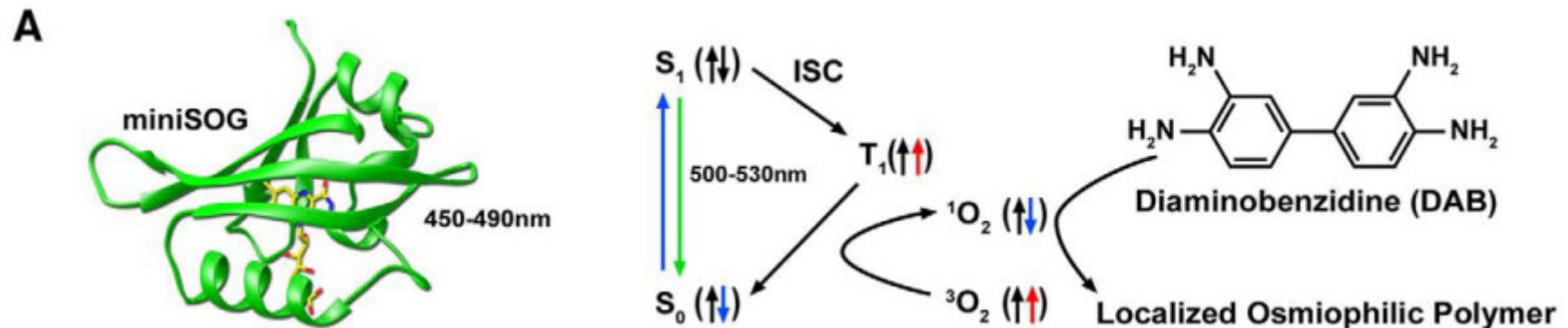
MiniSOG (Shu, Tsien PLoS Biol 2011)

Light causes generation of singlet oxygen  $\rightarrow$  DAB polymerized  $\rightarrow$  binds  $\text{OsO}_4$

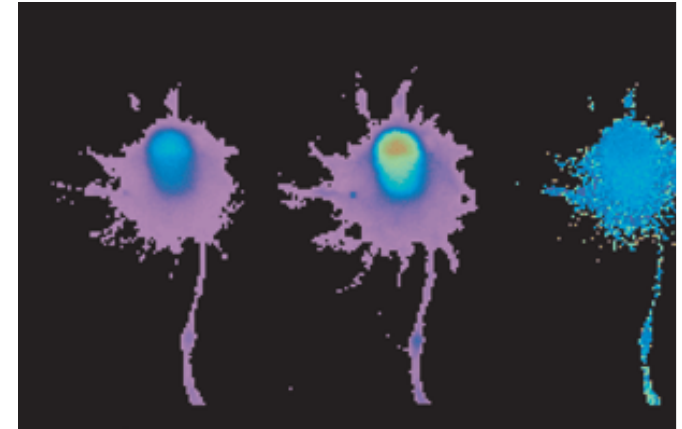
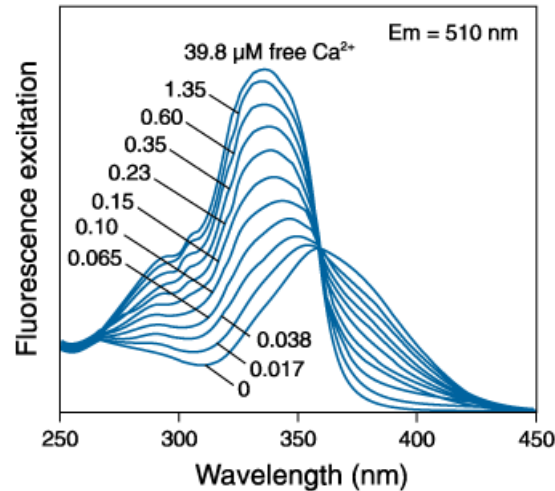
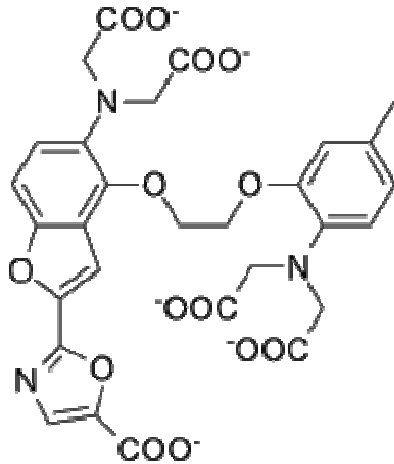
106 aa monomer, engineered from Arabidopsis LOV domain

tested in cell-lines, worms and transgenic mice

Overcomes trade-off between thorough fixation and penetration of labeling reagent



# Small molecule fluorescent sensors



Fura-2 sensing calcium

**Metal ions:** calcium, magnesium, zinc, sodium, potassium, chloride, mercury

**pH** (also dyes to conjugate to proteins, CyPher from GE, SNARF from Invitrogen)

**Reactive oxygen species, nitric oxide**

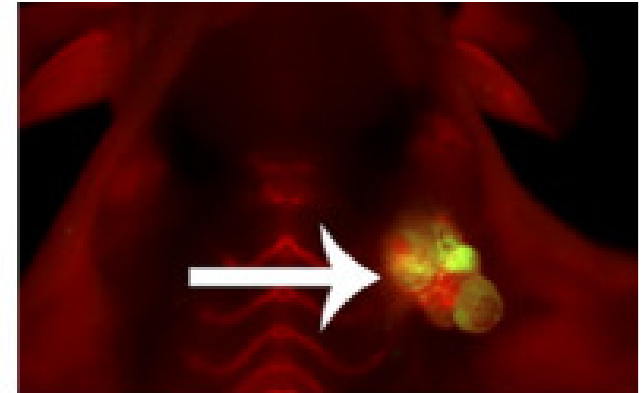
**Transmembrane potential**



# Why use small molecule rather than genetically-encoded probes?

## 1. No need to transfect

- hard for some organisms and primary cells
- easier to titrate
- potential clinical application- e.g. image-guided surgery



MMP-activated Cy5 peptide labels tumour (RY Tsien 2010)

## 2. Probes often brighter, with bigger signal to noise

- struggle to make GFP-based calcium reporter as good as fura-like dyes

## 3. Probes with entirely different fluorescent properties

- QD photostability, probes with long fluorescence lifetimes, photouncaging

## 4. Smaller

- e.g. calcium conc. right next to pore of ion channel

# References

## Fluorescence probes

Molecular Probes Handbook, free from Invitrogen.  
Principles of Fluorescence Spectroscopy 2<sup>nd</sup> edition,  
by Joseph R. Lakowicz.

## Protein modification

Bioconjugate Techniques, 2<sup>nd</sup> Edition  
by Greg T. Hermanson.  
Chemical labeling strategies for cell biology, Marks  
KM, Nolan GP. Nat Methods. 2006 Aug;3(8):591-6.

## Fluorescent proteins

(i) as labels: A guide to choosing fluorescent proteins.

Shaner NC, Steinbach PA, Tsien RY.

Nat Methods. 2005;2(12):905-9.

Poster: Fluorescent proteins illuminate cell biology

Lin M, Miyawaki A, Tsien RY.

<http://www.nature.com/nrm/posters/fluorescent/index.html>

(ii) as sensors: Creating new fluorescent probes for

cell biology. Zhang J, Campbell RE, Ting AY, Tsien

RY. Nat Rev Mol Cell Biol. 2002 Dec;3(12):906-18.



**nature REVIEWS MOLECULAR CELL BIOLOGY**

**Fluorescent proteins illuminate cell biology**  
Michael Z. Lin, Atsushi Miyawaki and Roger Y. Tsien

The ability to unveil the fine details of biological function has advanced remarkably. Green fluorescent protein (GFP) isolated from the jellyfish *Aequorea victoria* and GFP-like fluorescent proteins from other animals have become essential tools in the biological laboratory that have driven these advances. Inspired by the emergence of numerous specialized variants of fluorescent proteins in the past decade, an increasing number of researchers have been creating the development of a tool that enables the direct visualization of biological function. Although the more common application of fluorescent proteins is the imaging of gene expression and protein dynamics, biosensors have also been created using fluorescent proteins to image the concentration of ions and small molecules, enzymatic activity, protein-protein interactions and changes in protein conformation. In addition, an emerging property of some fluorescent proteins is that their fluorescence can be photo-modulated by illumination of specific wavelengths, which enables individual cells, organelles and proteins to be highlighted with high spatiotemporal resolution. This poster provides a broad perspective of the fluorescent proteins now available and their potential applications.

**Optical properties of bright fluorescent proteins**

Protein	Excitation (nm)	Emission (nm)	Quantum Yield	Stability
EGFP	365	510	0.52	High
EGFP-1	365	510	0.52	High
EGFP-2	365	510	0.52	High
EGFP-3	365	510	0.52	High
EGFP-4	365	510	0.52	High
EGFP-5	365	510	0.52	High
EGFP-6	365	510	0.52	High
EGFP-7	365	510	0.52	High
EGFP-8	365	510	0.52	High
EGFP-9	365	510	0.52	High
EGFP-10	365	510	0.52	High
EGFP-11	365	510	0.52	High
EGFP-12	365	510	0.52	High
EGFP-13	365	510	0.52	High
EGFP-14	365	510	0.52	High
EGFP-15	365	510	0.52	High
EGFP-16	365	510	0.52	High
EGFP-17	365	510	0.52	High
EGFP-18	365	510	0.52	High
EGFP-19	365	510	0.52	High
EGFP-20	365	510	0.52	High

**Bright monomers**  
Bright monomeric fluorescent proteins are used for labeling individual cells and proteins, and are best suited for applications requiring high brightness and low background. The development of these fluorescent proteins has been driven by the need for high brightness and low background. These proteins are well suited for labeling individual cells and proteins.

**Bright multimers**  
Certain monomeric fluorescent proteins are better suited for labeling cells and proteins. These proteins are well suited for labeling individual cells and proteins.

**Reversibly photoswitchable proteins**  
Certain fluorescent proteins can be reversibly switched between a fluorescent and a non-fluorescent state. These proteins are well suited for labeling individual cells and proteins.

**Irreversibly photoswitchable proteins**  
Certain fluorescent proteins can be irreversibly switched between a fluorescent and a non-fluorescent state. These proteins are well suited for labeling individual cells and proteins.

**Basic features of fluorescent proteins**  
Fluorescent proteins exhibit a variety of features that make them useful for labeling cells and proteins. These features include high brightness, low background, and stability.

**Chromophore-modulating sensors**  
Certain fluorescent proteins can be used as sensors for monitoring changes in the chromophore. These sensors are well suited for monitoring changes in the chromophore.

**Intensity-based single-chromophore sensors**  
Certain fluorescent proteins can be used as sensors for monitoring changes in intensity. These sensors are well suited for monitoring changes in intensity.

**Ratiometric single-chromophore sensors**  
Certain fluorescent proteins can be used as sensors for monitoring changes in the ratio of two different wavelengths. These sensors are well suited for monitoring changes in the ratio of two different wavelengths.

**FRET sensors**  
Certain fluorescent proteins can be used as sensors for monitoring changes in FRET efficiency. These sensors are well suited for monitoring changes in FRET efficiency.

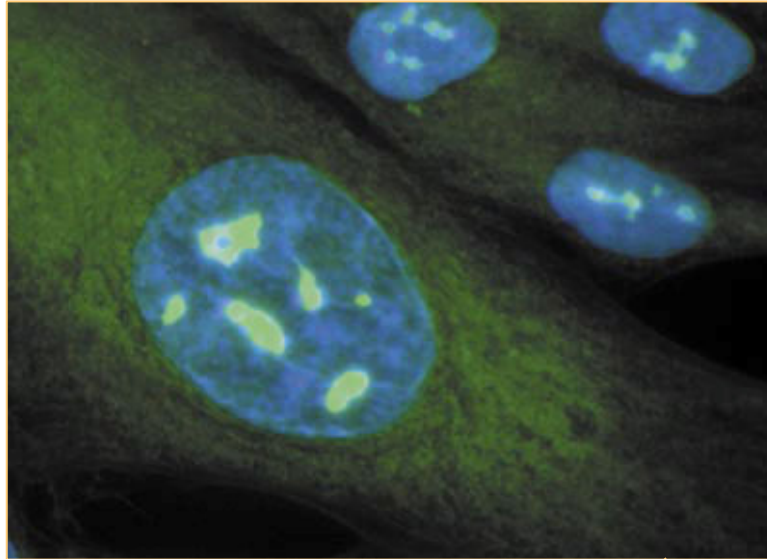
**Multiple sensors**  
Certain fluorescent proteins can be used as sensors for monitoring changes in multiple parameters. These sensors are well suited for monitoring changes in multiple parameters.

**Selected FRET sensor characteristics**

Sensor	Excitation (nm)	Emission (nm)	Quantum Yield	Stability
EGFP	365	510	0.52	High
EGFP-1	365	510	0.52	High
EGFP-2	365	510	0.52	High
EGFP-3	365	510	0.52	High
EGFP-4	365	510	0.52	High
EGFP-5	365	510	0.52	High
EGFP-6	365	510	0.52	High
EGFP-7	365	510	0.52	High
EGFP-8	365	510	0.52	High
EGFP-9	365	510	0.52	High
EGFP-10	365	510	0.52	High
EGFP-11	365	510	0.52	High
EGFP-12	365	510	0.52	High
EGFP-13	365	510	0.52	High
EGFP-14	365	510	0.52	High
EGFP-15	365	510	0.52	High
EGFP-16	365	510	0.52	High
EGFP-17	365	510	0.52	High
EGFP-18	365	510	0.52	High
EGFP-19	365	510	0.52	High
EGFP-20	365	510	0.52	High

**semrock**  
A Unit of **beck** Corporation

**ANDOR**  
TECHNOLOGY



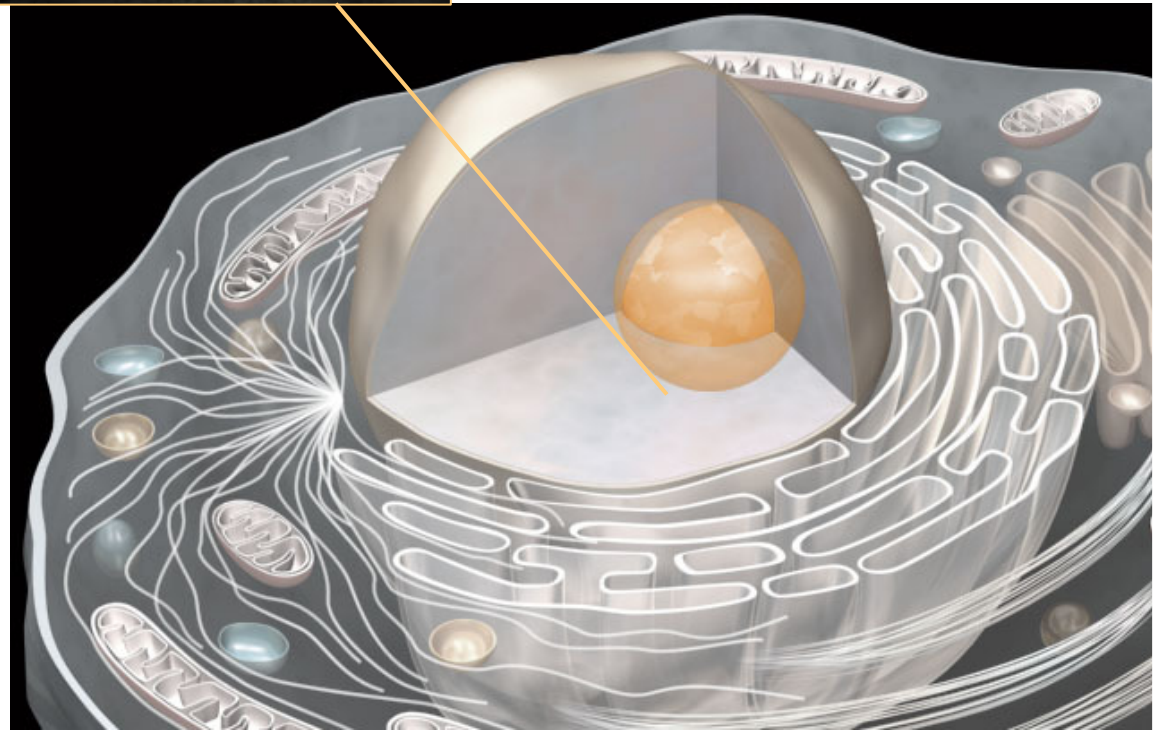
## Nucleoli

SYTO® RNASelect™

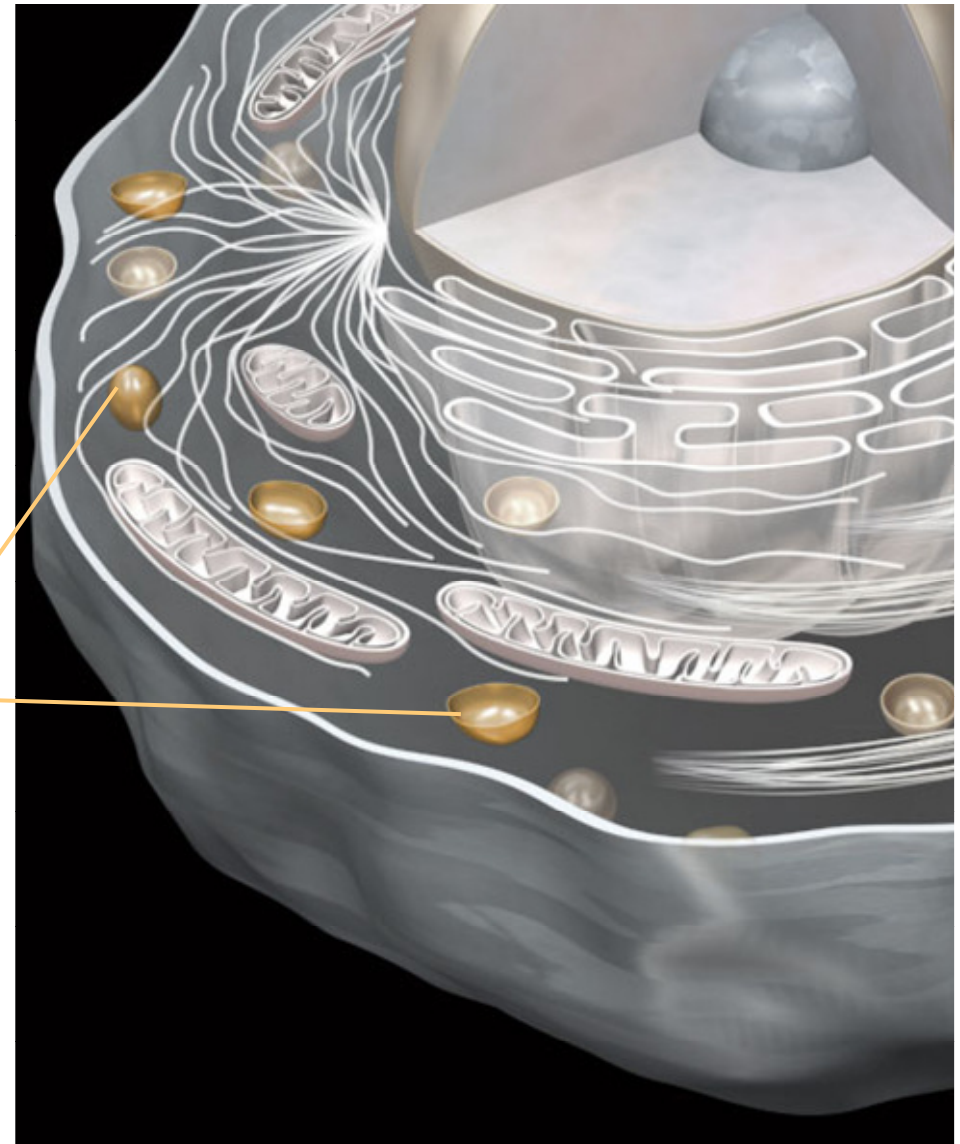
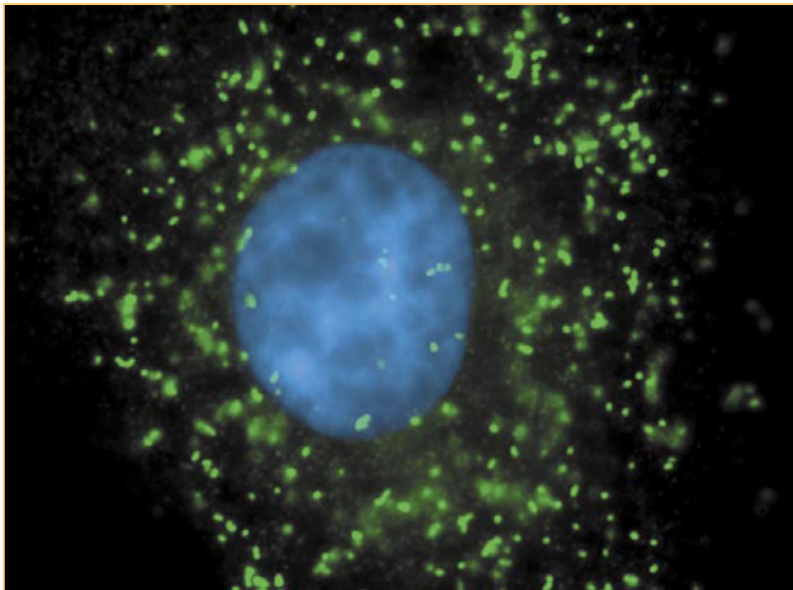
Live cells: GFP-Nopp140

Also note RNA-selective  
probe

RNA-selective, live cell imaging  
probes for studying  
nuclear structure and  
function. Chem. Biol.  
2006, 13, 615-623.





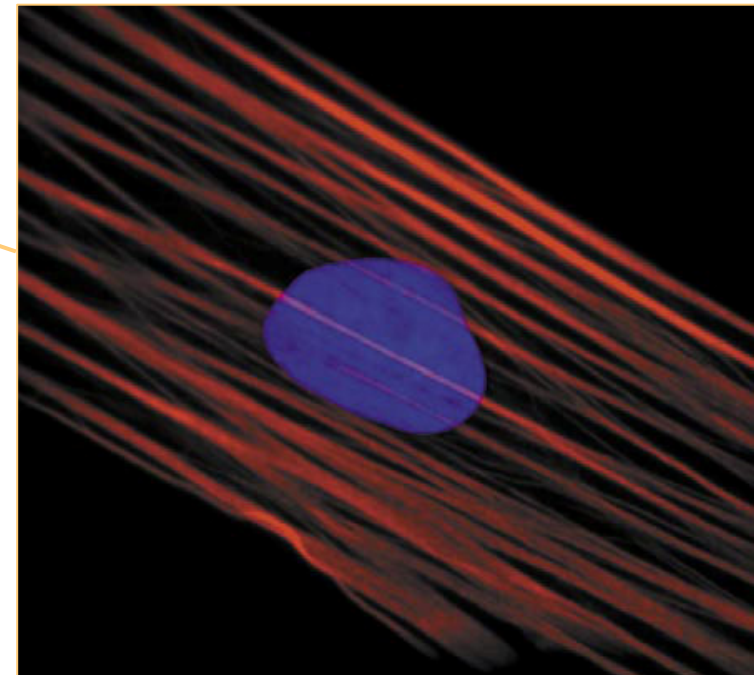
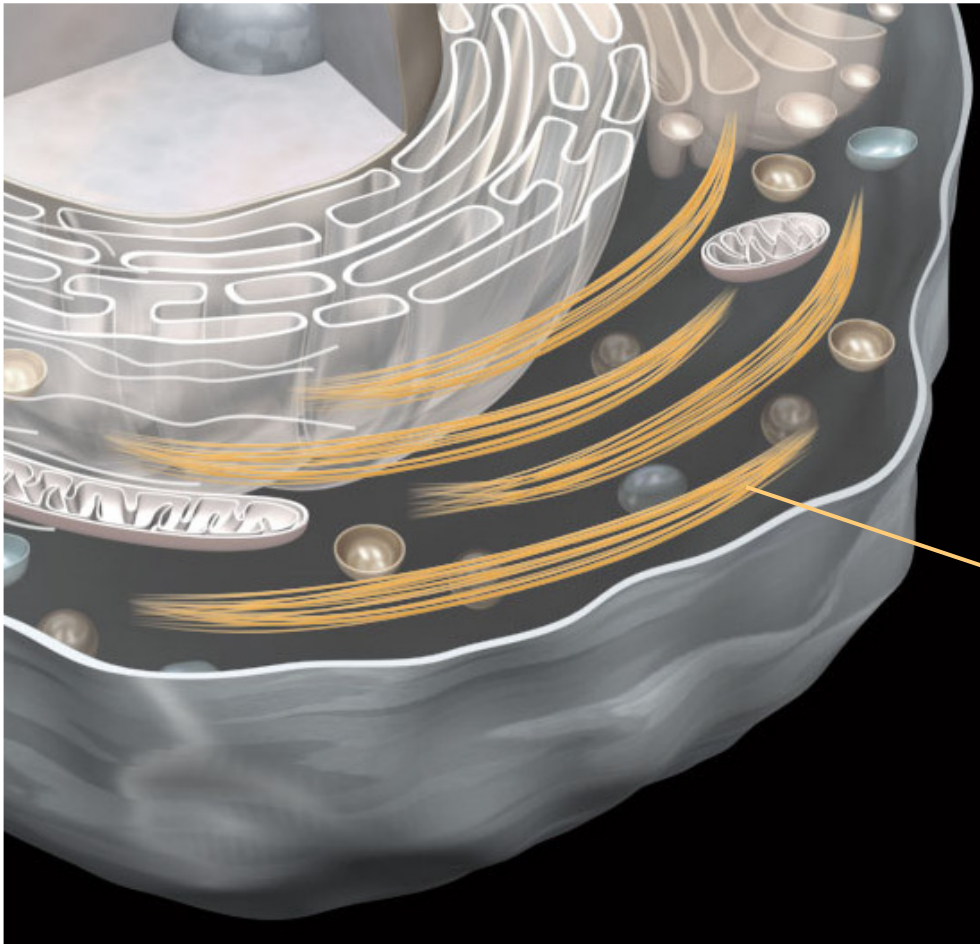


## Peroxisomes

SelectFX™ Alexa Fluor® 488 Peroxisome Labeling Kit

(antibody to Peroxisomal membrane protein 70)

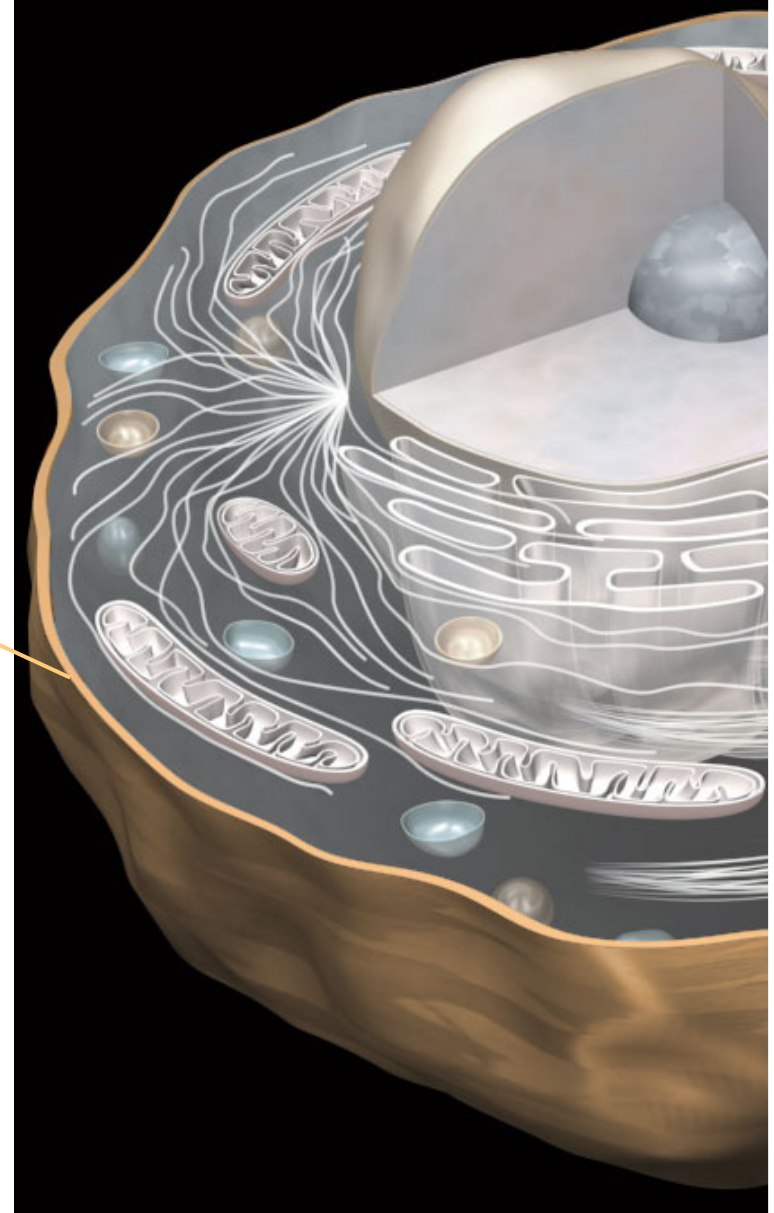
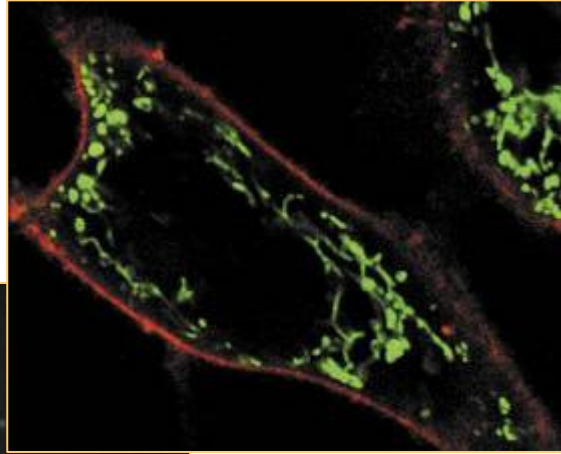
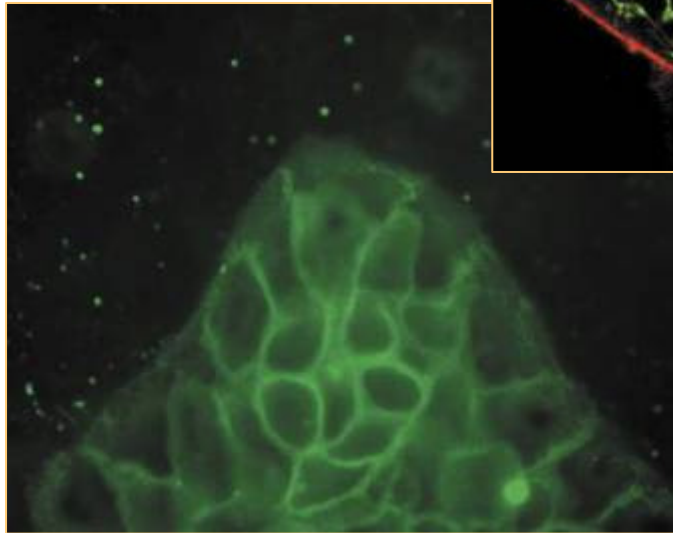
Live cells: GFP-SKL (tripeptide targeting sequence)



## Actin cytoskeleton

Fixed cells: Alexa Fluor-phalloidin

Live cells: Lifeact-GFP (17 aa peptide binding actin)



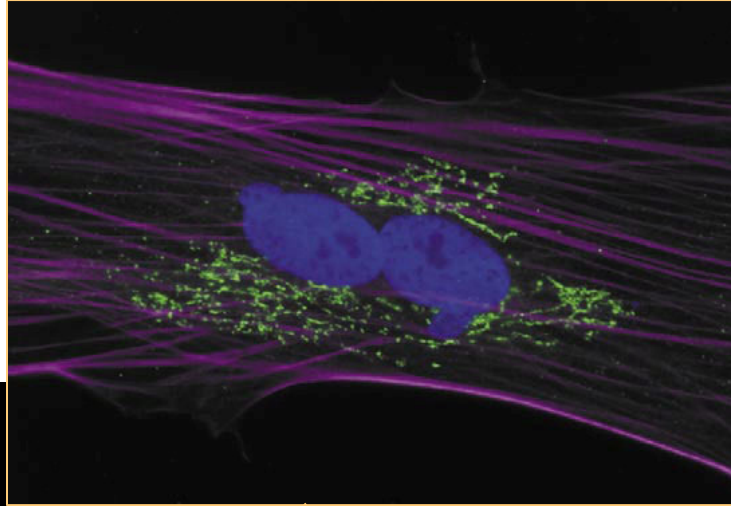
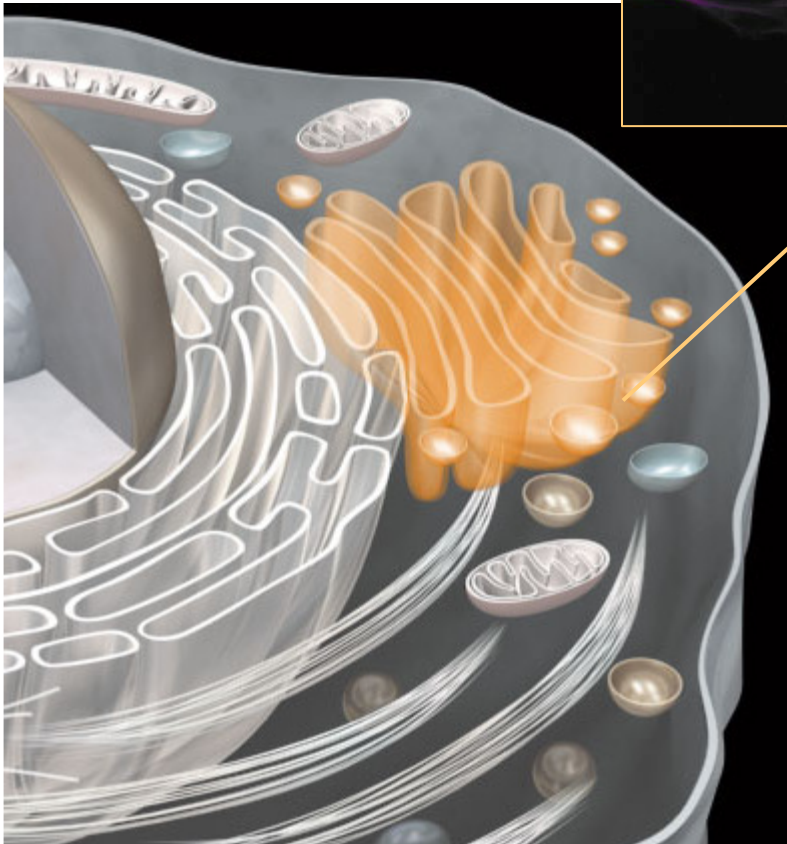
## Plasma Membrane

Wheat Germ Agglutinin

Live cells: FM dyes,

myristoyl+palmitoyl tagged-GFP





## Golgi

anti-golgin-97 antibody

NBD C<sub>6</sub>-ceramide complexed to BSA

Live cells: GalTase-GFP